

Validation of Bioequivalence Test Performed by BE R package

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1 Introduction

BE R package (Bae 2018) can analyze bioequivalence study data with industrial strength. The current version BE performs bioequivalency tests for several variables of a 2x2 crossover study in a data file. To establish BE, the calculated confidence interval should fall within a BE limit, usually 80-125% for the ratio of the product averages. (Drug Evaluation et al. 2001; Chow and Liu 2008; Hauschke, Steinijans, and Pigeot 2007) In this document, the author performed validation of bioequivalence test performed by BE R package.

2 Methods

The required packages are following.

```
library(BE)           # install.packages("BE", repos="http://r.acr.kr")
library(tidyverse)    # install.packages("tidyverse")
```

2.1 Calculation BE

BE::test2x2() function was used to calculate the 90% confidence interval.

```
Cmax_R_BE <- BE::test2x2(NCAREsult4BE, "Cmax")[[4]] %>%
  as.tibble() %>%
  mutate(Analysis = 'R: BE package') %>%
  select(Analysis, everything())

AUClast_R_BE <- BE::test2x2(NCAREsult4BE, "AUClast")[[4]] %>%
  as.tibble() %>%
  mutate(Analysis = 'R: BE package') %>%
  select(Analysis, everything())
```

2.2 Calculation SAS

SAS PROC GLM and SAS PROC MIXED in SAS version 9.0 with SEQ (sequence), TRT (treatment), SUBJ (subject), PRD (period) variables, and LNAUCL or LNCMAX denoting the response measure.

A part of exemplary SAS program statement is following and the full SAS scripts are appended in Appendix A.

```
PROC GLM DATA=BE OUTSTAT=STATRES; /* GLM use only complete subjects. */
CLASS SEQ PRD TRT SUBJ;
MODEL LNAUCL = SEQ SUBJ(SEQ) PRD TRT;
RANDOM SUBJ(SEQ)/TEST;
LSMEANS TRT /PDIFF=CONTROL('R') CL ALPHA=0.1 COV OUT=LSOUT;
```

```
PROC MIXED DATA=BE; /* MIXED uses all data. */
CLASS SEQ TRT SUBJ PRD;
MODEL LNAUCL = SEQ PRD TRT;
RANDOM SUBJ(SEQ);
ESTIMATE 'T VS R' TRT -1 1 /CL ALPHA=0.1;
ODS OUTPUT ESTIMATES=ESTIM COVPARMS=COVPAR;
```

A function, tab_sas_proc_results() reads SAS analysis results exported to Microsoft Excel files (.xls) and converted to comma separated version file (.csv). It returns a data frame of 90% confidence interval calculated either PROC GLM or PROC MIXED in SAS version 9.0.

```
tab_sas_proc_results <- function(filename, skip_no, analysis_name){
  read_lines(filename, skip = skip_no, n_max = 2) %>%
  paste(collapse='\n') %>% read_csv() %>%
  mutate(Analysis = analysis_name) %>%
  select(Analysis, `Lower Limit` = LL, `Point Estimate` = PE, `Upper Limit` = UL)
}
```

3 Results

3.1 C_{\max}

Dataset, NCAResult4BE is shown in Appendix 1.

Comparison between BE and SAS is shown in Table 1.

```
Cmax_proc_glm <- tab_sas_proc_results('sas/SAS_results_Cmax.csv',  
                                     skip = 206, 'SAS: PROC GLM')  
Cmax_proc_mixed <- tab_sas_proc_results('sas/SAS_results_Cmax.csv',  
                                       skip = 278, 'SAS: PROC MIXED')  
  
bind_rows(Cmax_R_BE, Cmax_proc_glm, Cmax_proc_mixed) %>%  
  kable(longtable = TRUE, booktabs = TRUE, caption = 'Cmax', digits = 5)
```

Table 1: Cmax

Analysis	Lower Limit	Point Estimate	Upper Limit
R: BE package	0.90136	0.97984	1.06515
SAS: PROC GLM	0.90136	0.97984	1.06515
SAS: PROC MIXED	0.90136	0.97984	1.06515

3.2 AUC_{last}

Comparison between BE and SAS is shown in Table 2.

```
AUClast_proc_glm <- tab_sas_proc_results('sas/SAS_results_AUClast.csv',  
                                         skip = 206, 'SAS: PROC GLM')  
AUClast_proc_mixed <- tab_sas_proc_results('sas/SAS_results_AUClast.csv',  
                                           skip = 278, 'SAS: PROC MIXED')  
  
bind_rows(AUClast_R_BE, AUClast_proc_glm, AUClast_proc_mixed) %>%  
  kable(longtable = TRUE, booktabs = TRUE, caption = 'AUClast', digits = 5)
```

Table 2: AUClast

Analysis	Lower Limit	Point Estimate	Upper Limit
R: BE package	0.88944	0.95408	1.02341
SAS: PROC GLM	0.88944	0.95408	1.02341
SAS: PROC MIXED	0.88944	0.95408	1.02341

4 Conclusion

There is no discrepancy between results from BE and SAS. We also performed multiple analyses with the real clinical trial datasets and have found no differences (data not shown: confidential). Noncompartmental analysis performed by the open-source R package, NonCompart can be **qualified and validated** enough to acquire the identical results of the commercial software, WinNonlin.

Please report issues regarding validation of the R package to <https://github.com/asancpt/NonCompart-tests/issues>.

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URL: www.github.com/shanmdphd

A Raw

Table 3: Description of settings for the noncompartmental analysis performed in WinNonlin and links to the raw data

SUBJ	GRP	PRD	TRT	AUClast	Cmax	Tmax
1	RT	1	R	5018.927	1043.13	1.04
1	RT	2	T	6737.507	894.21	1.03
2	TR	1	T	4373.970	447.26	1.01
2	TR	2	R	6164.276	783.92	1.98
4	TR	1	T	5592.993	824.42	1.97
4	TR	2	R	5958.160	646.31	0.97
5	TR	1	T	3902.590	803.70	0.80
5	TR	2	R	4620.156	955.30	0.74
6	RT	1	R	3735.274	995.34	1.02
6	RT	2	T	4257.802	816.33	1.00
7	RT	1	R	4314.993	608.99	0.95
7	RT	2	T	5030.372	806.57	0.74
8	RT	1	R	6053.098	1283.67	0.72
8	RT	2	T	5790.067	822.95	1.03
9	RT	1	R	4602.582	679.39	0.74
9	RT	2	T	6042.462	556.55	0.98
10	RT	1	R	8848.988	1136.91	1.03
10	RT	2	T	7349.822	1082.79	0.97
11	TR	1	T	3054.096	547.73	2.02
11	TR	2	R	4719.175	984.69	0.54
13	RT	1	R	4828.682	615.17	1.00
13	RT	2	T	4175.434	692.26	0.97
14	RT	1	R	4566.275	864.56	1.03
14	RT	2	T	5042.649	1122.75	0.75
15	TR	1	T	4950.980	719.40	0.97
15	TR	2	R	4959.554	660.17	0.96
16	RT	1	R	4577.432	609.64	3.01
16	RT	2	T	4773.723	807.65	1.01
17	RT	1	R	6462.652	861.56	2.02
17	RT	2	T	5246.032	1187.75	0.73
18	TR	1	T	4754.625	919.87	0.77
18	TR	2	R	3214.809	1042.84	0.53
19	TR	1	T	7619.304	1089.84	3.00
19	TR	2	R	5210.569	1127.94	2.04
20	TR	1	T	5063.471	1191.46	0.71
20	TR	2	R	6406.634	1069.19	1.00
21	RT	1	R	5580.289	742.67	0.97
21	RT	2	T	6304.119	447.85	0.99
22	RT	1	R	4398.887	682.73	2.02
22	RT	2	T	3760.359	669.01	1.04
23	TR	1	T	5141.165	937.02	0.51
23	TR	2	R	5835.275	894.72	1.04
24	TR	1	T	4343.439	713.57	1.03
24	TR	2	R	2848.448	811.83	0.71
25	TR	1	T	3983.260	1160.32	0.73
25	TR	2	R	3476.389	769.63	0.78
27	TR	1	T	5772.972	1219.56	0.99
27	TR	2	R	7673.260	1063.29	1.03

SUBJ	GRP	PRD	TRT	AUClast	Cmax	Tmax
28	RT	1	R	5679.039	650.24	1.00
28	RT	2	T	5160.875	891.63	1.05
29	TR	1	T	4800.455	770.63	2.02
29	TR	2	R	5772.925	738.17	1.04
30	RT	1	R	4722.324	1034.11	0.77
30	RT	2	T	2896.939	569.22	1.03
31	RT	1	R	8032.393	1043.82	1.98
31	RT	2	T	6076.359	1141.43	0.96
32	TR	1	T	4245.372	608.93	2.97
32	TR	2	R	4745.770	539.66	2.04
33	TR	1	T	3648.195	856.18	0.76
33	TR	2	R	3356.777	647.95	0.98
34	TR	1	T	5015.499	739.42	0.96
34	TR	2	R	6325.746	682.41	1.99
35	RT	1	R	6259.347	1020.55	1.96
35	RT	2	T	5802.468	835.87	2.04
36	RT	1	R	4669.384	682.87	3.01
36	RT	2	T	3783.584	729.63	1.00

B SAS Scripts and results

To run these scripts, the dataset NCAResult4BE should be exported from R by `write.csv()`.

B.1 C_{\max}

```
DATA BE; /* It will load 91 records. */
  INFILE 'c:\Users\mdlhs\asancpt\BEreport\sas\NCAResult4BE.csv' FIRSTOBS=2 DLM=",";
  INPUT SUBJ $ SEQ $ PRD $ TRT $ AUClast Cmax Tmax;
  IF CMAX =< 0 THEN DELETE;
  LNCMAX = LOG(Cmax);
  LNAUCL = LOG(AUClast );

PROC PRINT; RUN;

PROC GLM DATA=BE OUTSTAT=STATRES; /* GLM use only complete subjects. */
  CLASS SEQ PRD TRT SUBJ;
  MODEL LNCMAX = SEQ SUBJ(SEQ) PRD TRT;
  RANDOM SUBJ(SEQ)/TEST;
  LSMEANS TRT /PDIFF=CONTROL('R') CL ALPHA=0.1 COV OUT=LSOUT;
RUN;

PROC PRINT DATA=STATRES; RUN;
PROC PRINT DATA=LSOUT; RUN;

DATA STATRES;
  SET STATRES;
  IF _TYPE_='ERROR' THEN CALL SYMPUT('DF', DF);

DATA LSOUT;
  SET LSOUT;
  IF TRT='R' THEN CALL SYMPUT('GMR_R', LSMEAN);
  IF TRT='T' THEN CALL SYMPUT('GMR_T', LSMEAN);
  IF TRT='R' THEN CALL SYMPUT('V_R', COV1);
  IF TRT='T' THEN CALL SYMPUT('V_T', COV2);
  IF TRT='T' THEN CALL SYMPUT('COV', COV1);

DATA LSOUT2;
  LNPE = &GMR_T - &GMR_R;
  DF = &DF;
  SE = SQRT(&V_R + &V_T - 2*&COV);
  LNLM = TINV(0.95, DF)*SE;
  LNLL = LNPE - LNLM ;
  LNUL = LNPE + LNLM;
  PE = EXP(LNPE);
  LL = EXP(LNLL);
  UL = EXP(LNUL);
  WD = UL - LL;

PROC PRINT DATA=LSOUT2; RUN;

PROC MIXED DATA=BE; /* MIXED uses all data. */
  CLASS SEQ TRT SUBJ PRD;
  MODEL LNCMAX = SEQ PRD TRT;
```

```

RANDOM SUBJ(SEQ);
ESTIMATE 'T VS R' TRT -1 1 /CL ALPHA=0.1;
ODS OUTPUT ESTIMATES=ESTIM COVPARMS=COVPAR;
RUN;

DATA COVPAR;
SET COVPAR;
IF CovParm = 'Residual' THEN CALL SYMPUT('MSE', Estimate);

DATA ESTIM;
SET ESTIM;
MSE = &MSE;
LNLM = (Upper - Lower)/2;
PE = EXP(Estimate);
LL = EXP(Lower);
UL = EXP(Upper);
WD = UL - LL;

PROC PRINT Data=ESTIM; RUN;

read_csv('sas/SAS_results_Cmax.csv') %>%
  knitr::kable(longtable = TRUE, booktabs = TRUE, # format = "latex",
               caption = 'Cmax')

```

B.2 AUC_{last}

```

DATA BE; /* It will load 91 records. */
INFILE 'c:\Users\mdlhs\asancpt\BEreport\sas\NCAResult4BE.csv' FIRSTOBS=2 DLM=",";
INPUT SUBJ $ SEQ $ PRD $ TRT $ AUClast Cmax Tmax;
IF CMAX =< 0 THEN DELETE;
LNCMAX = LOG(Cmax);
LNAUCL = LOG(AUClast );

PROC PRINT; RUN;

PROC GLM DATA=BE OUTSTAT=STATRES; /* GLM use only complete subjects. */
CLASS SEQ PRD TRT SUBJ;
MODEL LNAUCL = SEQ SUBJ(SEQ) PRD TRT;
RANDOM SUBJ(SEQ)/TEST;
LSMEANS TRT /PDIFF=CONTROL('R') CL ALPHA=0.1 COV OUT=LSOUT;
RUN;

PROC PRINT DATA=STATRES; RUN;
PROC PRINT DATA=LSOUT; RUN;

DATA STATRES;
SET STATRES;
IF _TYPE_='ERROR' THEN CALL SYMPUT('DF', DF);

DATA LSOUT;
SET LSOUT;
IF TRT='R' THEN CALL SYMPUT('GMR_R', LSMEAN);
IF TRT='T' THEN CALL SYMPUT('GMR_T', LSMEAN);

```



```

IF TRT='R' THEN CALL SYMPUT('V_R', COV1);
IF TRT='T' THEN CALL SYMPUT('V_T', COV2);
IF TRT='T' THEN CALL SYMPUT('COV', COV1);

DATA LSOUT2;
  LNPE = &GMR_T - &GMR_R;
  DF = &DF;
  SE = SQRT(&V_R + &V_T - 2*&COV);
  LNLN = TINV(0.95, DF)*SE;
  LNLL = LNPE - LNLN ;
  LNUL = LNPE + LNLN;
  PE = EXP(LNPE);
  LL = EXP(LNLL);
  UL = EXP(LNUL);
  WD = UL - LL;

PROC PRINT DATA=LSOUT2; RUN;

PROC MIXED DATA=BE; /* MIXED uses all data. */
  CLASS SEQ TRT SUBJ PRD;
  MODEL LNAUCL = SEQ PRD TRT;
  RANDOM SUBJ(SEQ);
  ESTIMATE 'T VS R' TRT -1 1 /CL ALPHA=0.1;
  ODS OUTPUT ESTIMATES=ESTIM COVPARMS=COVPAR;
RUN;

DATA COVPAR;
  SET COVPAR;
  IF CovParm = 'Residual' THEN CALL SYMPUT('MSE', Estimate);

DATA ESTIM;
  SET ESTIM;
  MSE = &MSE;
  LNLN = (Upper - Lower)/2;
  PE = EXP(Estimate);
  LL = EXP(Lower);
  UL = EXP(Upper);
  WD = UL - LL;

PROC PRINT Data=ESTIM; RUN;

```

C Session Information

```
devtools::session_info()
```

```
## - Session info -----
## setting value
## version R version 3.5.1 (2018-07-02)
## os      Windows 7 x64 SP 1
## system  x86_64, mingw32
## ui      RTerm
## language (EN)
## collate Korean_Korea.949
## ctype   Korean_Korea.949
## tz      Asia/Seoul
## date    2018-10-23
##
## - Packages -----
## package      * version date      lib source
## assertthat    0.2.0  2017-04-11 [1] CRAN (R 3.5.0)
## backports     1.1.2  2017-12-13 [1] CRAN (R 3.5.0)
## base64enc     0.1-3  2015-07-28 [1] CRAN (R 3.5.0)
## BE            * 0.1.1  2018-07-19 [1] CRAN (R 3.5.1)
## bindr         0.1.1  2018-03-13 [1] CRAN (R 3.5.0)
## bindrcpp      0.2.2  2018-03-29 [1] CRAN (R 3.5.0)
## bookdown      0.7     2018-02-18 [1] CRAN (R 3.5.0)
## broom         0.5.0  2018-07-17 [1] CRAN (R 3.5.1)
## callr         3.0.0  2018-08-24 [1] CRAN (R 3.5.1)
## cellranger    1.1.0  2016-07-27 [1] CRAN (R 3.5.0)
## cli           1.0.1  2018-09-25 [1] CRAN (R 3.5.1)
## colorspace    1.3-2  2016-12-14 [1] CRAN (R 3.5.0)
## crayon        1.3.4  2018-06-08 [1] Github (gaborcsardi/crayon@3e751fb)
## debugme       1.1.0  2017-10-22 [1] CRAN (R 3.5.0)
## desc         1.2.0  2018-05-01 [1] CRAN (R 3.5.0)
## devtools      2.0.0  2018-10-19 [1] CRAN (R 3.5.1)
## digest        0.6.18 2018-10-10 [1] CRAN (R 3.5.1)
## dplyr         * 0.7.7  2018-10-16 [1] CRAN (R 3.5.1)
## evaluate      0.12    2018-10-09 [1] CRAN (R 3.5.1)
## forcats       * 0.3.0  2018-02-19 [1] CRAN (R 3.5.0)
## fs            1.2.6  2018-08-23 [1] CRAN (R 3.5.1)
## ggplot2       * 3.0.0  2018-07-03 [1] CRAN (R 3.5.1)
## glue          1.3.0  2018-07-17 [1] CRAN (R 3.5.1)
## gtable        0.2.0  2016-02-26 [1] CRAN (R 3.5.0)
## haven         1.1.2  2018-06-27 [1] CRAN (R 3.5.0)
## highr         0.7     2018-06-09 [1] CRAN (R 3.5.0)
## hms           0.4.2  2018-03-10 [1] CRAN (R 3.5.0)
## htmltools     0.3.6  2017-04-28 [1] CRAN (R 3.5.0)
## httr          1.3.1  2017-08-20 [1] CRAN (R 3.5.0)
## jsonlite      1.5     2017-06-01 [1] CRAN (R 3.5.0)
## knitr         * 1.20    2018-02-20 [1] CRAN (R 3.5.0)
## lattice       0.20-35 2017-03-25 [1] CRAN (R 3.5.0)
## lazyeval      0.2.1  2017-10-29 [1] CRAN (R 3.5.0)
## lubridate     1.7.4  2018-04-11 [1] CRAN (R 3.5.0)
## magrittr      1.5     2014-11-22 [1] CRAN (R 3.5.0)
## memoise       1.1.0  2017-04-21 [1] CRAN (R 3.5.0)
```

```
## modelr      0.1.2    2018-05-11 [1] CRAN (R 3.5.0)
## munsell     0.5.0    2018-06-12 [1] CRAN (R 3.5.0)
## nlme        3.1-137  2018-04-07 [2] CRAN (R 3.5.1)
## pillar      1.3.0    2018-07-14 [1] CRAN (R 3.5.1)
## pkgbuild    1.0.2    2018-10-16 [1] CRAN (R 3.5.1)
## pkgconfig   2.0.2    2018-08-16 [1] CRAN (R 3.5.1)
## pkgload     1.0.1    2018-10-11 [1] CRAN (R 3.5.1)
## plyr        1.8.4    2016-06-08 [1] CRAN (R 3.5.0)
## prettyunits 1.0.2    2015-07-13 [1] CRAN (R 3.5.0)
## processx    3.2.0    2018-08-16 [1] CRAN (R 3.5.1)
## ps          1.2.0    2018-10-16 [1] CRAN (R 3.5.1)
## purrr       * 0.2.5    2018-05-29 [1] CRAN (R 3.5.0)
## R.methodsS3 1.7.1    2016-02-16 [1] CRAN (R 3.5.0)
## R.oo         1.22.0   2018-04-22 [1] CRAN (R 3.5.0)
## R6           2.3.0    2018-10-04 [1] CRAN (R 3.5.1)
## Rcpp        0.12.19  2018-10-01 [1] CRAN (R 3.5.1)
## readr       * 1.1.1    2017-05-16 [1] CRAN (R 3.5.0)
## readxl      1.1.0    2018-04-20 [1] CRAN (R 3.5.0)
## remotes     2.0.1    2018-10-19 [1] CRAN (R 3.5.1)
## rlang       0.2.2    2018-08-16 [1] CRAN (R 3.5.1)
## rmarkdown   1.10     2018-06-11 [1] CRAN (R 3.5.0)
## rprojroot   1.3-2    2018-01-03 [1] CRAN (R 3.5.0)
## rstudioapi  0.8       2018-10-02 [1] CRAN (R 3.5.1)
## rtf         * 0.4-13   2018-05-17 [1] CRAN (R 3.5.1)
## rvest       0.3.2    2016-06-17 [1] CRAN (R 3.5.0)
## scales     1.0.0    2018-08-09 [1] CRAN (R 3.5.1)
## sessioninfo 1.1.0    2018-09-25 [1] CRAN (R 3.5.1)
## stringi     1.2.4    2018-07-20 [1] CRAN (R 3.5.1)
## stringr     * 1.3.1    2018-05-10 [1] CRAN (R 3.5.0)
## testthat    2.0.1    2018-10-13 [1] CRAN (R 3.5.1)
## tibble      * 1.4.2    2018-01-22 [1] CRAN (R 3.5.0)
## tidyr       * 0.8.1    2018-05-18 [1] CRAN (R 3.5.0)
## tidyselect  0.2.5    2018-10-11 [1] CRAN (R 3.5.1)
## tidyverse   * 1.2.1    2017-11-14 [1] CRAN (R 3.5.0)
## usethis     1.4.0    2018-08-14 [1] CRAN (R 3.5.1)
## withr       2.1.2    2018-03-15 [1] CRAN (R 3.5.0)
## xfun        0.3       2018-07-06 [1] CRAN (R 3.5.1)
## xml2        1.2.0    2018-01-24 [1] CRAN (R 3.5.0)
## yaml        2.2.0    2018-07-25 [1] CRAN (R 3.5.1)
##
## [1] C:/Users/mdlhs/Rlib
## [2] C:/Program Files/R/R-3.5.1/library
```

References

- Bae, Kyun-Seop. 2018. *BE: Bioequivalence Study Data Analysis*. <https://CRAN.R-project.org/package=BE>.
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- Drug Evaluation, Center for, Food Research (CDER), U.S. Department of Health Drug Administration, and Human Services. 2001. *Guidance for Industry Statistical Approaches to Establishing Bioequivalence*. <https://www.fda.gov/downloads/drugs/guidances/ucm070244.pdf>.

Hauschke, Dieter, Volker Steinijans, and Iris Pigeot. 2007. *Bioequivalence Studies in Drug Development: Methods and Applications*. Wiley.