

Density-dependence seedling mortality in Kadumane

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set up

```
library(tidyverse)
```

```
## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --
## v dplyr      1.1.4      v readr      2.1.5
## v forcats    1.0.0      v stringr    1.5.1
## v ggplot2    3.5.1      v tibble     3.2.1
```

```
## v lubridate 1.9.3      v tidyr      1.3.1
## v purrr      1.0.2
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()    masks stats::lag()
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to become errors
```

```
library(ggthemes)
library(knitr)

#library(vegan)
#library(grid)
#library(geosphere)
#library(mgcv)
library(glmmTMB)
library(DHARMA)
```

```
## This is DHARMA 0.4.6. For overview type '?DHARMA'. For recent changes, type news(package = 'DHARMA')
```

```
library(broom.mixed)
library(ggeffects)
library(sjPlot)
library(patchwork)
```

```
theme_set(theme_tufte())
```

Data

Load data

```
sp_codes <- read_csv("data/sp_codes.csv")
```

```
## Rows: 5 Columns: 4
## -- Column specification -----
## Delimiter: ","
## chr (4): code, genus, species, family
##
## i Use 'spec()' to retrieve the full column specification for this data.
## i Specify the column types or set 'show_col_types = FALSE' to quiet this message.
```

```
site_dat <- read_rds("data/kadumane_site_metadata.rds")
sdls <- read_rds("data/kadumane_seedlings.rds")
plot_dat <- read_rds("data/kadumane_plot_metadata.rds")
```

1. Join in site and plot meta-data.
2. Remove rows for species with no seedlings at start of the census in plot.
3. Select species that

- Are recorded in at least 5 plots.
- Vary in density among plots.
- Are groups of many (unidentified) species with the same code.

```
sdls <- sdls |> left_join(site_dat) |> left_join(plot_dat) |>
  filter(census.start > 0)
```

```
## Joining with 'by = join_by(site)'
## Joining with 'by = join_by(site, location, group, plot)'
```

```
## find species with enough individuals and variation in density to allow
## analysis
sp_list <- group_by(sdls, species) |>
  summarise(abund = sum(census.start) , ## total abund
            n = sum(census.start > 0), ## number of plots with the species
            sd_dens = sd(census.start[census.start > 0])) |> ##var in density
  filter(n > 1) |> arrange(n)
## only lose 8 species by restricting to 5 or more occurrences (instead of 1)
## Seems reasonable. Also removing species that were unreliably identified.
##
## @Ashwin - what was SC and Artoth?
sp_list <- filter(sp_list, n > 4, !(species %in% c("Palm", "Artoth", "SC")),
                 sd_dens > 0)
sdls <- filter(sdls, species %in% sp_list$species)
dim(sdls) ## 968 columns
```

```
## [1] 968 19
```

Rename columns, add total seedling density, scale and centre data.

```
## shorten names
sdls <- rename(sdls,
              "trt_F" = "treatment.fungicide",
              "trt_I" = "treatment.insecticide",
              "Pr_m" = "proportion.mortality",
              "gr" = "group",
              "loc" = "location")
## group causes problems with some helper funcs
## add total density
tot_dens <- sdls |> group_by(site, loc, gr, plot) |>
  summarise(tot_dens = sum(census.start))
```

```
## 'summarise()' has grouped output by 'site', 'loc', 'gr'. You can override using
## the '.groups' argument.
```

```
## add species mean density
## divide by total number of plots = 21 sites x 4 locs * 3 groups * 5 plots
sp_mean_dens <- sdls |> group_by(species) |>
  summarise(sp_mean_dens = sum(census.start)/(21*4*3*5),
            sp_mean_surv = sum(census.final)/sum(census.start))

sdls <- left_join(sdls, tot_dens, by = c("site", "loc", "gr", "plot")) |>
  left_join(select(sp_mean_dens, - sp_mean_surv))
```

```
## Joining with 'by = join_by(species)'

#scale density by mean, fix couple of NAs and calculate log density
sdls <- mutate(sdls,
  slope.degrees = replace_na(slope.degrees, 5),
  Pr_s = 1 - Pr_m,
  con_dens = census.start,
  con_dens_s = con_dens/sp_mean_dens,
  slope.degrees_s = as.vector(scale(slope.degrees)),
  trt_F = factor(trt_F, labels = c("0", "F")),
  trt_I = factor(trt_I, labels = c("0", "I"))
)
dim(sdls) ## 968 species x plot combinations

## [1] 968 25

sdls |> summarise(n_sdls = sum(census.start),
  n_survs = sum(census.final),
  n_species = n_distinct(species)) |> knitr::kable()
```

n_sdls	n_survs	n_species
6208	2680	26

Models

Raw conspecific density model

Fitting a model of mortality as a function of initial conspecific density, initial total density, biocide treatment and fragment area (and their interactions).

Also including slope of plot and random intercepts for plot, nested in location, nested in site. Random slopes and intercepts are included for each species to examine how density dependence varies among species.

Note that including the insecticide x fungicide interaction causes convergence problems and doesn't seem to be important either. Removing for practicality.

Model diagnostics

Take-homes

- Model diagnostics are generally good.
- Diagnostics are slightly better for non-logged version. Logged version has some evidence of quantiles deviating from expectations (removed logged version now).
- No evidence of trends between covariates and residuals.

The log density model has a slightly lower AIC. A little poking around (not shown) suggests that excluding the largest values reverses this order, so that the non-logged version is better.

Looks like the two models lead to roughly similar interpretation.

Inference

The fixed effects

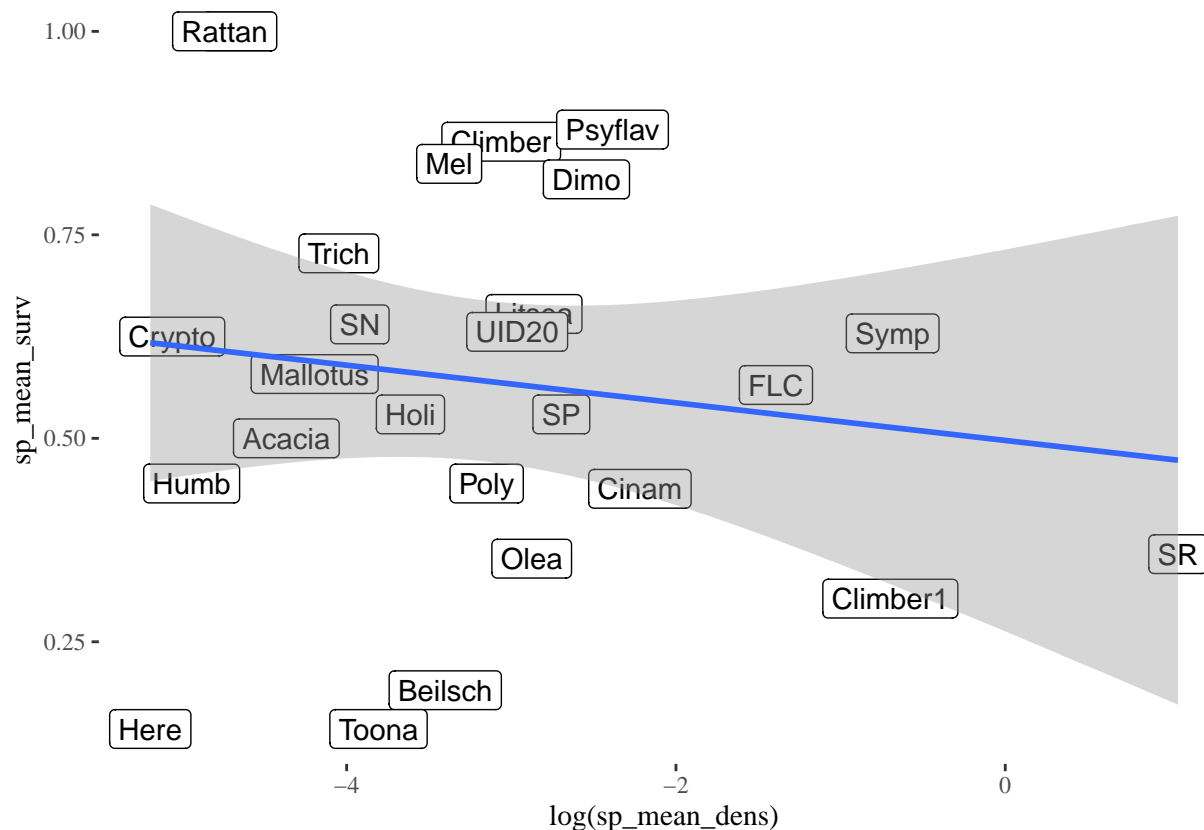
Main conclusions * Suppressing insects and fungi) increases survival. * Conspecific density reduces seedling survival. * The protection by fungicide is less effective at high conspecific density * There is a weak 3-way interaction between conspecific density, fragment size, and fungicide addition, so that the reduction in protection at high densities is more evident in larger fragments.

Scaled conspecific density models

The most abundant species initially will often have lower survival (fecundity/ survival trade-off). This could generate what looks like a density-dependent relationship when looking across species, even without a relationship within species (i.e., Simpson's paradox).

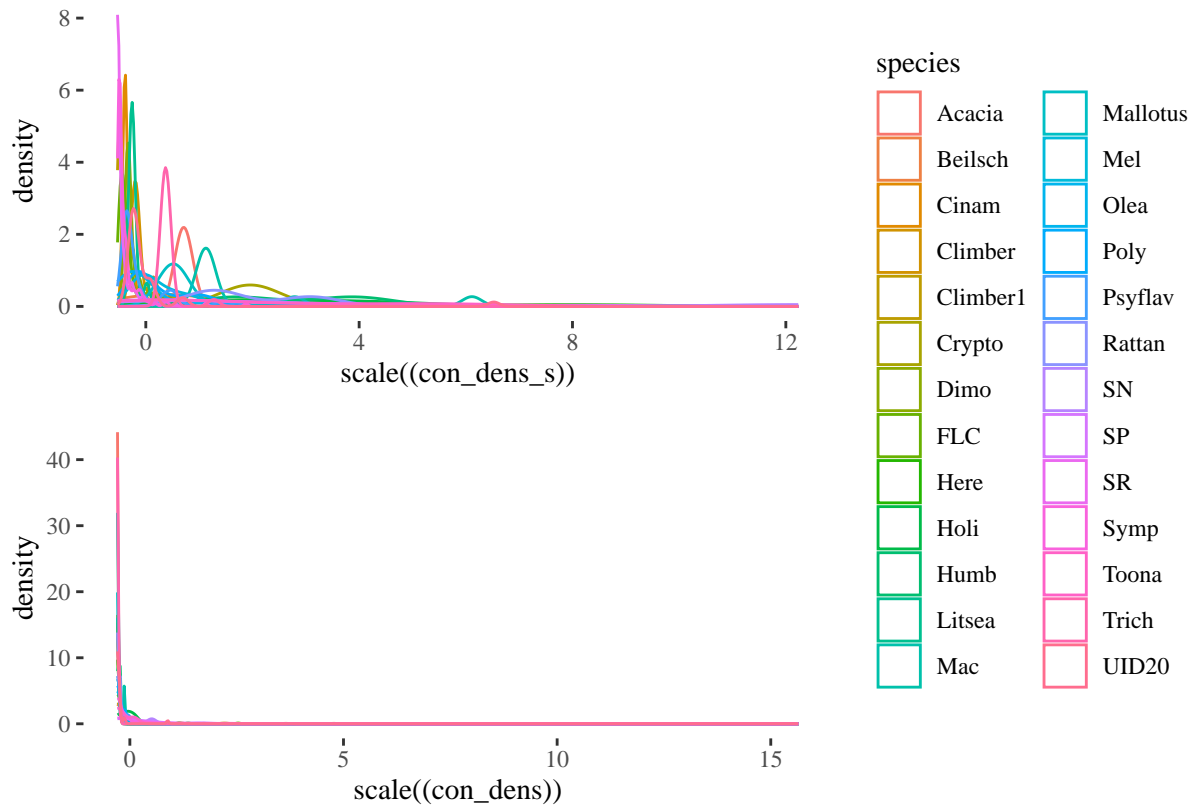
```
ggplot(sp_mean_dens, aes(x = log(sp_mean_dens), y = sp_mean_surv)) +  
  geom_label(aes(label= species)) + geom_smooth(method = "lm") + theme_tufte()
```

```
## 'geom_smooth()' using formula = 'y ~ x'
```



```
## not the clearest pattern, but worth accounting for.
```

```
(ggplot(sdl, aes(x = scale((con_dens_s)), colour = species)) + geom_density())/  
(ggplot(sdl, aes(x = scale((con_dens_s)), colour = species)) + geom_density()) + plot_layout(guides = "c
```



To account for this, we can scale conspecific density by dividing by mean density and refitting the models.

```
## models using species-scaled density
##
## Random intercept model
m_cdd_s_ri <- glmmTMB(Pr_s ~ slope.degrees_s +
  (scale(tot_dens) + scale(con_dens_s)) *
  (trt_I + trt_F) *
  scale(log(fragment.size)) +
  (1|species) +
  (1|site/loc/gr/plot),
  weights = census.start, data = sdls,
  family=binomial)
## Random intercept and slope model for species specific effects
m_cdd_s_ris <- glmmTMB(Pr_s ~ slope.degrees_s +
  (scale(tot_dens) + scale(con_dens_s)) *
  (trt_I + trt_F) * scale(log(fragment.size)) *
  (scale(con_dens_s) + scale(tot_dens)||species) +
  ## setting cor to 0 to converge
  (1|site/loc/gr/plot),
  weights = census.start, data = sdls,
  family=binomial)
anova(m_cdd_s_ri, m_cdd_s_ris) ## random slope *much* better
```

```
## Data: sdls
## Models:
```

```
## m_cdd_s_ri: Pr_s ~ slope.degrees_s + (scale(tot_dens) + scale(con_dens_s)) * , zi=~0, disp=~1
## m_cdd_s_ri:      (trt_I + trt_F) * scale(log(fragment.size)) + (1 | species) + , zi=~0, disp=~1
## m_cdd_s_ri:      (1 | site/loc/gr/plot), zi=~0, disp=~1
## m_cdd_s_ris: Pr_s ~ slope.degrees_s + (scale(tot_dens) + scale(con_dens_s)) * , zi=~0, disp=~1
## m_cdd_s_ris:      (trt_I + trt_F) * scale(log(fragment.size)) * (scale(con_dens_s) + , zi=~0, disp=~1
## m_cdd_s_ris:      scale(tot_dens) || species) + (1 | site/loc/gr/plot), zi=~0, disp=~1
##              Df      AIC      BIC logLik deviance Chisq Chi Df Pr(>Chisq)
## m_cdd_s_ri   24 2197.1 2314.1 -1074.6   2149.1
## m_cdd_s_ris  26 2187.2 2313.9 -1067.6   2135.2 13.954      2 0.0009331 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

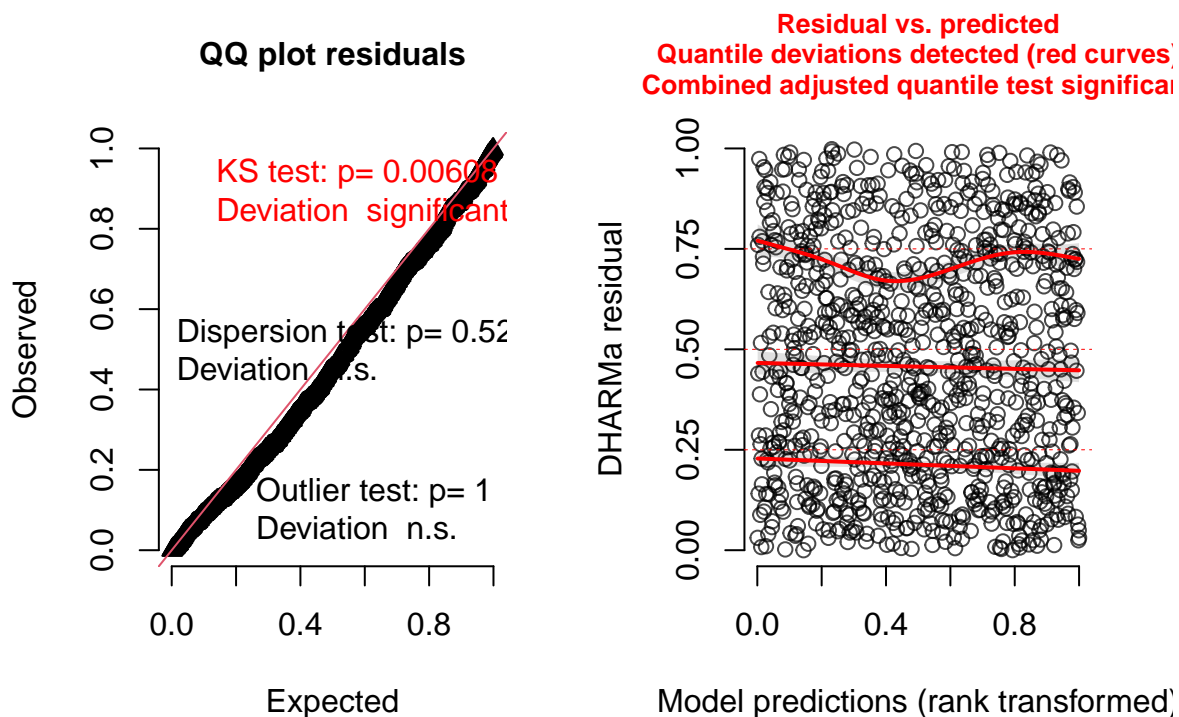
Models fit without issues as long as we fix the covariance between random slopes and intercepts to 0.

The improvment with the random slopes model suggests we need to look at individual species.

Diagnostics

```
res_s <- simulateResiduals(m_cdd_s_ri)
plot(res_s) ## ok - some deviation from ideal residual distribution, but
```

DHARMA residual



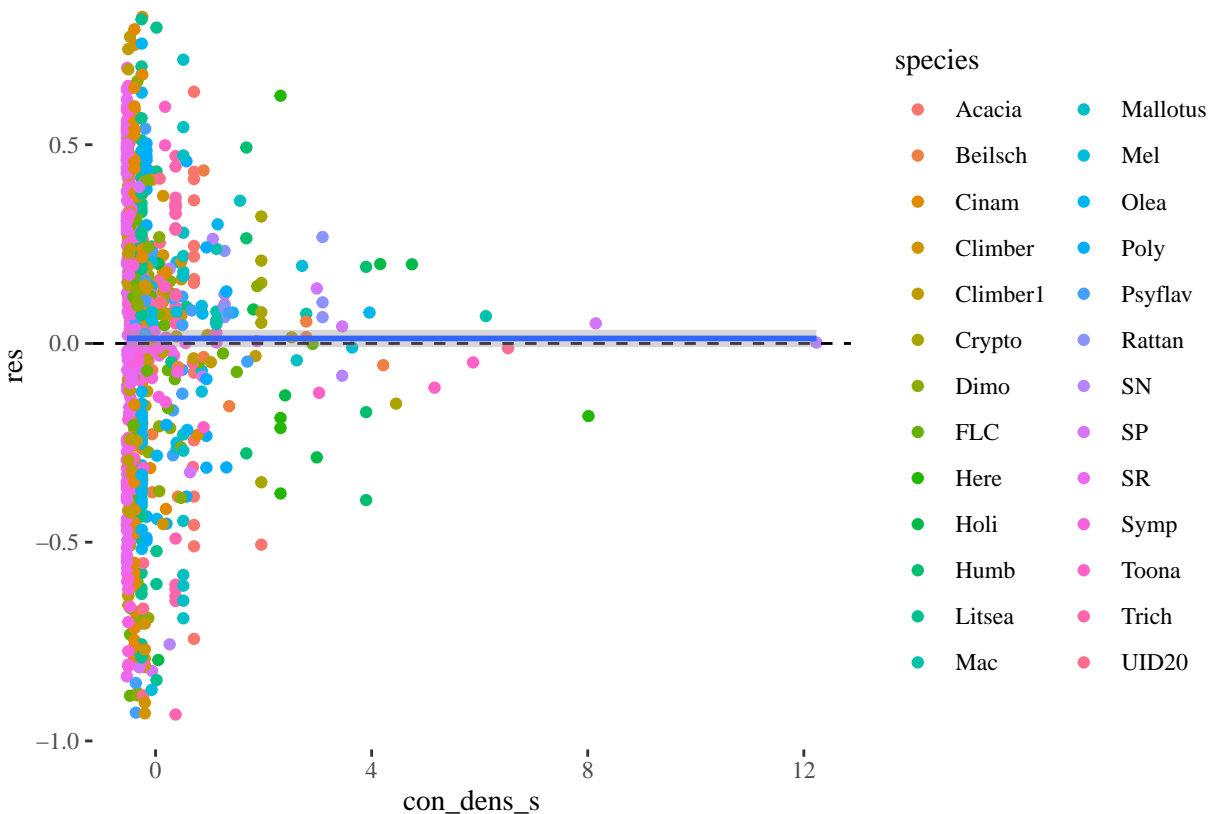
```
## acceptable.
## look at relationship with covariates
```

```
diag_dat <- data.frame(m_cdd_s_ri$frame, res = res_s$fittedResiduals)

diag_dat <- rename_with(diag_dat, ~ str_replace(.x, "scale\\.", "")) |>
  rename_with(~str_replace(.x, "\\.$", ""))

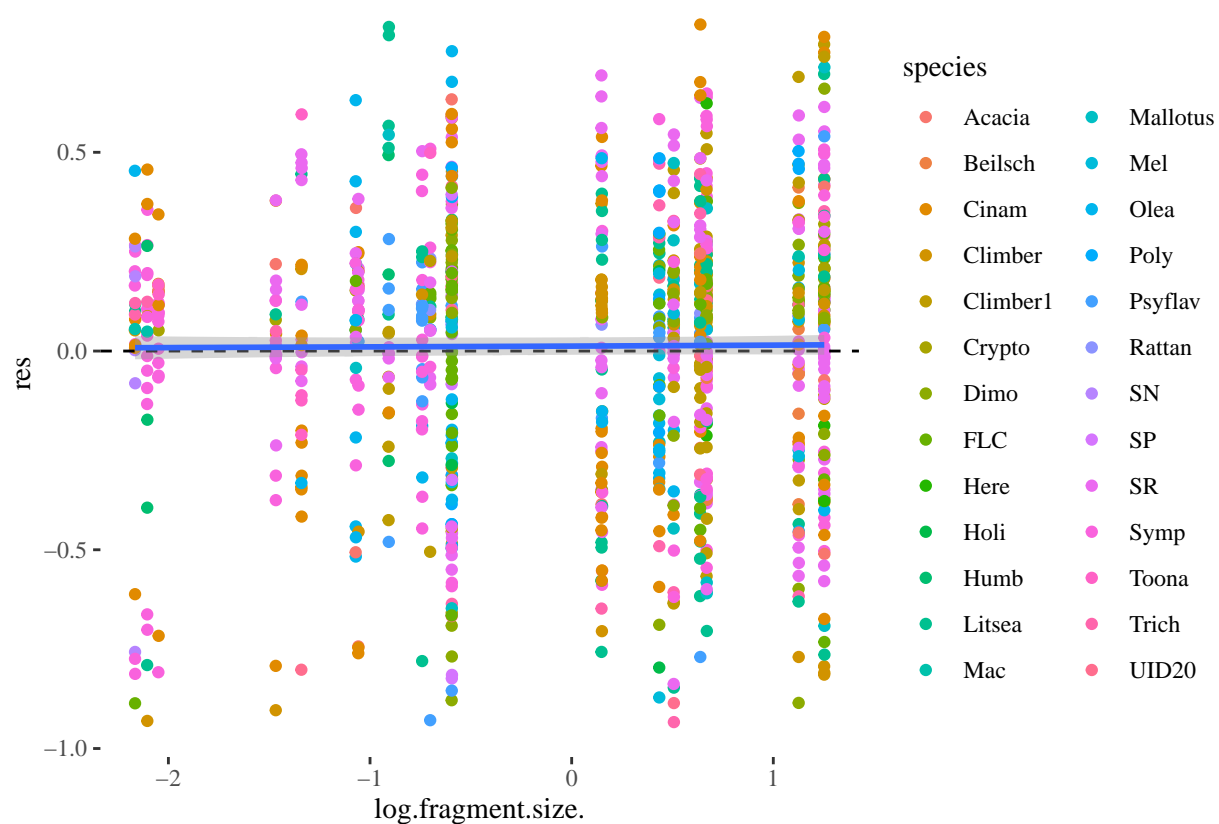
ggplot(diag_dat, aes(x = con_dens_s, y = res)) +
  geom_point(aes(colour = species)) +
  geom_hline(yintercept=0, linetype = "dashed") +
  geom_smooth(method="gam") ## no trend.
```

'geom_smooth()' using formula = 'y ~ s(x, bs = "cs")'



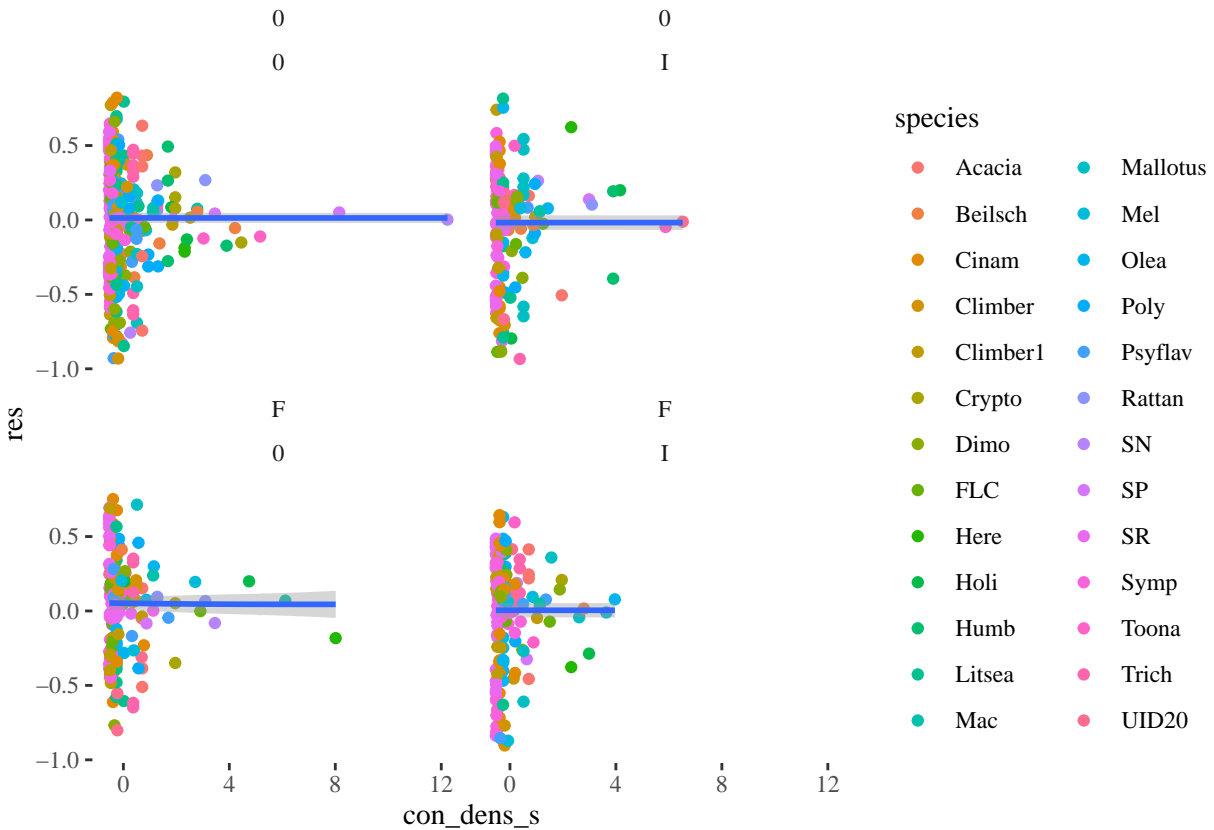
```
ggplot(diag_dat, aes(x = log.fragment.size., y = res)) +
  geom_point(aes(colour = species), position = "jitter") +
  geom_hline(yintercept=0, linetype = "dashed") +
  geom_smooth(method="gam") ## no trend.
```

'geom_smooth()' using formula = 'y ~ s(x, bs = "cs")'



```
ggplot(diag_dat, aes(x = con_dens_s, y = res)) +
  facet_wrap(~trt_F + trt_I ) +
  geom_point(aes(colour = species)) +
  geom_smooth(method="gam") ## no trend with treatment
```

```
## 'geom_smooth()' using formula = 'y ~ s(x, bs = "cs")'
```



The diagnostics aren't as perfect in the scaled case, but the scaled version does seem to be sensible given the massive differences in density.

Model inference

```
summary(m_cdd_s_ri)
```

```
## Family: binomial ( logit )
## Formula:
## Pr_s ~ slope.degrees_s + (scale(tot_dens) + scale(con_dens_s)) *
## (trt_I + trt_F) * scale(log(fragment.size)) + (1 | species) +
## (1 | site/loc/gr/plot)
## Data: sdls
## Weights: census.start
##
##      AIC      BIC   logLik deviance df.resid
##  2197.1   2314.1  -1074.6   2149.1     944
##
## Random effects:
##
## Conditional model:
## Groups          Name          Variance Std.Dev.
## species          (Intercept)  1.355e+00 1.1642158
## plot:gr:loc:site (Intercept)  2.340e-01 0.4836923
```

```

## gr:loc:site      (Intercept) 1.692e-01 0.4113449
## loc:site        (Intercept) 4.213e-01 0.6490432
## site            (Intercept) 3.512e-08 0.0001874
## Number of obs: 968, groups:
## species, 26; plot:gr:loc:site, 474; gr:loc:site, 110; loc:site, 37; site, 21
##
## Conditional model:
##
##                                Estimate Std. Error
## (Intercept)                   0.6755043  0.2920884
## slope.degrees_s               -0.1042895  0.0681512
## scale(tot_dens)               -0.3887989  0.0858523
## scale(con_dens_s)             -0.3193334  0.0792852
## trt_II                        0.4885509  0.1110445
## trt_FF                        0.0809017  0.1088436
## scale(log(fragment.size))     -0.4024701  0.1377352
## scale(tot_dens):trt_II        -0.1481742  0.1335183
## scale(tot_dens):trt_FF        0.0651851  0.1202666
## scale(con_dens_s):trt_II      -0.1235055  0.0920978
## scale(con_dens_s):trt_FF      0.2111937  0.0984225
## scale(tot_dens):scale(log(fragment.size)) 0.0611248  0.0755813
## scale(con_dens_s):scale(log(fragment.size)) -0.1211638  0.0509800
## trt_II:scale(log(fragment.size)) 0.2889510  0.1086788
## trt_FF:scale(log(fragment.size)) -0.0005297  0.1082614
## scale(tot_dens):trt_II:scale(log(fragment.size)) 0.0356476  0.1298870
## scale(tot_dens):trt_FF:scale(log(fragment.size)) 0.1362907  0.1049890
## scale(con_dens_s):trt_II:scale(log(fragment.size)) 0.0529964  0.0968246
## scale(con_dens_s):trt_FF:scale(log(fragment.size)) 0.1166648  0.0975779
##
##                                z value Pr(>|z|)
## (Intercept)                   2.313  0.02074 *
## slope.degrees_s               -1.530  0.12595
## scale(tot_dens)               -4.529 5.93e-06 ***
## scale(con_dens_s)             -4.028 5.63e-05 ***
## trt_II                        4.400 1.08e-05 ***
## trt_FF                        0.743  0.45731
## scale(log(fragment.size))     -2.922  0.00348 **
## scale(tot_dens):trt_II        -1.110  0.26710
## scale(tot_dens):trt_FF        0.542  0.58782
## scale(con_dens_s):trt_II      -1.341  0.17991
## scale(con_dens_s):trt_FF      2.146  0.03189 *
## scale(tot_dens):scale(log(fragment.size)) 0.809  0.41867
## scale(con_dens_s):scale(log(fragment.size)) -2.377  0.01747 *
## trt_II:scale(log(fragment.size)) 2.659  0.00784 **
## trt_FF:scale(log(fragment.size)) -0.005  0.99610
## scale(tot_dens):trt_II:scale(log(fragment.size)) 0.274  0.78374
## scale(tot_dens):trt_FF:scale(log(fragment.size)) 1.298  0.19424
## scale(con_dens_s):trt_II:scale(log(fragment.size)) 0.547  0.58414
## scale(con_dens_s):trt_FF:scale(log(fragment.size)) 1.196  0.23185
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

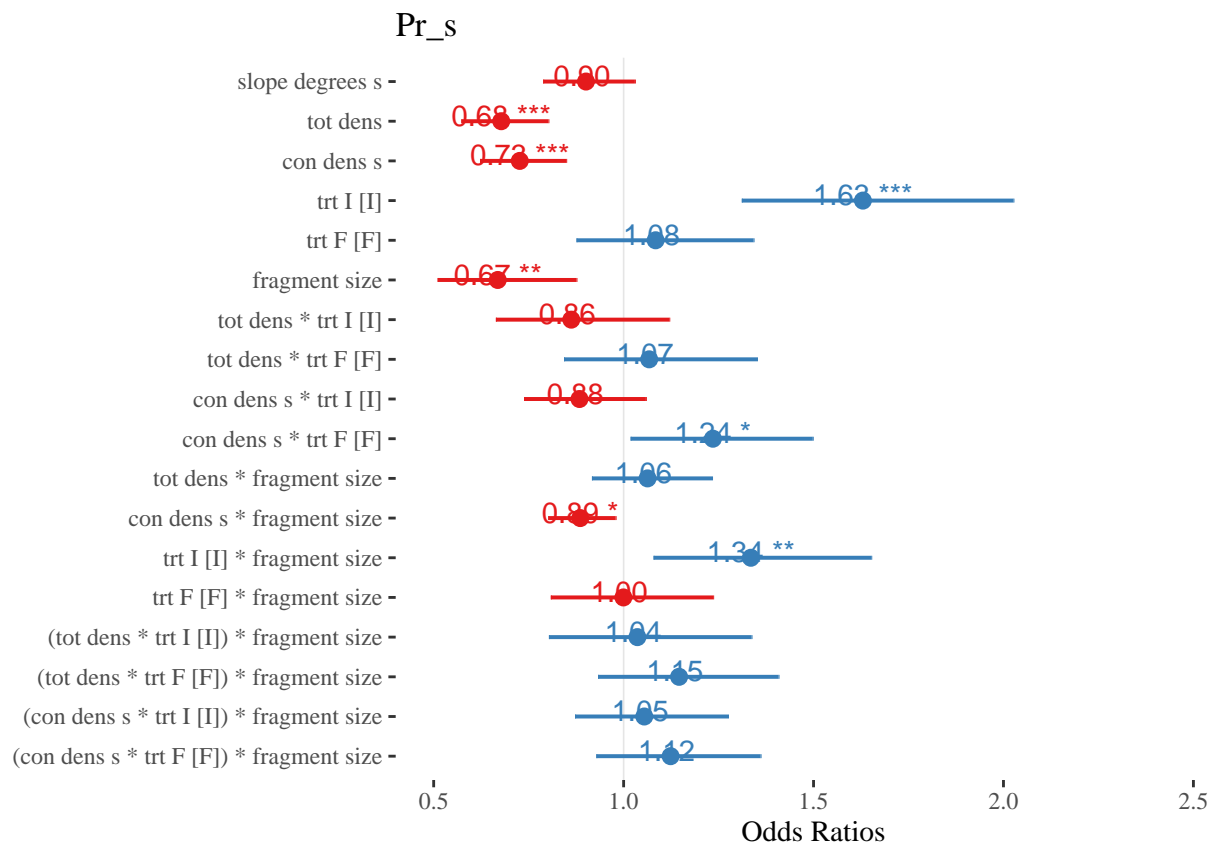
```

```
plot_model(m_cdd_s_ri, show.values=TRUE) + ylim(c(0.5, 2.5))
```

```

## Scale for y is already present.
## Adding another scale for y, which will replace the existing scale.

```



```
car::Anova(m_cdd_s_ri)
```

```
## Analysis of Deviance Table (Type II Wald chisquare tests)
##
## Response: Pr_s
##
##           Chisq Df Pr(>Chisq)
## slope.degrees_s      2.3417  1  0.125951
## scale(tot_dens)      23.3742  1  1.334e-06 ***
## scale(con_dens_s)     17.6388  1  2.671e-05 ***
## trt_I                 17.0703  1  3.602e-05 ***
## trt_F                  3.8922  1  0.048510 *
## scale(log(fragment.size))  5.2401  1  0.022072 *
## scale(tot_dens):trt_I    1.7183  1  0.189916
## scale(tot_dens):trt_F    6.5759  1  0.010337 *
## scale(con_dens_s):trt_I   2.4813  1  0.115208
## scale(con_dens_s):trt_F   3.5771  1  0.058580 .
## scale(tot_dens):scale(log(fragment.size))  3.8857  1  0.048700 *
## scale(con_dens_s):scale(log(fragment.size))  4.4018  1  0.035901 *
## trt_I:scale(log(fragment.size))  7.4137  1  0.006473 **
## trt_F:scale(log(fragment.size))  0.0772  1  0.781091
## scale(tot_dens):trt_I:scale(log(fragment.size))  0.0753  1  0.783738
## scale(tot_dens):trt_F:scale(log(fragment.size))  1.6852  1  0.194238
## scale(con_dens_s):trt_I:scale(log(fragment.size))  0.2996  1  0.584142
## scale(con_dens_s):trt_F:scale(log(fragment.size))  1.4295  1  0.231850
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
car::Anova(m_cdd_s_ris)
```

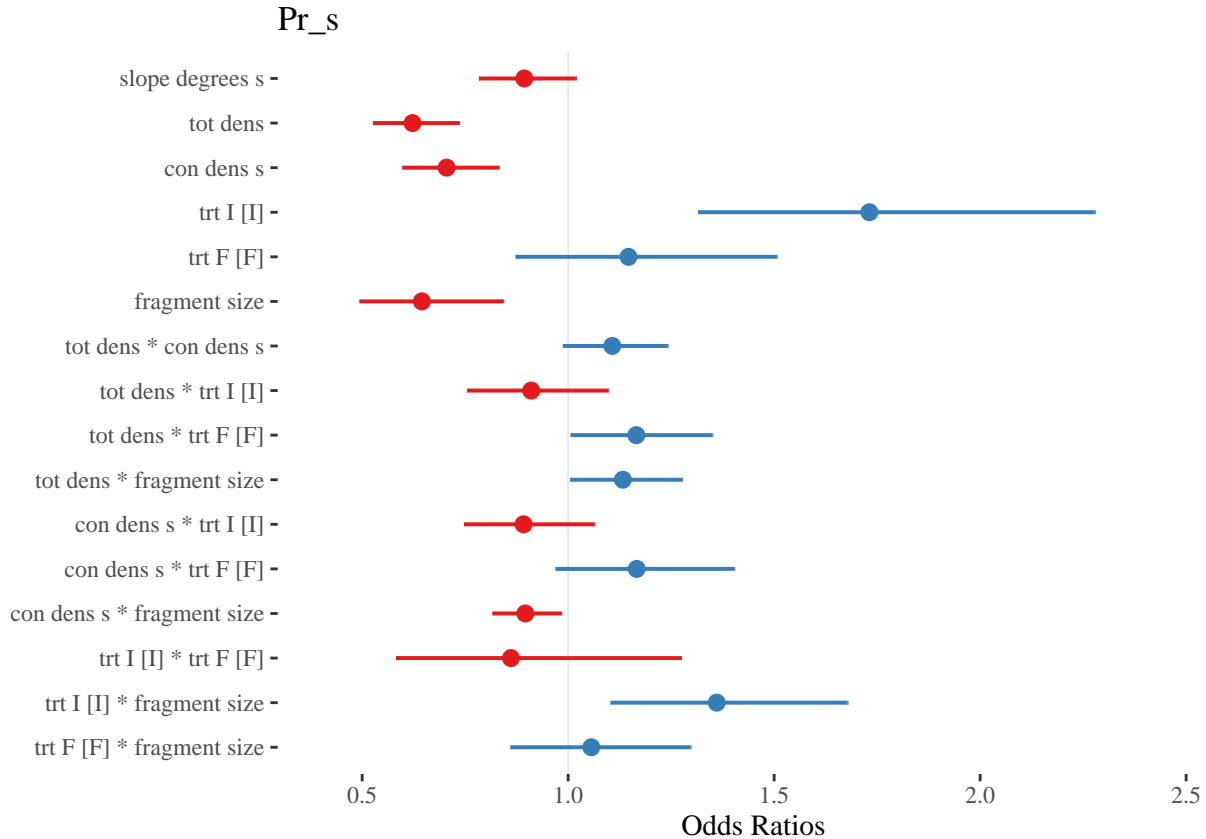
```
## Analysis of Deviance Table (Type II Wald chisquare tests)
##
## Response: Pr_s
##
##           Chisq Df Pr(>Chisq)
## slope.degrees_s      2.8454  1  0.0916364 .
## scale(tot_dens)       0.5362  1  0.4640207
## scale(con_dens_s)    11.0659  1  0.0008793 ***
## trt_I                17.7010  1  2.585e-05 ***
## trt_F                 3.5593  1  0.0592141 .
## scale(log(fragment.size)) 5.5929  1  0.0180337 *
## scale(tot_dens):trt_I    1.8384  1  0.1751423
## scale(tot_dens):trt_F    7.6943  1  0.0055396 **
## scale(con_dens_s):trt_I  0.6202  1  0.4309704
## scale(con_dens_s):trt_F  2.2386  1  0.1346005
## scale(tot_dens):scale(log(fragment.size)) 4.2905  1  0.0383262 *
## scale(con_dens_s):scale(log(fragment.size)) 0.2213  1  0.6380294
## trt_I:scale(log(fragment.size)) 8.1304  1  0.0043529 **
## trt_F:scale(log(fragment.size)) 0.0448  1  0.8324460
## scale(tot_dens):trt_I:scale(log(fragment.size)) 0.0813  1  0.7755017
## scale(tot_dens):trt_F:scale(log(fragment.size)) 0.9376  1  0.3328944
## scale(con_dens_s):trt_I:scale(log(fragment.size)) 0.3644  1  0.5460845
## scale(con_dens_s):trt_F:scale(log(fragment.size)) 1.9890  1  0.1584465
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
## Given lack of evidence for 3-way interactions, perhaps try only 2 way?
```

```
m_cdd_s_ri_2way <- glmmTMB(Pr_s ~ slope.degrees_s +
  ((scale(tot_dens) + scale(con_dens_s)) +
   (trt_I + trt_F) +
   scale(log(fragment.size)))^2 +
  (1|species) +
  (1|site/loc/gr/plot),
  weights = census.start, data = sdls,
  family=binomial)
plot_model(m_cdd_s_ri_2way) + ylim(c(0.4, 2.5))
```

```
## Scale for y is already present.
```

```
## Adding another scale for y, which will replace the existing scale.
```



Take-homes:

- Survival is negatively conspecific density dependent
- Fungicide reduces the density dependence, but the evidence is complex. In the full model with a 3 way CDD:frag_area:F interaction, the 2-way cdd:F interaction is significant ($P = 0.03$). Note that the magnitude of the effect (0.21) is 2/3rds of the overall conspecific density effect (-0.32). Worth remarking on in the results. However, the Type 2 anova suggests that the effect is conditional on the effect of fragment area - excluding the interaction weakens evidence for the 2-way interaction ($P = 0.059$)
- CDD strengthens significantly with fragment area ($P = 0.017$).
- The interaction between fungicide, fragment area and CDD is not significant, but its estimated magnitude (0.117) is almost equal (and opposite direction) to the interaction between fragment area and density (-0.121).
- Worth noting that survival also increases with total stem density, with a similar magnitude as with conspecific density.
- Survival also declines with fragment area.
- Insecticide increases survival independent of density, and its effect increases with fragment area.

Models split by categorical fragment size

The effect of fragment

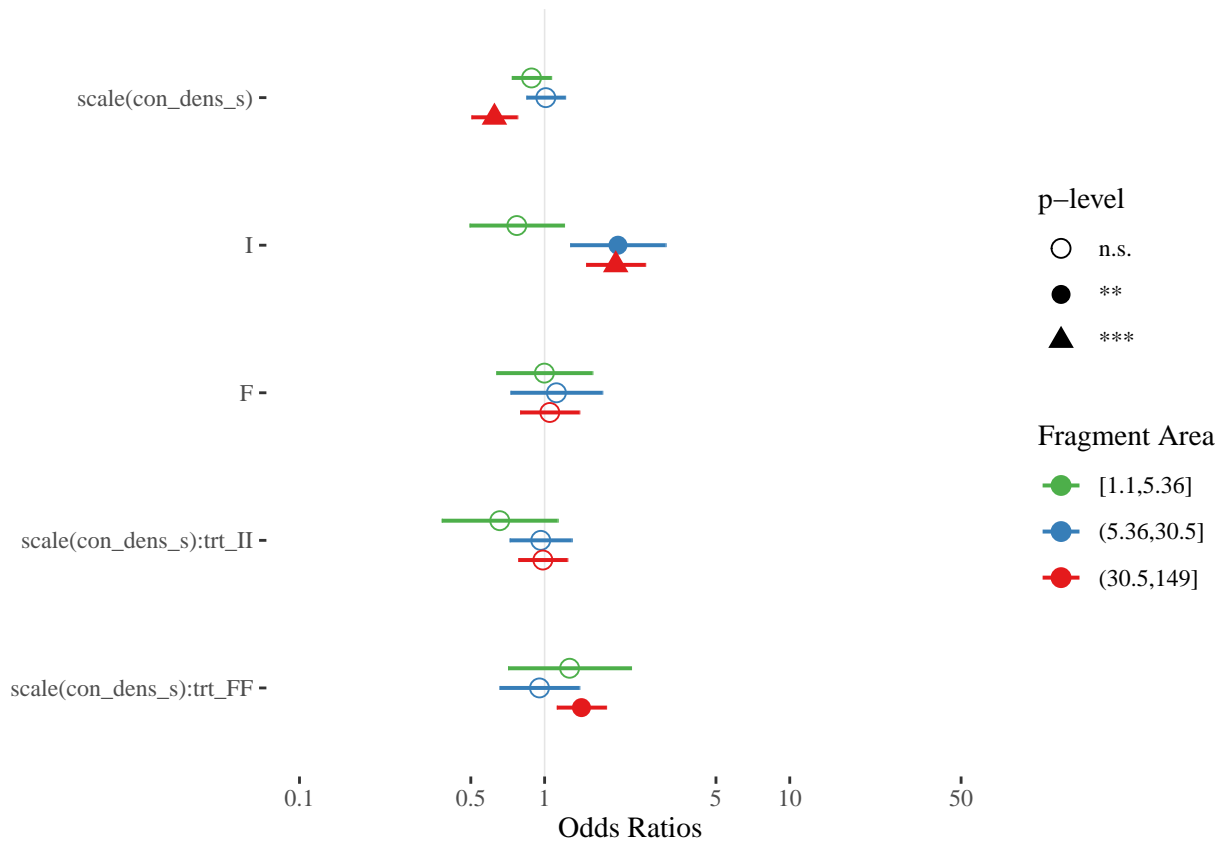
```

# Using 3 categories
sdls <- mutate(sdls,
               frag_sizeclass3 = cut(fragment.size,
                                     quantile(site_dat$fragment.size,
                                               c(0, 0.33, 0.66, 1)),
                                     include.lowest=TRUE))

m_cdd_s_ri_frag <- lapply(split(sdls, f= sdls$frag_sizeclass3),
                           function(d){
                             glmmTMB(Pr_s ~ slope.degrees_s +
                                       (scale(tot_dens) + scale(con_dens_s)) *
                                       (trt_I + trt_F) +
                                       (1|species) +
                                       ## setting cor to 0 to converge
                                       (1|site/loc/gr/plot),
                                       weights = census.start, data = d,
                                       family=binomial))
term_nms <- names(fixef(m_cdd_s_ri_frag[[1]])$cond)

plot_models(m_cdd_s_ri_frag,
            rm.terms=c("slope.degrees_s",
                      term_nms[str_detect(term_nms, "tot")]), ## declutter
            m.labels=names(m_cdd_s_ri_frag), p.shape=TRUE) +
  labs(colour = "Fragment Area")

```



```
map(m_cdd_s_ri_frag, summary)
```

```
## $'[1.1,5.36]'
```

```
## Family: binomial ( logit )
```

```
## Formula:
```

```
## Pr_s ~ slope.degrees_s + (scale(tot_dens) + scale(con_dens_s)) *
```

```
## (trt_I + trt_F) + (1 | species) + (1 | site/loc/gr/plot)
```

```
## Data: d
```

```
## Weights: census.start
```

```
##
```

	AIC	BIC	logLik	deviance	df.resid
	376.7	421.8	-173.4	346.7	134

```
##
```

```
## Random effects:
```

```
##
```

```
## Conditional model:
```

Groups	Name	Variance	Std.Dev.
species	(Intercept)	2.674e-01	5.171e-01
plot:gr:loc:site	(Intercept)	3.592e-09	5.994e-05
gr:loc:site	(Intercept)	9.554e-02	3.091e-01
loc:site	(Intercept)	3.052e-01	5.524e-01
site	(Intercept)	3.052e-01	5.524e-01

```
## Number of obs: 149, groups:
```

```
## species, 18; plot:gr:loc:site, 87; gr:loc:site, 20; loc:site, 7; site, 7
```

```
##
```

```
## Conditional model:
```

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	1.026283	0.417397	2.459	0.013941 *
slope.degrees_s	-0.147994	0.139908	-1.058	0.290148
scale(tot_dens)	-0.294738	0.086247	-3.417	0.000632 ***
scale(con_dens_s)	-0.123143	0.094292	-1.306	0.191562
trt_II	-0.261125	0.226313	-1.154	0.248573
trt_FF	-0.002225	0.230126	-0.010	0.992284
scale(tot_dens):trt_II	0.452427	0.181515	2.492	0.012685 *
scale(tot_dens):trt_FF	-0.121468	0.138459	-0.877	0.380331
scale(con_dens_s):trt_II	-0.421473	0.277932	-1.516	0.129404
scale(con_dens_s):trt_FF	0.234629	0.294453	0.797	0.425549

```
## ---
```

```
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
##
```

```
## $'[5.36,30.5]'
```

```
## Family: binomial ( logit )
```

```
## Formula:
```

```
## Pr_s ~ slope.degrees_s + (scale(tot_dens) + scale(con_dens_s)) *
```

```
## (trt_I + trt_F) + (1 | species) + (1 | site/loc/gr/plot)
```

```
## Data: d
```

```
## Weights: census.start
```

```
##
```

	AIC	BIC	logLik	deviance	df.resid
	629.2	683.9	-299.6	599.2	269

```
##
```

```
## Random effects:
```

```
##
```



```

## Conditional model:
## Groups          Name          Variance Std.Dev.
## species          (Intercept) 1.497e+00 1.223e+00
## plot:gr:loc:site (Intercept) 2.744e-01 5.238e-01
## gr:loc:site      (Intercept) 7.970e-09 8.928e-05
## loc:site         (Intercept) 1.955e-01 4.421e-01
## site            (Intercept) 3.622e-01 6.018e-01
## Number of obs: 284, groups:
## species, 21; plot:gr:loc:site, 124; gr:loc:site, 27; loc:site, 9; site, 7
##
## Conditional model:
##              Estimate Std. Error z value Pr(>|z|)
## (Intercept)      0.50437    0.45066   1.119  0.26307
## slope.degrees_s  -0.21107    0.11866  -1.779  0.07527 .
## scale(tot_dens)  -0.41809    0.13905  -3.007  0.00264 **
## scale(con_dens_s) 0.01107    0.09278   0.119  0.90501
## trt_II            0.68891    0.22859   3.014  0.00258 **
## trt_FF            0.11082    0.21994   0.504  0.61435
## scale(tot_dens):trt_II -0.63928  0.21656  -2.952  0.00316 **
## scale(tot_dens):trt_FF -0.02168  0.21259  -0.102  0.91877
## scale(con_dens_s):trt_II -0.03646  0.14890  -0.245  0.80659
## scale(con_dens_s):trt_FF -0.04826  0.19110  -0.252  0.80063
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## $'(30.5,149]
## Family: binomial ( logit )
## Formula:
## Pr_s ~ slope.degrees_s + (scale(tot_dens) + scale(con_dens_s)) *
##      (trt_I + trt_F) + (1 | species) + (1 | site/loc/gr/plot)
## Data: d
## Weights: census.start
##
##      AIC      BIC    logLik deviance df.resid
##  1195.2   1259.4   -582.6   1165.2     520
##
## Random effects:
##
## Conditional model:
## Groups          Name          Variance Std.Dev.
## species          (Intercept) 1.408e+00 1.1867797
## plot:gr:loc:site (Intercept) 1.392e-01 0.3731126
## gr:loc:site      (Intercept) 3.098e-01 0.5566388
## loc:site         (Intercept) 3.221e-01 0.5675476
## site            (Intercept) 1.671e-08 0.0001293
## Number of obs: 535, groups:
## species, 22; plot:gr:loc:site, 263; gr:loc:site, 63; loc:site, 21; site, 7
##
## Conditional model:
##              Estimate Std. Error z value Pr(>|z|)
## (Intercept)      0.53178    0.33873   1.570  0.11644
## slope.degrees_s  -0.02894    0.09389  -0.308  0.75788
## scale(tot_dens)  -0.32736    0.08167  -4.008 6.12e-05 ***
## scale(con_dens_s) -0.47153    0.10987  -4.292 1.77e-05 ***

```

```
## trt_II                0.66796      0.14118    4.731 2.23e-06 ***
## trt_FF                0.04856      0.14158    0.343 0.73158
## scale(tot_dens):trt_II -0.14344    0.10774   -1.331 0.18304
## scale(tot_dens):trt_FF  0.25646    0.08100    3.166 0.00155 **
## scale(con_dens_s):trt_II -0.01647    0.11729   -0.140 0.88830
## scale(con_dens_s):trt_FF  0.34613    0.11810    2.931 0.00338 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
## What about a single model with categorical fragment sizes
```

```
m_cdd_s_cat_ri <- glmmTMB(Pr_s ~ slope.degrees_s +
  (scale(tot_dens) + scale(con_dens_s)) *
  (trt_I + trt_F) *
  frag_sizeclass3 +
  (1|species) +
  ## setting cor to 0 to converge
  (1|site/loc/gr/plot),
  weights = census.start, data = sdls,
  family=binomial)
summary(m_cdd_s_cat_ri)
```

```
## Family: binomial ( logit )
## Formula:
## Pr_s ~ slope.degrees_s + (scale(tot_dens) + scale(con_dens_s)) *
## (trt_I + trt_F) * frag_sizeclass3 + (1 | species) + (1 |
## site/loc/gr/plot)
## Data: sdls
## Weights: census.start
##
##      AIC      BIC   logLik deviance df.resid
##  2190.2   2351.1  -1062.1   2124.2     935
##
## Random effects:
##
## Conditional model:
## Groups          Name          Variance Std.Dev.
## species          (Intercept)  1.288e+00 1.1347817
## plot:gr:loc:site (Intercept)  2.026e-01 0.4501594
## gr:loc:site      (Intercept)  1.529e-01 0.3909984
## loc:site         (Intercept)  3.730e-01 0.6107409
## site             (Intercept)  1.339e-08 0.0001157
## Number of obs: 968, groups:
## species, 26; plot:gr:loc:site, 474; gr:loc:site, 110; loc:site, 37; site, 21
##
## Conditional model:
##
##              Estimate Std. Error z value
## (Intercept)      1.85105    0.41523   4.458
## slope.degrees_s  -0.10584    0.06748  -1.568
## scale(tot_dens)  -0.48135    0.24136  -1.994
## scale(con_dens_s) -0.15125    0.08225  -1.839
## trt_II           -0.17653    0.25516  -0.692
## trt_FF           -0.12716    0.24995  -0.509
## frag_sizeclass3(5.36,30.5] -1.43466    0.42290  -3.392
```

```

## frag_sizeclass3(30.5,149] -1.43974 0.37518 -3.838
## scale(tot_dens):trt_II 0.37760 0.46687 0.809
## scale(tot_dens):trt_FF -0.12544 0.38609 -0.325
## scale(con_dens_s):trt_II -0.33583 0.21172 -1.586
## scale(con_dens_s):trt_FF 0.21189 0.22324 0.949
## scale(tot_dens):frag_sizeclass3(5.36,30.5] -0.17833 0.33211 -0.537
## scale(tot_dens):frag_sizeclass3(30.5,149] 0.18246 0.25002 0.730
## scale(con_dens_s):frag_sizeclass3(5.36,30.5] 0.13309 0.12092 1.101
## scale(con_dens_s):frag_sizeclass3(30.5,149] -0.38047 0.15135 -2.514
## trt_II:frag_sizeclass3(5.36,30.5] 0.75832 0.32129 2.360
## trt_II:frag_sizeclass3(30.5,149] 0.86138 0.29402 2.930
## trt_FF:frag_sizeclass3(5.36,30.5] 0.26949 0.31934 0.844
## trt_FF:frag_sizeclass3(30.5,149] 0.20441 0.29099 0.702
## scale(tot_dens):trt_II:frag_sizeclass3(5.36,30.5] -1.51532 0.59619 -2.542
## scale(tot_dens):trt_II:frag_sizeclass3(30.5,149] -0.48934 0.47641 -1.027
## scale(tot_dens):trt_FF:frag_sizeclass3(5.36,30.5] 0.12640 0.52109 0.243
## scale(tot_dens):trt_FF:frag_sizeclass3(30.5,149] 0.33064 0.39285 0.842
## scale(con_dens_s):trt_II:frag_sizeclass3(5.36,30.5] 0.27666 0.25709 1.076
## scale(con_dens_s):trt_II:frag_sizeclass3(30.5,149] 0.28871 0.25532 1.131
## scale(con_dens_s):trt_FF:frag_sizeclass3(5.36,30.5] -0.28808 0.28960 -0.995
## scale(con_dens_s):trt_FF:frag_sizeclass3(30.5,149] 0.20920 0.26489 0.790
## Pr(>|z|)
## (Intercept) 8.28e-06 ***
## slope.degrees_s 0.116773
## scale(tot_dens) 0.046115 *
## scale(con_dens_s) 0.065920 .
## trt_II 0.489036
## trt_FF 0.610925
## frag_sizeclass3(5.36,30.5] 0.000693 ***
## frag_sizeclass3(30.5,149] 0.000124 ***
## scale(tot_dens):trt_II 0.418640
## scale(tot_dens):trt_FF 0.745257
## scale(con_dens_s):trt_II 0.112701
## scale(con_dens_s):trt_FF 0.342540
## scale(tot_dens):frag_sizeclass3(5.36,30.5] 0.591297
## scale(tot_dens):frag_sizeclass3(30.5,149] 0.465529
## scale(con_dens_s):frag_sizeclass3(5.36,30.5] 0.271081
## scale(con_dens_s):frag_sizeclass3(30.5,149] 0.011942 *
## trt_II:frag_sizeclass3(5.36,30.5] 0.018264 *
## trt_II:frag_sizeclass3(30.5,149] 0.003393 **
## trt_FF:frag_sizeclass3(5.36,30.5] 0.398718
## trt_FF:frag_sizeclass3(30.5,149] 0.482395
## scale(tot_dens):trt_II:frag_sizeclass3(5.36,30.5] 0.011031 *
## scale(tot_dens):trt_II:frag_sizeclass3(30.5,149] 0.304352
## scale(tot_dens):trt_FF:frag_sizeclass3(5.36,30.5] 0.808345
## scale(tot_dens):trt_FF:frag_sizeclass3(30.5,149] 0.399979
## scale(con_dens_s):trt_II:frag_sizeclass3(5.36,30.5] 0.281872
## scale(con_dens_s):trt_II:frag_sizeclass3(30.5,149] 0.258149
## scale(con_dens_s):trt_FF:frag_sizeclass3(5.36,30.5] 0.319861
## scale(con_dens_s):trt_FF:frag_sizeclass3(30.5,149] 0.429671
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

```
car::Anova(m_cdd_s_cat_ri)
```

```
## Analysis of Deviance Table (Type II Wald chisquare tests)
##
## Response: Pr_s
##
##           Chisq Df Pr(>Chisq)
## slope.degrees_s      2.4601  1  0.116773
## scale(tot_dens)      24.0078  1  9.595e-07 ***
## scale(con_dens_s)     15.9951  1  6.351e-05 ***
## trt_I                17.6831  1  2.609e-05 ***
## trt_F                3.7025  1  0.054330 .
## frag_sizeclass3       8.3175  2  0.015627 *
## scale(tot_dens):trt_I  2.9478  1  0.085996 .
## scale(tot_dens):trt_F  7.5625  1  0.005959 **
## scale(con_dens_s):trt_I 1.3192  1  0.250742
## scale(con_dens_s):trt_F 5.0734  1  0.024295 *
## scale(tot_dens):frag_sizeclass3 8.3555  2  0.015333 *
## scale(con_dens_s):frag_sizeclass3 8.0497  2  0.017866 *
## trt_I:frag_sizeclass3 10.0242  2  0.006657 **
## trt_F:frag_sizeclass3  0.8234  2  0.662525
## scale(tot_dens):trt_I:frag_sizeclass3 8.6274  2  0.013384 *
## scale(tot_dens):trt_F:frag_sizeclass3 1.0077  2  0.604192
## scale(con_dens_s):trt_I:frag_sizeclass3 1.4575  2  0.482505
## scale(con_dens_s):trt_F:frag_sizeclass3 4.5986  2  0.100331
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Take-homes

The effect of fragment area is complex. * Density dependence is only present in large fragments. * Fungicide removes the effect in these large fragments.

Although the interaction between fragment size and cdd and fungicide is not significantly different from 0, this is owed more to its uncertainty than its magnitude.

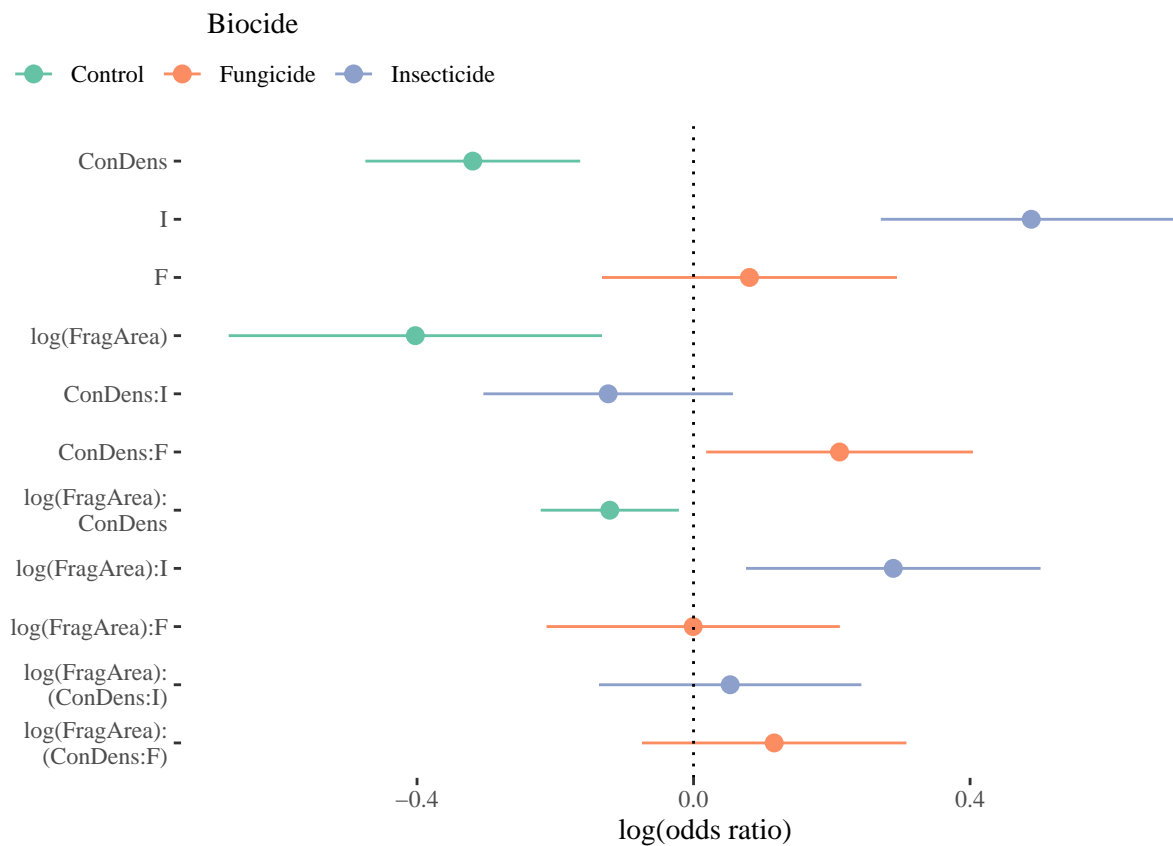
Graphics

```
labs <- c("ConDens", "I", "F", "log(FragArea)",
          "ConDens:I", "ConDens:F", "log(FragArea):\n ConDens",
          "log(FragArea):I" , "log(FragArea):F",
          "log(FragArea):\n (ConDens:I)",
          "log(FragArea):\n (ConDens:F)")
tp_cdd_s_ri <-
  tidy(m_cdd_s_ri, conf.int = TRUE) |>
  filter(effect == "fixed",
         !str_detect(term, "tot_dens"),
         !term %in% c("(Intercept)", "slope.degrees_s")) |>
  mutate(labels = factor(labs, levels = rev(labs)),
         Biocide = case_when(
           str_detect(term, "trt_II") ~ "Insecticide",
           str_detect(term, "trt_FF") ~ "Fungicide",
```

```

    .default = "Control")) |>
  ggplot(aes(y = labels, x = estimate, xmin = conf.low, xmax = conf.high,
    colour = Biocide)) +
  geom_pointrange() +
  geom_vline(xintercept=0, linetype = "dotted") +
  scale_colour_brewer(palette="Set2") +
  labs(y = NULL, x = "log(odds ratio)") +
  guides(colour = guide_legend(position = "top", direction = "horizontal",
    title.position = "top", title.hjust = 0.5)) +
  theme(
    legend.margin = margin(0, 0, 0, 0),
    legend.justification.top = "left",
    legend.location = "plot",
    plot.title.position = "plot"
  )
tp_cdd_s_ri

```



```

p_cdd_s_ri <- predict_response(m_cdd_s_ri,
  terms=c("con_dens_s[1:80, by = 1]",
    "trt_F", "trt_I",
    "fragment.size[5, 25, 125]"),
  margin="marginalmeans")

pr_cdd_s_ri <- residualize_over_grid(p_cdd_s_ri, m_cdd_s_ri) |>
  bind_cols(species = scls$species) |>

```

```

mutate(trt = case_when(
  group == "0" & facet == "0" ~ "Control",
  group == "F" & facet == "0" ~ "Fungicide",
  group == "0" & facet == "I" ~ "Insecticide",
  group == "F" & facet == "I" ~ "FI"),
  facet_lab = "Fragment Area (ha)") |>
rename("con_dens" = "x", "frag_area" = "panel") |>
group_by(con_dens, trt, frag_area, facet_lab, species) |>
summarise(predicted = mean(predicted), n = n())

```

'summarise()' has grouped output by 'con_dens', 'trt', 'frag_area',
 ## 'facet_lab'. You can override using the '.groups' argument.

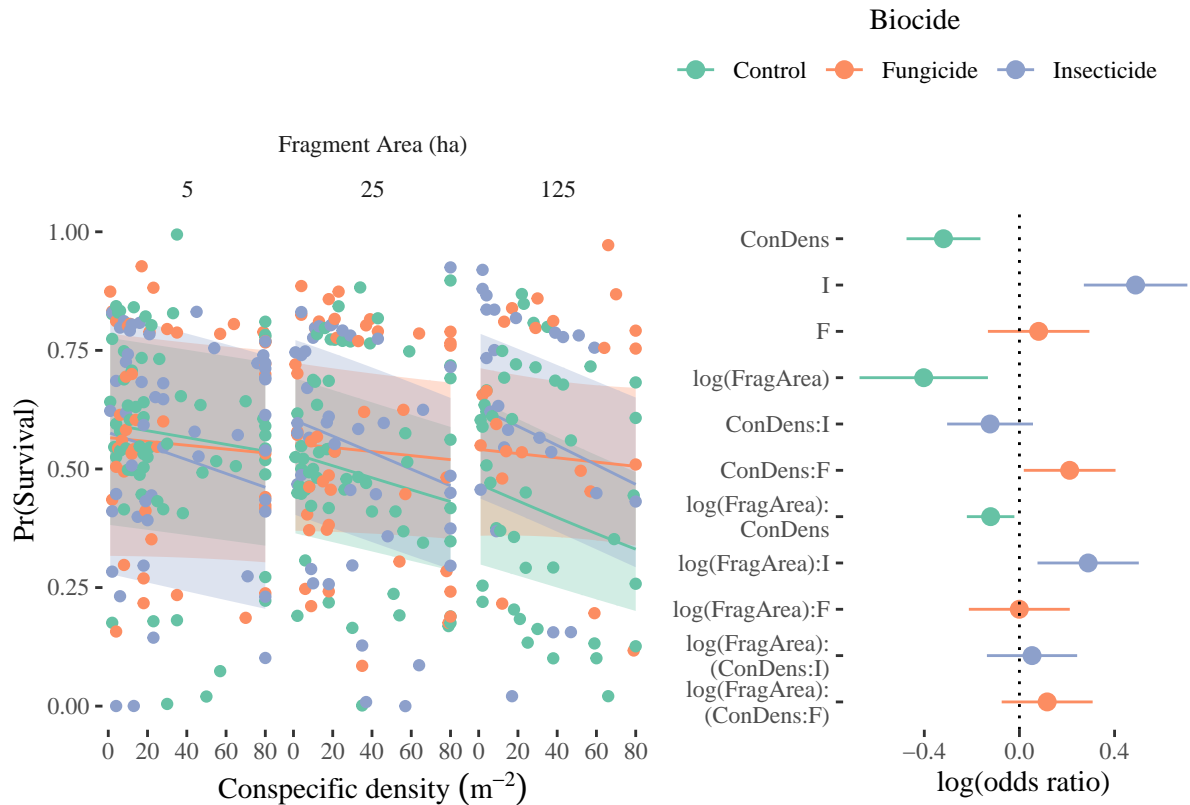
```

p_cdd_s_ri <- as.data.frame(p_cdd_s_ri) |>
mutate(trt = case_when(
  group == "0" & facet == "0" ~ "Control",
  group == "F" & facet == "0" ~ "Fungicide",
  group == "0" & facet == "I" ~ "Insecticide",
  group == "F" & facet == "I" ~ "FI"),
  facet_lab = "Fragment Area (ha)") |>
rename("con_dens" = "x", "frag_area" = "panel")

pl_cdd_s <- ggplot(filter(p_cdd_s_ri, trt != "FI"),
  aes(x=con_dens, y = predicted, colour = trt)) +
  ggh4x::facet_nested(~facet_lab + frag_area) +
  geom_ribbon(aes(ymin = conf.low, ymax = conf.high, fill = trt),
    colour = NA, alpha = 0.3) +
  geom_line() +
  geom_point(data = filter(pr_cdd_s_ri, trt != "FI")) +
  scale_colour_brewer(palette="Set2") +
  scale_fill_brewer(palette="Set2", guide = "none") +
  labs(x = expression(Conspecific~density~(m^-2)), y = "Pr(Survival)",
    colour = "Treatment") + theme(legend.position="none")

(pl_cdd_s | tp_cdd_s_ri ) +
  plot_layout(widths=c(0.6, 0.4))

```



Plotting effects of fragment area

The effects are complex and not terribly clear. To interrogate them more, here are some plots

```
## Using the model split by area to
sdl$ |> group_by(frag_sizeclass3) |> summarize(mean(fragment.size))
```

```
## # A tibble: 3 x 2
##   frag_sizeclass3 'mean(fragment.size)'
##   <fct>          <dbl>
## 1 [1.1,5.36]    2.64
## 2 (5.36,30.5]  12.2
## 3 (30.5,149]   85.3
```

```
## plot the effect of density over fragment size
int_data <- sdl$ |>
  mutate(con_dens_ss = scale(con_dens_s),
         tot_dens_s = scale(tot_dens),
         log_fragsize_s = scale(log(fragment.size)))

int_mod <- glmmTMB(Pr_s ~ slope.degrees_s +
  (tot_dens_s + con_dens_ss) *
  (trt_I + trt_F) * log_fragsize_s +
  (1 | species) +
```

```

      (1 | site/loc/gr/plot),
      weights = census.start, family = binomial,
      data = int_data)

preddat <- expand.grid(
  trt_F = levels(int_data$trt_F),
  con_dens_ss = 1,
  log_fragsize_s = seq(-2, 1.3, length = 20))

xmat <- model.matrix(~ con_dens_ss + con_dens_ss:(trt_F + log_fragsize_s) +
  con_dens_ss:trt_F:log_fragsize_s,
  preddat)[ , -1]
preddat$int_hat <- as.vector(xmat %*% fixef(int_mod)$cond[colnames(xmat)])
vmat <- (vcov(int_mod)$cond[colnames(xmat), colnames(xmat)])
preddat$int_se <- sqrt(diag(xmat %*% vmat %*% t(xmat)))

## Now insecticide
preddat_i <- expand.grid(
  trt_I = levels(int_data$trt_I),
  con_dens_ss = 1,
  log_fragsize_s = seq(-2, 1.3, length = 20))

xmat <- model.matrix(~ con_dens_ss + con_dens_ss:(trt_I + log_fragsize_s) +
  con_dens_ss:trt_I:log_fragsize_s,
  preddat_i)[ , -1]
preddat_i$int_hat <- as.vector(xmat %*% fixef(int_mod)$cond[colnames(xmat)])
vmat <- (vcov(int_mod)$cond[colnames(xmat), colnames(xmat)])
preddat_i$int_se <- sqrt(diag(xmat %*% vmat %*% t(xmat)))

preddat <- bind_rows(preddat |> rename("trt" = "trt_F"),
  filter(preddat_i, trt_I == "I") |> rename("trt" = "trt_I"))

preddat <- preddat |> mutate(.lower = int_hat - 1.96*int_se,
  .upper = int_hat + 1.96*int_se,
  frag_area = exp(log_fragsize_s*
    sd(log(sdls$fragment.size)) +
    mean(log(sdls$fragment.size))),
  trt = factor(trt,
    labels = c("Control", "Fungicide",
      "Insecticide")))

int_plot <- ggplot(preddat, aes(x = frag_area)) + facet_wrap( ~ trt) +
  geom_ribbon(aes(y = int_hat, ymin = .lower, ymax = .upper), alpha = 0.2, colour = NA) +
  geom_line(aes(y = int_hat)) + geom_hline(yintercept=0, linetype = "dotted") +
  #coord_trans(x = "log") +
  scale_x_continuous(trans = "log", breaks=c(2, 8, 32, 128)) +
  labs(x = "Fragment area (ha)",
    y = "Effect of conspecific density on survival")

sdls <- mutate(sdls, trt = case_when(
  trt_F == "F" ~ "F",
  trt_I == "I" ~ "I",
  .default = "C"),

```



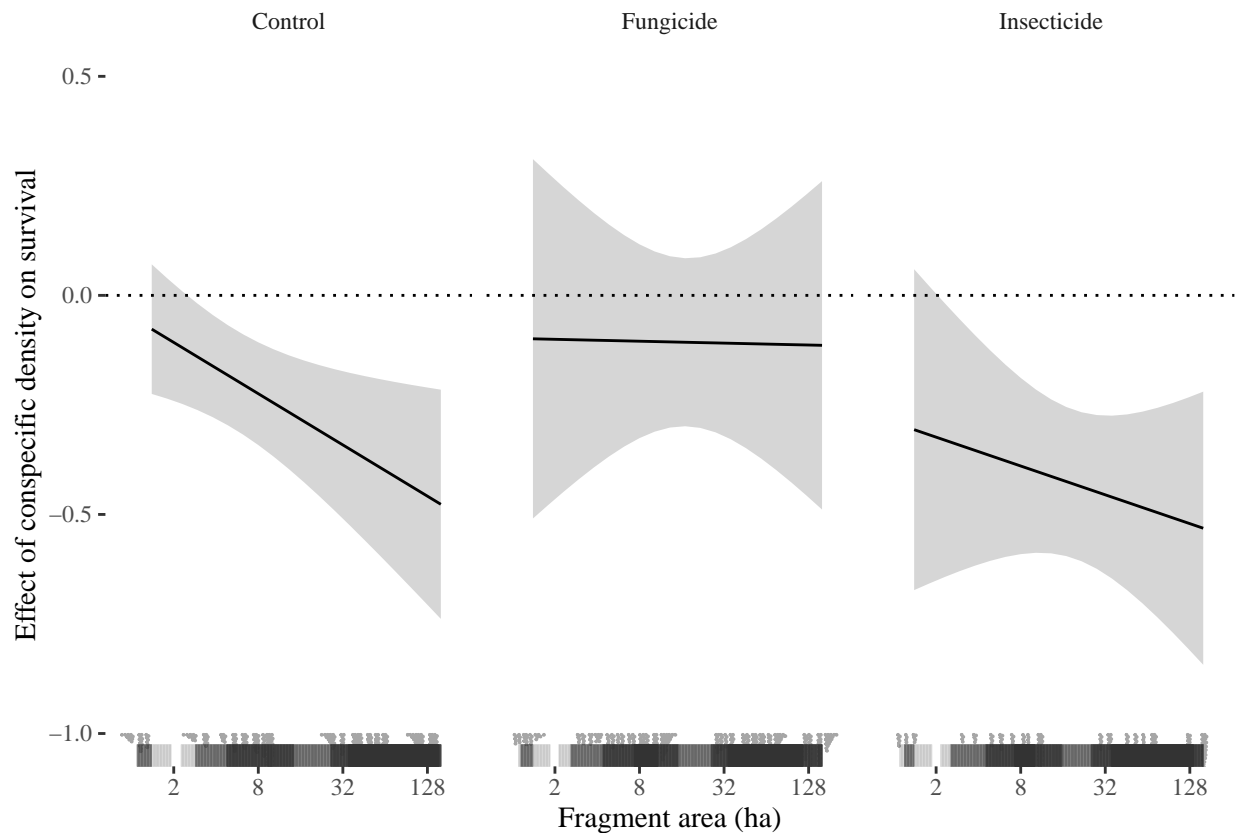
```

trt = factor(trt, labels = c("Control", "Fungicide", "Insecticide"))
library(ggdist)

int_plot <- int_plot + ggdist::geom_dots(data = sdls, aes(x = fragment.size), y = -1,
  smooth = smooth_unbounded(), layout = "swarm",
  side = "bottom", binwidth = 0.04, # alpha = 0.7,
  overflow = "compress") + ylim(c(-1, 0.5))

int_plot + ggdensity::geom_hdr_rug(data = sdls, aes(x = fragment.size)) +
  theme(legend.position="none")

```



```

## double checking results against interactions package (only works for
## lme4 and 2-way interactions
test_mod_C <- lme4::glmer(Pr_s ~ slope.degrees_s + trt_I +
  (tot_dens_s + con_dens_ss) *
  log_fragsize_s +
  (1|species) +
  ## setting cor to 0 to converge
  (1|site/loc/gr/plot),
  weights = census.start,
  data = filter(int_data, trt_F == "0") ,
  family=binomial)

```

```

## Warning in checkConv(attr(opt, "derivs"), opt$par, ctrl = control$checkConv, :
## Model failed to converge with max|grad| = 0.00859555 (tol = 0.002, component 1)

```

```
test_mod_F <- lme4::glmer(Pr_s ~ slope.degrees_s + trt_I +
  (tot_dens_s + con_dens_ss) *
  log_fragsize_s +
  (1|species) +
  ## setting cor to 0 to converge
  (1|site/loc/gr/plot),
  weights = census.start,
  data =filter(int_data, trt_F == "F"),
  family=binomial)
```

```
## boundary (singular) fit: see help('isSingular')
```

```
interactions::johnson_neyman(test_mod_C, pred = con_dens_ss,
  modx = log_fragsize_s)
```

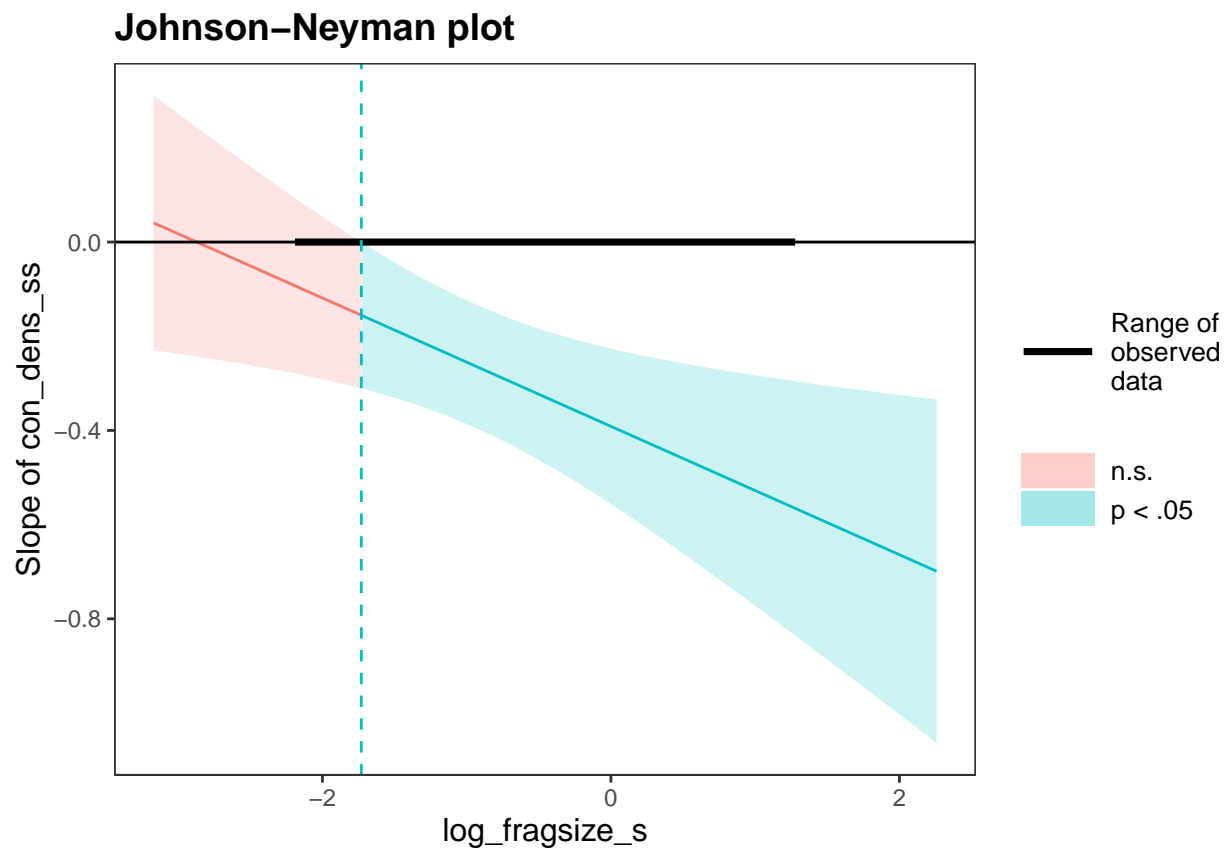
```
## JOHNSON-NEYMAN INTERVAL
```

```
##
```

```
## When log_fragsize_s is OUTSIDE the interval [-9.98, -1.73], the slope of
## con_dens_ss is p < .05.
```

```
##
```

```
## Note: The range of observed values of log_fragsize_s is [-2.17, 1.25]
```



```
exp(-1.73*sd(log(int_data$fragment.size)) + mean(log(int_data$fragment.size)))
```

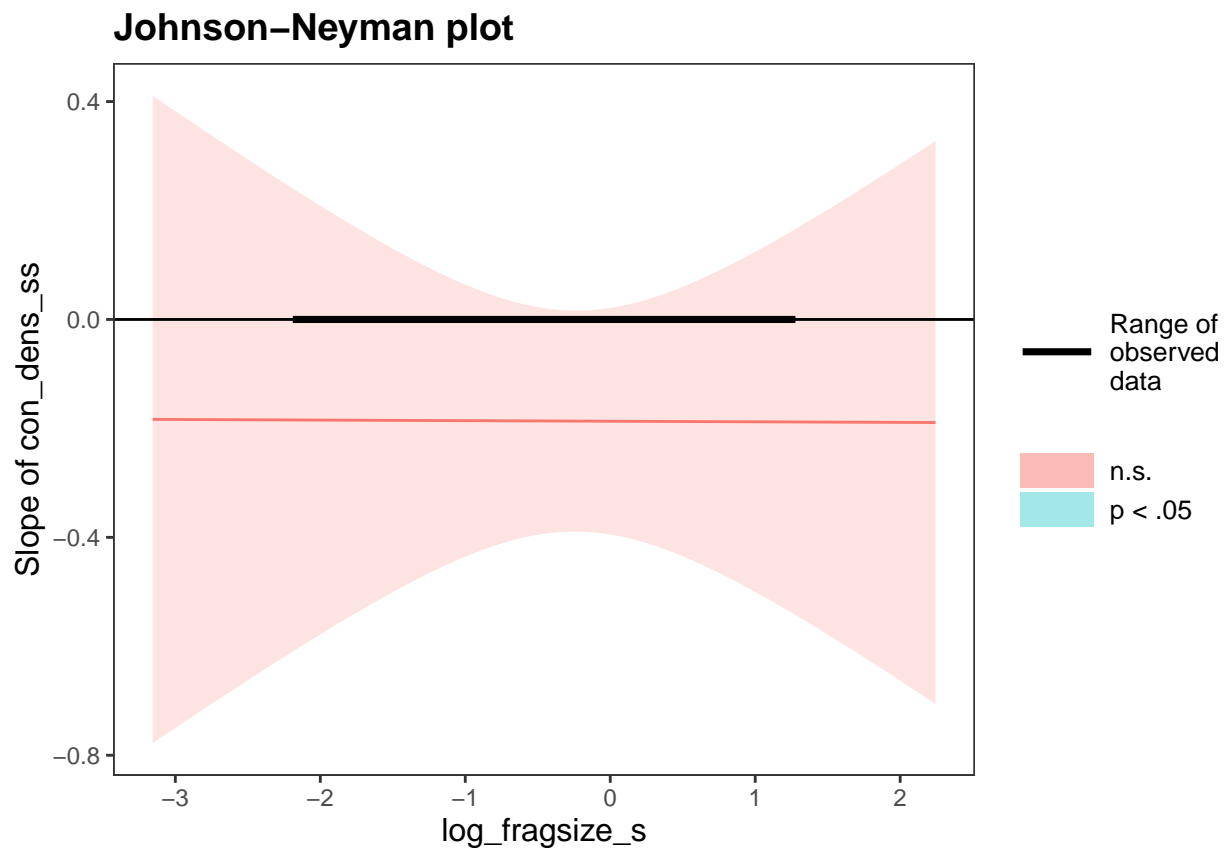
```
## [1] 2.055449
```

```
interactions::johnson_neyman(test_mod_F, pred = con_dens_ss,
                             modx = log_fragsize_s)
```

```
## JOHNSON-NEYMAN INTERVAL
```

```
##
```

```
## The Johnson-Neyman interval could not be found. Is the p value for your
## interaction term below the specified alpha?
```

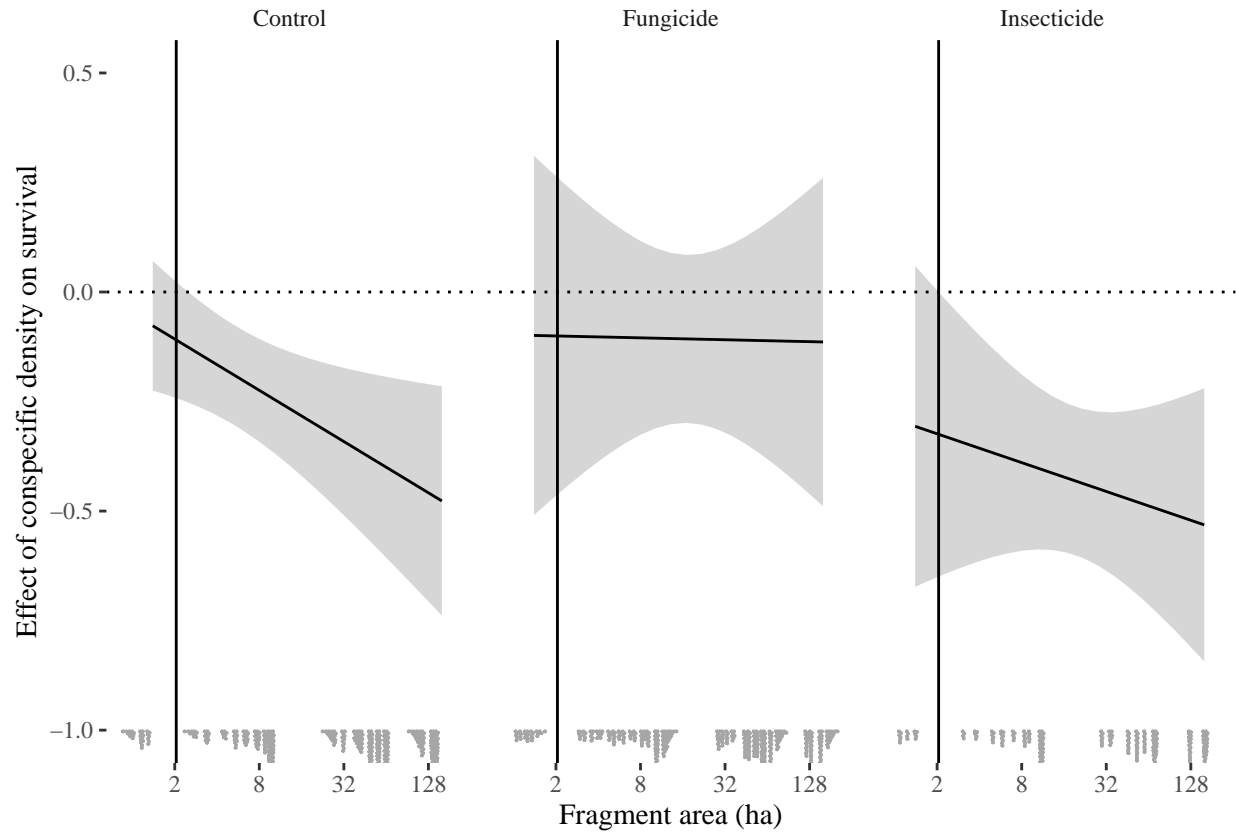


```
## basically the same result
```

```
int_plot + ggdist::geom_dots(data = sdds, aes(x = fragment.size), y = -1,
                             smooth = smooth_unbounded(), layout = "swarm",
                             side = "bottom", binwidth = 0.04, # alpha = 0.7,
                             overflow = "compress") + ylim(c(-1, 0.5)) +
  geom_vline(xintercept=2.05)
```

```
## Scale for y is already present.
```

```
## Adding another scale for y, which will replace the existing scale.
```

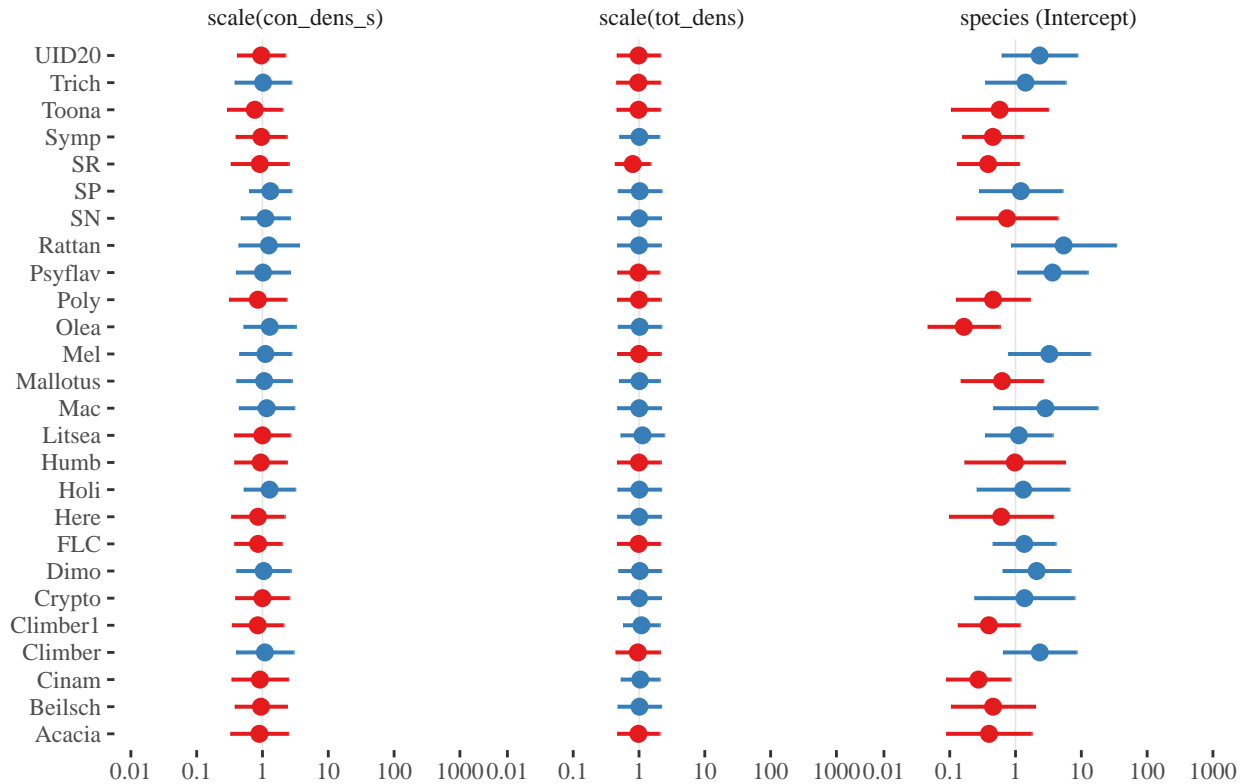


Species specific inferences.

Initial analyses suggested that some species were heavily influencing the results. The random slope should partly account for that.

```
sjPlot::plot_model(m_cdd_s_ris, type = "re", terms = "species", ri.nr = 1)
```

Random effects



very little variation among species in slopes, but perhaps obscured

by large variation in the intercept

```
sp_eff <- as.data.frame(ranef(m_cdd_s_ris, condVar = T)$cond$species)
```

```
sp_eff <- bind_cols(sp_eff,
                    t(apply(attr(sp_eff, "condVar"), 3, function(x)
                        sqrt(diag(x))))))
```

New names:

* ' -> '...4'

* ' -> '...5'

* ' -> '...6'

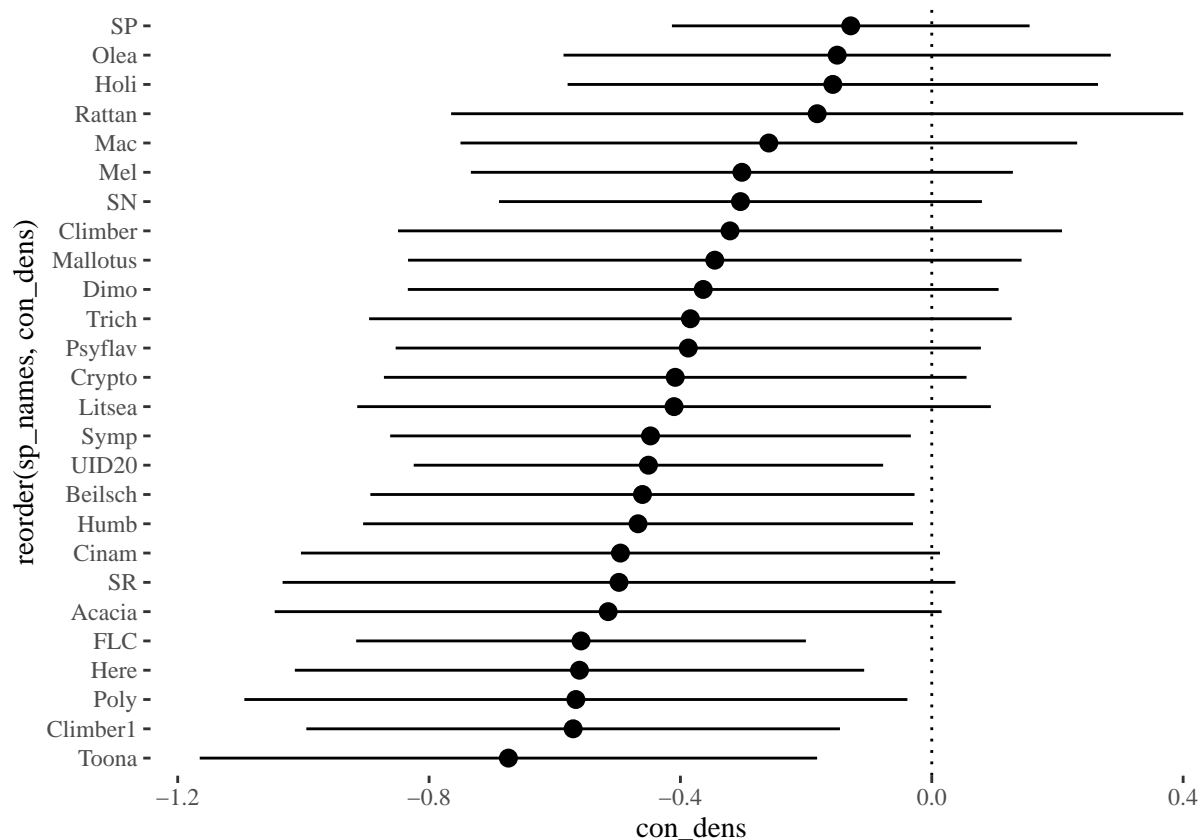
```
names(fixef(m_cdd_s_ris)$cond)
```

```
## [1] "(Intercept)"
## [2] "slope.degrees_s"
## [3] "scale(tot_dens)"
## [4] "scale(con_dens_s)"
## [5] "trt_II"
## [6] "trt_FF"
## [7] "scale(log(fragment.size))"
## [8] "scale(tot_dens):trt_II"
## [9] "scale(tot_dens):trt_FF"
## [10] "scale(con_dens_s):trt_II"
```

```
## [11] "scale(con_dens_s):trt_FF"
## [12] "scale(tot_dens):scale(log(fragment.size))"
## [13] "scale(con_dens_s):scale(log(fragment.size))"
## [14] "trt_II:scale(log(fragment.size))"
## [15] "trt_FF:scale(log(fragment.size))"
## [16] "scale(tot_dens):trt_II:scale(log(fragment.size))"
## [17] "scale(tot_dens):trt_FF:scale(log(fragment.size))"
## [18] "scale(con_dens_s):trt_II:scale(log(fragment.size))"
## [19] "scale(con_dens_s):trt_FF:scale(log(fragment.size))"
```

```
names(sp_eff) <- c("Intercept", "con_dens", "tot_dens",
                  "Intercept_se", "con_dens_se", "tot_dens_se")
sp_eff <- mutate(sp_eff, sp_names = row.names(sp_eff),
                 Intercept = Intercept + fixef(m_cdd_s_ris)$cond[1],
                 con_dens = con_dens + fixef(m_cdd_s_ris)$cond["scale(con_dens_s)"],
                 tot_dens = tot_dens + fixef(m_cdd_s_ris)$cond["scale(tot_dens)"])

sp_eff |> arrange(con_dens) |>
  ggplot(aes(y = reorder(sp_names, con_dens), x = con_dens,
                 xmin = con_dens - 2*con_dens_se,
                 xmax = con_dens + 2*con_dens_se)) +
  geom_vline(xintercept=0, linetype = "dotted") +
  geom_pointrange()
```



TBH not very illuminating.

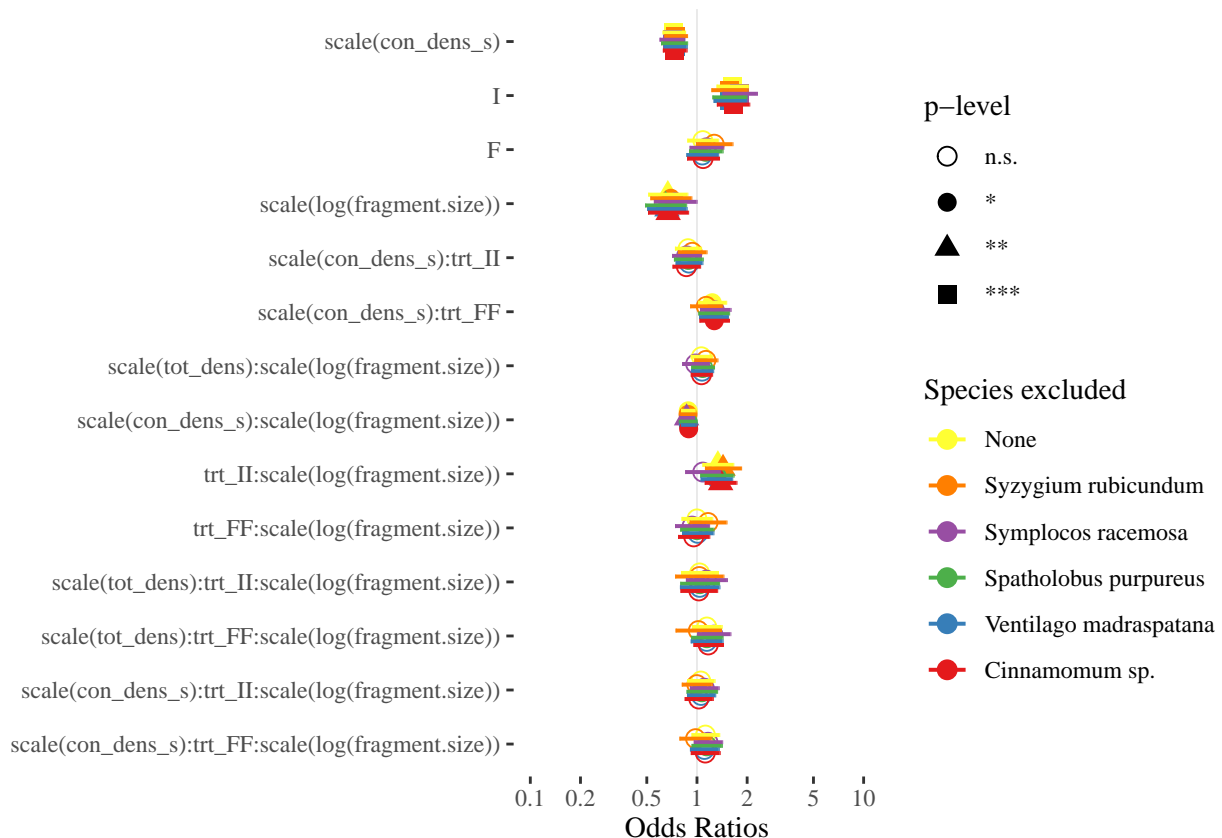
Little variation in cdd among species.

Perhaps remove most abundant species to check if patterns are robust.

```
sp_common <- sdls |> group_by(species) |> summarise(n = sum(census.start)) |>
  arrange(desc(n)) |> pull(species)

sp_mods <- lapply(c("none", as.character(sp_common[1:5])), function(i) {
  update(m_cdd_s_ri, data = filter(sdls, !species == i))})
sp_codes <- mutate(sp_codes, spbin = paste(genus, species))
term_nms <- names(fixef(m_cdd_s_ri_frag[[1]])$cond)

plot_models(sp_mods,
  rm.terms = c("slope.degrees_s",
    term_nms[str_detect(term_nms, "tot")]),
  m.labels = c("None",
    sp_codes$spbin[match(sp_common[1:5], sp_codes$code)]),
  p.shape=TRUE) + labs(colour = "Species excluded")
```



Only species that makes a difference is *S. rubicundum* - removing it dampens
interaction between density and fungicide. *Symplocos* affects insects a bit.

what about single species models?

```
## base model
## remove species random effect
names(sp_common) <- sp_common
single_sp_mods <- map(sp_common[1:5], function(i) {
  update(m_cdd_s_ri, data = filter(sdls, species == i)))
names(single_sp_mods) <- sp_codes$spbin[match(names(single_sp_mods), sp_codes$code)]
single_sp_mods$All <- m_cdd_s_ri

map(single_sp_mods, summary)
```

```
## $'Syzygium rubicundum'
## Family: binomial (logit)
## Formula:
## Pr_s ~ slope.degrees_s + (scale(tot_dens) + scale(con_dens_s)) *
## (trt_I + trt_F) * scale(log(fragment.size)) + (1 | species) +
## (1 | site/loc/gr/plot)
## Data: filter(sdls, species == i)
## Weights: census.start
##
##      AIC      BIC    logLik deviance df.resid
##    665.3    740.8   -308.6    617.3     148
##
## Random effects:
##
## Conditional model:
## Groups          Name          Variance Std.Dev.
## species          (Intercept)  2.446e-09 4.946e-05
## plot:gr:loc:site (Intercept)  2.332e-01 4.830e-01
## gr:loc:site      (Intercept)  2.258e-01 4.752e-01
## loc:site         (Intercept)  3.933e-01 6.272e-01
## site            (Intercept)  1.395e-01 3.735e-01
## Number of obs: 172, groups:
## species, 1; plot:gr:loc:site, 172; gr:loc:site, 61; loc:site, 30; site, 16
##
## Conditional model:
##
##                                     Estimate Std. Error z value
## (Intercept)                        -0.08341    0.23663   -0.352
## slope.degrees_s                     -0.26105    0.10750   -2.428
## scale(tot_dens)                     -0.97014    1.11892   -0.867
## scale(con_dens_s)                    0.26289    1.11257    0.236
## trt_II                              0.59067    0.19786    2.985
## trt_FF                             -0.14284    0.17851   -0.800
## scale(log(fragment.size))           -0.30395    0.23268   -1.306
## scale(tot_dens):trt_II               1.10639    1.40006    0.790
## scale(tot_dens):trt_FF               1.31054    1.61439    0.812
## scale(con_dens_s):trt_II             -1.09087    1.37685   -0.792
## scale(con_dens_s):trt_FF             -1.17973    1.53541   -0.768
## scale(tot_dens):scale(log(fragment.size)) 2.18684    1.33337    1.640
## scale(con_dens_s):scale(log(fragment.size)) -2.10714    1.32263   -1.593
## trt_II:scale(log(fragment.size))      0.13646    0.19755    0.691
## trt_FF:scale(log(fragment.size))     -0.24242    0.20876   -1.161
## scale(tot_dens):trt_II:scale(log(fragment.size)) -3.10582    2.10539   -1.475
## scale(tot_dens):trt_FF:scale(log(fragment.size)) -1.30978    2.73360   -0.479
```



```

## scale(con_dens_s):trt_II:scale(log(fragment.size)) 2.96720 2.00343 1.481
## scale(con_dens_s):trt_FF:scale(log(fragment.size)) 1.69460 2.59809 0.652
## Pr(>|z|)
## (Intercept) 0.72447
## slope.degrees_s 0.01516 *
## scale(tot_dens) 0.38593
## scale(con_dens_s) 0.81321
## trt_II 0.00283 **
## trt_FF 0.42361
## scale(log(fragment.size)) 0.19145
## scale(tot_dens):trt_II 0.42939
## scale(tot_dens):trt_FF 0.41692
## scale(con_dens_s):trt_II 0.42819
## scale(con_dens_s):trt_FF 0.44228
## scale(tot_dens):scale(log(fragment.size)) 0.10099
## scale(con_dens_s):scale(log(fragment.size)) 0.11113
## trt_II:scale(log(fragment.size)) 0.48973
## trt_FF:scale(log(fragment.size)) 0.24554
## scale(tot_dens):trt_II:scale(log(fragment.size)) 0.14017
## scale(tot_dens):trt_FF:scale(log(fragment.size)) 0.63184
## scale(con_dens_s):trt_II:scale(log(fragment.size)) 0.13859
## scale(con_dens_s):trt_FF:scale(log(fragment.size)) 0.51424
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## $'Symplocos racemosa'
## Family: binomial ( logit )
## Formula:
## Pr_s ~ slope.degrees_s + (scale(tot_dens) + scale(con_dens_s)) *
## (trt_I + trt_F) * scale(log(fragment.size)) + (1 | species) +
## (1 | site/loc/gr/plot)
## Data: filter(sdls, species == i)
## Weights: census.start
##
## AIC BIC logLik deviance df.resid
## 328.6 395.9 -140.3 280.6 98
##
## Random effects:
##
## Conditional model:
## Groups Name Variance Std.Dev.
## species (Intercept) 2.301e-10 1.517e-05
## plot:gr:loc:site (Intercept) 4.766e-02 2.183e-01
## gr:loc:site (Intercept) 2.541e-01 5.041e-01
## loc:site (Intercept) 2.110e-01 4.593e-01
## site (Intercept) 7.718e-09 8.785e-05
## Number of obs: 122, groups:
## species, 1; plot:gr:loc:site, 122; gr:loc:site, 53; loc:site, 24; site, 17
##
## Conditional model:
## Estimate Std. Error z value
## (Intercept) 0.56597 0.26352 2.148
## slope.degrees_s 0.04268 0.15992 0.267
## scale(tot_dens) 0.03218 0.43931 0.073

```

```

## scale(con_dens_s) -0.50090 0.43672 -1.147
## trt_II -0.19685 0.41647 -0.473
## trt_FF 0.34812 0.53743 0.648
## scale(log(fragment.size)) -0.82686 0.29724 -2.782
## scale(tot_dens):trt_II -2.34392 1.34650 -1.741
## scale(tot_dens):trt_FF 1.92338 1.65705 1.161
## scale(con_dens_s):trt_II 0.96019 1.11657 0.860
## scale(con_dens_s):trt_FF -1.32933 1.11749 -1.190
## scale(tot_dens):scale(log(fragment.size)) -0.06029 0.43948 -0.137
## scale(con_dens_s):scale(log(fragment.size)) -0.21580 0.38066 -0.567
## trt_II:scale(log(fragment.size)) 1.07587 0.51913 2.072
## trt_FF:scale(log(fragment.size)) -0.07420 0.62343 -0.119
## scale(tot_dens):trt_II:scale(log(fragment.size)) 1.45064 0.99840 1.453
## scale(tot_dens):trt_FF:scale(log(fragment.size)) 0.24135 1.66497 0.145
## scale(con_dens_s):trt_II:scale(log(fragment.size)) -0.38161 0.96850 -0.394
## scale(con_dens_s):trt_FF:scale(log(fragment.size)) -1.53899 1.19084 -1.292
## Pr(>|z|)
## (Intercept) 0.03173 *
## slope.degrees_s 0.78957
## scale(tot_dens) 0.94161
## scale(con_dens_s) 0.25139
## trt_II 0.63646
## trt_FF 0.51715
## scale(log(fragment.size)) 0.00541 **
## scale(tot_dens):trt_II 0.08173 .
## scale(tot_dens):trt_FF 0.24576
## scale(con_dens_s):trt_II 0.38982
## scale(con_dens_s):trt_FF 0.23422
## scale(tot_dens):scale(log(fragment.size)) 0.89089
## scale(con_dens_s):scale(log(fragment.size)) 0.57078
## trt_II:scale(log(fragment.size)) 0.03822 *
## trt_FF:scale(log(fragment.size)) 0.90526
## scale(tot_dens):trt_II:scale(log(fragment.size)) 0.14624
## scale(tot_dens):trt_FF:scale(log(fragment.size)) 0.88474
## scale(con_dens_s):trt_II:scale(log(fragment.size)) 0.69356
## scale(con_dens_s):trt_FF:scale(log(fragment.size)) 0.19623
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## $'Spatholobus purpureus'
## Family: binomial ( logit )
## Formula:
## Pr_s ~ slope.degrees_s + (scale(tot_dens) + scale(con_dens_s)) *
## (trt_I + trt_F) * scale(log(fragment.size)) + (1 | species) +
## (1 | site/loc/gr/plot)
## Data: filter(sdls, species == i)
## Weights: census.start
##
## AIC BIC logLik deviance df.resid
## 223.9 280.2 -88.0 175.9 53
##
## Random effects:
##
## Conditional model:

```

```

## Groups          Name          Variance Std.Dev.
## species         (Intercept) 6.017e-15 7.757e-08
## plot:gr:loc:site (Intercept) 8.422e-10 2.902e-05
## gr:loc:site      (Intercept) 5.644e-13 7.513e-07
## loc:site         (Intercept) 3.947e-12 1.987e-06
## site            (Intercept) 2.264e-12 1.505e-06
## Number of obs: 77, groups:
## species, 1; plot:gr:loc:site, 77; gr:loc:site, 30; loc:site, 15; site, 10
##
## Conditional model:
##
## Estimate Std. Error z value
## (Intercept) -0.69938 0.26370 -2.652
## slope.degrees_s -0.18592 0.16791 -1.107
## scale(tot_dens) -0.35819 0.34294 -1.044
## scale(con_dens_s) 0.30691 0.38895 0.789
## trt_II 0.46813 0.89568 0.523
## trt_FF 0.03359 0.35750 0.094
## scale(log(fragment.size)) 0.06661 0.37245 0.179
## scale(tot_dens):trt_II -0.70777 0.41933 -1.688
## scale(tot_dens):trt_FF 0.42425 0.42882 0.989
## scale(con_dens_s):trt_II -0.41483 1.81542 -0.228
## scale(con_dens_s):trt_FF -0.92405 0.54114 -1.708
## scale(tot_dens):scale(log(fragment.size)) 0.37376 0.60126 0.622
## scale(con_dens_s):scale(log(fragment.size)) -1.30508 1.05454 -1.238
## trt_II:scale(log(fragment.size)) 1.06328 1.95823 0.543
## trt_FF:scale(log(fragment.size)) 0.59247 0.55174 1.074
## scale(tot_dens):trt_II:scale(log(fragment.size)) 0.37598 0.71378 0.527
## scale(tot_dens):trt_FF:scale(log(fragment.size)) -0.48323 0.76957 -0.628
## scale(con_dens_s):trt_II:scale(log(fragment.size)) 1.42882 5.04251 0.283
## scale(con_dens_s):trt_FF:scale(log(fragment.size)) 2.10801 1.40836 1.497
## Pr(>|z|)
## (Intercept) 0.0080 **
## slope.degrees_s 0.2682
## scale(tot_dens) 0.2963
## scale(con_dens_s) 0.4301
## trt_II 0.6012
## trt_FF 0.9251
## scale(log(fragment.size)) 0.8581
## scale(tot_dens):trt_II 0.0914 .
## scale(tot_dens):trt_FF 0.3225
## scale(con_dens_s):trt_II 0.8193
## scale(con_dens_s):trt_FF 0.0877 .
## scale(tot_dens):scale(log(fragment.size)) 0.5342
## scale(con_dens_s):scale(log(fragment.size)) 0.2159
## trt_II:scale(log(fragment.size)) 0.5871
## trt_FF:scale(log(fragment.size)) 0.2829
## scale(tot_dens):trt_II:scale(log(fragment.size)) 0.5984
## scale(tot_dens):trt_FF:scale(log(fragment.size)) 0.5301
## scale(con_dens_s):trt_II:scale(log(fragment.size)) 0.7769
## scale(con_dens_s):trt_FF:scale(log(fragment.size)) 0.1345
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## $'Ventilago madraspatana'

```

```

## Family: binomial ( logit )
## Formula:
## Pr_s ~ slope.degrees_s + (scale(tot_dens) + scale(con_dens_s)) *
##      (trt_I + trt_FF) * scale(log(fragment.size)) + (1 | species) +
##      (1 | site/loc/gr/plot)
## Data: filter(sdls, species == i)
## Weights: census.start
##
##      AIC      BIC    logLik deviance df.resid
##    143.4    195.3    -47.7     95.4      40
##
## Random effects:
##
## Conditional model:
## Groups          Name          Variance Std.Dev.
## species          (Intercept) 3.615e-14 1.901e-07
## plot:gr:loc:site (Intercept) 2.882e-10 1.698e-05
## gr:loc:site      (Intercept) 1.192e-14 1.092e-07
## loc:site         (Intercept) 1.293e-14 1.137e-07
## site             (Intercept) 1.187e-13 3.445e-07
## Number of obs: 64, groups:
## species, 1; plot:gr:loc:site, 64; gr:loc:site, 27; loc:site, 16; site, 10
##
## Conditional model:
##
##                                     Estimate Std. Error z value
## (Intercept)                        1.17286    0.41678   2.814
## slope.degrees_s                    -0.40301    0.24721  -1.630
## scale(tot_dens)                     1.35978    1.17608   1.156
## scale(con_dens_s)                  -1.50109    0.86550  -1.734
## trt_II                             2.26471    0.83208   2.722
## trt_FF                             -0.72767    0.61846  -1.177
## scale(log(fragment.size))           0.08774    0.45545   0.193
## scale(tot_dens):trt_II              -1.51287    1.87200  -0.808
## scale(tot_dens):trt_FF              -1.71057    1.72319  -0.993
## scale(con_dens_s):trt_II            -2.06232    1.49050  -1.384
## scale(con_dens_s):trt_FF            2.43123    1.32627   1.833
## scale(tot_dens):scale(log(fragment.size)) 0.31136    1.35724   0.229
## scale(con_dens_s):scale(log(fragment.size)) 0.12988    1.08290   0.120
## trt_II:scale(log(fragment.size))      0.77671    0.91299   0.851
## trt_FF:scale(log(fragment.size))     -0.01336    0.74706  -0.018
## scale(tot_dens):trt_II:scale(log(fragment.size)) -2.45905    1.99499  -1.233
## scale(tot_dens):trt_FF:scale(log(fragment.size)) 0.58715    1.94177   0.302
## scale(con_dens_s):trt_II:scale(log(fragment.size)) -1.39202    1.61694  -0.861
## scale(con_dens_s):trt_FF:scale(log(fragment.size)) 0.19768    1.59299   0.124
##
##                                     Pr(>|z|)
## (Intercept)                        0.00489 **
## slope.degrees_s                     0.10306
## scale(tot_dens)                      0.24760
## scale(con_dens_s)                    0.08285 .
## trt_II                              0.00649 **
## trt_FF                              0.23936
## scale(log(fragment.size))            0.84725
## scale(tot_dens):trt_II                0.41900
## scale(tot_dens):trt_FF                0.32087

```

```

## scale(con_dens_s):trt_II                                0.16647
## scale(con_dens_s):trt_FF                                0.06678 .
## scale(tot_dens):scale(log(fragment.size))                0.81856
## scale(con_dens_s):scale(log(fragment.size))              0.90453
## trt_II:scale(log(fragment.size))                          0.39492
## trt_FF:scale(log(fragment.size))                          0.98573
## scale(tot_dens):trt_II:scale(log(fragment.size))         0.21772
## scale(tot_dens):trt_FF:scale(log(fragment.size))         0.76236
## scale(con_dens_s):trt_II:scale(log(fragment.size))       0.38930
## scale(con_dens_s):trt_FF:scale(log(fragment.size))       0.90124
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## $'Cinnamomum sp.'
## Family: binomial (logit)
## Formula:
## Pr_s ~ slope.degrees_s + (scale(tot_dens) + scale(con_dens_s)) *
##       (trt_I + trt_F) * scale(log(fragment.size)) + (1 | species) +
##       (1 | site/loc/gr/plot)
## Data: filter(sdls, species == i)
## Weights: census.start
##
##      AIC      BIC   logLik deviance df.resid
##    162.9    221.8   -57.5    114.9      62
##
## Random effects:
##
## Conditional model:
## Groups          Name          Variance Std.Dev.
## species          (Intercept)  6.917e-10 2.630e-05
## plot:gr:loc:site (Intercept)  7.773e-11 8.816e-06
## gr:loc:site      (Intercept)  2.035e+00 1.427e+00
## loc:site         (Intercept)  3.525e-08 1.878e-04
## site             (Intercept)  9.322e-01 9.655e-01
## Number of obs: 86, groups:
## species, 1; plot:gr:loc:site, 86; gr:loc:site, 42; loc:site, 23; site, 17
##
## Conditional model:
##
##                                     Estimate Std. Error z value
## (Intercept)                        -0.40851    0.58085  -0.703
## slope.degrees_s                     -0.32176    0.58540  -0.550
## scale(tot_dens)                     -0.97114    0.79709  -1.218
## scale(con_dens_s)                    0.35924    0.52315   0.687
## trt_II                              0.45861    0.87887   0.522
## trt_FF                              0.84066    1.79322   0.469
## scale(log(fragment.size))           -0.65747    0.54850  -1.199
## scale(tot_dens):trt_II               0.35824    2.25319   0.159
## scale(tot_dens):trt_FF               3.34970    5.22263   0.641
## scale(con_dens_s):trt_II             -0.03752    1.29818  -0.029
## scale(con_dens_s):trt_FF            -0.67189    1.35854  -0.495
## scale(tot_dens):scale(log(fragment.size)) 0.38663    0.59626   0.648
## scale(con_dens_s):scale(log(fragment.size)) 0.46242    0.47832   0.967
## trt_II:scale(log(fragment.size))     -1.04864    1.03031  -1.018
## trt_FF:scale(log(fragment.size))      1.67013    2.08919   0.799

```

```

## scale(tot_dens):trt_II:scale(log(fragment.size)) -1.68922 2.69246 -0.627
## scale(tot_dens):trt_FF:scale(log(fragment.size)) 1.56474 6.11823 0.256
## scale(con_dens_s):trt_II:scale(log(fragment.size)) 0.29282 1.16878 0.250
## scale(con_dens_s):trt_FF:scale(log(fragment.size)) 0.08156 1.51954 0.054
##
## Pr(>|z|)
## (Intercept) 0.482
## slope.degrees_s 0.583
## scale(tot_dens) 0.223
## scale(con_dens_s) 0.492
## trt_II 0.602
## trt_FF 0.639
## scale(log(fragment.size)) 0.231
## scale(tot_dens):trt_II 0.874
## scale(tot_dens):trt_FF 0.521
## scale(con_dens_s):trt_II 0.977
## scale(con_dens_s):trt_FF 0.621
## scale(tot_dens):scale(log(fragment.size)) 0.517
## scale(con_dens_s):scale(log(fragment.size)) 0.334
## trt_II:scale(log(fragment.size)) 0.309
## trt_FF:scale(log(fragment.size)) 0.424
## scale(tot_dens):trt_II:scale(log(fragment.size)) 0.530
## scale(tot_dens):trt_FF:scale(log(fragment.size)) 0.798
## scale(con_dens_s):trt_II:scale(log(fragment.size)) 0.802
## scale(con_dens_s):trt_FF:scale(log(fragment.size)) 0.957
##
## $All
## Family: binomial ( logit )
## Formula:
## Pr_s ~ slope.degrees_s + (scale(tot_dens) + scale(con_dens_s)) *
## (trt_I + trt_F) * scale(log(fragment.size)) * (scale(con_dens_s) +
## scale(tot_dens) || species) + (1 | site/loc/gr/plot)
## Data: sdls
## Weights: census.start
##
## AIC BIC logLik deviance df.resid
## 2187.2 2313.9 -1067.6 2135.2 942
##
## Random effects:
##
## Conditional model:
## Groups Name Variance Std.Dev. Corr
## species (Intercept) 1.135e+00 1.0653405
## scale(con_dens_s) 6.608e-02 0.2570611 0.00
## scale(tot_dens) 2.325e-02 0.1524655 0.00 0.00
## plot:gr:loc:site (Intercept) 2.424e-01 0.4923436
## gr:loc:site (Intercept) 1.804e-01 0.4247101
## loc:site (Intercept) 3.926e-01 0.6266100
## site (Intercept) 2.861e-08 0.0001691
## Number of obs: 968, groups:
## species, 26; plot:gr:loc:site, 474; gr:loc:site, 110; loc:site, 37; site, 21
##
## Conditional model:
## Estimate Std. Error z value
## (Intercept) 0.69057 0.27999 2.466

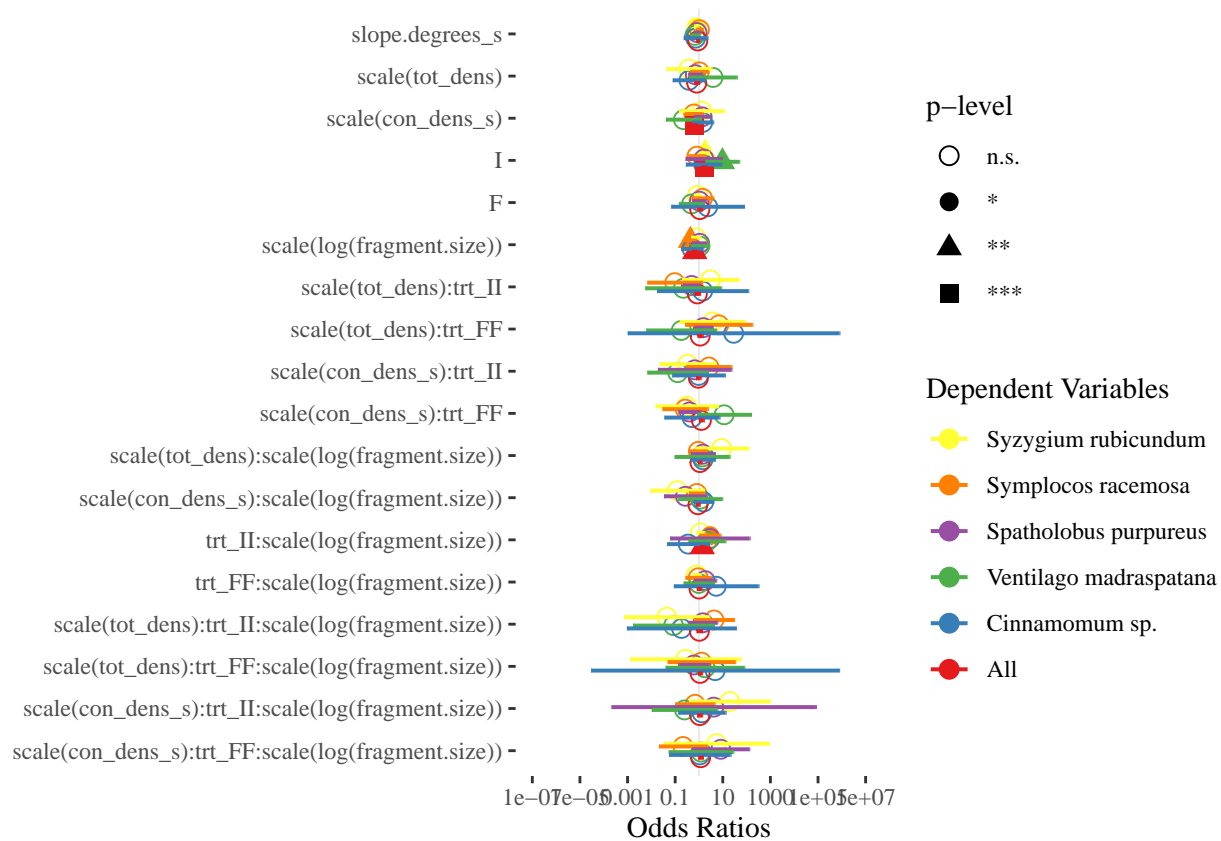
```

```

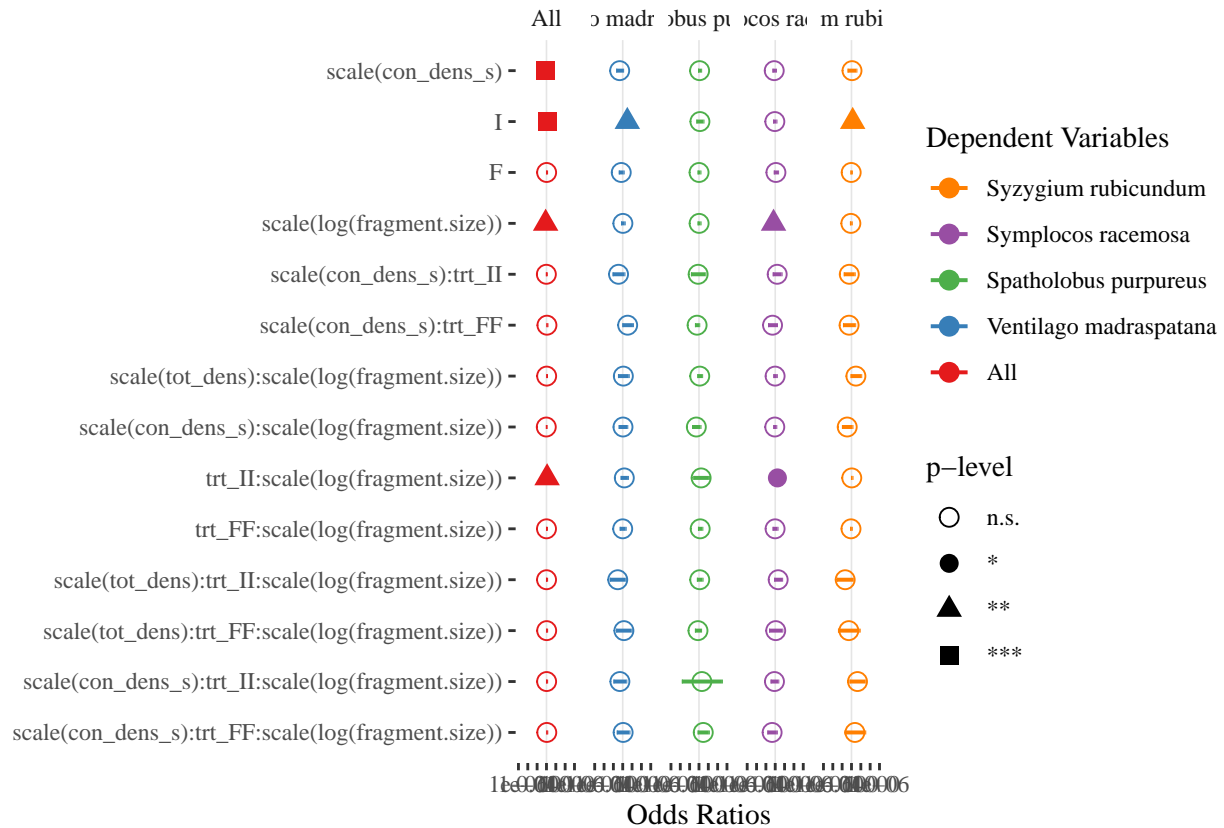
## slope.degrees_s -0.11658 0.06911 -1.687
## scale(tot_dens) -0.21421 0.12004 -1.785
## scale(con_dens_s) -0.40432 0.11910 -3.395
## trt_II 0.49115 0.11285 4.352
## trt_FF 0.08216 0.11073 0.742
## scale(log(fragment.size)) -0.41120 0.13689 -3.004
## scale(tot_dens):trt_II -0.15738 0.13792 -1.141
## scale(tot_dens):trt_FF 0.11644 0.12482 0.933
## scale(con_dens_s):trt_II -0.05721 0.11426 -0.501
## scale(con_dens_s):trt_FF 0.23160 0.12231 1.894
## scale(tot_dens):scale(log(fragment.size)) 0.09539 0.08419 1.133
## scale(con_dens_s):scale(log(fragment.size)) -0.09961 0.09165 -1.087
## trt_II:scale(log(fragment.size)) 0.31420 0.11084 2.835
## trt_FF:scale(log(fragment.size)) 0.00739 0.10952 0.067
## scale(tot_dens):trt_II:scale(log(fragment.size)) 0.03831 0.13434 0.285
## scale(tot_dens):trt_FF:scale(log(fragment.size)) 0.10500 0.10844 0.968
## scale(con_dens_s):trt_II:scale(log(fragment.size)) 0.07438 0.12322 0.604
## scale(con_dens_s):trt_FF:scale(log(fragment.size)) 0.16136 0.11441 1.410
## Pr(>|z|)
## (Intercept) 0.013646 *
## slope.degrees_s 0.091636 .
## scale(tot_dens) 0.074342 .
## scale(con_dens_s) 0.000686 ***
## trt_II 1.35e-05 ***
## trt_FF 0.458117
## scale(log(fragment.size)) 0.002666 **
## scale(tot_dens):trt_II 0.253811
## scale(tot_dens):trt_FF 0.350924
## scale(con_dens_s):trt_II 0.616592
## scale(con_dens_s):trt_FF 0.058277 .
## scale(tot_dens):scale(log(fragment.size)) 0.257195
## scale(con_dens_s):scale(log(fragment.size)) 0.277076
## trt_II:scale(log(fragment.size)) 0.004586 **
## trt_FF:scale(log(fragment.size)) 0.946205
## scale(tot_dens):trt_II:scale(log(fragment.size)) 0.775502
## scale(tot_dens):trt_FF:scale(log(fragment.size)) 0.332894
## scale(con_dens_s):trt_II:scale(log(fragment.size)) 0.546084
## scale(con_dens_s):trt_FF:scale(log(fragment.size)) 0.158447
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

```
plot_models(single_sp_mods, m.labels=names(single_sp_mods), p.shape=TRUE)
```



```
## Cinnamomum distorts scale, so dropping it
plot_models(single_sp_mods[c(1:4, 6)],
            rm.terms = c("slope.degrees_s",
                        term_nms[str_detect(term_nms, "tot")] ),
            m.labels=names(single_sp_mods)[c(1:4, 6)],
            p.shape = TRUE, grid=TRUE)
```

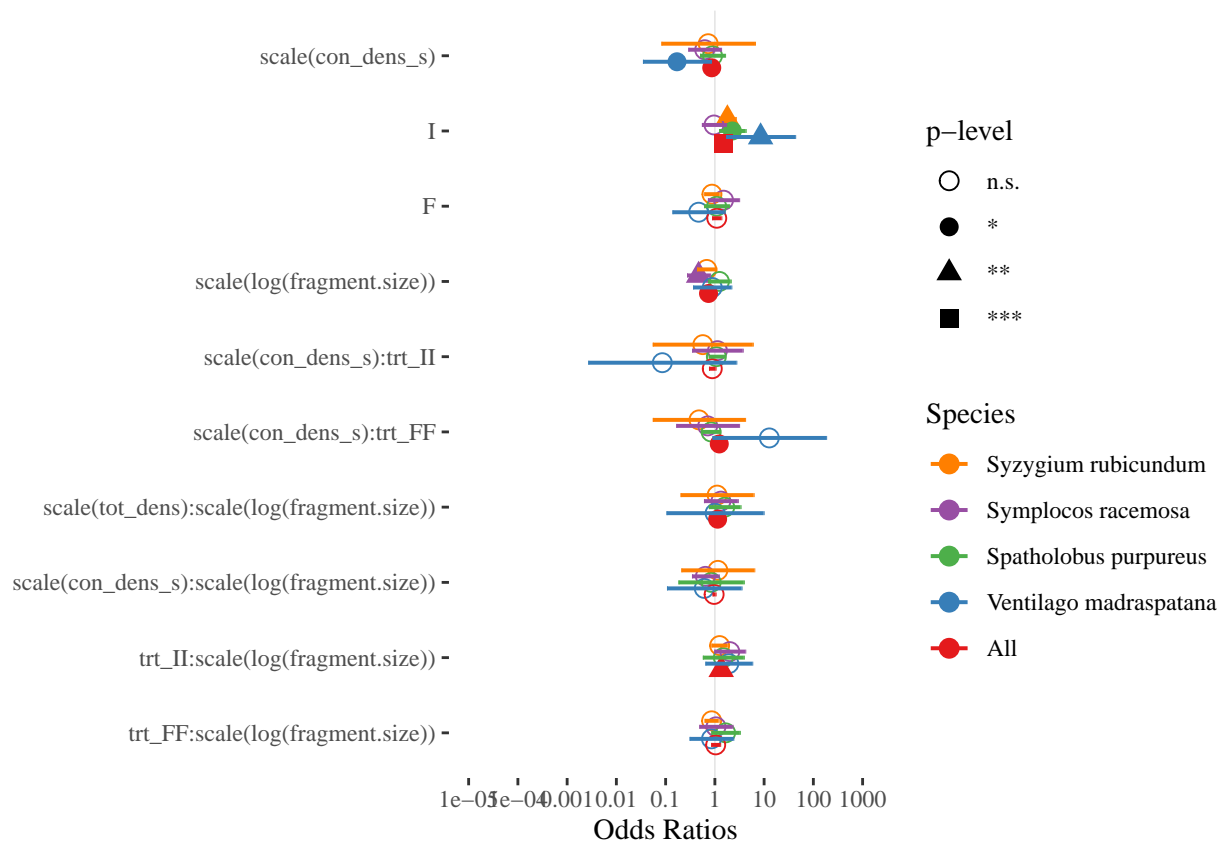



```
## perhaps too much complexity. Dropping 3-way interactions
##

m_cdd_s_ri_2way <- glmmTMB(Pr_s ~ slope.degrees_s +
  (scale(tot_dens) + scale(con_dens_s)) *
  (trt_I + trt_F) +
  scale(log(fragment.size)) *
  (scale(tot_dens) + scale(con_dens_s) + trt_I + trt_F) +
  (1|site/loc/gr/plot),
  weights = census.start, data = sdls,
  family=binomial)

single_sp_mods <- map(sp_common[1:4], function(i) {
  update(m_cdd_s_ri_2way, data = filter(sdls, species == i))})
names(single_sp_mods) <- sp_codes$spbin[match(names(single_sp_mods), sp_codes$code)]
single_sp_mods$All <- m_cdd_s_ri_2way

plot_models(single_sp_mods,
  rm.terms = c("slope.degrees_s",
    term_nms[str_detect(term_nms, "tot")]},
  m.labels=names(single_sp_mods),
  p.shape = TRUE, grid=FALSE) + labs(colour = "Species")
```



To be honest, probably not enough data for individual species. Dropping the 3-way interactions helps fit the models, but only notable effects are density dependence in Ventilago and effects of fragment size on symplocus.

Session Information

```
sessionInfo()
```

```
## R version 4.4.0 (2024-04-24 ucrt)
## Platform: x86_64-w64-mingw32/x64
## Running under: Windows 10 x64 (build 17763)
##
## Matrix products: default
##
## locale:
## [1] LC_COLLATE=English_United States.1252
## [2] LC_CTYPE=English_United States.1252
## [3] LC_MONETARY=English_United States.1252
## [4] LC_NUMERIC=C
## [5] LC_TIME=English_United States.1252
##
## time zone: America/New_York
```

```

## tzcode source: internal
##
## attached base packages:
## [1] stats      graphics  grDevices  utils      datasets  methods   base
##
## other attached packages:
## [1] ggdist_3.3.2      patchwork_1.2.0    sjPlot_2.8.16
## [4] ggeffects_1.6.0.2 broom.mixed_0.2.9.5 DHARMA_0.4.6
## [7] glmmTMB_1.1.9      knitr_1.47          ggthemes_5.1.0
## [10] lubridate_1.9.3    forcats_1.0.0      stringr_1.5.1
## [13] dplyr_1.1.4        purrr_1.0.2         readr_2.1.5
## [16] tidyr_1.3.1        tibble_3.2.1        ggplot2_3.5.1
## [19] tidyverse_2.0.0
##
## loaded via a namespace (and not attached):
## [1] Rdpack_2.6          rlang_1.1.3          magrittr_2.0.3
## [4] furrr_0.3.1         compiler_4.4.0        mgcv_1.9-1
## [7] vctrs_0.6.5         pkgconfig_2.0.3       crayon_1.5.2
## [10] fastmap_1.2.0       backports_1.5.0       labeling_0.4.3
## [13] pander_0.6.5        effectsize_0.8.8      utf8_1.2.4
## [16] promises_1.3.0      rmarkdown_2.27        tzdb_0.4.0
## [19] haven_2.5.4         nloptr_2.0.3          bit_4.0.5
## [22] xfun_0.44           highr_0.11            later_1.3.2
## [25] sjmisc_2.8.10       broom_1.0.6           parallel_4.4.0
## [28] R6_2.5.1            gap.datasets_0.0.6    stringi_1.8.4
## [31] qgam_1.3.4          RColorBrewer_1.1-3    parallely_1.37.1
## [34] car_3.1-2           boot_1.3-30           numDeriv_2016.8-1.1
## [37] estimability_1.5.1  Rcpp_1.0.12           iterators_1.0.14
## [40] parameters_0.21.7   httpuv_1.6.15         Matrix_1.7-0
## [43] splines_4.4.0       timechange_0.3.0      tidyselect_1.2.1
## [46] rstudioapi_0.16.0   abind_1.4-5           yaml_2.3.8
## [49] doParallel_1.0.17   TMB_1.9.11            codetools_0.2-20
## [52] sjlabelled_1.2.0    listenv_0.9.1          lattice_0.22-6
## [55] plyr_1.8.9          shiny_1.8.1.1         withr_3.0.0
## [58] bayestestR_0.13.2   coda_0.19-4.1         evaluate_0.23
## [61] future_1.33.2       jtools_2.2.2          pillar_1.9.0
## [64] gap_1.5-3           carData_3.0-5         foreach_1.5.2
## [67] insight_0.19.11     distributional_0.4.0   generics_0.1.3
## [70] vroom_1.6.5         hms_1.1.3             munsell_0.5.1
## [73] scales_1.3.0        minqa_1.2.7           globals_0.16.3
## [76] xtable_1.8-4        glue_1.7.0            emmeans_1.10.2
## [79] tools_4.4.0         lme4_1.1-35.3         mvtnorm_1.2-5
## [82] grid_4.4.0          rbibutils_2.2.16      datawizard_0.10.0
## [85] colorspace_2.1-0    nlme_3.1-164          performance_0.11.0
## [88] beeswarm_0.4.0      cli_3.6.2             fansi_1.0.6
## [91] ggdensity_1.0.0     interactions_1.1.5     sjstats_0.19.0
## [94] ggh4x_0.2.8         gtable_0.3.5          digest_0.6.35
## [97] farver_2.1.2        htmltools_0.5.8.1     lifecycle_1.0.4
## [100] mime_0.12           bit64_4.0.5           MASS_7.3-60.2

```