REVIEW ARTICLE

DISORDERS OF FLUIDS AND ELECTROLYTES

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Integration of Acid–Base and Electrolyte Disorders

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that incorporates insights from the traditional, bicarbonate-centered model and the Stewart (or strong ion) model (Table 1).¹-6 Acid-base balance and electrolyte homeostasis are intricately connected at the cellular level and in clinical disorders. This article emphasizes the integration of the principles of mass balance and electroneutrality — which are prominently featured in the strong ion model (also known as the physicochemical model) — for interpretation of acid-base phenomena. Most acid-base abnormalities can be diagnosed and interpreted with the use of the traditional approach. Why, then, should the strong ion theory be incorporated into teaching about acid-base balance? Although the Stewart model is not primarily a mathematical expression of a confirmed reality, it is relevant because it is a powerful construct that can shed light on an important biologic system.

Included in this article are several case vignettes that show the explanatory power of the strong ion approach in clinical practice. Some of these examples have been presented in a companion article on the physiological approach to acid–base balance by Berend et al.⁷ Other cases that are interpreted with a strong ion approach are included in the Supplementary Appendix, available with the full text of this article at NEJM.org. The more complex chemistry of the hydrogen-ion concentration in intracellular and extracellular fluid compartments is beyond the scope of this article.

BICARBONATE-CENTERED AND STRONG ION APPROACHES

The traditional model uses easily measured concentrations of blood carbon dioxide $[CO_2]$ and bicarbonate $[HCO_3^-]$.⁶ It is the basis of the Henderson–Hasselbalch equation:

pH=pK+log₁₀
$$\left(\frac{[HCO_3^-]}{0.03 (PacO_2)}\right)$$
, (1)

where pK is the acid dissociation constant, PacO₂ the partial pressure of arterial carbon dioxide, and 0.03 the solubility of CO₂ in blood.

The overall equilibrium between carbon dioxide and bicarbonate is shown below:

$$CO_2+H_2O\longleftrightarrow H_2CO_3\longleftrightarrow H^++HCO_3^-,$$
 (2)

where H₂CO₃ denotes carbonic acid, and H⁺ hydrogen.

In a teaching model, this relationship shows how alterations in the partial pressure of carbon dioxide (PCO₂) or levels of hydrogen or bicarbonate affect the other variables through mass balance. The fact that the hydrogen ion concentra-

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Traditional Approach Based on Bicarbonate-Carbon Dioxide	Physicochemical (Stewart) Approach†
CO_2/HCO_3^- equilibrium: $CO_2+H_2O \longleftrightarrow H_2CO_3 \longleftrightarrow H^+ + HCO_3^-$ and for $[CO_3^{-2}]$ $2[HCO_3^-] \longleftrightarrow [CO_3^{-2}] + [H_2O]$	CO_2/HCO_3^- equilibrium: $CO_2 + H_2O \longleftrightarrow H_2CO_3 \longleftrightarrow H^+ + HCO_3^-$ and for $[CO_3^{-2}]$ $2[HCO_3^-] \longleftrightarrow [CO_3^{-2}] + [H_2O] + [CO_2]$
Henderson–Hasselbalch equation: $pH = pK + log_{10} \left(\frac{[HCO_3^-]}{0.03 (PacO_2)} \right)$	Water dissociation: $K_w = \frac{[H^+][OH^-]}{H_2O}$ Weak acid (HA) dissociation: $[H^+][A^-] = K_A$ [HA]; weak acid conservation: $[A_{tot}] = [HA] + [A^-]$
Anion gap= $[Na^+]$ - $([Cl^-]+[HCO_3^-])$ mmol per liter	Strong ion difference (mmol per liter) = $[Na^+] + [K^+] + [Ca^{2+}] + [Mg^{2+}] - [Cl^-]$ [strong ion difference] - $[A^-] = [HCO_3^-] + [CO_3^{-2}] + [(OH^-)] - [H^+]$; or, strong ion difference - $[A^-] \approx [HCO_3^-]$
$Delta/delta = \frac{(anion gap - 12)}{(25 - [HCO_3])}$	
Example of expected compensation: $Pco_2 = 1.5 \ [HCO_3^-] + 8 \pm 2$ The Winters formula for respiratory compensation of metabolic acidosis, in which Pco_2 is dependent on the decrease in bicarbonate	Example of expected compensation: $Pco_2 = 1.5 \ [HCO_3^-] + 8 \pm 2$ The Winters formula for respiratory compensation of metabolic acidosis, in which Pco_2 is dependent on the decrease in bicarbonate

^{*} The traditional approach is based on bicarbonate-carbon dioxide. The physicochemical (Stewart) approach is dependent on the strong ion difference and Atot, the total content of albumin, phosphate, and circulating nonvolatile weak acids and their dissociated anions. H2CO3 denotes carbonic acid, and PaCO₂ partial pressure of arterial carbon dioxide.

tion is more than a million times lower than the bicarbonate level indicates that other forces are at work in the regulation of pH.

As in any chemical reaction in equilibrium, a change in the concentration of the reactant or product will move the reaction in the direction that would reestablish equilibrium (Le Châtelier's principle). If this principle is applied to equation 2, metabolic acidosis may be attributed to either the addition of hydrogen, with the consumption of bicarbonate as the reaction shifts to the left, or the removal of bicarbonate from the body, resulting in increased hydrogen as the reaction shifts to the right (from carbon dioxide) to replace lost bicarbonate. The observed relation between arterial PCO2 (PaCO2) and bicarbonate effectively predicts the direction but not the magnitude or time course of respiratory and renal compensations. Only empirical observations can determine the appropriate degree of compensation.

all unmeasured charged species (predominantly albumin) in plasma, is calculated below as

anion gap=
$$[Na^{+}]-([Cl^{-}]+[HCO_{3}^{-}]).$$
 (3)

The anion gap is used in the differential diagnosis of metabolic acidosis.8-10 It can suggest a cause for unmeasured anions. Possible causes include lactic acidosis, ketoacidosis, and uremic acidosis; ingestion of salicylate, methanol, ethylene glycol, or propylene glycol; and many inborn errors of metabolism. It is assumed that the unmeasured anion is added as the protonated acid (such as lactic acid). However, a severe form of metabolic acidosis results from treatment with sodium thiosulfate, a compound that has no hydrogen.11

In contrast to the anion gap shown in equation 3 above, the physicochemical model emphasizes that all cation and anion concentrations must balance, according to the laws of electro-The anion gap, consisting of the sum total of neutrality.³ The independent balance of each ion,

 $[\]dagger$ Relationships featuring electroneutrality are emphasized in the Stewart model. The equation for strong ion difference – [A $\bar{}$] can be reduced to approximately $[HCO_3^-]$ because $[HCO_3^-]$ is much greater than $[H^+]$, $[OH^-]$, and $[CO_3^{-2}]$. Stewart's equation (not shown), based on the relationships above, has been mathematically reduced to the Henderson-Hasselbalch equation (bicarbonate/CO2 method).

when disrupted, provides a mechanism for the acid-base condition. In their classic article, Peters and Van Slyke defined acid-base balance in the blood as the chemical state resulting from the balance between cations and anions. ¹² Carrying this idea to the extreme, one could view metabolic acid-base disorders as the predicted consequences of primary fluid and electrolyte imbalance.

Strong ions such as sodium and chloride are assumed to be completely dissociated in body water but can be lost or gained disproportionately. When the sum of all negatively charged ions (predominantly chloride) is subtracted from the sum of all positively charged strong ions, a value known as the strong ion difference (in millimoles per liter) is introduced. The strong ion difference is calculated as shown below:

strong ion difference= (4)
$$[Na^+]+[K^+]+[Ca^{2+}]+[Mg^{2+}]-[Cl^-],$$

where Ca²⁺ denotes calcium, and Mg²⁺ magnesium.

As shown in equation 5 below, the total content of albumin, phosphate, and circulating nonvolatile weak acids [HA] and their dissociated anions [A-] is referred to as [A_{tot}] in the Stewart model:

$$[A_{tot}] = [HA] + [A^{-}].$$
 (5)

As shown in equation 6 below, in which CO_3^{-2} denotes carbonate and OH⁻ hydroxide, an expression for remaining charged species, considered to be the dependent variables, is

[strong ion difference]–[A⁻]= (6)
[
$$HCO_3^-$$
]+[CO_3^-]+[(OH^-)]–[H^+],

in which the levels of carbonate, hydroxide, and hydrogen are much lower than the levels of bicarbonate. Any developed difference in the ionic charge, or strong ion difference, determines the bicarbonate concentration. Essential to this argument is that any difference in an unbalanced charge will immediately result in the appearance or disappearance of bicarbonate formed from ubiquitous and neutral carbon dioxide and water. It is also expected that the changes in the bicarbonate concentration will begin to occur at

a very minimal strong ion difference, since large charge separations are not possible. We can assume that electrostatic forces come into play until changes in bicarbonate concentrations match the charge separations among other ionic species. Clearly, electroneutrality in the macroenvironment always exists.

One drawback of using equation 6 in a clinical calculation of the hydrogen concentration is that the error in measurements of electrolytes in the millimolar range cannot allow for an accurate determination of the hydrogen level in nanomolar concentrations.

As shown in Table 1, the Stewart (or physicochemical) model of acid-base balance is quantitatively based on the view that the hydrogen and bicarbonate concentrations are not independently determined. Instead, they are dependent on the following: carbon dioxide (Paco₂) and its spontaneous relationship with hydrogen and bicarbonate, the dissociation of water (the abundant source of hydrogen within body fluids), the dissolved strong ions, the strong ion difference, and A_{tot}, which is the sum of all buffer pairs (mostly weak acids) that move toward equilibrium with a dissociated anion [A-] according to the dissociation constant for each (e.g., albumin with its net negative charge under physiological conditions).13

In keeping with the laws of electroneutrality,
all charged species must balance. This requires
that any change in the concentration of one of
the charged variables (the strong ion difference)
an must be matched by a change in the concentration of another charged species. According to
constraints in this internal system, the hydrogen
and bicarbonate concentrations are dependent
on the other variables, the total mass of which
is conserved.

The simultaneous mathematical solution of these reactions is complex and is not required to diagnose acid-base disorders. Furthermore, both experimental and clinical observations can be explained with the use of either model. Yet the physicochemical model is useful in revealing individual processes in the development of an acid-base disturbance because it associates the abnormality with specific electrolyte disturbances. The traditional model uses the calculated, and useful, anion gap to elucidate the pathophysiology of metabolic disorders. The usual calculation for the

Table 2. Acid-Base Disorders and Their Causes According to the Relationship between Gains and Losses of Circulating Cations or Anions.*

Metabolic alkalosis

Decrease (loss) of anion

Hypochloremic

Gastrointestinal

Vomiting

Chloridorrhea (villous adenoma, some chloride secretory diarrheas)

Renal

Chloruretic agents (loop diuretics, thiazides)

Chloride channelopathies (e.g., the Bartter syndrome, the Gitelman syndrome)

Hypokalemia leading to loss of chloride

Sweat

Cystic fibrosis

Hypoalbuminemic state¹³: malnutrition

Increase (gain) of cation

Sodium citrate, sodium lactate, sodium bicarbonate, sodium acetate

Hypernatremic

Hyperaldosteronism

Hypercalcemic

Milk alkali syndrome, calcium carbonate

Metabolic acidosis

Increase (gain) of anion

Hyperchloremic (potassium chloride, calcium chloride, hydrogen chloride, sodium chloride, arginine hydrochloride, lysine hydrochloride, ammonium chloride)

Anion-gap acidosis

Lactic acidosis

Diabetic ketoacidosis

Other unmeasured anions

Thiosulfate

Hyperphosphatemic

Decrease (loss) of cation (sodium and potassium)

Renal

Renal tubular acidosis

Natriuretic agents (e.g., amiloride, triamterene)

Sodium with anions in urine: ketoacids, D-lactate, hippurate

Hypoaldosteronism

Gastrointestinal

Diarrhea with bicarbonate or bacterial organic anions in stool

Vomiting pancreatic secretions

anion gap is shown in equation 3. For this equality to hold true, for electroneutrality purposes, the anion gap must be the net value for a complex mixture of all ionic species not included in the calculation, such as albumin, other proteins, calcium, magnesium, potassium, and phosphate, plus any additional anions such as lactate or acetoacetate. To illustrate the usefulness of a more inclusive approach in separating out various components of the anion gap, an anion gap hypothetically could be calculated as simply [Na⁺]–[HCO₃⁻]. From a charge point of view, it works out, but obviously, hyperchloremic acidosis could not be distinguished from an "anion gap" acidosis.

The anion-gap equation could be rearranged to solve for the bicarbonate concentration instead of unmeasured anions:

$$[Na^{+}]-([Cl^{-}]+[AG])=[HCO_{3}^{-}],$$

where AG denotes the anion gap.

This is analogous to the strong ion difference. The anion gap, which is usually calculated with the use of plasma bicarbonate, is useful clinically. If every charged species were known and measured, the equation could be rearranged to calculate the bicarbonate concentration, but often the unmeasured ion is unknown. How, then, does the strong ion difference increase or decrease? The answer lies in specific gains or losses of electrolytes such as sodium and chloride in a different proportion to each other than the proportion in the normal extracellular fluid. The first step in understanding how an acidbase disorder develops is to know or assume the specific electrolyte content of any gained fluids (e.g., intravenous fluids) or lost fluids (e.g., gastrointestinal fluids, sweat, or urinary fluids).

Since the normal concentration ratio of sodium to chloride in extracellular fluid is approximately 140:100, an increase in the sodium level, a decrease in the chloride level, or both will increase the strong ion difference and the bicarbonate concentration will increase (metabolic alkalosis), according to electroneutrality requirements.³ When the strong ion difference decreases, pH and the bicarbonate level will decrease (metabolic acidosis). The electroneutrality relationship in equation 6 can be useful in diagnosing the causes of metabolic alkalosis and metabolic acidosis shown in Table 2.

^{*} All metabolic acid-base disorders can be viewed in the context of the relative losses or gains of cations or anions in body fluids. Hypophosphatemia is not listed because the plasma phosphate level is normally low.

METABOLIC DISTURBANCES AND STRONG IONS

Acid—base balance is dependent on strong ions in the macroscopic sense because the same cellular mechanisms regulate acid—base homeostasis and electrolyte homeostasis. The following case vignette illustrates this point:

A 31-year-old woman with gastroenteritis had been vomiting for 2 days. She was weak and hypotensive. Laboratory tests revealed a sodium concentration of 125 mmol per liter, potassium 2.6 mmol per liter, chloride 72 mmol per liter, and bicarbonate 40 mmol per liter. The arterial pH was 7.54, the Paco₂ 48 mm Hg, and the urinary pH 5.0.

Depletion of the extracellular fluid from vomiting creates profound needs to conserve sodium and water and preserve potassium balance; these mechanisms are clearly obstacles to maintaining a normal blood pH and bicarbonate concentration. With volume and potassium depletion, metabolic alkalosis is maintained, not corrected, by the kidneys. 14,15 Low extracellular fluid volume and low blood pressure increase angiotensin II and aldosterone levels. Increased sodium reabsorption through proximal tubular sodiumhydrogen exchange and the collecting-duct sodium channel, accompanied by hydrogen secretion by the hydrogen ATPase and the potassiumhydrogen ATPase, in turn increases bicarbonate reabsorption until the urinary pH decreases as it becomes free of bicarbonate. This paradoxical aciduria in the midst of alkalemia is evidence that blood pH depends on strong ion balance. The alkalemia will be corrected only with sufficient replacement of sodium, chloride, and potassium. In this patient, after the administration of 0.9% normal saline with potassium chloride, the electrolyte status improved, and the urinary pH increased to 8.0 with the prompt excretion of sodium, potassium, and bicarbonate. In this clinical situation, the interdependency of acidbase and electrolyte balance is self-evident. If this were not the case, and kidney function instead focused on maintaining a normal acid-base balance, the bicarbonate generated by vomiting would result in the urinary loss of even larger quantities of sodium, potassium, and water, leading to life-threatening volume and potassium depletion.

The traditional acid–base approach tacitly overlaps with aspects of the strong ion theory (Table 1). Consider the familiar concept known as the "delta-delta" (Δ - Δ), the increase (Δ) in the anion gap versus the decrease (Δ) in the bicarbonate level.^{8,9}

All metabolic acid–base disorders are associated with either a change in the concentration of sodium, potassium, calcium, chloride, hydrogen phosphate, or albumin or a change in the anion gap. The normal anion gap can be adjusted for hypoalbuminemia by allowing for 2.5 mmol per liter of negative charge for each 1 g per deciliter of albumin concentration. The relative change in the bicarbonate level and the anion gap $(\Delta-\Delta)$ is only part of the electroneutrality requirement. The net sum of all cation and anion electrolyte charge gaps must cancel out. In the search for a " $\Delta-\Delta-\Delta-\Delta-\Delta-\Delta$," clues about any acid–base disorder will emerge.

A finding of an increase in the anion gap above the normal concentration (the Δ anion gap) that exceeds the decrease in the bicarbonate concentration (Δ bicarbonate) may indicate mixed metabolic acidosis and metabolic alkalosis. The following case shows that a ratio other than 1:1 is not pathognomonic for a mixed acid–base disturbance:

Before a cardiac arrest, a 67-year-old man with an acute myocardial infarction had normal levels of serum electrolytes (level of sodium 140 mmol per liter, potassium 4.0 mmol per liter, chloride 103 mmol per liter, and bicarbonate 25 mmol per liter). While he was anuric after the cardiac arrest, his laboratory tests showed a sodium level of 140 mmol per liter, potassium 5.0 mmol per liter, chloride 62 mmol per liter, and bicarbonate 5 mmol per liter. The arterial pH was 7.10, and the PacO₂ was 16 mm Hg. The lactate level was 60 mmol per liter, and the anion gap was 73 mmol per liter.

This patient had severe anion-gap metabolic acidosis due to lactate overproduction from tissue hypoperfusion. Since lactate production can result in blood lactate concentrations greater than a normal bicarbonate concentration, what

happens when the bicarbonate concentration decreases almost to zero? Hydrogen might be preferentially reactive with other tissue buffer systems, but a decrease in the chloride level is often observed, yielding a hypochloremic anion-gap acidosis. ^{16,17} With lactate acting as a strong ion, the strong ion difference [Na+]—[Cl-]—[lactate-] is decreased and the bicarbonate concentration decreases to achieve electroneutrality. Chloride moves into cells, probably in exchange for lactate or bicarbonate. In this case, there was no clinical evidence of superimposed metabolic alkalosis:

$\Delta [HCO_3^-] + \Delta [Cl^-] = \Delta [AG] = [lactate].$

Also, consider examples of metabolic alkalosis in which the increase in a strong cation is balanced by an increase in the bicarbonate concentration, as in the following case, which was also described by Berend et al.⁷:

A 50-year-old woman with a recent onset of hypertension had the following laboratory results: sodium level 150 mmol per liter, potassium 2.2 mmol per liter, chloride 103 mmol per liter, and bicarbonate 32 mmol per liter. The arterial pH was 7.50, and the Paco₂ was 43 mm Hg. She was found to have an aldosterone-secreting adrenal adenoma.

In this case, the change in the bicarbonate level was associated with an increased sodium concentration, which is often seen in primary hyperaldosteronism and is attributable to increased function of epithelial sodium channels in renal cortical collecting-duct principal cells. Mild hypernatremia probably occurred as a result of extracellular fluid expansion that decreased vasopressin release, with a consequent decrease in renal reabsorption of water. The plasma chloride level was not increased in proportion to the sodium level, which is consistent with less chloride than sodium retention in primary hyperaldosteronism.18 Loss of chloride, a feature of "aldosterone escape from edema," is linked to decreased sodium-chloride cotransport in the distal renal tubule.¹⁹ Hypokalemia in turn is associated with increased loss of urinary chloride. Thus,

 $\Delta [HCO_3^-] = \Delta [Na^+] + \Delta [K^+] - \Delta [Cl^-],$

with changes in each of these strong ions contributing to the alkalosis.

Treatment of the alkalosis in this patient with hyperaldosteronism will require replacement with potassium chloride. Administering chloride in the form of saline would worsen the hypokalemia, and administering potassium without chloride would not correct it; thus, the term "saline unresponsive" is more accurate than "chloride unresponsive" as a description of this type of alkalosis.

Hypercalcemia will increase the strong ion difference and is associated with metabolic alkalosis. The milk alkali syndrome, which is often caused by excessive ingestion of calcium-containing antacids, is characterized by alkalosis and hypercalcemia. In contrast, hypercalcemia in primary hyperparathyroidism is associated with a proximal renal tubular metabolic acidosis rather than metabolic alkalosis. This observation may be explained by the decrease in the strong ion difference due to losses of urinary sodium resulting from inhibition of proximal tubular sodium—hydrogen exchange by parathyroid hormone.

GASTROINTESTINAL LOSSES OF STRONG IONS

Losses of ions due to diarrhea are associated with the development of metabolic acidosis²² or metabolic alkalosis. Since depletion of extracellular volume can occur in cases of acidosis or alkalosis and may be initiated by losses of sodium and chloride in any ratio, the term "contraction alkalosis" is a misnomer. As shown in the following case, the relative content of the strong ions lost (sodium and potassium vs. chloride),²³ not the site of the loss, determines the acid–base disorder:

A 40-year-old woman who underwent a colonic resection for ulcerative colitis had excessive liquid drainage from an ileostomy. Her laboratory results revealed a plasma sodium level of 138 mmol per liter, potassium 5.0 mmol per liter, chloride 110 mmol per liter, and bicarbonate 15 mmol per liter. The arterial pH was 7.30, and the PaCO₂ was 32 mm Hg.

In this case, the loss of watery small-intestinal and pancreatic secretions, which have high sodium and bicarbonate levels and very low chloride levels, would result in the relative retention of more chloride than sodium in the extracellular fluid, causing hyperchloremic acidosis. In cases of colonic diarrhea, hyperchloremic acidosis may develop because of loss of sodium and potassium with organic anions of bacterial origin, such as acetate, rather than bicarbonate per se.²³

When diarrhea is the cause of metabolic alkalosis, rather than acidosis, the mechanism is determined by measuring the electrolyte content in stool. Large losses of chloride may occur in patients who have villous adenomas or other secretory diarrheas that cause depletion of chloride, as shown in the following case, described by Berend et al.⁷:

Large volumes of watery diarrhea from infectious gastroenteritis developed in a 22-year-old man. Laboratory tests revealed a plasma sodium concentration of 140 mmol per liter, potassium 3.0 mmol per liter, chloride 86 mmol per liter, and bicarbonate 38 mmol per liter. The arterial pH was 7.60, and the Paco₂ was 40 mm Hg.

In this case, the electrolyte concentrations in liquid stool, if measured, would probably show a charge gap, in which (Na⁺+K⁺)-Cl⁻ would be less than the normal plasma bicarbonate concentration. High losses of chloride in stool, like losses of chloride from vomiting or after the use of loop diuretics, cause hypochloremic alkalosis.

URINARY CHARGE GAP AND STRONG IONS

Measurements of urinary electrolyte concentrations and flow rate indicate renal acid-base function even without measurement of urinary bicarbonate. As shown in equation 7, the urinary net charge compares the loss of measured strong cations (sodium and potassium) with the loss of chloride²⁴:

Urinary net charge gap= $[U_{Na}^{+}]+[U_{K}^{+}]-[U_{Cl}^{-}]$. (7)

NEGATIVE URINARY CHARGE GAP

A negative value for the urinary net charge gap indicates the presence of the unmeasured cation, ammonium (excretion of ammonium chloride). The loss of ammonium chloride in the urine of a patient with metabolic acidosis is an appropriate

compensation, since the very process of excreting acid in this way has an alkalinizing effect on body fluids. The loss of net acid in the form of ammonium chloride is a normal renal response to nonrenal causes of metabolic acidosis, such as severe watery diarrhea. The losses of urinary chloride result in an increased plasma strong ion difference, which in turn permits the formation of more bicarbonate.

However, in a patient with metabolic alkalosis, a relative excess of chloride in the urine strongly suggests that the losses of urinary chloride cause the metabolic alkalosis by increasing the plasma strong ion difference. In the following case, such losses of urinary chloride led to hypochloremic alkalosis:

An 80-year-old man with congestive heart failure received furosemide until all peripheral edema disappeared. Laboratory tests revealed a sodium level of 130 mmol per liter, potassium 2.5 mmol per liter, chloride 80 mmol per liter, and bicarbonate 40 mmol per liter. The arterial pH was 7.50, and the Paco₂ was 53 mm Hg.

In this patient, the sodium-potassium-chloride cotransporter was inhibited by furosemide (in the thick ascending limb of the loop of Henle). Under these circumstances, the stoichiometric balance of sodium, potassium, and chloride was 1:1:2, and proportionately more chloride than sodium was lost in the urine. Thus, there is a direct explanation for the hypochloremic metabolic alkalosis in this patient. Inhibition of the sodium-chloride cotransporter of the distal tubule by thiazides (stoichiometric balance between sodium and chloride, 1:1) is also predictive of metabolic alkalosis because of the greater loss of chloride than sodium from the extracellular fluid. In addition to chloride-wasting diuretics, many hereditary disorders of sodium and chloride transport by the renal tubules (socalled channelopathies) may cause acid-base disorders, as shown in Table 2.

POSITIVE URINARY CHARGE GAP

A positive value for the urinary net charge gap indicates excretion of an unmeasured anion. The lost, unmeasured anion may be bicarbonate or nonbicarbonate anions such as ketones, lactate, L-lactate, D-lactate, and hippurate in persons who sniff glue. Such loss of anions will decrease

the plasma strong ion difference and acidify the extracellular fluid as the process returns chloride to the circulation. ^{10,25} If the urinary clearance of these nonchloride anions is high enough that they do not accumulate as a plasma anion gap, then the hyperchloremia may be mistaken for renal tubular acidosis. ²⁴ Without those nonbicarbonate anions, metabolic acidosis with the loss of urinary sodium and potassium and retention of chloride (the positive-charge gap) will result in a decreased plasma strong ion difference, constituting a renal cause of acidosis (e.g., carbonic anhydrase inhibition or renal tubular acidosis).

If metabolic alkalosis is present, a positive urinary gap suggests that the renal loss of strong cations (sodium and potassium) and conservation of chloride will acidify the extracellular fluid because of a decrease in the plasma strong ion difference and in the bicarbonate concentration.

The excretion patterns of urinary electrolytes reflect the ability of the kidney to counteract nonrenal acid-base disorders. The capacity of the kidney to excrete ammonium chloride in acidosis allows for elimination of anions with conservation of sodium for volume and potassium for potassium balance. This potassium-sparing effect of urinary ammonium is evident in hypokalemic stimulation of ammoniagenesis.

The traditional physiological approach interprets the urine electrolytes to deduce the presence of ammonium and bicarbonate with less emphasis on the strong ion pathogenesis of acid-base disorders. The physicochemical model emphasizes the relative losses of the actual measured quantities to determine the cause of the disturbance. Both perspectives are enlightening.

COMPENSATION FOR RESPIRATORY DISORDERS AND URINARY STRONG IONS

The ratio of bicarbonate to Paco₂ in the Henderson–Hasselbalch equation (equation 1) is a simple way to illustrate the initial disturbance and then the modulating effect of the compensatory response on pH. In respiratory conditions, the Paco₂ is the initial abnormality leading to sharp, sudden changes in pH, with little change in strong ion concentrations. Over time, however, the change in the level of chloride and reciprocal changes in the level of bicarbonate are the major factors that allow pH to return toward normal

Figure 1 (facing page). Renal Tubular Cells with Transporters That Are Targets of Hormones, Diuretics, and Mutations Affecting Acid–Base Balance.

Similar transporters in the gastrointestinal tract that are associated with disease are not shown. All cell transporters on the blood side interface with interstitial fluid (not shown) before transport into blood. AE1 denotes anion exchanger 1, ENaC epithelial sodium channel, NBC sodium bicarbonate cotransporter, NCC sodium chloride cotransporter, NHE3 sodium—hydrogen exchange, and NKCC sodium—potassium 2-chloride cotransporter.

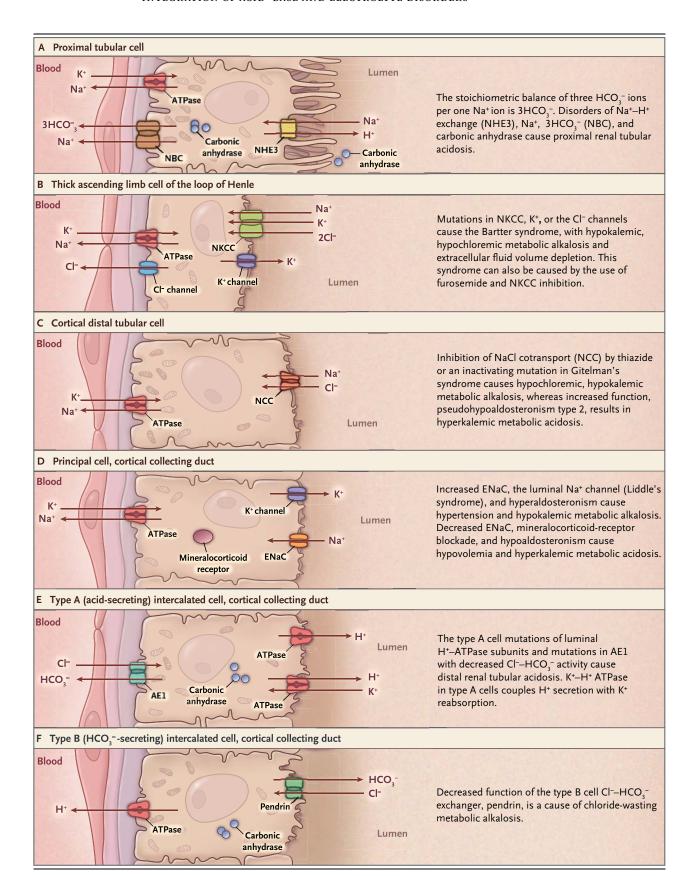
values. The hyperchloremic renal compensation for respiratory alkalosis is the excretion of filtered sodium and potassium with bicarbonate, because low Paco₂ decreases proximal and distal hydrogen secretion. As the plasma strong ion difference decreases, the plasma bicarbonate concentration will decrease.

In respiratory acidosis, high Paco₂ increases production of ammonia by the kidney, and the excretion of ammonium chloride with a negative urinary net charge, shown in equation 7, results in hypochloremia, an increased plasma strong ion difference, and an elevated plasma bicarbonate concentration. The elevated Paco₂ increases the renal reabsorption of sodium and bicarbonate, so the compensation is maintained. If the Paco₂ is abruptly lowered by means of a ventilator, the compensatory response transitions to posthypercapnic hypochloremic metabolic alkalosis, which will not resolve until the chloride that is lost as ammonium chloride is replenished.

INTRAVENOUS FLUIDS AND CONTENT OF STRONG IONS

The gain of fluids containing strong ions in ratios dissimilar to those in the extracellular fluid also affects acid–base balance, as shown in the following case, which was also described by Berend et al.⁷ in their article about the physiological approach:

A 22-year-old woman who had been injured in an accident received 6 liters of isotonic saline, after which the level of sodium was 135 mmol per liter, potassium 3.8 mmol per liter, chloride 115 mmol per liter, and bicarbonate 18 mmol per liter. The arterial pH was 7.28, and the Paco₂ was 39 mm Hg. The urinary sodium level was 65 mmol



per liter, potassium 15 mmol per liter, and chloride 110 mmol per liter.

This is an example of saline-induced acidosis, ²⁶ which develops because the infusion of a proportionately high sodium chloride-containing solution, one with a sodium-to-chloride ratio of less than 140:100, will decrease the plasma strong ion difference and the bicarbonate concentration. The insufficient urinary excretion of the extra chloride as ammonium chloride leads to metabolic acidosis. The infusion of saline with its 1:1 sodium-to-chloride ratio, resulting in hyperchloremic acidosis, is the converse of inhibition of the 1:1 sodium-to-chloride transport ratio in thiazide-induced diuresis and hypochloremic metabolic alkalosis.

Even Ringer's lactate, with a level of sodium of 130 mmol per liter, chloride 109 mmol per liter, and lactate 28 mmol per liter, can cause hyperchloremic acidosis because the ratio of sodium to chloride is smaller than the ratio of sodium to chloride in the normal extracellular fluid.^{26,27} Thus, what matters is the content and amount of infused fluids.

CONCLUSIONS

Clinical evidence can be interpreted with the use of both the strong ion theory and the traditional bicarbonate-centered approach to provide an optimal understanding of acid-base disorders. Electrolyte concentrations of plasma may be altered by the gains and losses associated with intravenous fluids and with urinary, intestinal, or sweat-gland secretions. An understanding of the consequences of these disturbances helps in the diagnosis and treatment of the associated acid-base disorders.

The evidence connecting acid-base balance with electrolyte balance is apparent at the cellular level (i.e., ion transporters, their stoichiometric balance, and the hormones that regulate them) (Fig. 1) and in clinical practice. The fact that transporters often couple a strong ion such as sodium or potassium with hydrogen, or chloride with bicarbonate, ²⁸⁻³⁰ suggests an ultimate coherence between the two approaches (Fig. 1). As more is learned about the molecular nature of disorders of epithelial-cell transport as well as about intracellular pH, it will become more important to understand interactions between carbon dioxide and bicarbonate with strong ions and cellular buffers in the body.³¹

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Disclosure forms provided by the author are available with the full text of this article at NEJM.org.

REFERENCES

- 1. Gamble JL Jr. Sodium and chloride and acid-base physiology. Bull Johns Hopkins Hosp 1960:107:247-54.
- 2. Kurtz I, Kraut J, Ornekian V, Nguyen MK. Acid-base analysis: a critique of the Stewart and bicarbonate-centered approaches. Am J Physiol Renal Physiol 2008;294:F1009-F1031.
- **3.** Stewart PA. Modern quantitative acidbase chemistry. Can J Physiol Pharmacol 1983;61:1444-61.
- **4.** Constable PD. A simplified strong ion model for acid-base equilibria: application to horse plasma. J Appl Physiol (1985) 1997;83:297-311.
- **5.** Fencl V, Rossing TH. Acid-base disorders in critical care medicine. Annu Rev Med 1989;40:17-29.
- **6.** Adrogué HJ, Gennari FJ, Galla JH, Madias NE. Assessing acid-base disorders. Kidney Int 2009;76:1239-47.
- **7.** Berend K, de Vries APJ, Gans ROB. Physiological approach to assessment of acid-base disturbances. N Engl J Med 2014;371:1434-45.
- **8.** Emmett M, Narins RG. Clinical use of the anion gap. Medicine (Baltimore) 1977; 56:38-54

- 9. Goodkin DA, Krishna GG, Narins RG. The role of the anion gap in detecting and managing mixed metabolic acid-base disorders. Clin Endocrinol Metab 1984; 13:333-49.
- **10.** Oh MS, Carroll HJ. The anion gap. N Engl J Med 1977;297:814-7.
- 11. Selk N, Rodby RA. Unexpectedly severe metabolic acidosis associated with sodium thiosulfate therapy in a patient with calcific uremic arteriolopathy. Semin Dial 2011:24:85-8
- 12. Carbonic acid and acid-base balance. In: Peters JP, Van Slyke DD. Quantitative clinical chemistry: interpretations. Baltimore: Williams & Wilkins, 1939:868-1018.
- **13.** McAuliffe JJ, Lind LJ, Leith DE, Fencl V. Hypoproteinemic alkalosis. Am J Med 1986:81:86-90.
- **14.** Seldin DW, Rector FC Jr. Symposium on acid-basis homeostasis: the generation and maintenance of metabolic alkalosis. Kidney Int 1972;1:306-21.
- **15.** Luke RG, Galla JH. It is chloride depletion alkalosis, not contraction alkalosis. J Am Soc Nephrol 2012;23:204-7.
- 16. Case Records of the Massachusetts

- General Hospital (Case 18-1968). N Engl J Med 1968;278:1004-13.
- 17. Madias NE, Homer SM, Johns CA, Cohen JJ. Hypochloremia as a consequence of anion gap metabolic acidosis. J Lab Clin Med 1984;104:15-23.
- **18.** Relman AS, Schwartz WB. The effect of DOCA on electrolyte balance in normal man and its relation to sodium chloride intake. Yale J Biol Med 1952:24:540-58.
- **19.** Wang XY, Masilamani S, Nielsen J, et al. The renal thiazide-sensitive Na-Cl cotransporter as mediator of the aldosterone-escape phenomenon. J Clin Invest 2001;108:215-22.
- **20.** Medarov BI. Milk-alkali syndrome. Mayo Clin Proc 2009:84:261-7.
- **21.** Kahn AM, Dolson GM, Hise MK, Bennett SC, Weinman EJ. Parathyroid hormone and dibutyryl cAMP inhibit Na+/H+ exchange in renal brush border vesicles. Am J Physiol 1985;248:F212-F218.
- **22.** Gennari FJ, Weise WJ. Acid-base disturbances in gastrointestinal disease. Clin J Am Soc Nephrol 2008;3:1861-8.
- 23. Fordtran JS. Organic anions in fecal contents. N Engl J Med 1971;284:329-30.

- **24.** Batlle DC, Hizon M, Cohen E, Gutterman C, Gupta R. The use of the urinary anion gap in the diagnosis of hyperchloremic metabolic acidosis. N Engl J Med 1988;318:594-9.
- **25.** Carlisle EJ, Donnelly SM, Vasuvattakul S, Kamel KS, Tobe S, Halperin ML. Glue-sniffing and distal renal tubular acidosis: sticking to the facts. J Am Soc Nephrol 1991;1:1019-27.
- **26.** Waters JH, Gottlieb A, Schoenwald P, Popovich MJ, Sprung J, Nelson DR. Nor-
- mal saline versus lactated Ringer's solution for intraoperative fluid management in patients undergoing abdominal aortic aneurysm repair: an outcome study. Anesth Analg 2001;93:817-22.
- **27.** Omron EM, Omron RM. A physicochemical model of crystalloid infusion on acid-base status. J Intensive Care Med 2010;25:271-80.
- **28.** Gennari FJ. Pathophysiology of metabolic alkalosis: a new classification based on the centrality of stimulated collecting
- duct ion transport. Am J Kidney Dis 2011; 58:626-36.
- **29.** Koeppen BM. The kidney and acidbase regulation. Adv Physiol Educ 2009; 33:275-81.
- **30.** Chadha V, Alon US. Hereditary renal tubular disorders. Semin Nephrol 2009; 29:399-411.
- **31.** Corey HE. Stewart and beyond: new models of acid-base balance. Kidney Int 2003;64:777-87.

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