

# Deep-learning based automatic segmentation of synaptic vesicles in cryo-electron tomograms

This manuscript ([permalink](#)) was automatically generated from [elatella/deep-prepyto-paper@67db72b](#) on January 10, 2022.

## Authors

---

- **Amin Khosrozadeh\***

 [XXXX-XXXX-XXXX-XXXX](#) ·  [ameen-khosrowzadeh](#)

Institute of Anatomy, University of Bern, Bern, Switzerland; Graduate School for Cellular and Biomedical Sciences, University of Bern · Funded by Grant XXXXXXXX

- **Raphaela Seeger\***

 [XXXX-XXXX-XXXX-XXXX](#) ·  [elatella](#)

Institute of Anatomy, University of Bern, Bern, Switzerland; Graduate School for Cellular and Biomedical Sciences, University of Bern

- **Julika Radecke**

 [0000-0002-5815-5537](#) ·  [julikaradecke](#)

Institute of Anatomy, University of Bern, Bern, Switzerland; Department of Neuroscience, Faculty of Health and Medical Science, 2200 Copenhagen N, University of Copenhagen, Copenhagen, Denmark; Diamond Light Source Ltd, Didcot, Oxfordshire, United Kingdom

- **Guillaume Witz**

 [0000-0003-1562-4265](#) ·  [guiwitz](#)

Science IT Service, University of Bern, Bern, Switzerland

- **Vladan Lučić**

 [0000-0003-3698-7436](#) ·  [vladanl](#)

Max-Planck-Institute of Biochemistry, Am Klopferspitz 18, 82152 Martinsried, Germany

- **Jakob B. Sørensen**

 [0000-0001-5465-3769](#) ·  [JBSorensen](#)

Department of Neuroscience, University of Copenhagen, Blegdamsvej 3B, 2200 Copenhagen N, Denmark · Funded by Novo Nordisk Fonden, NNF17OC0028516.; Carlsbergfondet, CF17-0875; Independent Research Fond Denmark, 8020-00228A; Lundbeckfonden, R277-2018-802

- **Benoît Zuber**✉

 [0000-0001-7725-5579](#) ·  [aseedb](#)

Institute of Anatomy, University of Bern, Bern, Switzerland · Funded by Swiss National Science Foundation, 179520; ERA-NET NEURON, NEURON-119

✉ Address correspondence to [benoit.zuber@ana.unibe.ch](mailto:benoit.zuber@ana.unibe.ch).

\* These authors contributed equally.

# Abstract

---

The communication between neurons is based on the exocytosis of neurotransmitter filled vesicles. Even small disbalances in this delicate system can result in different disorders. Numerous questions are still unanswered about synaptic vesicle exocytosis regulation as well as about which proteins affect this fundamental process. Hundreds to thousands of vesicles are present at the synapse and most of them are interconnected. There is a number of evidence suggesting that the connectivity between the vesicles influences synaptic vesicle exocytotic activity and that connectivity can be regulated. To address these issues, cryo-electron tomography is a method of choice. Yet inevitable quantitative studies require accurate segmentation of both synaptic vesicles and the interconnecting filaments.

This segmentation process has been widely recognized as a bottleneck by the community. When working with synapses, an incredibly tedious cellular feature to segment are synaptic vesicles. Even though they have a very homogeneous shape and size, which makes it easy to distinguish them from other cellular features, their high occurrence makes their segmentation challenging (one average-sized synapse or synaptosomes can easily contain between 300-500 synaptic vesicles).

Several attempts at utilizing machine learning algorithms to automatize the segmentation of synaptic vesicles yielded in unfavorable results. We analyzed these approaches and realized that most algorithms solely worked with the 2D image planes within the tomogram, but didn't tap into their 3D information and therefore increased the weight of the missing wedge.

We designed a framework that consists of two parts: 1) U-Net convolutional network to obtain mask for SVs following by 2) post-processing step which fine-tunes the mask and provides entire 3D-vesicles. A deep-learning model has been trained with manually segmented synaptic vesicles, which have been entirely annotated as 3D objects. By post-processing, we achieved to transfer the vesicle recognition through this pipeline. The evaluation metrics like the DICE value suggest a very high success rate.

## Introduction

---

The rise of machine learning is sweeping the technological landscape. Its use cases are highly diverse and are just starting to reach the scientific community. Especially the graphical deep-learning approaches have been quite popular [\[1\]](#). The graphical deep-learning approach is based around learning the description of the common attributes of a group or subset of a universe of objects [\[2\]](#). Describe the typical make up of a deeplearning approach But also the field of cryo electron microscopy has recently recognized the benefits of machine/deep-learning approaches [\[3\]](#).

The method of cryo-electron tomography (ET) utilises a series of tilted two-dimensional (2D) images to reconstruct a three-dimensional (3D) image of the imaged sample [\[4\]](#). It brings many advantages compared to other cryo-electron microscopy (EM) methods, as it yields images at in situ conditions. Therefore cryo ET can help to reveal the natural cellular environment of protein(-complexes), their function as well as their structure. In order to analyze the cellular context within a tomogram, cellular features are being segmented. Popular programs for manual segementation are IMOD and Amira [\[5,6\]](#). Currently manual segmentation is being considered the bottleneck in the workflow of cellular cryo-ET.

The communication between neurons is based on the exocytosis of neurotransmitter filled vesicles. The communication between CNS synapses depends on the very low basal vesicle fusion rate and the ability to rapidly upregulate that rate upon stimulation. The SNARE complex connects the synaptic

vesicles and the presynaptic cell membrane, mediating vesicle fusion. Their function and regulation still pose big questions. In addition, mutations of involved proteins have been linked to numerous neurological diseases. To address these issues, cryo-ET is the method of choice. To perform quantitative studies on the resulting tomograms, accurate segmentation of both synaptic vesicles and the interconnecting filaments are required. Segmenting the cellular features manually is a very time-intensive step. Among the different cell features, synaptic vesicles are the most time-consuming to segment. Despite their very homogeneous size and shape, their sheer number and finding their true center (due to the missing wedge problem) complicates their segmentation. Problem 2D and 3D

**Aim** Creation of a deep-learning pipeline which is taught to segment cellular features, starting with synaptic vesicles. We decided to teach our network with a real world dataset rather than using synthetic datasets, as it is often custom, as the synthetic datasets lack noise and could represent a complete 360° 3D object, which our datasets cannot due to the missing wedge problem in cryo-ET. proposed method provides the initial step for downstream segmentation of subcellular compartments such as synaptic connectors and tethers for which an extremely accurate vesicle segmentation is required

## Results

---

### Comparison of manual segmentation with automatic deep-learning based segmentation

Evaluation metric DICE for pixel/pixel analysis Global analysis

## Discussion

---

DICE

## Outlook

implement automatic cell-outline and active zone segmentation as deep learning workflow using UNet  
implement automatic connector and tether segmentation as a deep learning workflow using UNet

## Materials and methods

---

### Cryo-electron Tomography Datasets

Two datasets of different origin were used as input and test subjects for the automatic segmentation pipeline, rat synaptosomes as well as astrocytic and neural cell cultures derived from mice.

#### Rat synaptosomes

The preparation of the rat synaptosomes were previously described [7]. After the purification, the synaptosomes were incubated for 30min at room temperature. 1.3mM CaCl<sub>2</sub> and 10 nm gold fiducials were added (gold fiducials, #s10110/8. AURION Immuno Gold Reagents & Accessories. Wageningen, The Netherlands). The synaptosome solution was applied to a 200-mesh lacey finder carbon film grid (#AGS166-H2. Agar Scientific. Elektron Technology UK Ltd. Stansted, UK). Manual blotting was used to remove excess liquid with the aid of a filter paper. Thereafter the grid was immediately plunge frozen in liquid ethane using a homebuilt plunge freezer controlled with a LabView script (National

Onstruments Corporation. Mopac Expwy Austin, TX, USA). The grids coated with rat synaptosomes were mounted in a cryo-holder (Gatan, Pleasanton, CA, USA) and transferred to a Tecnai F20 (FEI, Eindhoven, The Netherlands) which was set to low dose conditions, operated at 200 kV, and equipped with a field emission gun. Images were recorded with a 2k x 2k CCD camera (Gatan) mounted after a GIF Tridiem post-column filter (Gatan) operated in zero-loss mode. The sample was kept at about -180°C. Tilt series were acquired using SerialEM [8] for automated acquisition recorded typically from -50° to 50° with a 2° angular increment and an unbinned pixel size of 0.75 or 1.2 nm. Due to sample thickness (400-700 nm), tomograms were usually not recorded with higher tilt angles. Defocus was set between -8 to -12  $\mu\text{m}$  and the total electron dose used was about 80-100  $\text{e}^-/\text{\AA}^2$ . Some tomograms were acquired at a Titan Krios equipped with a K2 direct electron detector (Gatan) without energy filter. The K2 camera was operated in superresolution counting mode and between 8-40 frames per tilt angle were taken. Tilt series were acquired using the Latitude software (Gatan) for automated acquisition recorded typically from -60° to 60° with a 2° angular increment and an unbinned pixel size of 0.6 nm. Defocus was set between -8 to -12  $\mu\text{m}$  and the total electron dose used was about 80-100  $\text{e}^-/\text{\AA}^2$ . Prior to image processing the frames at each tilt angle, frames were aligned and averaged in 2dx MC\_Automator [9] with motioncor [10]. 3D reconstruction was done in IMOD [5]. The alignments were done using the automated fiducial tracking function and the 3D reconstructions were done using the weighted back projection followed by a nonlinear anisotropic diffusion (NAD) filtering.

## Astrocytic and neuronal mouse culture

The preparation of astrocytic and neuronal culture has been published before [11]. After 12 to 14 days of incubation grids with mouse neurons were plunge frozen with a Vitrobot (ThermoFisher Scientific, Mark IV) with a blot time of 3 s and a blot force of -10. Wait time and drain time were not used. Humidity was set to 100% at 4°C. 4 undiluted 10 nm BSA gold tracer (Aurion) was added directly onto the grid prior to plunge freezing. Cultured mouse neurons tilt series were acquired at a Titan Krios, equipped with a Falcon 3 direct electron detector (ThermoFisher Scientific) without energy filter. The Falcon camera was operated in linear mode. Tilt series were acquired using the TEM Tomography software (TFS) for automated acquisition recorded typically from -60° to 60° with a 2° angular increment and an unbinned pixel size of 0.37 nm. Defocus was set between -6 to -10  $\mu\text{m}$  and the total electron dose used was about 80-100  $\text{e}^-/\text{\AA}^2$ . Tomogram reconstruction was done in the same way as for the synaptosome datasets.

## Manual segmentation procedures

Manual segmentation of SVs, mitochondria, and the active zone PM was done in IMOD (Figure S4A&B). The boundary marked the region to be analyzed by Pyto [12]. The analysis by Pyto was essentially the same as described previously [13] [12].

In short, the segmented area is divided in 1 voxel thick layers parallel to the active zone for distance calculations. A hierarchical connectivity segmentation detects densities interconnecting vesicles (so called connectors) and densities connecting vesicles to the active zone PM (so called tethers) (Figure S4C). Distance calculations are done with the center of the vesicle. Mainly default settings were used. The segmentation procedure is conservative and tends to miss some tethers and connectors because of noise. Consequently, the numbers of tethers and connectors should not be considered as absolute values but rather to compare experimental groups. As it was done before, an upper limit was set between 2100 and 3200  $\text{nm}^3$  on segment volume. The tomograms that were used for this analysis were binned by a factor of 2 to 3, resulting in voxel sizes between 2.1 and 2.4 nm.

## Description of Machine Learning: Training Stage

### Deep Model Training

Unet Training Datasets and Batch Generation

## **Transfer Learning**

### **Optimization / Postprocessing**

-Global Threshold -Mask Tuning -Compute Radial Profile -Radius Estimation (Cross Correlation through Radial Profile) -Remove Outlier Labels

## **Analysis of Results**

# References

---

1. **Machine Learning in Medicine**  
Alvin Rajkomar, Jeffrey Dean, Isaac Kohane  
*New England Journal of Medicine* (2019-04-04) <https://doi.org/gfzk8r>  
DOI: [10.1056/nejmra1814259](https://doi.org/10.1056/nejmra1814259) · PMID: [30943338](https://pubmed.ncbi.nlm.nih.gov/30943338/)
2. **A review of machine learning**  
David M Dutton, Gerard V Conroy  
*The Knowledge Engineering Review* (1997-12) <https://doi.org/dcswnd>  
DOI: [10.1017/s026988899700101x](https://doi.org/10.1017/s026988899700101x)
3. **Application of machine learning techniques to electron microscopic/spectroscopic image data analysis**  
Shunsuke Muto, Motoki Shiga  
*Microscopy* (2020-04-08) <https://doi.org/ggckm4>  
DOI: [10.1093/jmicro/dfz036](https://doi.org/10.1093/jmicro/dfz036) · PMID: [31682260](https://pubmed.ncbi.nlm.nih.gov/31682260/) · PMCID: [PMC7141894](https://pubmed.ncbi.nlm.nih.gov/PMC7141894/)
4. **Current data processing strategies for cryo-electron tomography and subtomogram averaging**  
Euan Pyle, Giulia Zanetti  
*Biochemical Journal* (2021-05-28) <https://doi.org/gkqqv6>  
DOI: [10.1042/bcj20200715](https://doi.org/10.1042/bcj20200715) · PMID: [34003255](https://pubmed.ncbi.nlm.nih.gov/34003255/) · PMCID: [PMC8133831](https://pubmed.ncbi.nlm.nih.gov/PMC8133831/)
5. **Computer Visualization of Three-Dimensional Image Data Using IMOD**  
James R Kremer, David N Mastronarde, JRichard McIntosh  
*Journal of Structural Biology* (1996-01) <https://doi.org/d9nfzw>  
DOI: [10.1006/jsbi.1996.0013](https://doi.org/10.1006/jsbi.1996.0013) · PMID: [8742726](https://pubmed.ncbi.nlm.nih.gov/8742726/)
6. **amira: A Highly Interactive System for Visual Data Analysis**  
Detlev Stalling, Malte Westerhoff, Hans-Christian Hege  
*Visualization Handbook* (2005) <https://doi.org/dj723z>  
DOI: [10.1016/b978-012387582-2/50040-x](https://doi.org/10.1016/b978-012387582-2/50040-x) · ISBN: 9780123875822
7. **A rapid Percoll gradient procedure for preparation of synaptosomes**  
Peter R Dunkley, Paula E Jarvie, Phillip J Robinson  
*Nature Protocols* (2008-11) <https://doi.org/b7zwh8>  
DOI: [10.1038/nprot.2008.171](https://doi.org/10.1038/nprot.2008.171) · PMID: [18927557](https://pubmed.ncbi.nlm.nih.gov/18927557/)
8. **Automated electron microscope tomography using robust prediction of specimen movements**  
David N Mastronarde  
*Journal of Structural Biology* (2005-10) <https://doi.org/ff7gzx>  
DOI: [10.1016/j.jsb.2005.07.007](https://doi.org/10.1016/j.jsb.2005.07.007) · PMID: [16182563](https://pubmed.ncbi.nlm.nih.gov/16182563/)
9. **2dx\_automator: Implementation of a semiautomatic high-throughput high-resolution cryo-electron crystallography pipeline**  
Sebastian Scherer, Julia Kowal, Mohamed Chami, Venkata Dandey, Marcel Arheit, Philippe Ringler, Henning Stahlberg  
*Journal of Structural Biology* (2014-05) <https://doi.org/f522h3>  
DOI: [10.1016/j.jsb.2014.03.016](https://doi.org/10.1016/j.jsb.2014.03.016) · PMID: [24680783](https://pubmed.ncbi.nlm.nih.gov/24680783/)
10. **Electron counting and beam-induced motion correction enable near-atomic-resolution single-particle cryo-EM**

Xueming Li, Paul Mooney, Shawn Zheng, Christopher R Booth, Michael B Braunfeld, Sander Gubbens, David A Agard, Yifan Cheng  
*Nature Methods* (2013-06) <https://doi.org/f4zpjf>  
DOI: [10.1038/nmeth.2472](https://doi.org/10.1038/nmeth.2472) · PMID: [23644547](https://pubmed.ncbi.nlm.nih.gov/23644547/) · PMCID: [PMC3684049](https://pubmed.ncbi.nlm.nih.gov/PMC3684049/)

11. **Preparation of Primary Neurons for Visualizing Neurites in a Frozen-hydrated State Using Cryo-Electron Tomography**

Sarah H Shahmoradian, Mauricio R Galiano, Chengbiao Wu, Shurui Chen, Matthew N Rasband, William C Mobley, Wah Chiu  
*Journal of Visualized Experiments* (2014-02-12) <https://doi.org/gmh9w3>  
DOI: [10.3791/50783](https://doi.org/10.3791/50783) · PMID: [24561719](https://pubmed.ncbi.nlm.nih.gov/24561719/) · PMCID: [PMC4089403](https://pubmed.ncbi.nlm.nih.gov/PMC4089403/)

12. **Hierarchical detection and analysis of macromolecular complexes in cryo-electron tomograms using Pyto software**

Vladan Lučić, Rubén Fernández-Busnadiego, Ulrike Laugks, Wolfgang Baumeister  
*Journal of Structural Biology* (2016-12) <https://doi.org/f9d5t2>  
DOI: [10.1016/j.jsb.2016.10.004](https://doi.org/10.1016/j.jsb.2016.10.004) · PMID: [27742578](https://pubmed.ncbi.nlm.nih.gov/27742578/)

13. **Quantitative analysis of the native presynaptic cytomatrix by cryoelectron tomography**

Rubén Fernández-Busnadiego, Benoît Zuber, Ulrike Elisabeth Maurer, Marek Cyrklaff, Wolfgang Baumeister, Vladan Lučić  
*Journal of Cell Biology* (2010-01-11) <https://doi.org/b9c26b>  
DOI: [10.1083/jcb.200908082](https://doi.org/10.1083/jcb.200908082) · PMID: [20065095](https://pubmed.ncbi.nlm.nih.gov/20065095/) · PMCID: [PMC2812849](https://pubmed.ncbi.nlm.nih.gov/PMC2812849/)