

Deep-learning based automatic segmentation of vesicles in cryo-electron tomograms

This manuscript ([permalink](#)) was automatically generated from [elatella/deep-prepyto-paper@7ce4142](#) on June 21, 2022.

Authors

- **Amin Khosrozadeh***

 [XXXX-XXXX-XXXX-XXXX](#) ·  [ameen-khosrowzadeh](#)

Institute of Anatomy, University of Bern, Bern, Switzerland; Graduate School for Cellular and Biomedical Sciences, University of Bern · Funded by Grant XXXXXXXX

- **Raphaela Seeger***

 [XXXX-XXXX-XXXX-XXXX](#) ·  [elatella](#)

Institute of Anatomy, University of Bern, Bern, Switzerland; Graduate School for Cellular and Biomedical Sciences, University of Bern

- **Julika Radecke**

 [0000-0002-5815-5537](#) ·  [julikaradecke](#)

Institute of Anatomy, University of Bern, Bern, Switzerland; Department of Neuroscience, Faculty of Health and Medical Science, 2200 Copenhagen N, University of Copenhagen, Copenhagen, Denmark; Diamond Light Source Ltd, Didcot, Oxfordshire, United Kingdom

- **Guillaume Witz**

 [0000-0003-1562-4265](#) ·  [guiwitz](#)

Science IT Service, University of Bern, Bern, Switzerland; Microscopy Imaging Center, University of Bern, Bern, Switzerland

- **Jakob B. Sørensen**

 [0000-0001-5465-3769](#) ·  [JBSorensen](#)

Department of Neuroscience, University of Copenhagen, Blegdamsvej 3B, 2200 Copenhagen N, Denmark · Funded by Novo Nordisk Fonden, NNF17OC0028516.; Carlsbergfondet, CF17-0875; Independent Research Fond Denmark, 8020-00228A; Lundbeckfonden, R277-2018-802

- **Benoît Zuber[✉]**

 [0000-0001-7725-5579](#) ·  [aseedb](#)

Institute of Anatomy, University of Bern, Bern, Switzerland · Funded by Swiss National Science Foundation, 179520; ERA-NET NEURON, NEURON-119

✉ Address correspondence to benoit.zuber@ana.unibe.ch.

* These authors contributed equally.

Abstract

Cryo-electron Tomography (Cryo-ET) has the potential to reveal cell structure down to atomic resolution. Nevertheless, cellular cryo-ET data is often highly complex and visualization, as well as quantification, of subcellular structures require image segmentation. Due to a relatively high level of noise and to anisotropic resolution in cryo-ET data, automatic segmentation based on classical computer vision approaches usually does not perform satisfyingly. For this reason, cryo-ET researchers have mostly performed manual segmentation.

Communication between neurons rely on neurotransmitter-filled synaptic vesicle (SV) exocytosis. Recruitment of SVs to the plasma membrane is an important means of regulating exocytosis and is influenced by interactions between SVs. Cryo-ET study of the spatial organization of SVs and of their interconnections allows a better understanding of the mechanisms of exocytosis regulation. To obtain a faithful representation of SV connectivity state, an absolutely vital prerequisite is an extremely accurate SV segmentation. Hundreds to thousands of SVs are present in a typical synapse, and their manual segmentation is a burden. Typically accurately segmenting all SVs in one synapse takes between 3 to 8 days. This segmentation process has been widely recognized as a bottleneck by the community.

Several attempts to automate vesicle segmentation by classical computer vision or machine learning algorithms have not yielded very robust results. We addressed this problem by designing a workflow consisting of a U-Net convolutional network followed by post-processing steps. This combination yields highly accurate results. Furthermore, we provide an interactive tool for accurately segmenting spherical vesicles in a fraction of the time required by available manual segmentation methods. This tool can be used to segment vesicles that were missed by the fully automatic procedure or to quickly segment a handful of vesicles, while bypassing the fully automatic procedure. Our pipeline can in principle be used to segment any spherical vesicle in any cell type as well as extracellular vesicles.

Introduction

The fine architecture of cells can be investigated by cryo-electron tomography (cryo-ET) [1]. Cellular structures are preserved down to the atomic scale through vitrification and observation of the samples in a fully hydrated state. When a macromolecule is present in a sufficient number of copies in the cells imaged by cryo-ET, it is possible to obtain its atomic structure in situ using subtomogram averaging [2,3]. Cellular cryo-ET datasets are usually extremely complex, making them difficult to analyze. This is aggravated by the sensitivity of biological samples to electron radiation, which limits the signal-to-noise ratio in cryo-ET datasets [4]. Tomographic reconstructions are generated from a series of images of the sample acquired at different viewing angles. The geometry of the samples prevents acquisition at certain angles, resulting in anisotropic spatial coverage. The resolution in the directions close to the axis of the electron beam incident on the untilted sample is strongly reduced. This effect, commonly referred to as the missing-wedge artifact, further complicates data analysis. In particular, organelles fully bounded by a membrane appear to have holes at their top and bottom (relative to the electron beam axis) [4].

The synapse is the functional cellular contact at which information is transmitted from a neuron to another. The former neuron is called presynaptic and the latter is postsynaptic. In most cases, the signal is transmitted by the release of neurotransmitters into the intercellular space. Neurotransmitters are stored in SVs and are released following the fusion of a vesicle with the presynaptic plasma membrane. A synapse contains hundreds of SVs and their mobility and recruitability for neurotransmitter release depends on inter-vesicle interactions through so-called connector structures [5]. The characterization of these interactions can be performed automatically

with the pyto software, which implements a hierarchical connectivity approach to detect and annotate connectors [6]. For accurate connector segmentation, an exceptionally precise segmentation of SVs is prerequisite. To date, this SV segmentation has been achieved manually, but given the massive number of SVs per dataset, it is an extremely time-consuming process. Typically, one person spends 3 to 8 working days to segment a single dataset. Attempts to perform this task automatically based on classical computer vision algorithms have not yielded sufficiently accurate performance [7]. To alleviate this situation, we considered applying deep learning methods.

Convolutional neural networks (CNN) have been successfully employed to segment cryo-ET data [8]. Although entirely satisfying for visualization purposes, this approach has not met the requirements of pyto. A recent publication described accurate SV segmentation of transmission electron microscopy images using CNN, but it is limited to 2-dimensional images of resin-embedded synapses [9]. For our use-case, a common issue is that the input data consists of 2-dimensional images. In the first study, cryo-ET data are decomposed in individual 2-dimensional slices, which are handed as separate input to the CNN. The independent output prediction images are re-assembled in a 3-dimensional stack.[8] As discussed above, membranes oriented approximately parallel to the plane of the 2-dimensional tomographic images are not resolved. In the absence of contextual knowledge of the other 2-dimensional images, the CNN fails to segment these regions of the vesicles. Hence, spherical vesicles appear open, whereas we expect closed spherical objects. To overcome this limitation, we used a U-Net CNN that takes 3-dimensional images as input. U-Net architecture has been introduced in 2015 by Ronenberger et al. [10]. It consists of a contracting path, typical of CNN, and a symmetric expanding path. At each expansion step, the correspondingly cropped feature map of the contracting path is concatenated. The contracting path captures context, while the expanding path coupled with concatenation enables precise localization. The U-Net architecture was developed to achieve a fast and accurate segmentation of biomedical two-dimensional images, with the requirement of only a small fraction of training data in comparison to previous CNNs. It was then extended to segment 3-dimensional biomedical images (3D U-Net) [11]. Weigert et al. [12] implemented a U-Net for content-aware restoration (CARE) of 3-dimensional fluorescence microscopy datasets. They showed that it can restore information from anisotropic and very noisy datasets.

We implemented a 3D U-Net based on CARE building blocks and trained it with manually segmented datasets. This method provided good accuracy and was only slightly affected by the missing wedge artifact. Nevertheless, it was not quite sufficient for our downstream pyto analysis. Hence, we developed a post-processing method, which transforms the segmented objects into spheres and refines their radius and center location. This leads to a substantial accuracy improvement, which are reflected in better pyto performance. Additionally, we designed a multivariate ranking procedure, highlighting possibly wrongly segmented SVs. We also introduce a semi-automatic method to very quickly fix wrongly segmented and missed SVs.

Although our set of procedures was developed with the use case of SV segmentation in mind, it can be used to segment any other types of biological spherical vesicles, such as transport vesicles, secretory vesicles, endocytic vesicles, and extracellular vesicles.

Results

To automatize the tracing of cellular features in tomograms, also called segmentation, we developed a deep-learning pipeline, which segments 3D objects. -> similar approaches in 2D The manual segmentation of synaptic vesicles is one of the most time-intensive steps, when segmenting tomograms of presynaptic terminals. Synaptic vesicles also constitute a large, homogeneous group, which would create a large training set for deep learning applications. Therefore, we decided to develop the automatic segmentation for synaptic vesicles in an initial step. The used tomograms were previously manually segmented with IMOD [13]. In a next step, filaments connecting the synaptic

vesicles with each other (connectors) and to the active zone (AZ) were automatically segmented with the algorithm application Pyto [6].

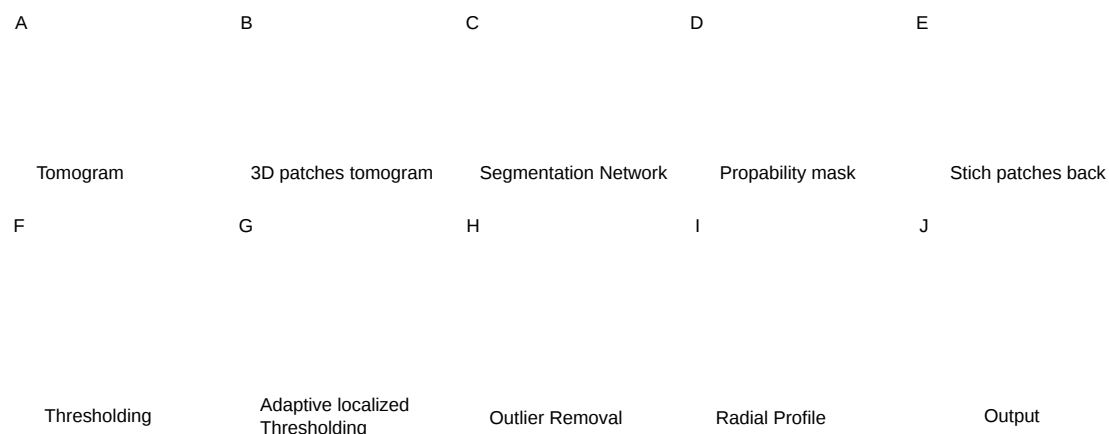


Figure 1: Pipeline of automatic segmentation. a) tomograms b) patchify the tomograms into 3D patches c) Segmentation Network d) probability masks e) stitching patches back f) thresholding g) adaptive localized thresholding h) outlier removal i) radial profile

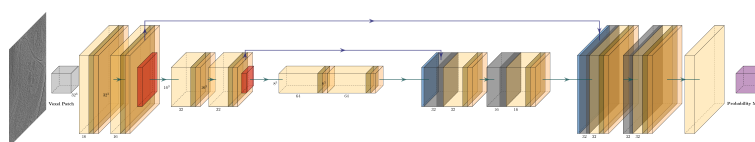


Figure 2: Segmentation Network: UNET. Input Size is 32^3 , in each resolution we have two convolution layer followed by batch normalization layer and relu activation function. Intermediate sizes are written on top of arrows, number of convolution filters is written bottom of boxes. Skip connections shows concatenation of the features from contracting path (left side of the network) and expansive path (right side of the network).

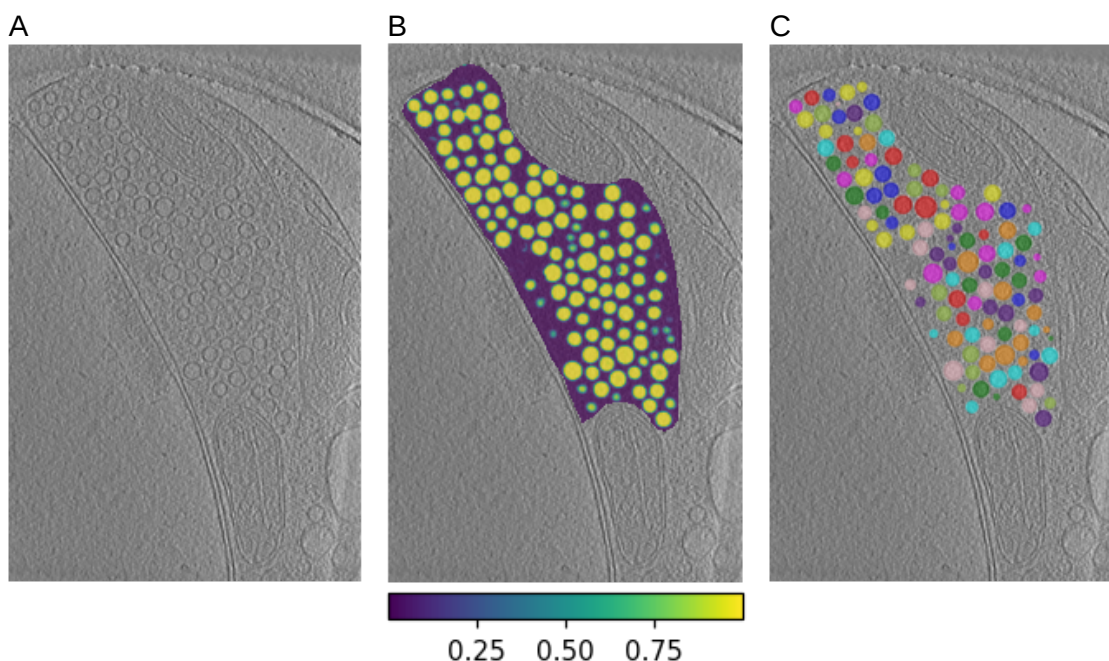


Figure 3: 2D Slices A) a section from z axis of a tomogram's presynaptic terminal of a neuron B) instance mask of the vesicles after post processing C) predicted probability mask by the segmentation network

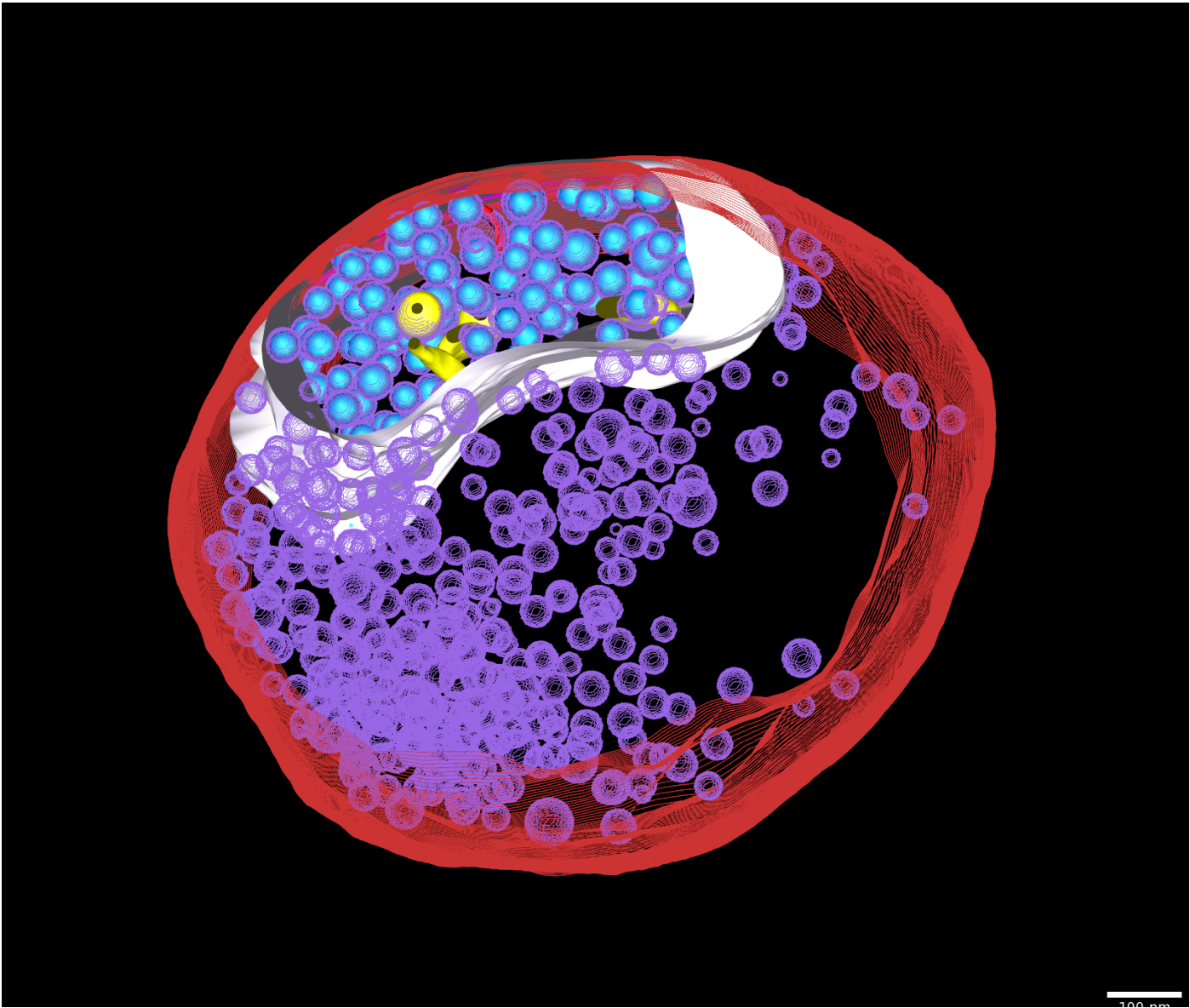
Fig 4- Dice improvements after post processing of initial predicted mask (different colors correspond to different tomograms): a) training datasets b) synaptosome test datasets c) Neuron test datasets

[Vesicle radius and position through radial profile and cross-correlation Radial Profile Refinement A) couple of vesicles are not centered B) Radial Profile. Blue range is from membrane

center to outer white halo center, this is the search range for the optimal radius. (smoothed by gaussian filtering) C) second derivative of radial profile E, F, H, G) Same as above columns after refinement.] (images/radial_avg_115-099.svg){#fig:radial_profile width="15cm"}

Fig 6- Splitting adjacent vesicles. A) Examples of tomogram, no labels; B) raw label with connected vesicle-labels; C) modified label with seperated vesicles —> for software: IMOD

Comparison of manual segmentation with automatic deep-learning based segmentation



3D model of manual segmented and automatically segmented synaptosome.

Evaluation metric DICE for pixel/pixel analysis

Table 1- Evaluation of the segmentation- MDICE: Mask Dice coefficient for the predicted mask PDICE: Dice coefficient after post-processing SIGMA-d: diameter error on correctly detected vesicle, DELTA-c: average error center (nm) #Vesicles: number of expected vesicles TP: True Positive FN: False Negative FP: False Positive

Global analysis

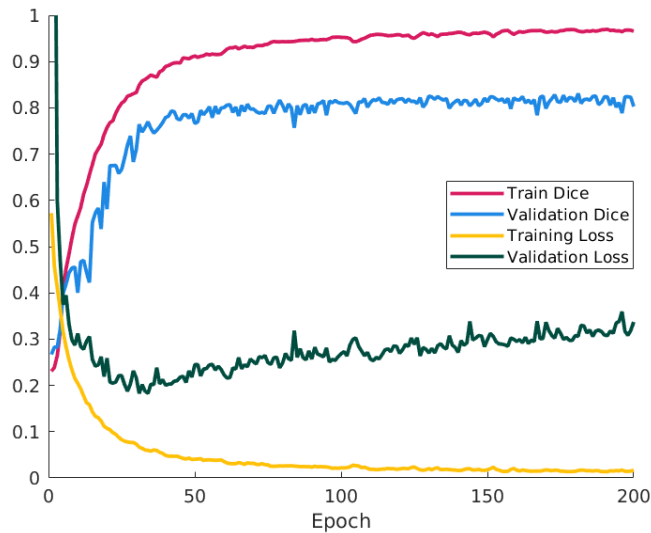


Figure 4: Dice coefficient and loss value for training and validation set.

3d unet good for 3D processing recent Nature methods paper by Ben Engel, DeepFinder -> Relion for STA creates mask to find more using dl -what are they doing, maybe compare that in the text, different aims; we might compare results we achieve (keep as bonus, revision)

3 different types of dl: classification, localization and segmentation We specialize in accurately segmenting 3D svcs -> results through the network, output is a mask, smooth DICE/ binary DICE = which one for our mask? one function for both? -> Amin will check when we encounter error until now we remove sv(?), bug inside function -> fix bug for better performance, how many fail and why? evaluation: objectwise evaluation for I/O, radius, center (according to nature paper) -> compare with manual seg other common evaluation tool other than DICE (Amin wants to check)

Discussion

DICE

Outlook

implement automatic cell-outline and active zone segmentation as deep learning workflow using UNet
implement automatic connector and tether segmentation as a deep learning workflow using UNet

Materials and methods

Benoît doing some tests.

Cryo-electron Tomography Datasets

Two datasets of different origin were used as input and test subjects for the automatic segmentation pipeline, rat synaptosomes as well as astrocytic and neural cell cultures derived from mice. `_cite`
Vladan, segmentation write a bit more*

Rat synaptosomes

The preparation of the rat synaptosomes were previously described [14]. After the purification, the synaptosomes were incubated for 30min at room temperature. 1.3mM CaCl₂ and 10 nm gold fiducials were added (gold fiducials, #s10110/8. AURION Immuno Gold Reagents & Accessories. Wageningen, The Netherlands). The synaptosome solution was applied to a 200-mesh lacey finder carbon film grid (#AGS166-H2. Agar Scientific. Elektron Technology UK Ltd. Stansted, UK). Manual blotting was used to remove excess liquid with the aid of a filter paper. Thereafter the grid was immediately plunge frozen in liquid ethane using a homebuilt plunge freezer controlled with a LabView script (National Instruments Corporation. Mopac Expwy Austin, TX, USA). The grids coated with rat synaptosomes were mounted in a cryo-holder (Gatan, Pleasanton, CA, USA) and transferred to a Tecnai F20 (FEI, Eindhoven, The Netherlands) which was set to low dose conditions, operated at 200 kV, and equipped with a field emission gun. Images were recorded with a 2k x 2k CCD camera (Gatan) mounted after a GIF Tridiem post-column filter (Gatan) operated in zero-loss mode. The sample was kept at about -180°C. Tilt series were acquired using SerialEM [15] for automated acquisition recorded typically from -50° to 50° with a 2° angular increment and an unbinned pixel size of 0.75 or 1.2 nm. Due to sample thickness (400-700 nm), tomograms were usually not recorded with higher tilt angles. Defocus was set between -8 to -12 µm and the total electron dose used was about 80-100 e⁻/Å². Some tomograms were acquired at a Titan Krios equipped with a K2 direct electron detector (Gatan) without energy filter. The K2 camera was operated in superresolution counting mode and between 8-40 frames per tilt angle were taken. Tilt series were acquired using the Latitude software (Gatan) for automated acquisition recorded typically from -60° to 60° with a 2° angular increment and an unbinned pixel size of 0.6 nm. Defocus was set between -8 to -12 µm and the total electron dose used was about 80-100 e⁻/Å². Prior to image processing the frames at each tilt angle, frames were aligned and averaged in 2dx MC_Automator [16] with motioncor [17]. 3D reconstruction was done in IMOD [13]. The alignments were done using the automated fiducial tracking function and the 3D reconstructions were done using the weighted back projection followed by a nonlinear anisotropic diffusion (NAD) filtering.

Astrocytic and neuronal mouse culture

The preparation of astrocytic and neuronal culture has been published before [18]. After 12 to 14 days of incubation grids with mouse neurons were plunge frozen with a Vitrobot (ThermoFisher Scientific, Mark IV) with a blot time of 3 s and a blot force of -10. Wait time and drain time were not used. Humidity was set to 100% at 4°C. 4 undiluted 10 nm BSA gold tracer (Aurion) was added directly onto the grid prior to plunge freezing. Cultured mouse neurons tilt series were acquired at a Titan Krios, equipped with a Falcon 3 direct electron detector (ThermoFisher Scientific) without energy filter. The Falcon camera was operated in linear mode. Tilt series were acquired using the TEM Tomography software (TFS) for automated acquisition recorded typically from -60° to 60° with a 2° angular increment and an unbinned pixel size of 0.37 nm. Defocus was set between -6 to -10 µm and the total electron dose used was about 80-100 e⁻/Å². Tomogram reconstruction was done in the same way as for the synaptosome datasets.

Manual segmentation procedures

Manual segmentation of SVs, mitochondria, and the active zone PM was done in IMOD (Figure S4A&B). The boundary marked the region to be analyzed by Pyto [6]. The analysis by Pyto was essentially the same as described previously [5] [6].

In short, the segmented area is divided in 1 voxel thick layers parallel to the active zone for distance calculations. A hierarchical connectivity segmentation detects densities interconnecting vesicles (so called connectors) and densities connecting vesicles to the active zone PM (so called tethers) (Figure S4C). Distance calculations are done with the center of the vesicle. Mainly default settings were used. The segmentation procedure is conservative and tends to miss some tethers and connectors because of noise. Consequently, the numbers of tethers and connectors should not be considered as absolute values but rather to compare experimental groups. As it was done before, an upper limit was set

between 2100 and 3200 nm³ on segment volume. The tomograms that were used for this analysis were binned by a factor of 2 to 3, resulting in voxel sizes between 2.1 and 2.4 nm.

Pre-processing of manual segmentation outputs from IMOD for further use (jupyter notebook pre-pyto)

Description of Machine Learning: Training Stage

Deep Model Training

Unet Training Datasets and Batch Generation

Transfer Learning

Optimization / Postprocessing

-Global Threshold

-Mask Tuning

-Compute Radial Profile

-Radius Estimation (Cross Correlation through Radial Profile) -Remove Outlier Labels

Analysis of Results

References

1. **Towards Visual Proteomics at High Resolution**
Felix JB B  uerlein, Wolfgang Baumeister
Journal of Molecular Biology (2021-10) <https://doi.org/gn9t3v>
DOI: [10.1016/j.jmb.2021.167187](https://doi.org/10.1016/j.jmb.2021.167187) · PMID: [34384780](https://pubmed.ncbi.nlm.nih.gov/34384780/)
2. **Exploring high-resolution cryo-ET and subtomogram averaging capabilities of contemporary DEDs**
Martin Obr, Wim JH Hagen, Robert A Dick, Lingbo Yu, Abhay Kotecha, Florian KM Schur
Molecular Biology (2022-01-10) <https://doi.org/gn92pd>
DOI: [10.1101/2022.01.10.475481](https://doi.org/10.1101/2022.01.10.475481)
3. **High-resolution in situ structure determination by cryo-electron tomography and subtomogram averaging using emClarity**
Tao Ni, Thomas Frosio, Luiza Mendon  a, Yuewen Sheng, Daniel Clare, Benjamin A Himes, Peijun Zhang
Nature Protocols (2022-02) <https://doi.org/gn92pc>
DOI: [10.1038/s41596-021-00648-5](https://doi.org/10.1038/s41596-021-00648-5) · PMID: [35022621](https://pubmed.ncbi.nlm.nih.gov/35022621/)
4. **STRUCTURAL STUDIES BY ELECTRON TOMOGRAPHY: From Cells to Molecules**
Vladan Lu  i  , Friedrich F  rster, Wolfgang Baumeister
Annual Review of Biochemistry (2005-06-01) <https://doi.org/cfd27f>
DOI: [10.1146/annurev.biochem.73.011303.074112](https://doi.org/10.1146/annurev.biochem.73.011303.074112) · PMID: [15952904](https://pubmed.ncbi.nlm.nih.gov/15952904/)
5. **Quantitative analysis of the native presynaptic cytomatrix by cryoelectron tomography**
Rub  n Fern  ndez-Busnadiego, Beno  t Zuber, Ulrike Elisabeth Maurer, Marek Cyrklaff, Wolfgang Baumeister, Vladan Lu  i  
Journal of Cell Biology (2010-01-11) <https://doi.org/b9c26b>
DOI: [10.1083/jcb.200908082](https://doi.org/10.1083/jcb.200908082) · PMID: [20065095](https://pubmed.ncbi.nlm.nih.gov/20065095/) · PMCID: [PMC2812849](https://pubmed.ncbi.nlm.nih.gov/PMC2812849/)
6. **Hierarchical detection and analysis of macromolecular complexes in cryo-electron tomograms using Pyto software**
Vladan Lu  i  , Rub  n Fern  ndez-Busnadiego, Ulrike Laugks, Wolfgang Baumeister
Journal of Structural Biology (2016-12) <https://doi.org/f9d5t2>
DOI: [10.1016/j.jsb.2016.10.004](https://doi.org/10.1016/j.jsb.2016.10.004) · PMID: [27742578](https://pubmed.ncbi.nlm.nih.gov/27742578/)
7. **Robust membrane detection based on tensor voting for electron tomography**
Antonio Martinez-Sanchez, Inmaculada Garcia, Shoh Asano, Vladan Lucic, Jose-Jesus Fernandez
Journal of Structural Biology (2014-04) <https://doi.org/f5zkj8>
DOI: [10.1016/j.jsb.2014.02.015](https://doi.org/10.1016/j.jsb.2014.02.015) · PMID: [24625523](https://pubmed.ncbi.nlm.nih.gov/24625523/)
8. **Convolutional neural networks for automated annotation of cellular cryo-electron tomograms**
Muyuan Chen, Wei Dai, Stella Y Sun, Darius Jonasch, Cynthia Y He, Michael F Schmid, Wah Chiu, Steven J Ludtke
Nature Methods (2017-10) <https://doi.org/gkpi62>
DOI: [10.1038/nmeth.4405](https://doi.org/10.1038/nmeth.4405) · PMID: [28846087](https://pubmed.ncbi.nlm.nih.gov/28846087/) · PMCID: [PMC5623144](https://pubmed.ncbi.nlm.nih.gov/PMC5623144/)
9. **Automated Detection and Localization of Synaptic Vesicles in Electron Microscopy Images**
Barbara Imbrosci, Dietmar Schmitz, Marta Orlando
eneuro (2022-01) <https://doi.org/gn9n8k>
DOI: [10.1523/eneuro.0400-20.2021](https://doi.org/10.1523/eneuro.0400-20.2021) · PMID: [34983830](https://pubmed.ncbi.nlm.nih.gov/34983830/) · PMCID: [PMC8805189](https://pubmed.ncbi.nlm.nih.gov/PMC8805189/)

10. **U-Net: Convolutional Networks for Biomedical Image Segmentation**
Olaf Ronneberger, Philipp Fischer, Thomas Brox
arXiv (2015-05-19) <https://arxiv.org/abs/1505.04597>
11. **3D U-Net: Learning Dense Volumetric Segmentation from Sparse Annotation**
Özgün Çiçek, Ahmed Abdulkadir, Soeren S Lienkamp, Thomas Brox, Olaf Ronneberger
arXiv (2016-06-22) <https://arxiv.org/abs/1606.06650>
12. **Content-aware image restoration: pushing the limits of fluorescence microscopy**
Martin Weigert, Uwe Schmidt, Tobias Boothe, Andreas Müller, Alexandr Dibrov, Akanksha Jain, Benjamin Wilhelm, Deborah Schmidt, Coleman Broadus, Siân Culley, ... Eugene W Myers
Nature Methods (2018-12) <https://doi.org/gfkkfd>
DOI: [10.1038/s41592-018-0216-7](https://doi.org/10.1038/s41592-018-0216-7) · PMID: [30478326](https://pubmed.ncbi.nlm.nih.gov/30478326/)
13. **Computer Visualization of Three-Dimensional Image Data Using IMOD**
James R Kremer, David N Mastronarde, JRichard McIntosh
Journal of Structural Biology (1996-01) <https://doi.org/d9nfzw>
DOI: [10.1006/jsbi.1996.0013](https://doi.org/10.1006/jsbi.1996.0013) · PMID: [8742726](https://pubmed.ncbi.nlm.nih.gov/8742726/)
14. **A rapid Percoll gradient procedure for preparation of synaptosomes**
Peter R Dunkley, Paula E Jarvie, Phillip J Robinson
Nature Protocols (2008-11) <https://doi.org/b7zwh8>
DOI: [10.1038/nprot.2008.171](https://doi.org/10.1038/nprot.2008.171) · PMID: [18927557](https://pubmed.ncbi.nlm.nih.gov/18927557/)
15. **Automated electron microscope tomography using robust prediction of specimen movements**
David N Mastronarde
Journal of Structural Biology (2005-10) <https://doi.org/ff7gzx>
DOI: [10.1016/j.jsb.2005.07.007](https://doi.org/10.1016/j.jsb.2005.07.007) · PMID: [16182563](https://pubmed.ncbi.nlm.nih.gov/16182563/)
16. **2dx_automator: Implementation of a semiautomatic high-throughput high-resolution cryo-electron crystallography pipeline**
Sebastian Scherer, Julia Kowal, Mohamed Chami, Venkata Dandey, Marcel Arheit, Philippe Ringler, Henning Stahlberg
Journal of Structural Biology (2014-05) <https://doi.org/f522h3>
DOI: [10.1016/j.jsb.2014.03.016](https://doi.org/10.1016/j.jsb.2014.03.016) · PMID: [24680783](https://pubmed.ncbi.nlm.nih.gov/24680783/)
17. **Electron counting and beam-induced motion correction enable near-atomic-resolution single-particle cryo-EM**
Xueming Li, Paul Mooney, Shawn Zheng, Christopher R Booth, Michael B Braunfeld, Sander Gubbens, David A Agard, Yifan Cheng
Nature Methods (2013-06) <https://doi.org/f4zpjf>
DOI: [10.1038/nmeth.2472](https://doi.org/10.1038/nmeth.2472) · PMID: [23644547](https://pubmed.ncbi.nlm.nih.gov/23644547/) · PMCID: [PMC3684049](https://pubmed.ncbi.nlm.nih.gov/pmc/PMC3684049/)
18. **Preparation of Primary Neurons for Visualizing Neurites in a Frozen-hydrated State Using Cryo-Electron Tomography**
Sarah H Shahmoradian, Mauricio R Galiano, Chengbiao Wu, Shurui Chen, Matthew N Rasband, William C Mobley, Wah Chiu
Journal of Visualized Experiments (2014-02-12) <https://doi.org/gmh9w3>
DOI: [10.3791/50783](https://doi.org/10.3791/50783) · PMID: [24561719](https://pubmed.ncbi.nlm.nih.gov/24561719/) · PMCID: [PMC4089403](https://pubmed.ncbi.nlm.nih.gov/pmc/PMC4089403/)