

Management of Patients with Sickle Cell Disease Using Transfusion Therapy Guidelines and Complications



Stella T. Chou, MD^{a,*}, Ross M. Fasano, MD^{b,*}

KEYWORDS

• Sickle cell disease • Red blood cell transfusion • Alloimmunization • Iron overload

KEY POINTS

- Urgent or emergent red blood cell transfusion is indicated for acute ischemic stroke, acute chest syndrome, splenic or hepatic sequestration, transient aplastic crisis, multisystem organ failure, intrahepatic cholestasis, or obstetric complications in patients with sickle cell disease (SCD).
- Chronic transfusion therapy is indicated for primary and secondary stroke prevention and short-term for prevention of splenic sequestration recurrence.
- Patients with SCD should receive red cells antigen matched for C, E, and K to reduce alloimmunization risk.
- The iron status of chronically transfused patients with SCD should be closely monitored and iron chelation therapy and/or erythrocytapheresis implemented to maintain iron balance.

INTRODUCTION

Over the past few decades, significant advances in the care of patients with sickle cell disease (SCD) have led to improvements in morbidity and survival. The average life span of patients with SCD has increased from 14 years in 1973 to more than 50 years.¹ A key component in the management of patients with SCD is red blood cell (RBC)

^a Department of Pediatrics, Abramson Research Center, The Children's Hospital of Philadelphia, Perelman School of Medicine, University of Pennsylvania, 316D, 3615 Civic Center Boulevard, Philadelphia, PA 19104, USA; ^b Transfusion, Tissue, & Apheresis, Children's Healthcare of Atlanta and Grady Health System Transfusion Services, Departments of Clinical Pathology and Pediatric Hematology, Emory University School of Medicine, 7105B Woodruff Memorial Building, 101 Woodruff Circle, Atlanta, GA 30322, USA

* Corresponding author. Department of Clinical Pathology, Emory University School of Medicine, 7105B Woodruff Memorial Building, 101 Woodruff Circle, Atlanta, GA 30322

E-mail addresses: chous@email.chop.edu; ross.fasano@emory.edu

transfusion therapy. The major goals of RBC transfusions are relief of anemia, reduction of circulating sickle hemoglobin (HbS) erythrocytes, and improvement in oxygen-carrying capacity.² Although transfusion can be lifesaving, it is not without adverse effects. Using evidence-based transfusion policies can minimize transfusion-related complications. This review addresses RBC transfusion methods, indications (Table 1), and complications.

METHODS OF TRANSFUSION THERAPY

RBC transfusions can be administered by simple or exchange transfusion. Exchange transfusion is preferably performed by automated erythrocytapheresis but can be performed manually. Simple transfusions are dosed in units (1–3 units for adults) or

Table 1 Indications for transfusion therapy in adults and children with sickle cell disease	
Transfusion Indication	Transfusion Method
Generally accepted indications for transfusion	
Acute ischemic stroke	Exchange transfusion preferred
Primary stroke prevention	Chronic simple or exchange transfusion ^a
Secondary stroke prevention	Chronic simple or exchange transfusion ^a
Acute chest syndrome (acute)	Simple or exchange transfusion ^a
Acute splenic sequestration	Simple transfusion
Acute splenic sequestration, recurrence	Chronic simple transfusion (before splenectomy) ^b
Preoperative (when general anesthesia required)	Simple transfusion
Transient aplastic crisis	Simple transfusion
Acute multisystem organ failure	Simple or exchange transfusion ^c
Acute hepatic sequestration	Simple or exchange transfusion ^c
Acute intrahepatic cholestasis	Simple or exchange transfusion ^c
Acute sickle or obstetric complications during pregnancy	Simple or exchange transfusion ^c
Controversial indications for transfusion	
Acute chest syndrome (recurrent)	Chronic simple or exchange transfusion ^c
Vasooclusive painful episode (recurrent)	Chronic simple or exchange transfusion ^c
Pulmonary hypertension	Chronic simple or exchange transfusion ^c
Transfusion generally not indicated	
Uncomplicated vasoocclusive painful episode	NA
Priapism	NA
Uncomplicated pregnancy	NA
Leg ulcers	NA
Nonsurgically managed avascular necrosis	NA

Abbreviation: NA, not applicable.

^a Exchange transfusion may be preferred in rapidly deteriorating patients when emergent HbS reduction is needed or when there are concerns for post-transfusion hyperviscosity due to a high pretransfusion hemoglobin (ie, >9 g/dL).

^b Chronic transfusion may be used to delay but not prevent the need for splenectomy in very young children (ie, <2 years) who are at increased risk for invasive pneumococcal infections.

^c Exchange transfusion may be preferred in patients with iron overload.

volume (10–20 mL/kg for children). Exchange transfusion requires a higher volume of RBCs administered ($1.0\text{--}1.5 \times$ patients' RBC volume) but simultaneously removes patients' RBCs. Erythrocytapheresis offers the advantage of rapidly reducing HbS independent of the hematocrit and minimizes iron accumulation. Exchange transfusion is often preferred for emergent HbS reduction in patients with higher pretransfusion hemoglobin (Hb) (>9 g/dL) because of hyperviscosity concerns and to prevent iron overload. The decision to use simple versus exchange transfusion depends on specific clinical needs and availability of resources, including apheresis equipment and technical support, adequate supply of antigen-negative donor units, and the potential need for central venous access.^{3–5} Partial manual exchange (PME) is an alternative method for patients with a higher Hb (>8.5 g/dL) and involves phlebotomy of 5 to 10 mL/kg (depending on patients' baseline Hb and tolerance) immediately before transfusion. PME has been used to slow progression of transfusional iron overload when used as a chronic transfusion regimen.⁶

INDICATIONS

Acute Splenic Sequestration

RBC transfusion is indicated for acute exacerbation of anemia occurring with splenic sequestration. Because splenic involution is usually complete by 5 years of age in hemoglobin SS and S β^0 thalassemia, acute splenic sequestration most commonly affects young children. The spleen becomes acutely engorged with sequestered blood and may result in a precipitous decrease in Hb level. Severe episodes may lead to hypovolemic shock and death from cardiovascular collapse within hours. Immediate RBC transfusion will correct the anemia and hypovolemia, but patients should be transfused cautiously to prevent hyperviscosity after splenic sequestration resolves. Aliquots of 5 mL/kg may be administered along with close monitoring of the spleen size, Hb level, and cardiovascular status. In cases of severe sequestration and anemia with hypovolemic shock, initial transfusion with 10 mL/kg packed RBCs (PRBCs) is appropriate. Relapse of acute splenic sequestration is frequent, with 50% to 75% of patients experiencing recurrent episodes.⁷ Chronic RBC transfusion to prevent recurrence has not been prospectively studied but is often used to delay definitive treatment of splenectomy in very young children. In a retrospective multicenter study of 190 children with hemoglobin-SS or S β^0 disease, 29% were managed with a blood transfusion program and, overall, 37% ultimately required splenectomy.⁷ However, 54% of patients were managed with close monitoring and without prophylactic blood transfusion or splenectomy, of which 59% did not experience a recurrent episode.

Transient Aplastic Crisis

Human parvovirus B19 infects erythrocyte precursors and temporarily suppresses erythropoiesis that can result in severe anemia given the shortened life span of RBCs in patients with SCD. In a single-institution observational study of parvovirus B19-induced red cell aplasia, the median nadir Hb was 4.8 g/dL and 49 of 68 pediatric patients (72%) received a transfusion.⁸ The need for transfusion depends on the severity of the anemia, whether they are in the reticulocytopenia stage, and the clinical status of patients. Because anemia associated with red cell aplasia is subacute, patients are typically euvolemic and physiologically compensated. RBC transfusion should be administered slowly with serial small aliquots to prevent congestive heart failure. Parvovirus aplastic crisis typically does not recur because of long-term humoral immunity.

Acute Chest Syndrome

Acute chest syndrome (ACS) describes a new pulmonary infiltrate and respiratory findings, including cough, dyspnea, or new-onset hypoxia, in patients with SCD and is often accompanied by fever. Triggers include infection, pulmonary fat embolism, hypoventilation/atelectasis, and bronchospasm. ACS is the leading cause of death and second most common cause of hospitalization among patients with SCD. The management is primarily supportive and includes respiratory therapy, antibiotics, and, often, RBC transfusion. There have been no randomized controlled trials comparing either simple or exchange transfusion versus no transfusion for ACS. However, in a large epidemiologic study of ACS, transfusion was associated with a shorter duration of hospitalization, suggesting an association with clinical improvement.⁹ A difference in efficacy of simple transfusion compared with exchange transfusion as measured by length of hospital stay was not detected in a small study of 20 patients with ACS.¹⁰ In practice, simple transfusion should be considered for any patient with ACS and hypoxemia or acute exacerbation of anemia. Exchange transfusion is typically reserved for patients who are not sufficiently anemic to accommodate a simple transfusion or those with progressive respiratory decline or persistent hypoxia despite oxygen supplementation or simple transfusion.

No prospective randomized trial has been performed to determine the efficacy of chronic transfusion therapy to prevent recurrent ACS. Chronic transfusion therapy is sometimes offered, particularly to individuals who experienced a severe or life-threatening episode. A dramatic reduction in hospitalization for ACS was observed in children undergoing chronic transfusion for primary stroke prevention compared with the observed group, suggesting chronic transfusions may prevent recurrent episodes.¹¹ In one single-institution study, chronic transfusion therapy reduced the incidence of ACS events among patients with recurrent ACS but did not significantly impact episode severity.¹² Although hydroxyurea is indicated for the prevention of recurrent ACS,¹³ future studies are needed to compare chronic transfusion therapy and hydroxyurea for recurrent ACS prevention.

Acute Sickle Hepatopathy

Acute hepatic sequestration and sickle cell intrahepatic cholestasis (SCIC) are severe forms of hepatic injury from vascular occlusion of liver sinusoids.¹⁴ Acute hepatic sequestration manifests as painful enlarging hepatomegaly with a concurrent decrease in hematocrit, reticulocytosis, direct hyperbilirubinemia, and mild transaminitis. Although acute hepatic sequestration rarely results in end organ failure, simple or exchange transfusion is often necessary to resolve the hepatopathy and anemia.¹⁵ However, overly aggressive simple transfusion should be avoided to prevent acute hyperviscosity syndrome resulting from rapid release of intrahepatic RBCs back into the circulation with resolved sequestration.

SCIC is the most severe form of acute sickle hepatopathy, with an overall mortality rate of 40% to 50% in adults and 30% in children due to uncontrolled bleeding and fulminant liver failure.¹⁴ SCIC manifests with sudden onset of right upper quadrant abdominal pain, significantly elevated transaminases (>1000 mg/dL), severe hyperbilirubinemia (total serum bilirubin often >50 mg/dL), coagulopathy, hepatomegaly, renal insufficiency, and acute liver failure in severe cases.¹⁶ Patients can have recurrent episodes, and a subset develop chronic progressive disease that evolves into progressive liver failure.¹⁷ Limited reports describe outcomes based on transfusion management. Ahn and colleagues¹⁸ reviewed 22 cases (7 pediatric, 15 adults) of

severe sickle hepatopathy in the literature that met criteria for SCIC; 7 of 9 patients who received exchange transfusion survived compared with 1 of 13 patients who did not receive erythrocytapheresis. Although this data set is limited, it supports a role for exchange transfusion in the management of SCIC, particularly acute SCIC.^{15,17} Correction of coagulopathies with plasma, cryoprecipitate, and platelet transfusions is also often necessary. Maintaining HbS levels less than 30% by chronic erythrocytapheresis has been proposed for patients with recurrent episodes of acute SCIC or chronic progressive hepatopathy.¹⁷

Multisystem Organ Failure

Multisystem organ failure (MSOF) is a life-threatening complication of SCD resulting from diffuse microvascular occlusion, which usually develops several days after a severe vasoocclusive crisis (VOC). MSOF is characterized by rapid lung, liver, and/or kidney dysfunction and is typically accompanied by a precipitous decrease in the Hb level and platelet count, fever, encephalopathy, and rhabdomyolysis. In addition to broad-spectrum antibiotics, mechanical ventilation, pharmacologic and/or mechanical hemodynamic support, and renal replacement therapy, the use of RBC transfusion can be life saving. The largest retrospective report of SCD-associated MSOF included 17 episodes occurring after unusually severe VOCs.¹⁹ All patients except one recovered with aggressive transfusion support via either multiple simple transfusions or an exchange transfusion. The National Heart, Lung, and Blood Institute (NHLBI)-appointed SCD expert panel recommends immediate simple or exchange transfusion for MSOF because of the gravity of this severe sickle-related manifestation.¹⁵

Preoperative Transfusion Management

Perioperative conditions, including suboptimal hydration, poor oxygenation, and acidemia, can lead to SCD-related complications, such as ACS, painful VOC, and infections. The Transfusion Alternatives Preoperatively in SCD (TAPS) trial demonstrated that preoperative transfusion is associated with decreased perioperative complications.²⁰ The TAPS trial compared outcomes of preoperative transfusion versus no transfusion in patients with HbSS or undergoing low-risk or medium-risk surgery. The study was terminated early because of an imbalance of adverse events occurring in the no-preoperative-transfusion arm, with 13 of 33 (39%) individuals experiencing a clinically important complication compared with 5 of 34 (15%) patients who were transfused preoperatively. Ten of 11 serious adverse events were ACS: 9 in the no transfusion and one in the transfusion group. The trial did not include individuals with hemoglobin SC or S β^+ thalassemia, and poor enrollment of patients requiring low-risk procedures hampered the determination of optimal management for this surgical category.²⁰ A prior randomized control trial showed that preoperative simple transfusion to achieve a Hb of 10 g/dL is equally effective in preventing postoperative complications compared with erythrocytapheresis to decrease the HbS level to less than 30%.²¹ Taken together, patients with SCD should receive a simple transfusion preoperatively to increase the Hb to 10 g/dL for medium- to high-risk surgery.

Neurologic Complications

In children with SCD, the routine use of transcranial Doppler (TCD) screening coupled with chronic transfusion therapy has decreased the prevalence of overt stroke from 11% to 1%.²² However, neurologic complications in children and adults with SCD remain a major cause of long-term morbidity. Acute ischemic stroke is managed

with RBC transfusion to reduce the HbS level to less than 30% to prevent progression of cerebral ischemia. Exchange transfusion at the time of stroke presentation may be associated with a lower risk of subsequent stroke compared with simple transfusion,²³ but no prospective study has directly addressed this question. In practice, erythrocytapheresis is the preferred transfusion method for initial treatment of an acute stroke. A simple transfusion may be considered for immediate treatment because it may require several hours to establish central venous access, crossmatch multiple PRBC units, and mobilize the apheresis team for erythrocytapheresis.

An evidence-based approach to the management of acute hemorrhagic stroke in patients with SCD is lacking.^{22,24} In the largest case series, 15 adults with SCD with subarachnoid hemorrhage from ruptured intracranial aneurysms were treated with partial exchange transfusion in the acute setting before cerebral angiography.²⁵ Given the lack of formal studies to guide management of hemorrhagic strokes in individuals with SCD, transfusion in the acute setting to decrease the HbS to less than 30% is recommended in addition to therapy for acute intracranial hemorrhage for the general population, including blood pressure management and seizure control.^{22,24}

Stroke recurs in 60% to 90% of patients without therapeutic intervention²⁶ and decreases to approximately 20% with chronic RBC transfusions when the HbS is maintained at less than 30%.^{27,28} Although a controlled clinical trial is lacking, standard care for secondary stroke prevention is chronic transfusion therapy. Indefinite therapy is recommended, as discontinuation after short-term or long-term prophylactic transfusions has led to recurrent stroke, even with transition to hydroxyurea.²⁹ The randomized phase 3 trial, Stroke With Transfusions Changing to Hydroxyurea, addressed transition to hydroxyurea for patients with a history of stroke and iron overload. The trial was closed because of statistical futility on the composite end point of iron overload resolution and stroke prevention. At the time of study closure, no strokes occurred in patients receiving transfusions with chelation, but 7 patients (10%) receiving hydroxyurea and phlebotomy had a new stroke. Moreover, patients receiving chronic transfusion for secondary stroke prevention are still at risk for silent cerebral infarcts (SCIs),^{27,30} and cerebral vasculopathy progression.^{27,31,32} Taken together, transfusion remains the optimal choice for managing individuals with SCD and stroke.

Children with SCD at the highest risk of overt stroke can be identified by abnormally high blood flow velocities on TCD ultrasound. The Stroke Prevention Trial for sickle cell anemia study (STOP) demonstrated that chronic transfusion therapy decreased the rate of initial stroke in children with an abnormal TCD by 92% compared with the observation arm.³³ Transfusion reduces cerebral blood-flow velocities, in part because of correction of the anemia, which contributes to lower stroke risk. Universal adoption of routine TCD screening and primary prophylactic transfusion therapy for at-risk patients has resulted in significantly decreased rates of first stroke.^{34–36} The STOP2 trial supported the use of chronic transfusion indefinitely, as discontinuation after 30 months resulted in an increased rate of abnormal TCD conversion and overt stroke as well as a higher occurrence of SCIs.^{37,38}

Based on previous studies demonstrating that hydroxyurea can lower TCD velocities in patients with SCD,^{39,40} the Transfusions Changing to Hydroxyurea (TWITCH) trial (ClinicalTrials.gov: NCT01425307) aimed to compare the efficacy of hydroxyurea with transfusion therapy for children with abnormal TCDs but no primary stroke. The TWITCH study ended in 2014 at the first interim analysis after data indicated the trial had reached its primary end point; preliminary results showed that hydroxyurea is not inferior to chronic RBC transfusion in lowering TCD velocities in children with abnormal

TCDs. Given the burden of chronic transfusion therapy and because most children with abnormal TCDs will not have a stroke if untreated, the ability to prevent stroke with hydroxyurea is a significant advance.

SCI is more common than overt stroke and is associated with increased risk of overt stroke, new or enlarged SCIs, poor academic achievement, and lower IQ.^{41–43} The Silent Infarct Transfusion trial demonstrated that chronic transfusion therapy significantly reduced the incidence of recurrent cerebral infarcts in patients with SCIs at baseline, no history of overt stroke, and normal TCD velocities.⁴⁴ Notably, 6 of 99 children (6%) assigned to transfusions had an event (1 stroke, 5 new or enlarged SCIs) compared with 14 of 97 children (14%) in the observation group (7 strokes, 7 new or enlarged SCIs). Because approximately 25% of children with SCD have SCIs,^{42,45} the resources needed to provide chronic transfusions to this group would be immense. Whether hydroxyurea can prevent new SCIs or overt stroke in patients with SCIs is not known. Future studies are also needed to determine whether patients with magnetic resonance angiography–defined vasculopathy but normal TCD velocities would benefit from RBC transfusions to prevent vasculopathy progression and/or silent or overt strokes.

CONTROVERSIAL INDICATIONS

Pulmonary Hypertension

Pulmonary arterial hypertension (PAH) is a common complication in adults with SCD and imposes an increased risk of death. The incidence of PAH diagnosed by right heart catheterization (RHC) is between 6% and 11% in adults with SCD.^{46–48} Mortality in these patients is approximately 35% to 40% at 3 to 6 years from diagnosis.^{47,48} RHC is the gold standard for diagnosing PAH; but noninvasive evaluations, including tricuspid regurgitant jet velocity (TRV) by Doppler echocardiography, serum N-terminal pro–brain natriuretic peptide (NT-pro-BNP), and the 6-minute walk distance, can be used to predict the presence of PAH and estimate mortality risk.

There are no randomized control trials of hydroxyurea or transfusion therapy for patients with SCD and PAH. The American Thoracic Society (ATS) recommends hydroxyurea for all patients with HbSS who have a TRV of 3.0 m/s or greater alone or TRV of greater than 2.5 m/s with either an elevated NT-pro-BNP level or RHC-confirmed PAH based on studies showing the benefits of hydroxyurea in reducing morbidity and mortality in SCD.^{49,50} Transfusion may prevent chronic regional pulmonary hypoxia by reducing the frequency of ACS and decreasing nitric oxide depletion, which results from chronic hemolysis, both of which are risk factors for PAH in SCD.⁵¹ Because transfusion may reverse or stabilize PAH in its early stages by mitigating these pulmonary endothelial effects, the ATS recommends that chronic transfusion therapy be offered to patients who are not responsive to or not candidates for hydroxyurea.^{51,52} A recent cross-sectional study of children and young adults with HbSS or HbSβ° supports this recommendation by demonstrating a protective effect of chronic transfusions on pulmonary vascular disease defined by lower TRV measurements.⁵³ Further investigation is needed to determine if either chronic transfusions or hydroxyurea impact mortality.

Pregnancy in Sickle Cell Disease

Pregnant women with SCD have a higher risk of obstetric complications, including preeclampsia/eclampsia, venous thromboembolism, intrauterine fetal demise, preterm birth, intrauterine growth restriction, and SCD-related complications, including

prepartum and postpartum VOC and ACS. Greater than 50% of patients with SCD have a pain crisis during pregnancy,⁵⁴ and 28% will have VOC at the time of delivery.⁵⁵ Additionally, approximately 20% of women develop ACS during pregnancy.⁵⁶ Furthermore, the maternal mortality rate is 16 times higher and fetal death at delivery is twice as likely in pregnant women with SCD.⁵⁷

Because hydroxyurea may be teratogenic, RBC transfusion is the only disease-modifying therapy for pregnant women with SCD. Transfusion is indicated for women with acute medical or obstetric complications, but conflicting data exist regarding the benefit of regular prophylactic transfusion during pregnancy. Prophylactic transfusion has been proposed to prevent SCD-related and obstetric complications toward the end of pregnancy when they are most frequent. Transfusion may be initiated at 28 weeks' gestation with the goal of maintaining an HbS less than 35% and an Hb of 10 to 11 g/dL.⁵⁸ This prophylactic transfusion strategy significantly reduced pain crises and symptomatic anemia but had no effect on pregnancy outcomes in 2 small trials involving 98 women with SCD conducted in the 1980s.^{59,60} Recently, prophylactic erythrocytapheresis initiated either in the second or third trimester has been shown in a small single-center retrospective cross-sectional study to be safe and effective in prevention of SCD-related events.⁶¹ Future studies are required to determine if prophylactic transfusions during pregnancy can decrease maternal and perinatal morbidity and mortality associated with SCD. Currently, most obstetricians provide selective transfusion to address specific complications that arise during pregnancy for women with SCD.

Leg Ulcers

Leg ulcers are a common complication of SCD and increase in occurrence with advancing age. By their fourth decade of life, 22% of patients with HbSS and 9% of patients with HbSC will report a history of leg ulcers.⁶² Standard treatment of leg ulcers is debridement, wet to dry dressings, topical agents, and prompt treatment of soft tissue infection and/or osteomyelitis.¹⁵ There have been anecdotal reports of transfusion stimulating ulcer healing. Although there are no studies to support chronic transfusion therapy for long-term management, RBC transfusion may be beneficial for patients undergoing surgical debridement, skin grafts, or muscle flaps to promote healing.⁶³

Vasoocclusive Crisis

Acute VOC is the most common cause of hospital admissions among patients with SCD. In the Cooperative Study of SCD, mean frequencies of pain episodes for patients with HbSC or HbSβ+, HbSS, and HbSβ° were 0.4, 0.8, and 1.0 episodes per year, respectively.⁶⁴ The average hospital stay for VOC is 7.5 days for adults and 4.4 days for children. Treatment of severe VOC requiring hospitalization is pain management with nonsteroidal antiinflammatory drugs and parenteral opioids, intravenous (IV) hydration, and incentive spirometry to minimize the risk of ACS. During acute uncomplicated VOC, mild exacerbation of anemia is common but does not require transfusion. There are no data to support RBC transfusion to manage acute uncomplicated VOC, and transfusion in this setting is associated with a higher risk of RBC alloimmunization.⁶⁵ A subset of patients have unusually frequent and severe pain episodes, associated with a poor quality of life. There is empirical evidence that chronic transfusions may reduce acute painful episodes,^{11,66} but a concurrent multidisciplinary pain program and periodic assessment for treatment response is required.

Priapism

Priapism affects approximately 35% of boys and men with SCD.⁶⁷ Prolonged and recurrent episodes of priapism are associated with tissue necrosis and fibrosis leading to erectile dysfunction. Initial management consists of IV fluids and analgesics; subsequent interventions include penile aspiration and corporal irrigation using α -adrenergic agents. The use of oral pseudoephedrine and low-dose phosphodiesterase type 5 inhibitors has been reported to have prophylactic benefit in SCD-associated priapism, and limited experience suggests hydroxyurea may prevent recurrence.⁶⁸ Surgical shunting procedures and penile prosthesis implantation are used when conservative measures fail.⁶⁹

The benefit of RBC transfusion for acute priapism in SCD is controversial. A systematic literature review of patients with SCD-associated priapism reported a mean time to detumescence of 8.0 days in 16 patients who did not receive transfusion compared with 11 days in 26 individuals who received transfusion. The review identified 9 cases of serious neurologic sequelae, including obtundation, seizures, and stroke following transfusion consistent with Association of Sickle cell disease, Priapism, Exchange transfusion, and Neurologic events (ASPEN) syndrome. Most resolved completely but left severe neurologic deficits in some.⁷⁰ Given the risk of ASPEN, blood transfusion is not typically recommended to treat acute priapism in patients with SCD. Chronic transfusion therapy is sometimes used to prevent recurrent intermittent priapism, but there is no published evidence of its efficacy. The NHLBI-appointed SCD expert panel recommends against the use of transfusion for the treatment of priapism.¹⁵ **Table 1** summarizes indications for and against transfusion therapy in patients with SCD.

PREVENTION OF COMPLICATIONS

Alloimmunization

RBC alloimmunization is a major complication associated with transfusions in SCD. Alloantibodies to the Rh (primarily C and E) and Kell (typically K) systems compose more than two-thirds of the antibodies detected. The high rate of alloimmunization is multifactorial, but discordance of blood group antigen expression on donor and patient RBCs is likely the major contributing factor. Prevention has focused on prophylactic RBC antigen matching, either limited C, E, K matching or extended antigen matching to include the Duffy, Kidd, and MNS systems (Fy^a, Fy^b, Jk^a, Jk^b, S). Alloimmunization prevalence in patients with SCD ranges from 27% to 75% with ABO-D matching alone, 5% to 14% with limited C, E, and K matching, and 0% to 7% for extended RBC antigen matching (**Table 2**). The rate of alloantibody formation ranges from 1.7 to 3.9 antibodies formed per 100 units transfused with ABO-D matching, 0.26 to 0.50 with limited C, E, K matching, and 0 to 0.10 with extended antigen matching (see **Table 2**).

Limited C-, E-, and K-matched RBCs can reduce alloimmunization (see **Table 2**). Despite consensus in the United States to provide C-, E-, and K-matched RBCs for patients with SCD,¹⁵ this has not been universally practiced.^{71,72} Some transfusion services provide extended matched RBCs after patients with SCD have formed one alloantibody, recognizing these patients as “immune responders.”⁷³ In addition to C-, E-, and K-matched RBCs, recruitment of dedicated donors to limit donor exposure for a given patient⁷⁴ or providing RBCs from African American donors who are more likely to have similar Jk, Fy, or S antigen status⁷⁵ have been alternative strategies used for SCD. In a single-institution study, C-, E-, and K-matched RBCs from African American donors resulted in no cases of anti-K formation and a low rate of anti-Jk, -Fy, and -S antibodies; but Rh immunization remained problematic (**Fig. 1**).⁷⁵

Table 2 Red blood cell antigen matching and alloimmunization					
Study	N	Total Units Transfused	Matching	Patients with Alloantibodies (%)	Rate (Alloantibody per 100 Units)
Ambruso et al, ⁸⁷ 1987	12	492	ABO, D	75	3.5
Rosse et al, ⁸⁸ 1990	1044	n/a	ABO, D	27	n/a
Vichinsky et al, ⁸⁹ 1990	107	1711	ABO, D	30	3.9
Aygun et al, ⁹⁰ 2002	140	3239	ABO, D	37	n/a
Castro et al, ⁹¹ 2002	351	8939	ABO, D	29	n/a
Sakhalkar et al, ⁹² 2005	387	14,263	ABO, D	31	1.7
Vichinsky et al, ⁹³ 2001	61	1830	Limited (C, E, K)	11	0.50
Sakhalkar et al, ⁹² 2005	113	2354	Limited (C, E, K)	5	0.26
O'Suoji et al, ⁹⁴ 2013	180	n/a	Limited (C, E, K)	14	n/a
DeBaun et al, ⁴⁴ 2014	90	3236	Limited (C, E, K)	4.5	0.28
Tahhan et al, ⁹⁵ 1994	40	608	Extended matching ^a	0	0
Lasalle-Williams et al, ⁹⁶ 2011	99	6946	Extended matching ^b	7	0.10
Chou et al, ⁷⁵ 2013	182	44,482	Limited (C, E, K) from African American donors	44	0.33

Abbreviation: n/a, not available.

^a C/c, E/e, K, Fya, Fyb, S.

^b C/c, E/e, K, Fya, Jka, Jkb.

Data from Refs.^{44,75,87–96}

Inheritance of variant *RHD* and *RHCE* genes encoding amino acid changes in the Rh antigens (D, C, c, E, e) is common in Africans, and its contribution to alloimmunization in patients with SCD despite Rh-matching programs has been increasingly appreciated.^{75,76} Many *RH* variants encode partial Rh antigens, and individuals become immunized when exposed to epitopes that their own cells lack.^{75,77} *RH* alleles that encode altered D, C, and e antigens often underlie the complex Rh alloantibody and apparent autoantibody specificities found in patients with SCD. Patients form anti-D, -C, and/or -e despite their own RBCs testing positive for those antigens and may represent Rh alloantibodies rather than autoantibodies. These antibodies can be clinically significant, and future transfusions should be with antigen-negative RBCs. This requirement poses significant challenges to transfusion services in providing compatible RBCs to these patients, taking care to avoid additional alloantibody development.

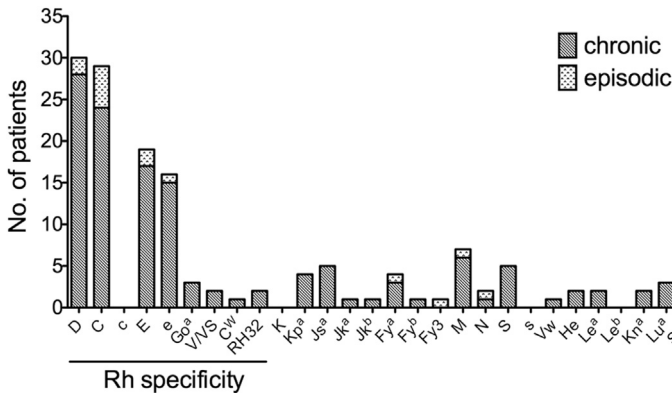


Fig. 1. Antibody specificities in patients with SCD transfused with Rh D-, C-, E-, and K-matched RBCs from African American donors. One hundred forty-six specific antibodies in 123 chronically and 59 episodically transfused patients identified at a single institution over 15 years. (From Chou ST, Jackson T, Vege S, et al. High prevalence of red blood cell alloimmunization in sickle cell disease despite transfusion from Rh-matched minority donors. *Blood* 2013;122(6):1064; with permission. Copyright © the American Society of Hematology.)

Patients with SCD benefit from having an extended RBC antigen phenotype (C/c, E/e, K/k, Fy^a/Fy^b, Jk^a/Jk^b, M/N, S/s, and Le^a/Le^b) completed in the first year of life. Knowledge of RBC antigen status facilitates antibody evaluation and identification of compatible RBC units in patients with a newly formed antibody. RBCs should be matched for C, E, and K antigens at a minimum.¹⁵ Extended antigen-matched RBCs to minimize alloimmunization against Kidd, Duffy, and S antigens is preferable; but finding sufficient RBC units for chronically transfused patients may be challenging for some regions. Overall transfusion cost increases with higher levels of antigen matching, which is prohibitive for many providers. Using molecular blood typing can augment serologic typing and may allow the ability to provide highly antigen-matched products in the future and potentially be cost saving.^{78,79}

Altered *RH* alleles in patients with SCD suggest an important emerging role for molecular methods to improve RBC matching in the Rh system.^{75,80} Prevention of Rh alloantibody formation in patients with partial antigens is a challenge. Providing antigen-negative RBCs for individuals predicted to exclusively express partial Rh antigens may be performed on an individual case basis, with consideration of their extended RBC antigen profile, alloimmunization history and specificities, and the predicted availability of appropriate donors. For individuals with the relatively common hybrid *RHD*DIlla-CE(4-7)-D* gene that encodes a partial C antigen, providing C-negative RBCs to those lacking an *RHCE* gene encoding conventional C can minimize anti-C alloimmunization and is consistent with a prophylactic C matching policy.⁸¹ For patients whose *RH* genotypes predict exclusive expression of partial D and/or e antigens, selection of appropriate RBC units is more complex. For example, D-negative units are more likely to be identified among European donors. Providing D-negative RBCs to patients with partial D antigen status may increase alloimmunization risk to other antigens (Fya, Jkb, and S) that are more common in Europeans compared with Africans. As sequencing platforms increasingly improve and become cost-effective, *RH* genotyping can complement serologic testing for both typing and matching donors and patients.

Monitoring and Treatment of Iron Overload

Patients with SCD on chronic or recurrent episodic transfusions are at risk for iron overload and subsequent liver, endocrine, and/or cardiac dysfunction.⁸² Recommendations for monitoring and treatment of iron overload in SCD are largely based on thalassemia literature. Transfusional iron burden should be monitored with serum ferritin levels and liver and cardiac MRI. Liver iron quantification by R2 and R2* MRI techniques correlates well with liver iron content (LIC) determined by liver biopsy.⁸³ Although less common than comparably transfused patients with thalassemia major, cardiac iron overload occurs in a small proportion of patients with SCD with exceptionally poor control of iron status.⁸⁴ In such patients, cardiac T2* MRI can predict the risk of developing heart disease, allowing intensification of therapy.⁸⁵ LIC should be measured by R2-MRI every 1 to 2 years while on chronic transfusions and/or if serum ferritin remains greater than 1000 ng/mL. Iron-chelating agents are typically initiated within 1 to 2 years of instituting chronic transfusions, after 10 to 20 cumulative RBC units (~120 mL/kg), or when LIC is 7 mg/g or greater dry weight.⁸⁶ Chelation is titrated to maintain serum ferritin less than 1000 ng/mL and LIC less than 7 mg/g liver dry weight, extrapolated from the thalassemia data.²

Iron chelation therapy can maintain a negative iron balance in most compliant patients requiring chronic transfusions. Deferoxamine (25–40 mg/kg/d SQ) and deferasirox (20–40 mg/kg/d by mouth) have been licensed for the treatment of iron overload in SCD, but the safety and efficacy of deferiprone (75–100 mg/kg/d by mouth) has not been established for SCD in the United States. Although deferiprone has been available for patients with SCD outside the United States and Canada since 1995, experience is limited.^{2,82}

Unique to SCD, the iron-loading rate of transfusion therapy depends on the modality used (simple vs exchange), the type of exchange (partial manual vs automated), and the target Hb and percentage of HbS.⁸² The net iron balance is also influenced by dose and compliance with iron chelation therapy. Patients with severe iron overload may benefit from erythrocytapheresis combined with chelation and by adjusting automated erythrocytapheresis parameters to achieve minimal iron loading. **Tables 3** and **4** provide calculations of transfusion-associated iron accumulation and balance with different transfusion regimens.

Table 3 Calculations of iron accumulation from transfusion	
RBC Transfusion Volumes	Amount of Fe Accumulation
1 mL of erythrocytes	Approximately 1 mg of Fe
Fe content of transfused blood	Approximately 0.75 mg/mL (225 mg Fe per 300 mL unit) ^a
Approximately 2 units of RBCs	Approximately 0.5 g of Fe accumulation within the body ^b
10 mL/kg of RBCs	Approximately 7.5 mg Fe/kg
15 mL/kg of RBCs	Approximately 11.3 mg Fe/kg
For chronic transfusions	
10 mL/kg RBCs every 4 weeks	Approximately 0.25 mg/kg/d
15 mL/kg RBCs every 4 weeks	Approximately 0.40 mg/kg/d
15 mL/kg RBCs every 3 weeks	Approximately 0.54 mg/kg/d

Abbreviation: Fe, iron.
^a This value assumes a red cell unit hematocrit of 65%.⁶
^b Adults normally have approximately 4 to 5 g of total body iron.

Table 4
Iron balance with transfusion regimens

Transfusion Modality	Fe Accumulation (mg/kg/d) ^a	DFX Dosage to Balance Input	DFO Dosage to Balance Input
Simple transfusion			
Target HbS <30%	0.42	20 mg/kg/d	40 mg/kg × 5/wk
Target HbS <50%	0.32	16 mg/kg/d	32 mg/kg × 5/wk
Partial Manual Exchange ^b			
Target HbS <30%	0.36	~ 18 mg/kg/d	37 mg/kg × 5/wk
Erythrocytapheresis			
Target HbS <30%	0.04–0.08	<5 mg/kg/d	<10 mg/kg × 5/wk
Target HbS <50%	≤0.052	<5 mg/kg/d	<10 mg/kg × 5/wk

Abbreviations: DFO, deferiprone; DFX, deferoxamine; Fe, iron.

^a These calculations are approximations. The rate of iron accumulation varies within the transfusion modality used based on transfusion volume (10 vs 15 mL/kg), frequency of transfusions (every 3 weeks vs every 4 weeks), phlebotomy volume (partial manual exchange), and after exchange target Hb (erythrocytapheresis).

^b Fe accumulation approximation for partial manual exchange extrapolated from data calculations from reference.⁶

Adapted from Porter J, Garbowski M. Consequences and management of iron overload in sickle cell disease. *Hematology Am Soc Hematol Educ Program* 2013;2013:448.

SUMMARY

RBC transfusion is a critical component in the treatment of SCD. Clinicians should be knowledgeable of evidence-based or expert panel-based consensus of transfusion indications and strategies to prevent and manage transfusion-related complications.

REFERENCES

1. Quinn CT. Sick cell disease in childhood: from newborn screening through transition to adult medical care. *Pediatr Clin North Am* 2013;60(6):1363–81.
2. Smith-Whitley K, Thompson AA. Indications and complications of transfusions in sickle cell disease. *Pediatr Blood Cancer* 2012;59(2):358–64.
3. Wahl S, Quirolo KC. Current issues in blood transfusion for sickle cell disease. *Curr Opin Pediatr* 2009;21(1):15–21.
4. Josephson CD, Su LL, Hillyer KL, et al. Transfusion in the patient with sickle cell disease: a critical review of the literature and transfusion guidelines. *Transfus Med Rev* 2007;21(2):118–33.
5. Wanko SO, Telen MJ. Transfusion management in sickle cell disease. *Hematol Oncol Clin North Am* 2005;19(5):803–26, v-vi.
6. Savage WJ, Reddoch S, Wolfe J, et al. Partial manual exchange reduces iron accumulation during chronic red cell transfusions for sickle cell disease. *J Pediatr Hematol Oncol* 2013;35(6):434–6.
7. Brousse V, Elie C, Benkerrou M, et al. Acute splenic sequestration crisis in sickle cell disease: cohort study of 190 paediatric patients. *Br J Haematol* 2012;156(5):643–8.
8. Smith-Whitley K, Zhao H, Hodinka RL, et al. Epidemiology of human parvovirus B19 in children with sickle cell disease. *Blood* 2004;103(2):422–7.

9. Vichinsky EP, Neumayr LD, Earles AN, et al. Causes and outcomes of the acute chest syndrome in sickle cell disease. National Acute Chest Syndrome Study Group. *N Engl J Med* 2000;342(25):1855–65.
10. Turner JM, Kaplan JB, Cohen HW, et al. Exchange versus simple transfusion for acute chest syndrome in sickle cell anemia adults. *Transfusion* 2009;49(5):863–8.
11. Miller ST, Wright E, Abboud M, et al. Impact of chronic transfusion on incidence of pain and acute chest syndrome during the Stroke Prevention Trial (STOP) in sickle-cell anemia. *J Pediatr* 2001;139(6):785–9.
12. Hankins J, Jeng M, Harris S, et al. Chronic transfusion therapy for children with sickle cell disease and recurrent acute chest syndrome. *J Pediatr Hematol Oncol* 2005;27(3):158–61.
13. Miller ST. How I treat acute chest syndrome in children with sickle cell disease. *Blood* 2011;117(20):5297–305.
14. Banerjee S, Owen C, Chopra S. Sickle cell hepatopathy. *Hepatology* 2001;33(5):1021–8.
15. Yawn BP, Buchanan GR, Afenyi-Annan AN, et al. Management of sickle cell disease: summary of the 2014 evidence-based report by expert panel members. *JAMA* 2014;312(10):1033–48.
16. Papafragkakis H, Ona MA, Changela K, et al. Acute liver function decompensation in a patient with sickle cell disease managed with exchange transfusion and endoscopic retrograde cholangiography. *Therap Adv Gastroenterol* 2014;7(5):217–23.
17. Gardner K, Suddle A, Kane P, et al. How we treat sickle hepatopathy and liver transplantation in adults. *Blood* 2014;123(15):2302–7.
18. Ahn H, Li CS, Wang W. Sickle cell hepatopathy: clinical presentation, treatment, and outcome in pediatric and adult patients. *Pediatr Blood Cancer* 2005;45(2):184–90.
19. Hassell KL, Eckman JR, Lane PA. Acute multiorgan failure syndrome: a potentially catastrophic complication of severe sickle cell pain episodes. *Am J Med* 1994;96(2):155–62.
20. Howard J, Malfroy M, Llewelyn C, et al. The Transfusion Alternatives Preoperatively in Sickle Cell Disease (TAPS) study: a randomised, controlled, multicentre clinical trial. *Lancet* 2013;381(9870):930–8.
21. Vichinsky EP, Haberkern CM, Neumayr L, et al. A comparison of conservative and aggressive transfusion regimens in the perioperative management of sickle cell disease. The Preoperative Transfusion in Sickle Cell Disease Study Group. *N Engl J Med* 1995;333(4):206–13.
22. Kassim AA, Galadanci NA, Pruthi S, et al. How I treat and manage strokes in sickle cell disease. *Blood* 2015;125(22):3401–10.
23. Hulbert ML, Scothorn DJ, Panepinto JA, et al. Exchange blood transfusion compared with simple transfusion for first overt stroke is associated with a lower risk of subsequent stroke: a retrospective cohort study of 137 children with sickle cell anemia. *J Pediatr* 2006;149(5):710–2.
24. Strouse JJ, Lanzkron S, Urrutia V. The epidemiology, evaluation and treatment of stroke in adults with sickle cell disease. *Expert Rev Hematol* 2011;4(6):597–606.
25. Oyesiku NM, Barrow DL, Eckman JR, et al. Intracranial aneurysms in sickle-cell anemia: clinical features and pathogenesis. *J Neurosurg* 1991;75(3):356–63.
26. Powars D, Wilson B, Imbus C, et al. The natural history of stroke in sickle cell disease. *Am J Med* 1978;65(3):461–71.
27. Hulbert ML, McKinstry RC, Lacey JL, et al. Silent cerebral infarcts occur despite regular blood transfusion therapy after first strokes in children with sickle cell disease. *Blood* 2011;117(3):772–9.

28. Scothorn DJ, Price C, Schwartz D, et al. Risk of recurrent stroke in children with sickle cell disease receiving blood transfusion therapy for at least five years after initial stroke. *J Pediatr* 2002;140(3):348–54.
29. Ware RE, Helms RW, Investigators SW. Stroke With Transfusions Changing to Hydroxyurea (SWITCH). *Blood* 2012;119(17):3925–32.
30. Gyang E, Yeom K, Hoppe C, et al. Effect of chronic red cell transfusion therapy on vasculopathies and silent infarcts in patients with sickle cell disease. *Am J Hematol* 2011;86(1):104–6.
31. Bishop S, Matheus MG, Abboud MR, et al. Effect of chronic transfusion therapy on progression of neurovascular pathology in pediatric patients with sickle cell anemia. *Blood Cells Mol Dis* 2011;47(2):125–8.
32. Brousse V, Hertz-Pannier L, Consigny Y, et al. Does regular blood transfusion prevent progression of cerebrovascular lesions in children with sickle cell disease? *Ann Hematol* 2009;88(8):785–8.
33. Adams RJ, McKie VC, Hsu L, et al. Prevention of a first stroke by transfusions in children with sickle cell anemia and abnormal results on transcranial Doppler ultrasonography. *N Engl J Med* 1998;339(1):5–11.
34. Enninfu-Eghan H, Moore RH, Ichord R, et al. Transcranial Doppler ultrasonography and prophylactic transfusion program is effective in preventing overt stroke in children with sickle cell disease. *J Pediatr* 2010;157(3):479–84.
35. McCarville MB, Goodin GS, Fortner G, et al. Evaluation of a comprehensive transcranial Doppler screening program for children with sickle cell anemia. *Pediatr Blood Cancer* 2008;50(4):818–21.
36. McCavit TL, Xuan L, Zhang S, et al. National trends in incidence rates of hospitalization for stroke in children with sickle cell disease. *Pediatr Blood Cancer* 2013;60(5):823–7.
37. Adams RJ, Brambilla D, Optimizing Primary Stroke Prevention in Sickle Cell Anemia (STOP 2) Trial Investigators. Discontinuing prophylactic transfusions used to prevent stroke in sickle cell disease. *N Engl J Med* 2005;353(26):2769–78.
38. Abboud MR, Yim E, Musallam KM, et al. Discontinuing prophylactic transfusions increases the risk of silent brain infarction in children with sickle cell disease: data from STOP II. *Blood* 2011;118(4):894–8.
39. Zimmerman SA, Schultz WH, Burgett S, et al. Hydroxyurea therapy lowers transcranial Doppler flow velocities in children with sickle cell anemia. *Blood* 2007;110(3):1043–7.
40. Gulbis B, Haberman D, Dufour D, et al. Hydroxyurea for sickle cell disease in children and for prevention of cerebrovascular events: the Belgian experience. *Blood* 2005;105(7):2685–90.
41. Miller ST, Macklin EA, Pegelow CH, et al. Silent infarction as a risk factor for overt stroke in children with sickle cell anemia: a report from the Cooperative Study of Sickle Cell Disease. *J Pediatr* 2001;139(3):385–90.
42. Pegelow CH, Macklin EA, Moser FG, et al. Longitudinal changes in brain magnetic resonance imaging findings in children with sickle cell disease. *Blood* 2002;99(8):3014–8.
43. Schatz J, Brown RT, Pascual JM, et al. Poor school and cognitive functioning with silent cerebral infarcts and sickle cell disease. *Neurology* 2001;56(8):1109–11.
44. DeBaun MR, Gordon M, McKinstry RC, et al. Controlled trial of transfusions for silent cerebral infarcts in sickle cell anemia. *N Engl J Med* 2014;371(8):699–710.
45. Kwiatkowski JL, Zimmerman RA, Pollock AN, et al. Silent infarcts in young children with sickle cell disease. *Br J Haematol* 2009;146(3):300–5.

46. Parent F, Bachir D, Inamo J, et al. A hemodynamic study of pulmonary hypertension in sickle cell disease. *N Engl J Med* 2011;365(1):44–53.
47. Fonseca GH, Souza R, Salemi VM, et al. Pulmonary hypertension diagnosed by right heart catheterisation in sickle cell disease. *Eur Respir J* 2012;39(1):112–8.
48. Mehari A, Gladwin MT, Tian X, et al. Mortality in adults with sickle cell disease and pulmonary hypertension. *JAMA* 2012;307(12):1254–6.
49. Charache S, Terrin ML, Moore RD, et al. Effect of hydroxyurea on the frequency of painful crises in sickle cell anemia. Investigators of the Multicenter Study of Hydroxyurea in Sickle Cell Anemia. *N Engl J Med* 1995;332(20):1317–22.
50. Voskaridou E, Christoulas D, Bilalis A, et al. The effect of prolonged administration of hydroxyurea on morbidity and mortality in adult patients with sickle cell syndromes: results of a 17-year, single-center trial (LaSHS). *Blood* 2010;115(12):2354–63.
51. Klings ES, Machado RF, Barst RJ, et al. An official American Thoracic Society clinical practice guideline: diagnosis, risk stratification, and management of pulmonary hypertension of sickle cell disease. *Am J Respir Crit Care Med* 2014;189(6):727–40.
52. Hayes MM, Vedamurthy A, George G, et al. Pulmonary hypertension in sickle cell disease. *Ann Am Thorac Soc* 2014;11(9):1488–9.
53. Detterich JA, Kato RM, Rabai M, et al. Chronic transfusion therapy improves but does not normalize systemic and pulmonary vasculopathy in sickle cell disease. *Blood* 2015;126(6):703–10.
54. Powars DR, Sandhu M, Niland-Weiss J, et al. Pregnancy in sickle cell disease. *Obstet Gynecol* 1986;67(2):217–28.
55. Alayed N, Kezouh A, Oddy L, et al. Sickle cell disease and pregnancy outcomes: population-based study on 8.8 million births. *J Perinat Med* 2014;42(4):487–92.
56. Okusanya BO, Oladapo OT. Prophylactic versus selective blood transfusion for sickle cell disease in pregnancy. *Cochrane Database Syst Rev* 2013;(12):CD010378.
57. Barfield WD, Barradas DT, Manning SE, et al. Sickle cell disease and pregnancy outcomes: women of African descent. *Am J Prev Med* 2010;38(4 Suppl):S542–9.
58. ACOG Committee on Obstetrics. ACOG practice bulletin No. 78: hemoglobinopathies in pregnancy. *Obstet Gynecol* 2007;109(1):229–37.
59. Koshy M, Burd L, Wallace D, et al. Prophylactic red-cell transfusions in pregnant patients with sickle cell disease. A randomized cooperative study. *N Engl J Med* 1988;319(22):1447–52.
60. Koshy M, Burd L, Dorn L, et al. Frequency of pain crisis during pregnancy. *Prog Clin Biol Res* 1987;240:305–11.
61. Asma S, Kozanoglu I, Tarim E, et al. Prophylactic red blood cell exchange may be beneficial in the management of sickle cell disease in pregnancy. *Transfusion* 2015;55(1):36–44.
62. Minniti CP, Taylor JG 6th, Hildesheim M, et al. Laboratory and echocardiography markers in sickle cell patients with leg ulcers. *Am J Hematol* 2011;86(8):705–8.
63. Minniti CP, Kato GJ. How we treat sickle cell patients with leg ulcers. *Am J Hematol* 2016;91(1):22–30.
64. Platt OS, Thorington BD, Brambilla DJ, et al. Pain in sickle cell disease. Rates and risk factors. *N Engl J Med* 1991;325(1):11–6.
65. Fasano RM, Booth GS, Miles M, et al. Red blood cell alloimmunization is influenced by recipient inflammatory state at time of transfusion in patients with sickle cell disease. *Br J Haematol* 2015;168(2):291–300.

66. Styles LA, Vichinsky E. Effects of a long-term transfusion regimen on sickle cell-related illnesses. *J Pediatr* 1994;125(6 Pt 1):909–11.
67. Olujobungbe AB, Adeyoju A, Yardumian A, et al. A prospective diary study of stuttering priapism in adolescents and young men with sickle cell anemia: report of an international randomized control trial—the priapism in sickle cell study. *J Androl* 2011;32(4):375–82.
68. Kato GJ. Priapism in sickle-cell disease: a hematologist's perspective. *J Sex Med* 2012;9(1):70–8.
69. Anele UA, Le BV, Resar LM, et al. How I treat priapism. *Blood* 2015;125(23):3551–8.
70. Merritt AL, Haiman C, Henderson SO. Myth: blood transfusion is effective for sickle cell anemia-associated priapism. *CJEM* 2006;8(2):119–22.
71. Afenyi-Annan A, Willis MS, Konrad TR, et al. Blood bank management of sickle cell patients at comprehensive sickle cell centers. *Transfusion* 2007;47(11):2089–97.
72. Osby M, Shulman IA. Phenotype matching of donor red blood cell units for non-alloimmunized sickle cell disease patients: a survey of 1182 North American laboratories. *Arch Pathol Lab Med* 2005;129(2):190–3.
73. Karafin MS, Shirey RS, Ness PM, et al. Antigen-matched red blood cell transfusions for patients with sickle cell disease at the Johns Hopkins Hospital. *Immunohematology* 2012;28(1):3–6.
74. Roberts DO, Covert B, Lindsey T, et al. Directed blood donor program decreases donor exposure for children with sickle cell disease requiring chronic transfusion. *Immunohematology* 2012;28(1):7–12.
75. Chou ST, Jackson T, Vege S, et al. High prevalence of red blood cell alloimmunization in sickle cell disease despite transfusion from Rh-matched minority donors. *Blood* 2013;122(6):1062–71.
76. Noizat-Pirenne F, Tournamille C. Relevance of RH variants in transfusion of sickle cell patients. *Transfus Clin Biol* 2011;18(5–6):527–35.
77. Chou ST, Westhoff CM. The role of molecular immunohematology in sickle cell disease. *Transfus Apher Sci* 2011;44(1):73–9.
78. Casas J, Friedman DF, Jackson T, et al. Changing practice: red blood cell typing by molecular methods for patients with sickle cell disease. *Transfusion* 2015;55(6 Pt 2):1388–93.
79. Wilkinson K, Harris S, Gaur P, et al. Molecular blood typing augments serologic testing and allows for enhanced matching of red blood cells for transfusion in patients with sickle cell disease. *Transfusion* 2012;52(2):381–8.
80. Kappler-Gratias S, Auxerre C, Dubeaux I, et al. Systematic RH genotyping and variant identification in French donors of African origin. *Blood Transfus* 2014;12(Suppl 1):S264–72.
81. Tournamille C, Meunier-Costes N, Costes B, et al. Partial C antigen in sickle cell disease patients: clinical relevance and prevention of alloimmunization. *Transfusion* 2010;50(1):13–9.
82. Porter J, Garbowski M. Consequences and management of iron overload in sickle cell disease. *Hematol Am Soc Hematol Educ Program* 2013;2013:447–56.
83. Wood JC, Enriquez C, Ghugre N, et al. MRI R2 and R2* mapping accurately estimates hepatic iron concentration in transfusion-dependent thalassemia and sickle cell disease patients. *Blood* 2005;106(4):1460–5.
84. Meloni A, Puliyl M, Pepe A, et al. Cardiac iron overload in sickle-cell disease. *Am J Hematol* 2014;89(7):678–83.

85. Wood JC. Magnetic resonance imaging measurement of iron overload. *Curr Opin Hematol* 2007;14(3):183–90.
86. Inati A, Khoriaty E, Musallam KM, et al. Iron chelation therapy for patients with sickle cell disease and iron overload. *Am J Hematol* 2010;85(10):782–6.
87. Ambruso DR, Githens JH, Alcorn R, et al. Experience with donors matched for minor blood group antigens in patients with sickle cell anemia who are receiving chronic transfusion therapy. *Transfusion* 1987;27:94–8.
88. Rosse WF, Gallagher D, Kinney TR, et al. Transfusion and alloimmunization in sickle cell disease. The Cooperative Study of Sickle Cell Disease. *Blood* 1990;76:1431–7.
89. Vichinsky EP, Earles A, Johnson RA, et al. Alloimmunization in sickle cell anemia and transfusion of racially unmatched blood. *N Engl J Med* 1990;322:1617–21.
90. Aygun B, Padmanabhan S, Paley C, et al. Clinical significance of RBC alloantibodies and autoantibodies in sickle cell patients who received transfusions. *Transfusion* 2002;42:37–43.
91. Castro O, Sandler SG, Houston-Yu P, et al. Predicting the effect of transfusing only phenotype-matched RBCs to patients with sickle cell disease: theoretical and practical implications. *Transfusion* 2002;42:684–90.
92. Sakhalkar VS, Roberts K, Hawthorne LM, et al. Allosensitization in patients receiving multiple blood transfusions. *Ann N Y Acad Sci* 2005;1054:495–9.
93. Vichinsky EP, Luban NL, Wright E, et al. Prospective RBC phenotype matching in a stroke-prevention trial in sickle cell anemia: a multicenter transfusion trial. *Transfusion* 2001;41:1086–92.
94. O'Suoji C, Liem RI, Mack AK, et al. Alloimmunization in sickle cell anemia in the era of extended red cell typing. *Pediatr Blood Cancer* 2013;60:1487–91.
95. Tahhan HR, Holbrook CT, Braddy LR, et al. Antigen-matched donor blood in the transfusion management of patients with sickle cell disease. *Transfusion* 1994;34:562–9.
96. Lasalle-Williams M, Nuss R, Le T, et al. Extended red blood cell antigen matching for transfusions in sickle cell disease: a review of a 14-year experience from a single center (CME). *Transfusion* 2011;51:1732–9.