

Class8

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Background

The goal of this mini-project is for you to explore a complete analysis using the unsupervised learning techniques covered in class. You'll extend what you've learned by combining PCA as a preprocessing step to clustering using data that consist of measurements of cell nuclei of human breast masses. This expands on our RNA-Seq analysis from last day.

The data itself comes from the Wisconsin Breast Cancer Diagnostic Data Set first reported by K. P. Benne and O. L. Mangasarian: "Robust Linear Programming Discrimination of Two Linearly Inseparable Sets".

Data Import

Here we downloaded the file to our laptops then added it to our BIMM 143 folder. Then it will appear in the "Files" on the right and can be added into wisc.df.

```
wisc.df <- read.csv("WisconsinCancer.csv", row.names=1)
```

Now let's view some of our data.

```
head(wisc.df)
```

	diagnosis	radius_mean	texture_mean	perimeter_mean	area_mean
842302	M	17.99	10.38	122.80	1001.0
842517	M	20.57	17.77	132.90	1326.0
84300903	M	19.69	21.25	130.00	1203.0
84348301	M	11.42	20.38	77.58	386.1
84358402	M	20.29	14.34	135.10	1297.0
843786	M	12.45	15.70	82.57	477.1

	smoothness_mean	compactness_mean	concavity_mean	concave.points_mean
842302	0.11840	0.27760	0.3001	0.14710
842517	0.08474	0.07864	0.0869	0.07017
84300903	0.10960	0.15990	0.1974	0.12790
84348301	0.14250	0.28390	0.2414	0.10520
84358402	0.10030	0.13280	0.1980	0.10430
843786	0.12780	0.17000	0.1578	0.08089

	symmetry_mean	fractal_dimension_mean	radius_se	texture_se	perimeter_se
842302	0.2419	0.07871	1.0950	0.9053	8.589
842517	0.1812	0.05667	0.5435	0.7339	3.398
84300903	0.2069	0.05999	0.7456	0.7869	4.585
84348301	0.2597	0.09744	0.4956	1.1560	3.445
84358402	0.1809	0.05883	0.7572	0.7813	5.438
843786	0.2087	0.07613	0.3345	0.8902	2.217

	area_se	smoothness_se	compactness_se	concavity_se	concave.points_se
842302	153.40	0.006399	0.04904	0.05373	0.01587
842517	74.08	0.005225	0.01308	0.01860	0.01340
84300903	94.03	0.006150	0.04006	0.03832	0.02058
84348301	27.23	0.009110	0.07458	0.05661	0.01867
84358402	94.44	0.011490	0.02461	0.05688	0.01885
843786	27.19	0.007510	0.03345	0.03672	0.01137

	symmetry_se	fractal_dimension_se	radius_worst	texture_worst
842302	0.03003	0.006193	25.38	17.33
842517	0.01389	0.003532	24.99	23.41
84300903	0.02250	0.004571	23.57	25.53
84348301	0.05963	0.009208	14.91	26.50
84358402	0.01756	0.005115	22.54	16.67
843786	0.02165	0.005082	15.47	23.75

	perimeter_worst	area_worst	smoothness_worst	compactness_worst
842302	184.60	2019.0	0.1622	0.6656
842517	158.80	1956.0	0.1238	0.1866
84300903	152.50	1709.0	0.1444	0.4245
84348301	98.87	567.7	0.2098	0.8663

84358402	152.20	1575.0	0.1374	0.2050
843786	103.40	741.6	0.1791	0.5249
	concavity_worst	concave.points_worst	symmetry_worst	
842302	0.7119	0.2654	0.4601	
842517	0.2416	0.1860	0.2750	
84300903	0.4504	0.2430	0.3613	
84348301	0.6869	0.2575	0.6638	
84358402	0.4000	0.1625	0.2364	
843786	0.5355	0.1741	0.3985	
	fractal_dimension_worst			
842302	0.11890			
842517	0.08902			
84300903	0.08758			
84348301	0.17300			
84358402	0.07678			
843786	0.12440			

Let's remove the first column from the data.

```
wisc.data <- wisc.df[, -1]
```

Let's create a vector for later that contains the diagnosis column data.

```
diagnosis<-factor(wisc.df$diagnosis)
```

Q1. How many observations are in the diagnosis dataset?

```
length(wisc.df$diagnosis)
```

```
[1] 569
```

Q2. How many observations are malignant?

```
table(wisc.df$diagnosis)
```

```

  B    M
357 212

```

Q3. How many variables/features in the data are suffixed with `_mean`?

```
allcolnames<-colnames(wisc.data)
grep("_mean",allcolnames,value=TRUE)
```

```
[1] "radius_mean"          "texture_mean"        "perimeter_mean"
[4] "area_mean"           "smoothness_mean"     "compactness_mean"
[7] "concavity_mean"      "concave.points_mean" "symmetry_mean"
[10] "fractal_dimension_mean"
```

Principal Component Analysis (PCA)

We need to determine if we should perform a PCA on the data. If units are different across the data it will likely be beneficial to scale the data.

```
colMeans(wisc.data)
```

radius_mean	texture_mean	perimeter_mean
1.412729e+01	1.928965e+01	9.196903e+01
area_mean	smoothness_mean	compactness_mean
6.548891e+02	9.636028e-02	1.043410e-01
concavity_mean	concave.points_mean	symmetry_mean
8.879932e-02	4.891915e-02	1.811619e-01
fractal_dimension_mean	radius_se	texture_se
6.279761e-02	4.051721e-01	1.216853e+00
perimeter_se	area_se	smoothness_se
2.866059e+00	4.033708e+01	7.040979e-03
compactness_se	concavity_se	concave.points_se
2.547814e-02	3.189372e-02	1.179614e-02
symmetry_se	fractal_dimension_se	radius_worst
2.054230e-02	3.794904e-03	1.626919e+01
texture_worst	perimeter_worst	area_worst
2.567722e+01	1.072612e+02	8.805831e+02
smoothness_worst	compactness_worst	concavity_worst
1.323686e-01	2.542650e-01	2.721885e-01
concave.points_worst	symmetry_worst	fractal_dimension_worst
1.146062e-01	2.900756e-01	8.394582e-02

```
apply(wisc.data,2,sd)
```

radius_mean	texture_mean	perimeter_mean
-------------	--------------	----------------

3.524049e+00	4.301036e+00	2.429898e+01
area_mean	smoothness_mean	compactness_mean
3.519141e+02	1.406413e-02	5.281276e-02
concavity_mean	concave.points_mean	symmetry_mean
7.971981e-02	3.880284e-02	2.741428e-02
fractal_dimension_mean	radius_se	texture_se
7.060363e-03	2.773127e-01	5.516484e-01
perimeter_se	area_se	smoothness_se
2.021855e+00	4.549101e+01	3.002518e-03
compactness_se	concavity_se	concave.points_se
1.790818e-02	3.018606e-02	6.170285e-03
symmetry_se	fractal_dimension_se	radius_worst
8.266372e-03	2.646071e-03	4.833242e+00
texture_worst	perimeter_worst	area_worst
6.146258e+00	3.360254e+01	5.693570e+02
smoothness_worst	compactness_worst	concavity_worst
2.283243e-02	1.573365e-01	2.086243e-01
concave.points_worst	symmetry_worst	fractal_dimension_worst
6.573234e-02	6.186747e-02	1.806127e-02

As we can see it is worth performing a PCA.

```
wisc.pr <- prcomp(wisc.data, scale=TRUE)
summary(wisc.pr)
```

Importance of components:

	PC1	PC2	PC3	PC4	PC5	PC6	PC7
Standard deviation	3.6444	2.3857	1.67867	1.40735	1.28403	1.09880	0.82172
Proportion of Variance	0.4427	0.1897	0.09393	0.06602	0.05496	0.04025	0.02251
Cumulative Proportion	0.4427	0.6324	0.72636	0.79239	0.84734	0.88759	0.91010
	PC8	PC9	PC10	PC11	PC12	PC13	PC14
Standard deviation	0.69037	0.6457	0.59219	0.5421	0.51104	0.49128	0.39624
Proportion of Variance	0.01589	0.0139	0.01169	0.0098	0.00871	0.00805	0.00523
Cumulative Proportion	0.92598	0.9399	0.95157	0.9614	0.97007	0.97812	0.98335
	PC15	PC16	PC17	PC18	PC19	PC20	PC21
Standard deviation	0.30681	0.28260	0.24372	0.22939	0.22244	0.17652	0.1731
Proportion of Variance	0.00314	0.00266	0.00198	0.00175	0.00165	0.00104	0.0010
Cumulative Proportion	0.98649	0.98915	0.99113	0.99288	0.99453	0.99557	0.9966
	PC22	PC23	PC24	PC25	PC26	PC27	PC28
Standard deviation	0.16565	0.15602	0.1344	0.12442	0.09043	0.08307	0.03987
Proportion of Variance	0.00091	0.00081	0.0006	0.00052	0.00027	0.00023	0.00005
Cumulative Proportion	0.99749	0.99830	0.9989	0.99942	0.99969	0.99992	0.99997

	PC29	PC30
Standard deviation	0.02736	0.01153
Proportion of Variance	0.00002	0.00000
Cumulative Proportion	1.00000	1.00000

Q4. What proportion of the original variance is captured by the first principal components?

Answer: 0.4427

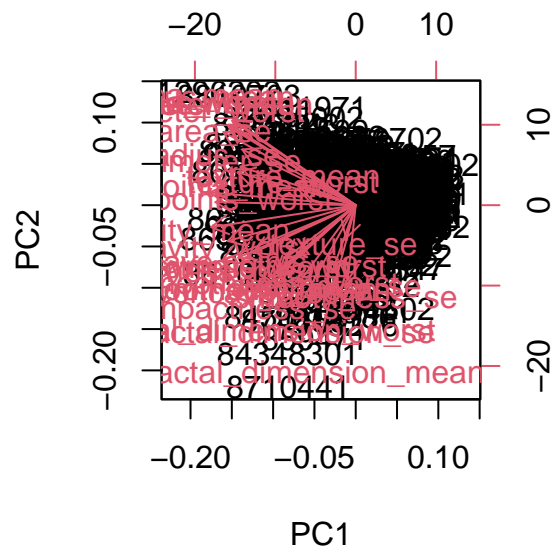
Q5. How many principal components (PCs) are required to describe at least 70% of the original variance in the data?

Answer: For this we look at cumulative proportion. PC3 contains 0.72636 of the original variance.

Q6. How many principal components (PCs) are required to describe at least 90% of the original variance in the data?

Answer: PC7.

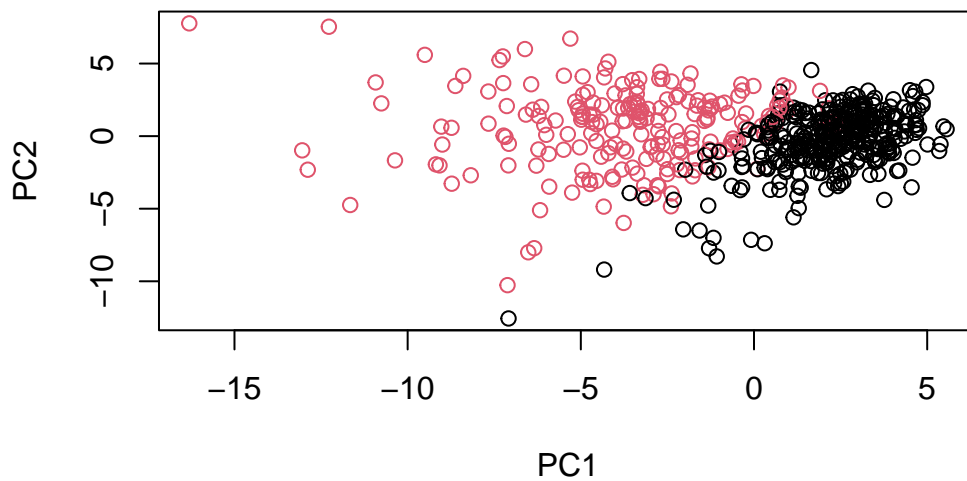
```
biplot(wisc.pr)
```



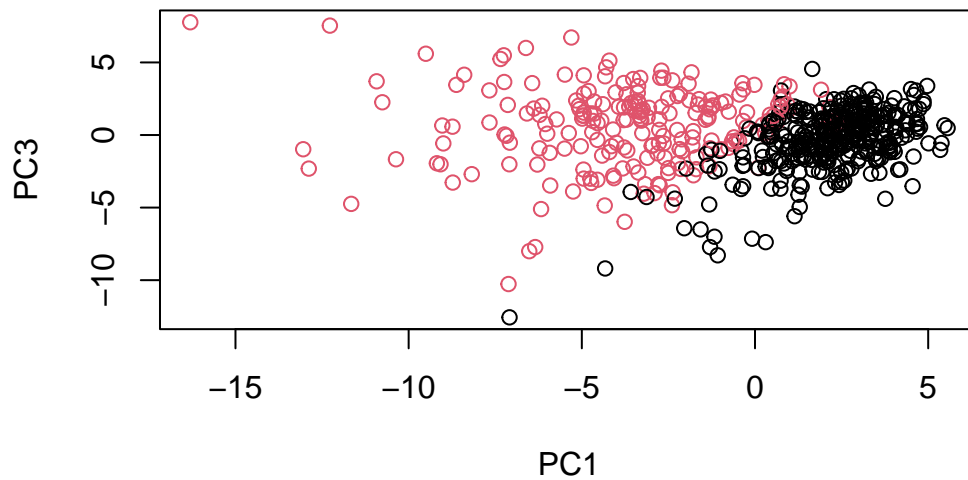
Q7. What stands out to you about this plot? Is it easy or difficult to understand? Why?

Answer: What stands out is how much information is shown at once making it hard for me to understand. There are numbers on all four sides of the graph and too much overlap at data at once.

```
plot( wisc.pr$x[, 1:2] , col = diagnosis ,  
      xlab = "PC1", ylab = "PC2")
```



```
plot(wisc.pr$x[, 1:2 ], col = diagnosis,  
      xlab = "PC1", ylab = "PC3")
```



Q8. Generate a similar plot for principal components 1 and 3. What do you notice about these plots?

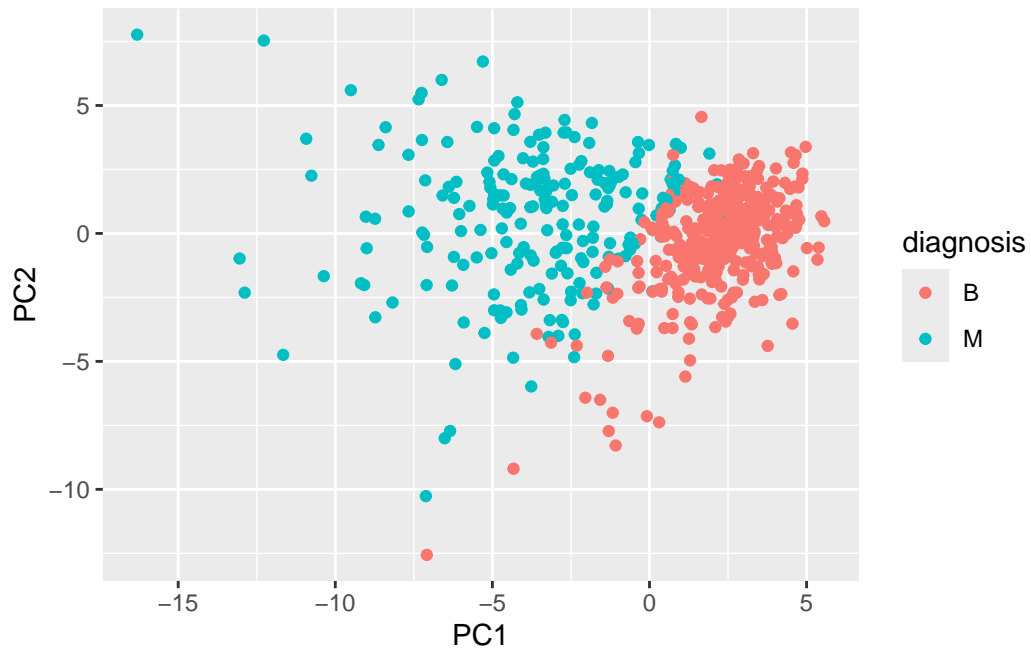
Answer: They are separating the spread between malignant and benign tumors.

The main PCA result figure is called a “score plot” or “PC plot” or “ordination plot”...

```
library(ggplot2)
```

Lets plot PC1 vs PC2.

```
ggplot(wisc.pr$x)+  
  aes(PC1,PC2,col=diagnosis)+  
  geom_point()
```

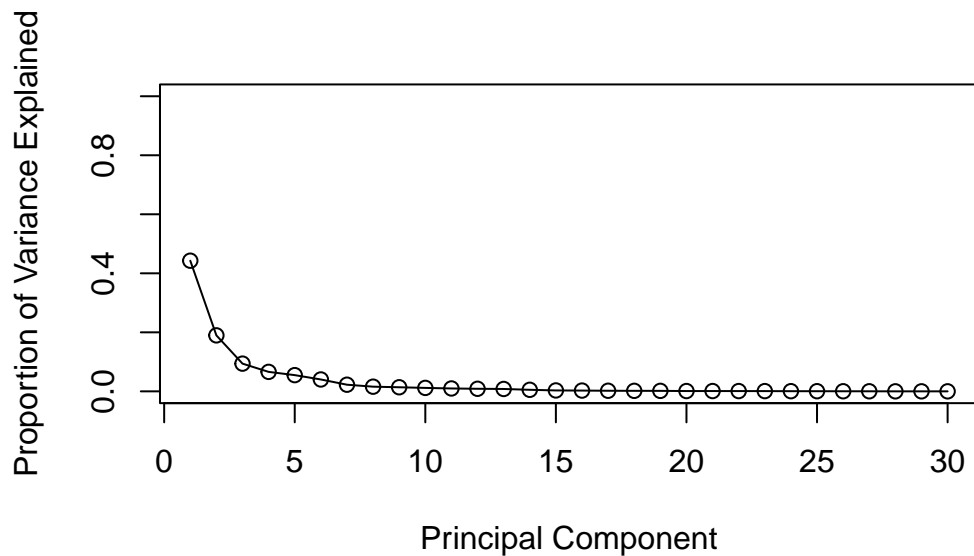



Let's calculate the variance of each component in wisc.pr.

```
pr.var <- wisc.pr$sdev^2  
head(pr.var)
```

```
[1] 13.281608  5.691355  2.817949  1.980640  1.648731  1.207357
```

```
pve <- pr.var / sum(pr.var)  
  
# Plot variance explained for each principal component  
plot(pve, xlab = "Principal Component",  
      ylab = "Proportion of Variance Explained",  
      ylim = c(0, 1), type = "o")
```



Communicating PCA results

Q9. For the first principal component, what is the component of the loading vector (i.e. `wisc.pr$rotation[,1]`) for the feature `concave.points_mean`?

Answer: -0.2609

```
summary(wisc.pr$rotation["concave.points_mean", "PC1"])
```

Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
-0.2609	-0.2609	-0.2609	-0.2609	-0.2609	-0.2609

Q10. What is the minimum number of principal components required to explain 80% of the variance of the data?

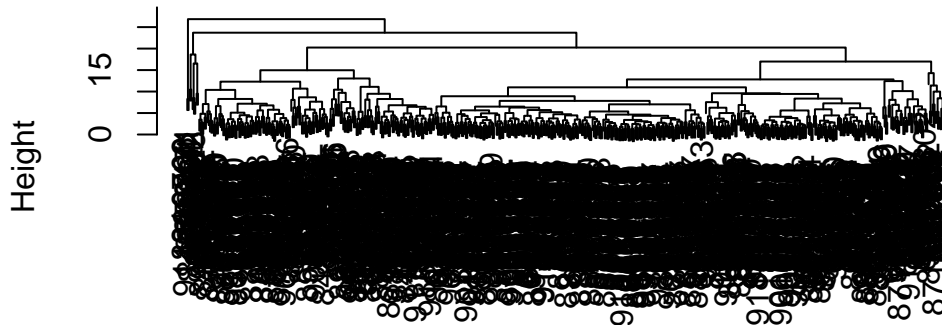
Answer: PC5

Hierarchical clustering

Just clustering the original data is not very informative or helpful.

```
data.scaled<-scale(wisc.data)
data.dist<-dist(data.scaled)
wisc.hclust<-hclust(data.dist)
plot(wisc.hclust)
```

Cluster Dendrogram



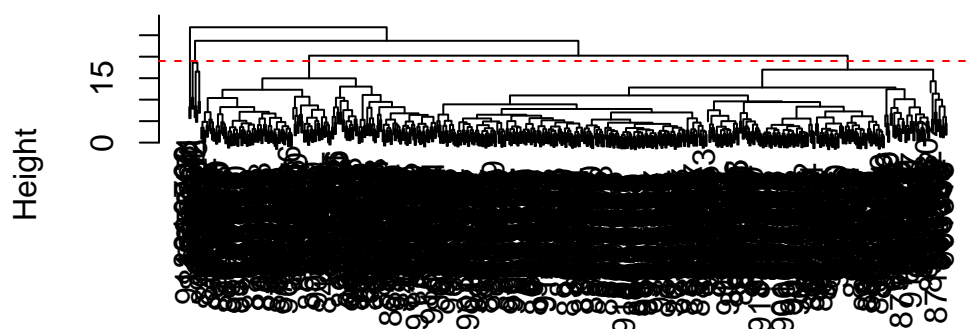
```
data.dist
hclust (*, "complete")
```

Q11. Using the plot() and abline() functions, what is the height at which the clustering model has 4 clusters?

Answer: h=19

```
plot(wisc.hclust)
abline(h=19, col="red", lty=2)
```

Cluster Dendrogram



data.dist
hclust (*, "complete")

```
wisc.hclust.clusters<-cutree(wisc.hclust,k=2)
table(cutree(wisc.hclust,k=2))
```

```
1  2
567 2
```

```
table(wisc.hclust.clusters,diagnosis)
```

```
          diagnosis
wisc.hclust.clusters  B  M
1  357 210
2    0   2
```

Q12. Can you find a better cluster vs diagnoses match by cutting into a different number of clusters between 2 and 10?

Answer: Two clusters provides a more even distribution between benign and malignant.

Q13. Which method gives your favorite results for the same data.dist dataset? Explain your reasoning.

Answer: I prefer ward.D2 the most because the plots themselves are very busy with data and this method most clearly conveys the high amount of information.

```
wisc.km <- kmeans(wisc.data, centers= 2, nstart= 20)
```

```
table(wisc.km$cluster, diagnosis)
```

	diagnosis	
	B	M
1	356	82
2	1	130

Q14. How well does k-means separate the two diagnoses? How does it compare to your hclust results?

Answer: HClust once adjusted to two clusters does a slightly better job in my opinion of separating the results into a more even distribution while k-means does a decent job but not as well. HClust separated benign and malignant into roughly 350 and 200 while the distribution for k-means is about 350 to 130.

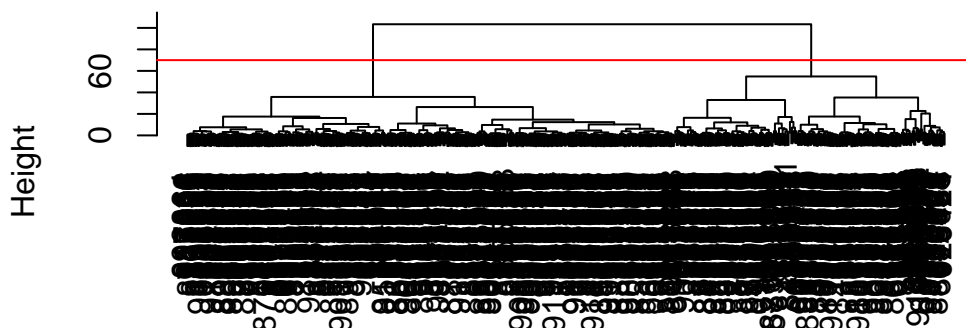
Combining methods (PCA and Clustering)

Clustering the original data was not productive. The PCA results looked promising. Here we combine these methods by clustering from our PCA results. In other words “clustering in PC space”.

```
## Take the first 3 PCs
dist.pc<-dist(wisc.pr$x[,1:3])
wisc.pr.hclust<-hclust(dist.pc,method="ward.D2")
```

```
plot(wisc.pr.hclust)
abline(h=70,col="red")
```

Cluster Dendrogram



```
dist.pc
hclust (*, "ward.D2")
```

To get our clustering membership vector (i.e. our main clustering result) we “cut” the tree at a desired height or to yield a desired number of “k” groups.

This is the code being used but for the sake of not adding 5 pages it will not be executed.

```
cutree(wisc.pr.hclust,k=2)
```

```
grps<-cutree(wisc.pr.hclust,h=70)
table(grps)
```

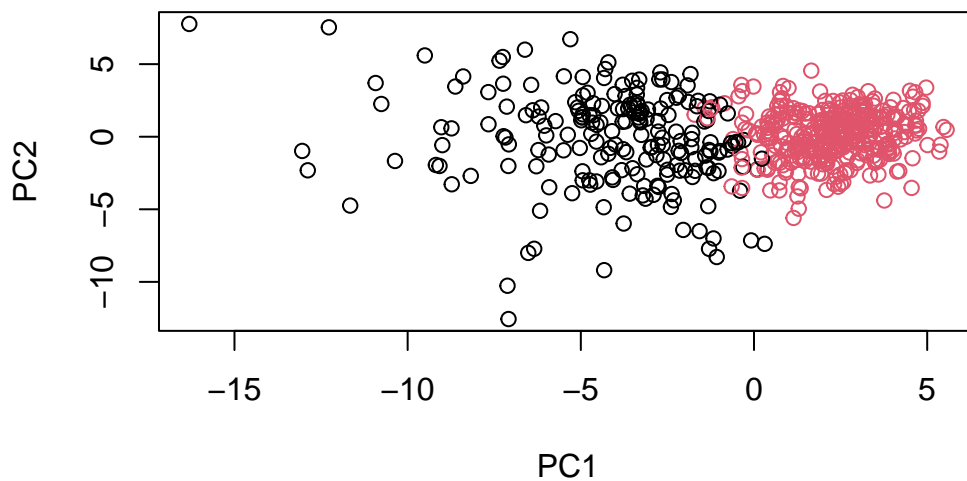
```
grps
  1  2
203 366
```

How does this clustering grps compare to the expert?

```
table(grps,diagnosis)
```

```
      diagnosis
grps   B    M
  1   24 179
  2  333   33
```

```
plot(wisc.pr$x[,1:2], col=grps)
```



```
wisc.pr.dist <- dist(wisc.pr$x[, 1:7])
wisc.pr.hclust <- hclust(wisc.pr.dist, method="ward.D2")
wisc.pr.hclust.clusters <- cutree(wisc.pr.hclust, k=4)
table(wisc.pr.hclust.clusters,diagnosis)
```

	diagnosis	
wisc.pr.hclust.clusters	B	M
1	0	45
2	2	77
3	26	66
4	329	24

Q15. How well does the newly created model with four clusters separate out the two diagnoses?

Answer: The new model doesn't do the best job of separating the two diagnoses with four clusters as they are very skewed.

Q16. How well do the k-means and hierarchical clustering models you created in previous sections (i.e. before PCA) do in terms of separating the diagnoses? Again,

use the `table()` function to compare the output of each model (`wisc.km$cluster` and `wisc.hclust.clusters`) with the vector containing the actual diagnoses.

Answer: In this case the k-means method provides a better separation of the data because the data is more evenly distributed between the clusters.

```
table(wisc.km$cluster, diagnosis)
```

```
      diagnosis
      B      M
1 356    82
2   1   130
```

```
table(wisc.hclust.clusters, diagnosis)
```

```
      diagnosis
wisc.hclust.clusters  B      M
1 357   210
2   0     2
```

##Sensitivity

Sensitivity: $TP/(TP+FN)$ Specificity: $TN/(TN+FN)$

K-means: Sensitivity: $175/(175+37)=0.825$ Specificity: $343/(343+357)=0.961$

Hierarchical: Sensitivity: $172/(172+40)=0.811$ Specificity: $343/(343+14)=0.961$

Q17. Which of your analysis procedures resulted in a clustering model with the best specificity? How about sensitivity?

Answer: The K-means has slightly higher sensitivity but both have equal specificity.

7. Prediction

We can use our PCA model for prediction with new input patient samples.

```
url <- "https://tinyurl.com/new-samples-CSV"
new <- read.csv(url)
npc <- predict(wisc.pr, newdata=new)
npc
```


	PC1	PC2	PC3	PC4	PC5	PC6	PC7
[1,]	2.576616	-3.135913	1.3990492	-0.7631950	2.781648	-0.8150185	-0.3959098
[2,]	-4.754928	-3.009033	-0.1660946	-0.6052952	-1.140698	-1.2189945	0.8193031
	PC8	PC9	PC10	PC11	PC12	PC13	PC14
[1,]	-0.2307350	0.1029569	-0.9272861	0.3411457	0.375921	0.1610764	1.187882
[2,]	-0.3307423	0.5281896	-0.4855301	0.7173233	-1.185917	0.5893856	0.303029
	PC15	PC16	PC17	PC18	PC19	PC20	
[1,]	0.3216974	-0.1743616	-0.07875393	-0.11207028	-0.08802955	-0.2495216	
[2,]	0.1299153	0.1448061	-0.40509706	0.06565549	0.25591230	-0.4289500	
	PC21	PC22	PC23	PC24	PC25	PC26	
[1,]	0.1228233	0.09358453	0.08347651	0.1223396	0.02124121	0.078884581	
[2,]	-0.1224776	0.01732146	0.06316631	-0.2338618	-0.20755948	-0.009833238	
	PC27	PC28	PC29	PC30			
[1,]	0.220199544	-0.02946023	-0.015620933	0.005269029			
[2,]	-0.001134152	0.09638361	0.002795349	-0.019015820			

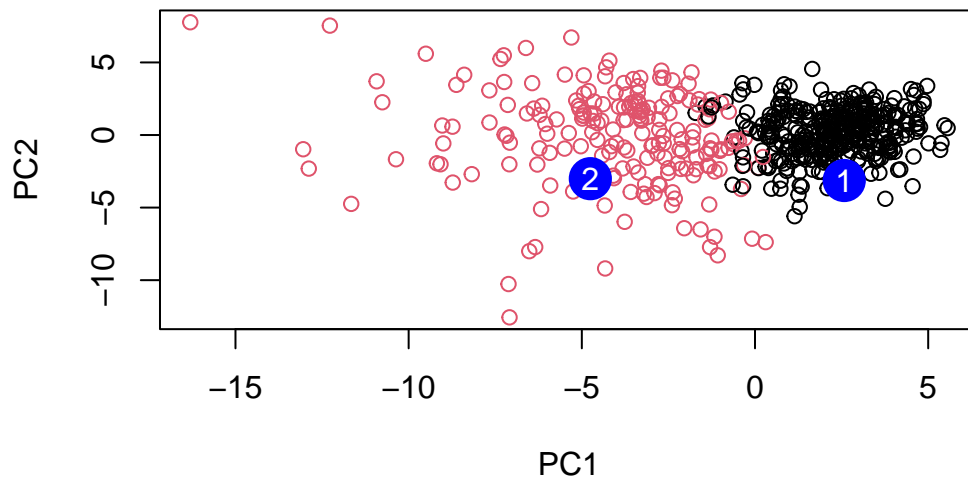
```
g <- as.factor(grps)
levels(g)
```

```
[1] "1" "2"
```

```
g <- relevel(g,2)
levels(g)
```

```
[1] "2" "1"
```

```
plot(wisc.pr$x[,1:2], col=g)
points(npc[,1], npc[,2], col="blue", pch=16, cex=3)
text(npc[,1], npc[,2], c(1,2), col="white")
```



Q18. Which of these new patients should we prioritize for follow up based on your results?

Answer: Patient 2 should be followed up with because malignant tumors have been seen on the lower end of PC1.

```
sessionInfo()
```

```
R version 4.5.1 (2025-06-13)
Platform: aarch64-apple-darwin20
Running under: macOS Tahoe 26.0.1
```

```
Matrix products: default
```

```
BLAS:   /Library/Frameworks/R.framework/Versions/4.5-arm64/Resources/lib/libRblas.0.dylib
LAPACK: /Library/Frameworks/R.framework/Versions/4.5-arm64/Resources/lib/libRlapack.dylib;
```

```
locale:
```

```
[1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
```

```
time zone: America/Los_Angeles
```

```
tzcode source: internal
```

```
attached base packages:
```

```
[1] stats      graphics  grDevices  utils      datasets  methods   base
```

other attached packages:

```
[1] ggplot2_4.0.0
```

loaded via a namespace (and not attached):

```
[1] digest_0.6.37      labeling_0.4.3      RColorBrewer_1.1-3  R6_2.6.1
[5] fastmap_1.2.0      xfun_0.53           farver_2.1.2        gtable_0.3.6
[9] glue_1.8.0         knitr_1.50          htmltools_0.5.8.1   rmarkdown_2.30
[13] lifecycle_1.0.4    cli_3.6.5           S7_0.2.0            scales_1.4.0
[17] vctrs_0.6.5        grid_4.5.1          withr_3.0.2         compiler_4.5.1
[21] tools_4.5.1        evaluate_1.0.5      yaml_2.3.10         rlang_1.1.6
[25] jsonlite_2.0.0
```