



Deep Learning for Malaria Detection

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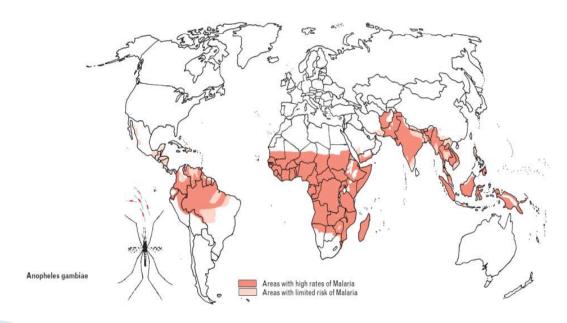
Objective

Problem Statement

- Malaria remains a global health concern
- Microscopic diagnosis, though highly accurate, is tedious, time consuming, and highly dependent on technician's expertise
- Early and accurate detection can dramatically improve morbidity and mortality rates

Solution

 Develop a CNN (Convolution Neural Network) model to analyze blood smear images; and improve efficiency, accuracy, and scalability of malaria diagnosis



Source - https://www.iamat.org/risks/malaria

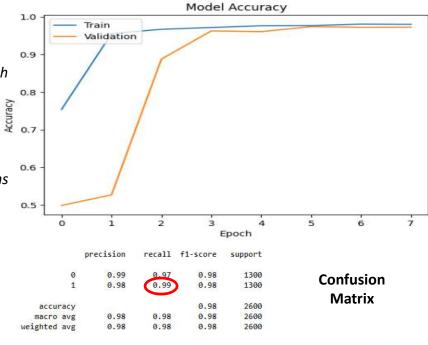
Observations & Next Steps

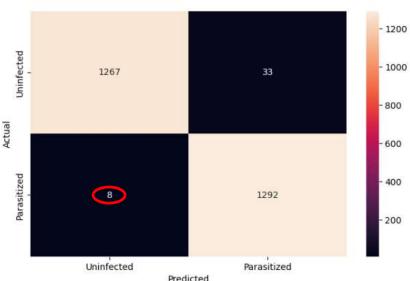
Observations:

- High Accuracy Compared to the other models, this model achieved high accuracy of 98%, and classified only 8 false positives (predicted = uninfected, actual = parasitized) driving the recall to 0.99
- Generalization The convergence between training and validation accuracy (98%) indicates the model has generalized well to the variations in the dataset
- Training time The model took only 8 epochs to train, even with 4 convolution layers and ~0.6M parameters

Next Steps

- Data Quality/Bias Training the models with high-quality labeled datasets that have diversity in geographic regions, patient demographics, and malaria species would be essential to limit bias and improve generalization of the model
- Explainability In order to make the model more interpretable explore what features are being learnt and how they influence the classification.
 This can be done through SHAP and/or plotting the features being learned by each layer





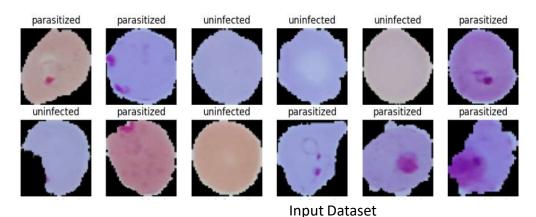
Solution Architecture

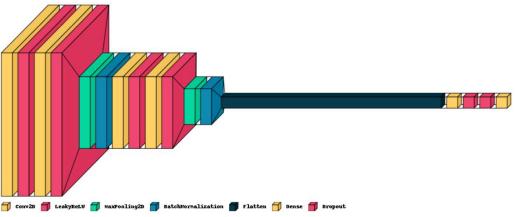
Dataset:

- Train 25K labeled RGB images (12.6K parasitized, 12.4K uninfected)
- Test 2.6K labeled RGB images (1.3K parasitized, 1.3K uninfected)

Model:

- 4 convolution 2D layers of 16, 32, 32, 64 filters respectively to learn features such as what species of parasite is present, what is the stage of infection, what are the changes in RBC morphology, etc.
- LeakyReLU activation function with a negative slope of 0.1 to capture complex relationships within the data by avoiding dead neurons
- Max pooling with filter size 2x2 to extract dominant features
- Regularization
 - Batch Normalization to fasten training and reduce sensitivity to initialization
 - Dropout rate of 0.5 to reduce overfitting
- Output layer sigmoid activation for binary classification





Malaria Detection CNN Model Architecture

Things to Consider

Deployment –

- Model conversion Convert the model to a format compatible with the desired deployment environment. This may involve converting to TensorFlow Lite for mobile deployment or ONNX for cloud deployment
- API development Develop an API that allows users to submit blood smear images and receive predictions from the model
- Integration Integrate the API into the desired platform (web app, mobile app, etc.)

Other factors:

- o Computation resources Training deep CNN is computationally expensive and can be a challenge if hardware resources are limited
- Model drift Keeping up with new parasites strains and lab practices would be key to the long term efficacy of the CNN model, which could be cost prohibitive
- Accessibility CNN model when deployed in areas with constrained healthcare personnel resources, can increase accessibility and provide significant public health benefits

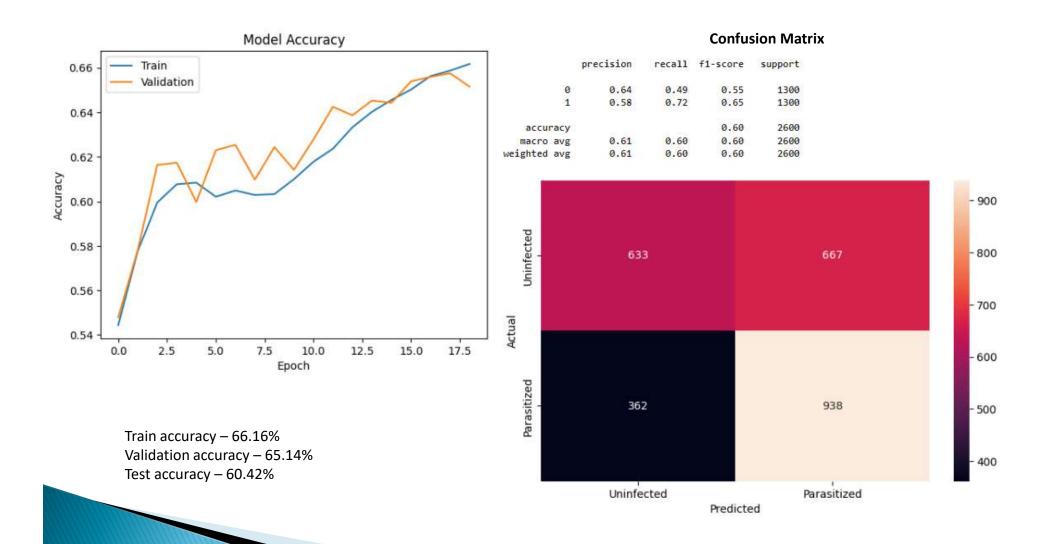
Appendix

Model Comparison

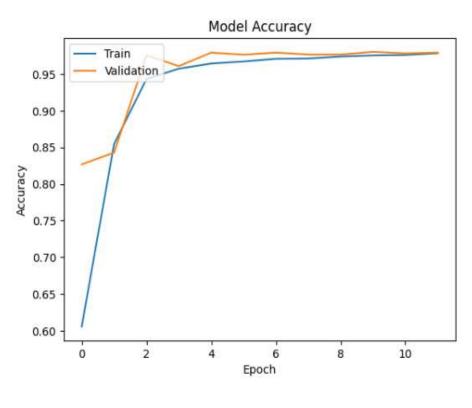
Model Name	Model Technique	Activation	Optimizer	Epoch	Batch Size	Image type	Trainable Params	Non-Trainable Params	Ttl Params	Train-Time (# of Epochs)	Train Accuracy	Val Accuracy	Test Accuracy	F1 score	Precision	Recall
model_base	2 CONV2D + Maxpooling + Flatten + Dense + Output	relu	SGD	20	64	BRG normalized	1,053,762	0	1,053,762	20	66%	65%	60%	0.65	0.58	0.72
model1	2 CONV2D + Maxpooling + 2 CONV2D + Maxpooling + Flatten + Dense + Output	relu	Adam	30	128	BRG normalized	557,218	0	557,218	12	98%	98%	99%	0.99	0.98	0.99
model2	2 CONV2D + Maxpooling + BatchNormalization + 2 CONV2D + Maxpooling + BatchNormalization + Flatten + Dense + Droput (0.5) + Output	LeakyReLU(0.1)	Adam	30	128	BRG normalized	557,410	192	557,602	8	98%	97%	98%	0.98	0.98	0.99
model3	2 CONV2D + Maxpooling + BatchNormalization + 2 CONV2D + Maxpooling + BatchNormalization + Flatten + Dense + Droput (0.5) + Output	LeakyReLU(0.1)	Adam	30	128	Image Data Generator	557,410	192	557,602	8	92%	96%	96%	0.96	0.94	1.00
model4	VGG16 + Flatten + Output		Adam	30	128	BRG normalized	1,026	14,714,688	14,715,714	30	94%	94%	91%	0.91	0.91	0.92
model_hsv	2 CONV2D + Maxpooling + Flatten + Dense + Droput (0.4) + Output	LeakyReLU(0.1)	Adam	30	128	HSV normalized	926,242	0	926,242	4	98%	98%	99%	0.99	0.98	0.99
model_gb	2 CONV2D + Maxpooling + BatchNormalization + 2 CONV2D + Maxpooling + BatchNormalization + 2 CONV2D + Maxpooling + BatchNormalization + Flatten + Dense + Droput (0.4) + Output	LeakyReLU(0.1)	Adam	30	128	Gausian Blur normalized	926,946	704	927,650	22	98%	98%	98%	0.98	0.97	0.99

Final Model

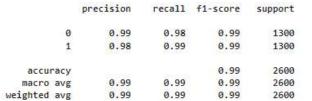
Base Model – Training Accuracy & Confusion Matrix

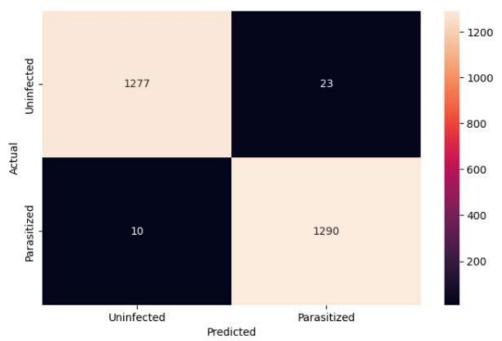


Model1 - Training Accuracy & Confusion Matrix



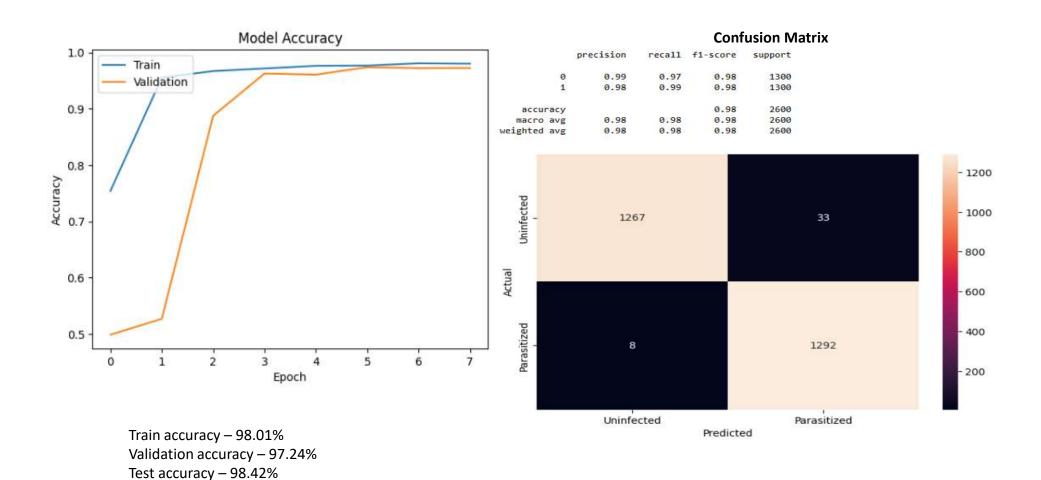
Train accuracy – 97.85% Validation accuracy – 97.92% Test accuracy – 98.73%



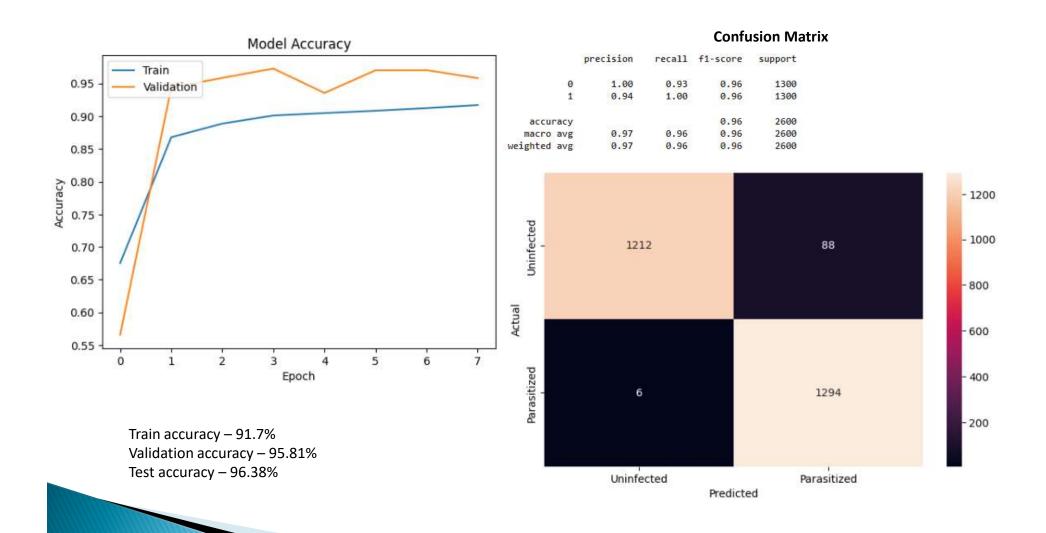


Confusion Matrix

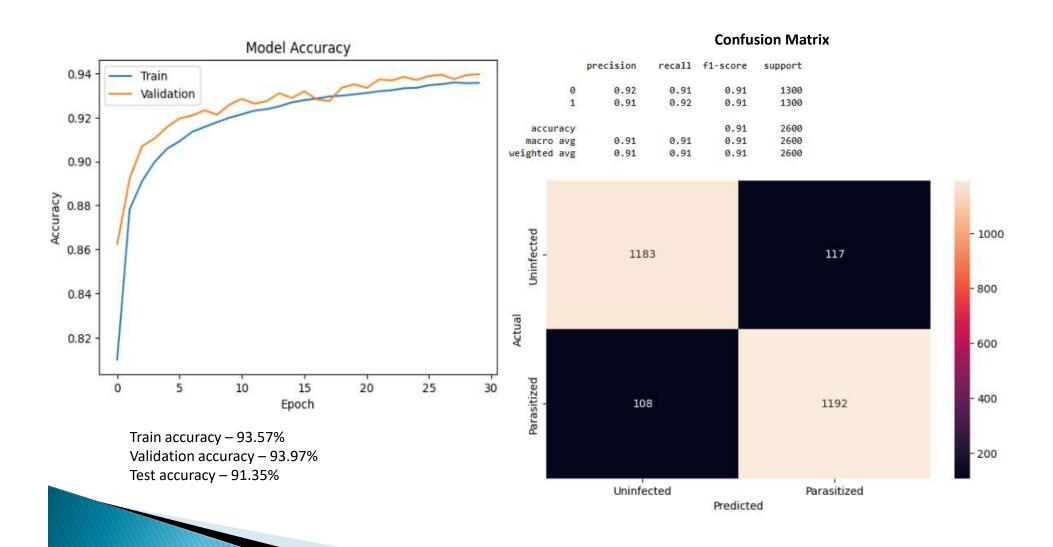
Model2 - Training Accuracy & Confusion Matrix



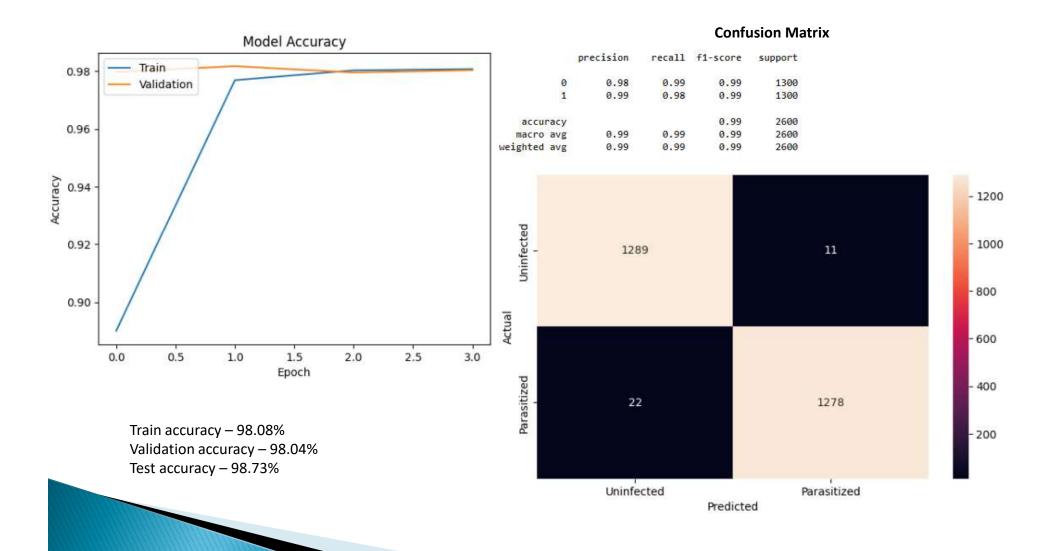
Model3 - Training Accuracy & Confusion Matrix



Model4 - Training Accuracy & Confusion Matrix



Model_HSV - Training Accuracy & Confusion Matrix



Model_GB - Training Accuracy & Confusion Matrix

