

AARF: Any-Angle Routing for Flow-Based Microfluidic Biochips

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Abstract—Flow-based microfluidic biochips are promising with significant applications for automating and miniaturizing laboratory procedures in biochemistry. Automated design methods for flow-based microfluidic biochips are becoming increasingly important due to the advancement in both integration scale and design complexity for complicated biochemical applications. Though the multilayer soft lithography fabrication provides flexibility to route both flow and control channels in any angle, existing routing algorithms still adopt Manhattan routing metrics, which design channel in either vertical or horizontal direction only. Moreover, based on the computational fluid dynamics (CFD) analysis, rectilinear channels with 90° bends have the following issues: (1) reduced the fluidic flow rate, which degrades the performance of the biochip and may even result in the erroneous outcome of the whole procedure, and (2) increased pressure at the right-angle bend, which negatively affects the reliability of the biochip. To fully utilize the routing flexibility, this paper proposes the first any-angle routing algorithm for flow-based microfluidic biochip, called AARF. Computational simulation results show that compared with traditional Manhattan routing method, the proposed AARF significantly improves the total wirelength and total effective wirelength (considering the turning angles) by 19.28% and 53.04%, respectively, which prove the effectiveness of the AARF routing flow.

Index Terms—Flow-based microfluidic biochips, Any-angle routing, Dynamic programming, Computational fluidic dynamics (CFD) simulation

I. INTRODUCTION

Microfluidic biochip, also known as Lab-on-a-Chip (LoC), has emerged as a revolutionary technique toward miniaturization and automation of the laboratory procedures in biochemistry [1], [2], [3]. LoC integrates different biochemical analysis modules, such as mixer, separator, chamber, etc., into a single chip, and hence reduces samples/reagents to

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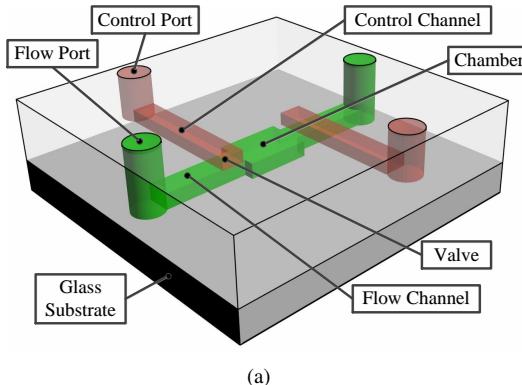
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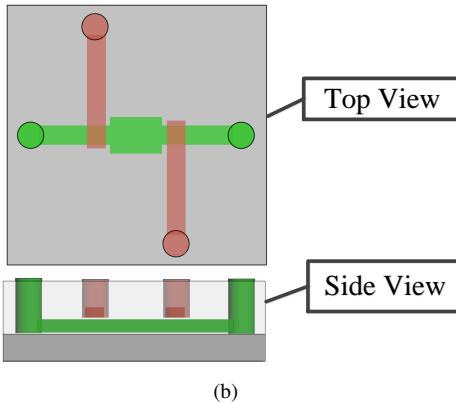
microliter or even nanoliter scale [4], [5]. Noticeable merits of microfluidic biochips over traditional laboratory platforms include: (1) greatly saving the assay cost by reducing expensive samples/reagents to nano-liter or pico-liter volume, (2) effectively integrating the automatic control logic for reduced human intervention and labor cost, (3) significantly increasing sensitivity, accuracy and throughput, (4) essentially facilitating portability for point-of-care diagnostics, and (5) naturally enabling microscale assays (e.g., single-cell culture, capture and analysis) that are infeasible by traditional macroscale approaches. According to Research and Markets, the global biochips market is expected to grow at a compound annual growth rate (CAGR) of 22.8% from 2013 to 2019, and will reach \$52 Billion by 2019 [6]. Applications of biochips cover many different fields, such as diagnostics and treatment, drug discovery and development, biological research, forensic analysis, agriculture, environmental sensors, food inspection, etc. For example, microfluidic biochips are recently designed for point-of-care HIV/AIDS and cancer diagnosis [7], [8].

Flow-based microfluidic biochip is one of the most commonly used microfluidic biochips, which is fabricated using *multilayer soft lithography* (MSL) technology. In flow-based microfluidic biochips, microfluidic functional units are fabricated by elastomer material (polydimethylsiloxane, PDMS) [4], [9]. Figure 1 shows the schematic of flow-based microfluidic biochip, which consists of two functional layers [10]. In the figure, the 3D view of a demonstrative chamber for cell culture is shown in Figure 1(a), and the corresponding top and side views are shown in Figure 1(b). On top of the glass substrate, there are two PDMS layers: (1) *flow layer* for fluidic sample/reagent transportation, and (2) *control layer* for enforcing the controlling logic through control channels. Both control and flow channels are connected to input/output ports, which are punched holes through the PDMS material. A *microvalve* (*valve* for short) is formed between control and flow channels at their intersection area. To switch the valve, external pressure is injected through *control port* (also called *control pin*). Then hydraulic or pneumatic pressure (~10 psi) will be conducted through control channel to the valve, squeezing the elastomer down to seal the flow channel. When external pressure is released, the elastomer will restore back, and the fluidic flow will resume due to external pressure from the *flow port*.

Different complex units can be formed in the biochip by combining several valves, such as micropumps, mixers, multiplexers, etc [5]. By controlling these units through different actuation patterns on valves, fluids (i.e., samples/reagents) are manipulated to transport in flow channels for different oper-



(a)

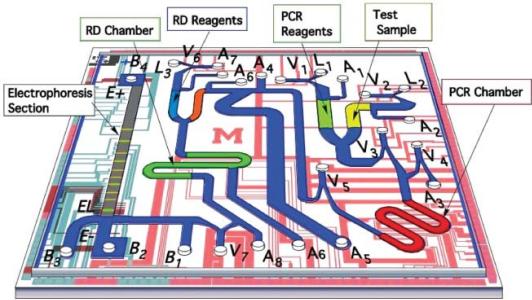


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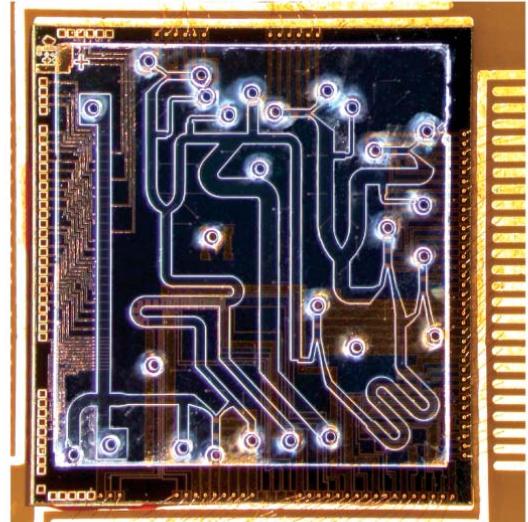
Fig. 1. Schematic of flow-based biochip with different perspective views. (a) 3D view, and (b) Top and side views.

ations, e.g., splitting, mixing, filtering, metering, etc. Thus, an assay can be realized by executing the sequential fluidic operations automatically in a programmed way [13]. With the advancements of fabrication techniques in multilayer soft lithography, the size of valves has been reduced to $6 \times 6\mu\text{m}^2$ [14], [15]. And the valve density has reached 1 million valves per cm^2 , which makes manual design impossible for the microfluidic very large-scale integration (mVLSI). Due to the ever increasing complexity in both bioassays and integration scale, computer-aided design (CAD) methods are becoming necessary for reliable biochip designs to achieve correct functionality and reduced turnaround time.

Though PDMS is a flexible material for molding, pressure propagation through the control channel is very slow from the control pin to the corresponding valve(s) [16]. Moreover, the propagation time will significantly increase due to reduced driving force especially in portable microfluidic devices. This critical issue also existed in flow channels. Therefore, flow-based microfluidic biochips typically adopt any-angle flow channels for better performance in the assay experiments. Figure 2(a)(b) shows an example of an integrated microfluidic device with any-angle flow channels for influenza and genetic analyses in both schematic and photographic view [17]. However, existing automated design methods for flow-based microfluidic biochips are still based on Manhattan metrics, which compute Manhattan channels on both flow and control layers. In this paper, we propose the first any-angle routing method, called **AARF**, which obtains shorter



(a)



(b)

Fig. 2. Microfluidic chip example with any-angle flow channels. (a) Schematic representation, and (b) Photograph of the assembled device.

channel lengths compared with Manhattan routing. Moreover, the fluidic flow rate is further enhanced by the proposed dynamic programming approach. From the computational fluidic dynamics (CFD) analysis, the computed any-angle channel has much better performance compared with the corresponding Manhattan channel.

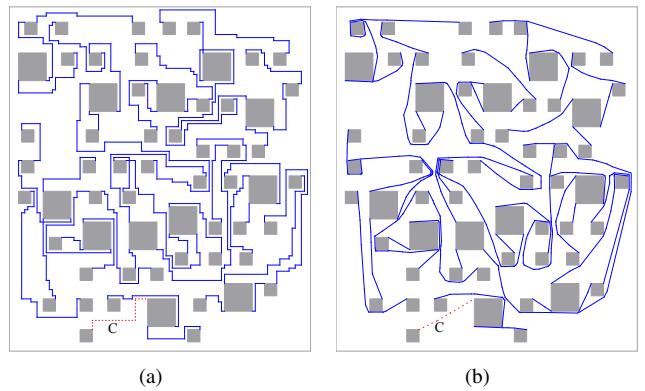


Fig. 3. Routing result of (a) Manhattan routing, and (b) AARF.

Figure 3(a) shows the rectilinear channels designed using

the traditional Manhattan routing method, and Figure 3(b) shows the any-angle routing result from our AARF router. From CFD simulation reports by ANSYS Fluent software [18], the velocity magnitude (i.e., flow rate) of channel C by Manhattan routing is 0.0053 m/s (see Figure 3(a)), whereas that of any-angle routing from AARF is 0.0316 m/s. That is, around $6\times$ improvements in flow rate can be obtained using the proposed any-angle routing method. To the best of our knowledge, this paper is the first work on any-angle routing considering the flow rate for flow-based microfluidic biochips. In the past decade, noticeable advances have been made in CAD methodology for digital microfluidic biochips, including resource binding, operation scheduling, module placement, and droplet routing [21], [22], [23], [24], [25]. However, only a few works addressed the CAD problems for flow-based biochips. Minhass et al. presented a constraint programming based synthesis method for flow-based biochips [26]. Lin et al. presented a flow channel routing method, which simultaneously minimizes the total length and the maximum length of flow channel [27]. Amin et al. presented the first control-layer routing method for flow-based microfluidic biochips [28]. Minhass et al. presented a control synthesis method, which addresses the assignment from valves to control pins for minimized number of control pins [12]. In [29], a control-layer routing algorithm is presented to minimize the pressure-propagation delay of the control signals. In [30], the length-matching routing constraints are enforced during the routing for control-layer channels. In [31], placement and routing methods for flow-based microfluidic biochips are proposed based on the classic sequence-pair-based representation. However, no existing works consider any-angle routing for flow-based microfluidic biochip to significantly enhance flow rate.

In this paper, we propose the first any-angle routing method for flow-based microfluidic biochips, called **AARF**, which designs the non-rectilinear flow channels for enhanced fluidic flow rate. With the constrained Delaunay triangulation structure, we proposed the dynamic-programming-based approach to further enhance the flow rate in non-rectilinear channels based on CFD metrics. Moreover, the total channel lengths are also significantly reduced compared with the Manhattan routing results. Major contributions of the paper are listed as follows.

- The first any-angle routing method is proposed with enhanced fluidic flow rate and minimized total effective channel length, i.e., weighted wirelength considering the turning angles of non-rectilinear channels.
- Constrained Delaunay triangulation structure is computed for the coarse path searching procedure, and iteratively updated with newly routed flow channels.
- Dynamic-programming-based routing approach is proposed to optimize the total effective wirelength, which significantly improves the flow rate.
- A theoretical model is adopted in any-angle routing process to efficiently compute the effective wirelength from the turning angles in the non-rectilinear flow channels.
- CFD simulations are performed to accurately evaluate the flow rates of the non-rectilinear channels obtained

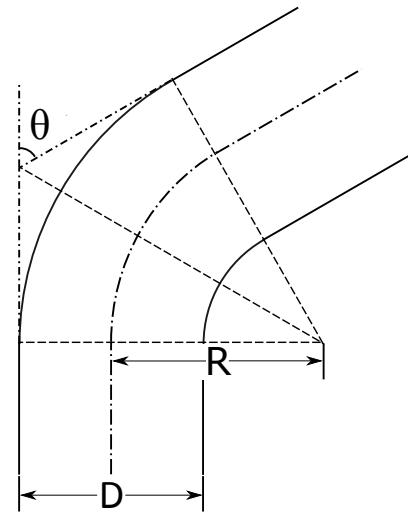


Fig. 4. Definition of a turning angle formed by two channel segments along with an arc channel.

by AARF vs. the rectilinear channels from the existing Manhattan routing method.

The remainder of this paper is organized as follows. Section II presents the problem formulation. Section III presents the overall flow of the proposed any-angle routing method AARF. Section III-A presents the details of constrained Delaunay triangulation graph. Section III-B presents the coarse path searching process on the related search graph. Section III-C presents the dynamic-programming-based routing method for minimized total effective wirelength. Section IV presents the pseudocode of AARF and the analysis of its runtime complexity. Section V presents and discusses the computational simulation results. Finally, conclusion is drawn in Section VI.

II. PRELIMINARIES AND PROBLEM FORMULATION

A. Effective Channel Length

According to simulation results of computational fluid dynamics (CFD), the flow rate is mainly affected by layout pattern of the channel, especially the turns along the channel. Here we define *turning angle* as supplement of the angle formed by two segments at the turn. Figure 4 shows an example, where angle α is the turning angle of the elbow. Generally, the more turns a channel contains, the slower the flow rate would be; and the larger the turning angle is, the slower the flow rate would be. Besides the influence on the flow rate, a large turning angle also increases the pressure against the channel at the elbow, which is similar to the river channel erosion at the river bend. Such increased pressure against the channel's sidewall is hazardous to the lifetime of the PDMS channel in flow-based microfluidic biochips. Therefore, in this paper we propose the first any-angle routing method, AARF, to avoid elbows with large turning angles in the computed flow channels.

In AARF, we adopt a classical method in the CFD field to convert the turning angles into effective channel lengths such that the impact of turning angles could be efficiently evaluated.

The effective channel length L' of a turning angle is defined as follows:

$$L' = \frac{\xi}{\lambda} \cdot D \quad (1)$$

where D denotes the width of the channel. λ is the *coefficient of frictional resistance*, which is determined by the Reynolds number, the status of the fluid, and the roughness of the channel. And ξ is the *coefficient of local resistance*, which is affected by the turning angle θ and is computed as follows:

$$\xi = [\alpha + \beta \cdot (\frac{D}{R})^\gamma] \cdot \frac{\theta}{90} \quad (2)$$

where θ is the turning angle, D is the width of the channel, R is the radius of curvature of the turn (see Figure 4). α , β , and γ are experimentally determined parameters. When the effective channel lengths are computed for the turns along a channel, the *total effective wirelength* represents sum of the actual channel lengths and the effective channel lengths of the turns. During the routing process, the total effective wirelength is used to measure the solution quality of the computed channels with elbows. In this way, the proposed routing method can design the channels with relatively shorter length and elbows of smaller turning angles.

B. Problem Formulation

Considering the effective channel length for enhanced fluidic flow rate, the any-angle routing problem for flow-based microfluidic biochips can be formulated as follows:

Input: A list of nets (pairs of routing terminals) to be connected, a set of functional modules with routing terminals, a set of rectilinear routing blockages, a set of design rules for routing channels, and the biochip's design boundary.

Objective: Find the channel routing solution for all the nets with minimized the total effective wirelength computed by Equation (1).

Constraint: (1) The computed flow channel can be in any angle rather than just Manhattan. (2) The minimum channel width and the minimum channel spacing constraints must be satisfied.

III. ALGORITHM OVERVIEW

Figure 5 shows the overall flow of the proposed any-angle router named AARF, which is based on the Constrained Delaunay Triangulation Graph (CDT). Initially, the CDT is constructed on the routing terminals and routing obstacles. Then The flow channels for the nets are routed on the CDT one by one until all channels are finished. Once a flow channel is routed, the CDT will be updated accordingly. For each channel, the routing process consists of two steps: (1) finding a coarse routing solution, and (2) determining the physical routing channel. In the first step, a set of sequential CDT edges are computed by the Dijkstra shortest path algorithm, which will be crossed by the routing channel. In the second step, a detailed routing algorithm based on dynamic programming is performed to find the routing channel crossing the computed CDT edges. As the elbows in a flow channel can severely slow

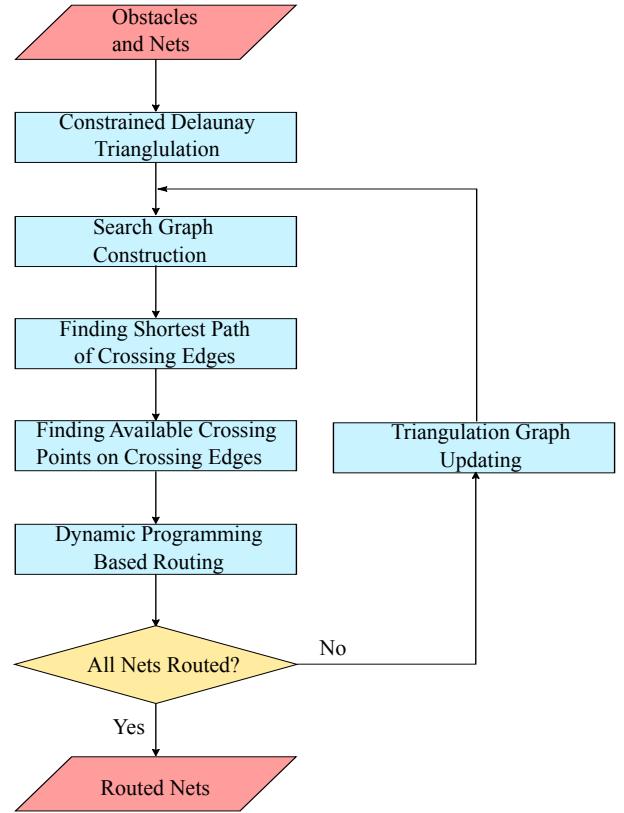


Fig. 5. The proposed AARF routing flow.

down the flow rate, the dynamic-programming-based routing method is proposed to significantly improvement the flow rate, and thus improve the lifetime of the flow-based microfluidic biochip. The intrinsic idea of the dynamic-programming-based routing method is to assign cross points to the CDT edges of the coarse routing solution, and then optimally determine the cross points along the CDT edges for maximized flow rate. The notations used in the paper are summarized in Table I.

A. Constrained Delaunay Triangulation Graph

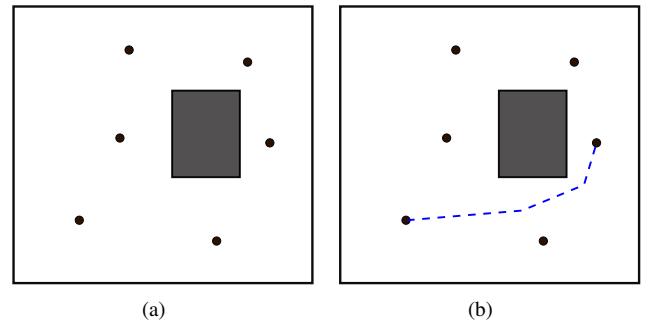


Fig. 6. Example of a non-rectilinear flow channel obtained by the any-angle routing method. (a) Input terminals and routing obstacle. (b) Any-angle routing channel.

As mentioned in previous sections, the proposed any-angle routing algorithm is based on the CDT, which consists of the *constrained edges*. Here, constrained edges refer to those edges that need to be contained in the final Delaunay triangulation

TABLE I
NOTATIONS USED IN THE PROPOSED ALGORITHMS.

Notations	Meaning
N	Lists of input nets
N_i	The i^{th} input net
V_i	The i^{th} vertex of an input net
w	The minimum width of channels
s	The minimum space between channels
CDT	Constrained Delaunay triangulation graph
SG	Search graph
Te_i	The i^{th} triangle edge in CDT
T_i	The i^{th} triangle in CDT
e_i	The i^{th} neutrality edge of a triangle
E	The set of triangle edges the routed channel crosses
E_i	The i^{th} triangle edge the routed channel crosses
m	The precision of dynamic programming, which means the number of a subproblem's alternative solutions
F_{ij}	The j^{th} possible point of interception of the routed channel and E_i
C_i	The i^{th} routed channel
\overline{PQ}	The edge between vertex P and vertex Q
$len(\overline{PQ})$	The distance between vertex P and vertex Q
\overline{PQR}	The elbow shaped by \overline{PQ} and \overline{QR}
$eqv(\overline{PQR})$	The effective length of the turning angle of elbow \overline{PQR}
$\min\{a_1, a_2, \dots, a_n\}$	The smallest element among a_1, a_2, \dots, a_n

result. Using the constrained Delaunay triangulation algorithm, the set of constrained edges can be specified as input. Given the routing terminals and obstacles (see Figure 6), we construct the CDT as follows:

- 1) The input vertices consist of all the routing terminals, the vertices of the rectilinear routing blockages, and the vertices of the biochip's design boundary.
- 2) The constrained edges consist of the edges of the biochip's design boundary, as well as the edges of the routing obstacles.
- 3) After one round of constrained Delaunay triangulation followed by dynamic-programming-based routing process, the corresponding routing segments of the routed channel are inserted into the constrained edges, and the corner points between the segments are inserted into the input vertices.

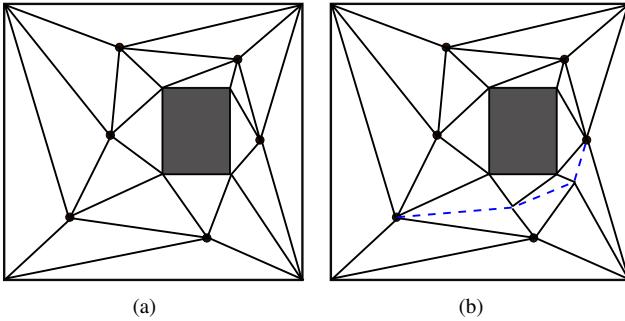


Fig. 7. Example of the constrained Delaunay triangulation graph: (a) initial constrained Delaunay triangulation graph, and (b) updated constrained Delaunay triangulation graph with a routed channel.

Figure 7(a) shows an example of the CDT on the initial

input. And Figure 7(b) shows the updated CDT with a routed flow channel. In Figure 7(b), all the segments of the routed flow channel are inserted into the CDT as constrained edges. In real biochip designs, the routing terminals are mostly located at the boundaries of the functional modules. Here, for easily understanding the proposed routing method, the routing terminals in the illustrating figures are floating, i.e., the corresponding functional modules are not given. We claim that the basic any-angle routing algorithm remains the same when the routing terminals are attached to the functional modules. In the experiments, most routing terminals are attached to functional modules, and the proposed routing algorithm works well. In the updated CDT, besides insertion of the new edges, certain edges in the original CDT may be removed or replaced by new edges for incorporating the routed channel and maintaining the Delaunay Triangulation.

B. Coarse Path Searching

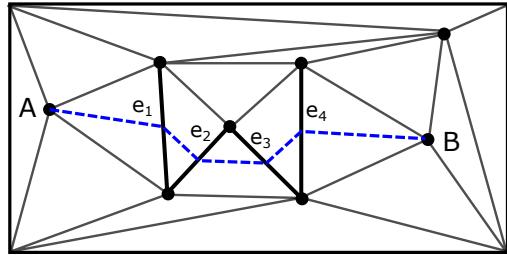


Fig. 8. Example of coarse path searching on the CDT.

After the above introduction of the CDT representation, we now describe the first step of routing, i.e., coarse routing, to search for a set of edges over the CDT edges which routing path crosses. After coarse routing for one net, the obtained routing solution is a set of sequential CDT edges. The flow channel to be computed should cross these edges one by one. To estimate the length of the routing path, we define the *pseudo cross point* on a CDT edge as the midpoint of the edge. Then the length of the coarse routing path is estimated as the sum of Euclidean distances between the source terminal and the pseudo cross point, between the target terminal and the pseudo cross point, and between adjacent pseudo cross points. Figure 8 shows an example, where the set of bold edges (i.e., e_1, e_2, e_3 , and e_4) form the coarse routing path between terminals A and B . The length of the coarse routing path is estimated as the total length of the dash segments. As shown in Figure 8, this routing stage obtains a set of CDT edges, which provides a coarse routing result. Therefore, in the proposed routing flow, there are two major stages: (1) the *coarse routing* stage, which obtains the coarse routing solution, and (2) the *detailed routing* stage, which computes the exact routing path crossing the CDT edges based on the predetermined coarse routing solution.

During coarse routing stage, the Dijkstra's shortest-path algorithm is performed to find the routing path. To perform the Dijkstra's shortest-path algorithm, we define a *search graph* (SG), which is an undirected graph formed by the source and target terminals of the net to be routed, the midpoints of the

CDT edges, the *neutrality edges* over the CDT (i.e., the line segments between midpoints of a triangle edges in the CDT), and certain triangle medians corresponding to source/target terminals. A search graph is constructed as follows:

- 1) The vertices of SG consist of the source and target terminal of the net to be routed, and all midpoints of all the triangle edges except those on the design's boundary, the routing blockage's boundary, and the routed channel.
- 2) Each neutrality edge of a triangle corresponds to an SG edge, unless it violates the design rules, or at least one terminal of the neutrality edge does not correspond to the SG vertex.
- 3) A triangle median should be added into SG if and only if its vertex corresponds to the source/target terminal.

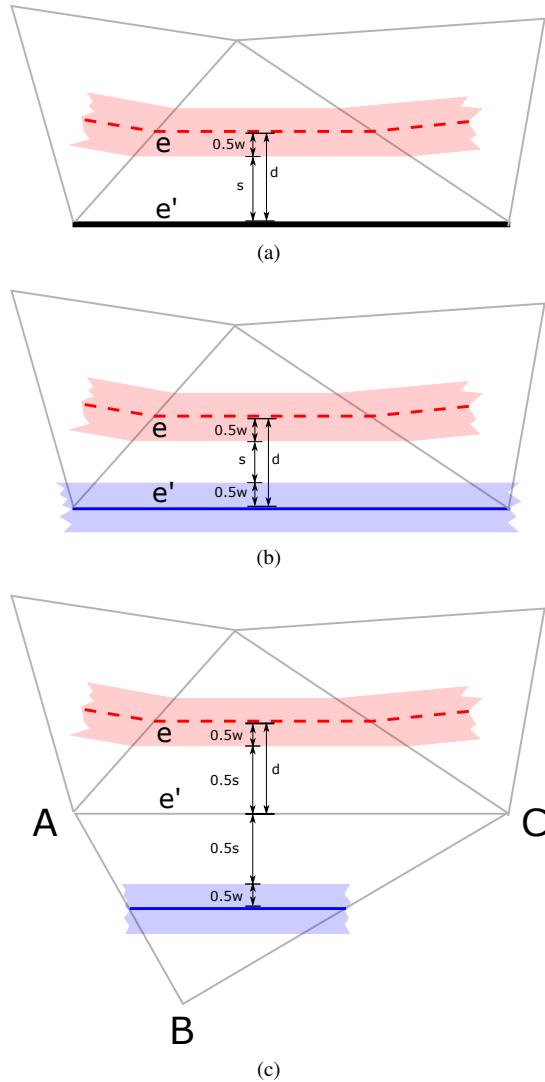


Fig. 9. Example of the design rule violation: (a) If e' is an edge of boundary or obstacles, e breaks the design rule when $d < 0.5w + s$; (b) If e' is an edge of a routed channel, e breaks the design rule when $d < w + s$; (c) If e' is a normal triangle edge, e breaks design rule when $d < 0.5(w + s)$.

To determine whether a neutrality edge is an edge on the search graph, we should verify whether it violates the design rule. As shown in Figure 9, while checking neutrality edge e in a triangle, there are three different cases according to the

triangle edge e' parallel to e . For each case, whether e breaks design rules depends on the distance d between e and e' . e will be included as an edge in SG as long as the design rules are observed. Figure 9(a) shows the first case where e' is an edge of the design's boundary or obstacles. In this case, e breaks the design rule if and only if $d < 0.5w + s$, where w means the minimum width of channels, and s means the minimum spacing between channels. By enforcing the above design rule, the spacing between the channel's boundary and the design's or obstacle's boundary is guaranteed to be not less than the required spacing s . Figure 9(b) shows the second case where e' is an edge from a routed channel. In this case, e breaks the design rule if and only if $d < w + s$. Figure 9(c) shows the third case where a flow channel goes through triangle \overline{ABC} . Obviously, the distance between that channel and e should be greater than $w + s$. Hence, d is enforced to be larger than $0.5(w + s)$.

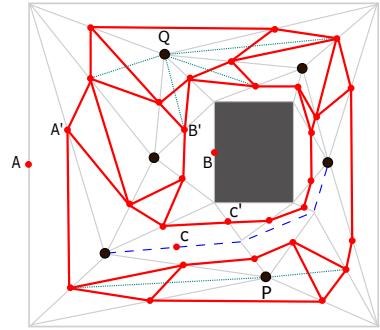


Fig. 10. Example of the search graph.

Figure 10 shows an example of the search graph SG. Edges in black form the CDT, and edges in red form its corresponding search graph SG. In Figure 10, points in red are midpoints of the triangle edges, which correspond to the vertices of SG. Please note that points A , B , and C are not contained in SG, because A is on the design boundary, B is on the blockage boundary, and C is on a routed flow channel. Therefore, edges $\overline{AA'}$, $\overline{BB'}$ and $\overline{CC'}$ are not contained in SG.

To find a shortest channel on SG, several additional edges connecting CDT and SG are in need because the terminals of a channel are triangle vertices, which are on CDT, and the segments of a channel are neutrality edges, which are on SG. Intuitively, the medians of triangles can serve as additional edges. When we are finding the channel between two given terminals, for example vertex P and Q in Figure 10, a triangle median has to be added into SG as an additional edge once its vertices include either P or Q (the dash line segments). Thus, we can find the shortest channel between any given pair of terminals on SG with additional edges using Dijkstra shortest path algorithm.

The result of the coarse routing stage is a path between two terminals, which crosses a set of triangle edges at their midpoints. With the coarse routing solution, we can move on to the next routing stage, i.e., finding the detailed routing path. As the total effective wirelength of the coarse solution is the shortest among all possible coarse routing paths, the total effective wirelength of the detailed channel based on the

In this way, we can route all the nets in N one by one using the proposed routing algorithm.

B. Complexity Analysis

Now we analyze the runtime complexity of Algorithm 3. Assume the sum of number of vertices of the design boundary, the obstacles and the routing pins is $O(M)$, a routed channel has $O(L)$ terminals in average, the number of nets is $O(N)$, and during dynamic-programming-based routing we insert $O(m)$ crosspoints on the CDT edge. Constructing constrained Delaunay triangulation graph takes $O(M \log(M))$ time [32]. Then we analyze the complexity of routing one channel. Suppose we have already routed x flow channels. Thence, the total number of vertices in CDT is $O(M + xL)$, so is the number of triangle edges. Thus, constructing the search graph on the CDT takes $O(M + xL)$ time, and the search graph contains $O(M + xL)$ edges and points. So the Dijkstra algorithm for searching the coarse routing solution on the search graph takes $O((M + xL)^2)$ time. Since there are $O(L)$ sub-problems corresponding to the $O(L)$ terminals on the CDT edges along the coarse routing solution, and each of them has $O(m)$ solutions, the dynamic programming stage for computing the detailed routing solution takes $O(Lm)$ time. Finally, we need to insert all segments of the current routed flow channel into the CDT, and this step takes $O(L(M + xL)^{\frac{4}{3}})$ time using the jump-and-walk approach for point location [33]. Nonetheless, if we reconstruct the CDT instead of inserting constrained edges, it takes $O((M + xL)\log(M + xL))$ time, which is more efficient. Therefore, we prefer to rebuild the CDT after each round of routing rather than directly inserting constrained edges. As a result, it takes $O((M + xL)^2 + Lm + (M + xL)\log(M + xL))$ time to route one channel. By sum of progression, our algorithm takes $O(NM^2 + N^2LM + N^3L^2 + NLm + N(M + NL)\log(M + NL))$ time. In most cases, NL is larger than M and m , so the overall runtime complexity is approximately $O(N^3L^2)$. Therefore, the longer the distance between a pair of terminals to be routed, the more crosspoints/terminals the routed channel is expected to have. And the more obstacles the chip contains, the more crosspoints the routed channel will have. Therefore, the time complexity depends mostly on the number of nets and the layout of the chip (i.e., number of obstacles, the distance between the pair of terminals in a net). Other factors, such as the precision of dynamic programming m , may slightly influence the runtime, but do not have any major impact.

V. EXPERIMENTAL RESULTS

We have implemented the proposed AARF algorithm in C++ programming language on a 2.60GHz 32-core Intel Xeon Linux workstation with 132GB memory. Only a single thread is used for the experiments. The CGAL library is used for the implementation of the constrained Delaunay triangulation algorithm [34]. A typical Manhattan routing algorithm for flow-based microfluidic biochips is tested to verify the proposed approach [30]. Six flow-based microfluidic biochip benchmarks are used in the experiments to verify the performance of the proposed AARF method. Each benchmark has 13 to 45 flow channels to be routed. Moreover, CFD

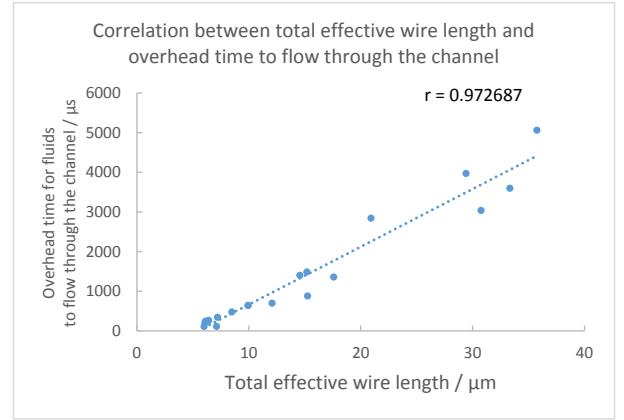


Fig. 12. Computational simulation results on different total effective wirelengths of channels.

simulations are performed using ANSYS Fluent to accurately verify the effectiveness and validity of our computational model to calculate the effective length of turning angles [18].

A. Validity Test of the Computational Model

Our first experiment is designed to prove the validity of our computational model for simulating the flow rate in different channel conditions. Different turning angles affect the flow rate and hence affect the flow transmission time of fluids through the flow channel. The idea of converting the turning angle into effective transmission length is a classic method to evaluate the effect of different turning angles on the fluidic transmission time. Thus, the total effective wirelength calculated by the computational model can be used to estimate transmission time for fluids through the channel. In this experiment, we randomly select 20 out of total 45 channels obtained by our AARF algorithm and Manhattan routing. The total effective wirelength of each channel is calculated using our CFD model. To validating the model of total effective wirelength, we obtain the flow rate of each channel through CFD simulation. With the flow rate from CFD simulation and the actual channel length, the transmission time of fluids through the flow channel can be accurately computed. Figure 12 shows the computational simulation results of the selected channels, which indicate the correlation between the transmission time and the total effective wirelength. We compute the correlation coefficient between the transmission time and the total effective wirelength as follows:

$$r = \frac{E(X \cdot Y) - E(X) \cdot E(Y)}{\sigma(X) \cdot \sigma(Y)} \quad (5)$$

where $E(X)$ means the mathematical expectation of variable X , and $\sigma(X)$ means the standard deviation of X . Here, Y and X stand for the transmission time and the total effective wirelength, respectively. According to the computational results, the correlation coefficient equals to 0.97, which verifies the effectiveness of the proposed model of total effective wirelength. Typically, fluids spend more time flowing through a channel with longer total effective wirelength. Therefore, our proposed total effective wirelength model can effectively

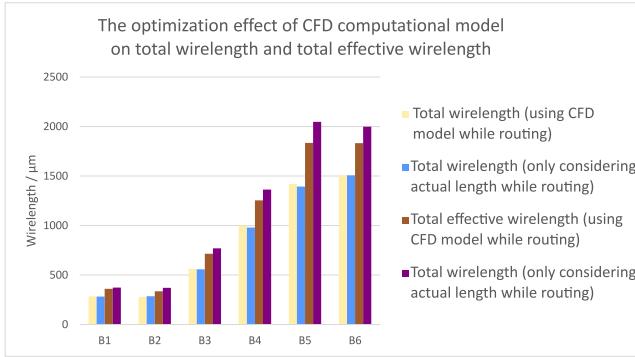


Fig. 13. Comparison of two different routing methods showing the optimization of applying CFD model in routing process.

estimate the impact of turning angles on fluidic transmission time.

B. Effect of the CFD Model

In the second experiment, we route all the six benchmarks with our AARF routing algorithm. Each benchmark is routed twice using different strategies to verify the effectiveness of the routing algorithm considering the total effective wirelength. In strategy I, we compute the total effective wirelength considering the turning angles of elbows during routing, and find the routing solution with minimized total effective wirelength. In strategy II, we only minimize the channel's wirelength without considering the turning angles of elbows. Ideally, the total effective wirelength of strategy I should be shorter than strategy II, and the actually total wirelength may be slightly longer. From the previous experiment, channels with shorter total effective wirelength perform better. Therefore, using CFD metrics to compute the effective length of turning angles, strategy I optimizes the routing result considering the fluidic transmission time.

As shown in Figure 13, we compute the total wirelength and total effective wirelength of each benchmark, and compare the statistics of the two routing results. From the figure, the total wirelengths obtained by the two routing strategies are almost the same. Nevertheless, the routing result obtained by strategy I has shorter total effective wirelength. As the total effective wirelength grows, the disparity of the two routing strategies tends to increase. This is because the longer a channel is, the more elbows it tends to have, with the larger solution space for optimization. From the results, we conclude that using our CFD model during routing can optimize the routing result, especially for biochips with long flow channels.

C. Effectiveness Test of the Precision of Dynamic Programming

In the third experiment, we test the effectiveness of the *precision of dynamic programming* m . We route each benchmark using our AARF algorithm for several times with different values of m . This experiment verifies that the proposed dynamic programming method can approach the optimal solution. Figure 14 and Figure 15 show the experimental results. Figure 14

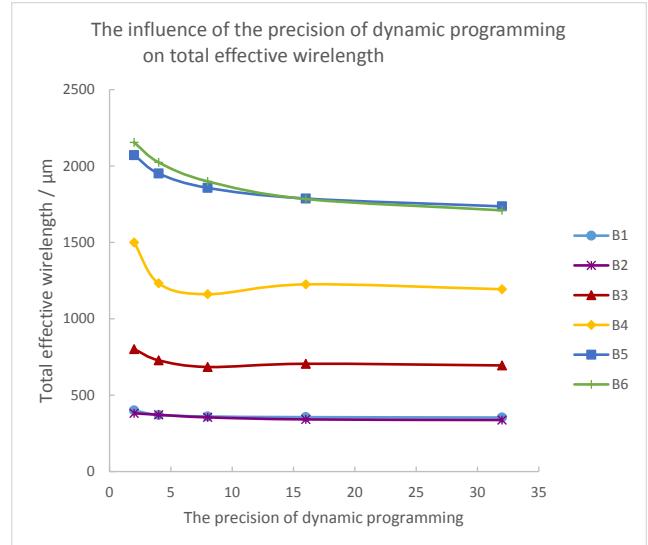


Fig. 14. Relationship between the precision of dynamic programming and the sum of total effective wirelength of one chip.

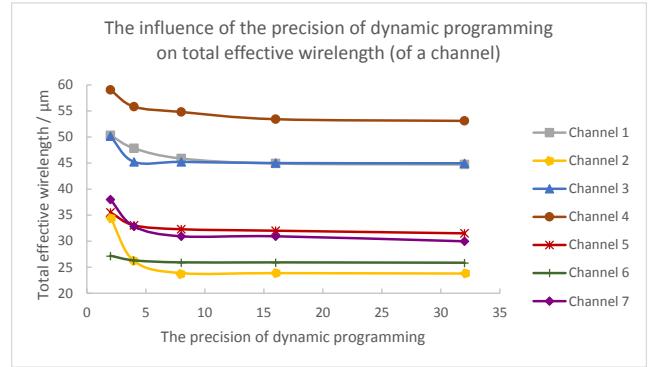


Fig. 15. Relationship between the precision of dynamic programming and the total effective wirelength of one specific flow channel.

shows the relationship between the precision of dynamic programming and the sum of total effective wirelength, whereas figure 15 shows the relationship between the precision of dynamic programming and the total effective wirelength of a specific flow channel. Both figures show that the total effective wirelength decreases as the precision of dynamic programming grows. And the total effective wirelength tends to converge when m is large enough. Therefore, our dynamic programming approach can practically shorten the total effective wirelength, and the routed channel approaches the optimal one when m is large enough. Moreover, large m does not severely increase the runtime. In Figure 14 and 15, the total effective wirelength converges when m is 20 or 30, which is sufficiently small without significantly affecting the runtime of the overall routing algorithm.

D. Comparison Between AARF and Manhattan Routing

Finally, we compare our AARF algorithm with a typical Manhattan routing algorithm [30]. The precision of dynamic programming is set to be 10 in this experiment. Table II shows the details of the benchmarks, where “Size” represents the size of the chip, “#Block” gives the number of blockages, “#Net”



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