

# Supplementary Materials II – Example Data and Code for QSM Reconstruction

Version, v0.2.1

## Introduction

This supplementary material provides an overview of the QSM reconstruction processing, from scanner-provided data to QSM maps, based on the recommendations present in the main text. This document has two main purposes:

- (1) Allowing readers to reproduce the results shown throughout the paper;
- (2) Providing readers with the means to reconstruct their own data using the recommended processing with data acquired on any of the 3 major MR providers broadly following the recommendations.

Full datasets, results and processing scripts are available on Zenodo:  
<https://doi.org/10.5281/zenodo.7410455>

Example data of version v0.2.1 were used in this paper.

## Data availability

Data are available from three vendors: GE, SIEMENS and PHILIPS, using the recommended acquisition in the main text. For each vendor, both monopolar and bipolar readout strategies were used to acquire the data for demonstration purposes. The data from GE and SIEMENS are not pre-scan normalized (which does not follow the recommendation), while the PHILIPS data have two normalization methods applied. In this way, we demonstrate the robustness of the proposed pipeline to a variety of implementations of the recommended protocol.

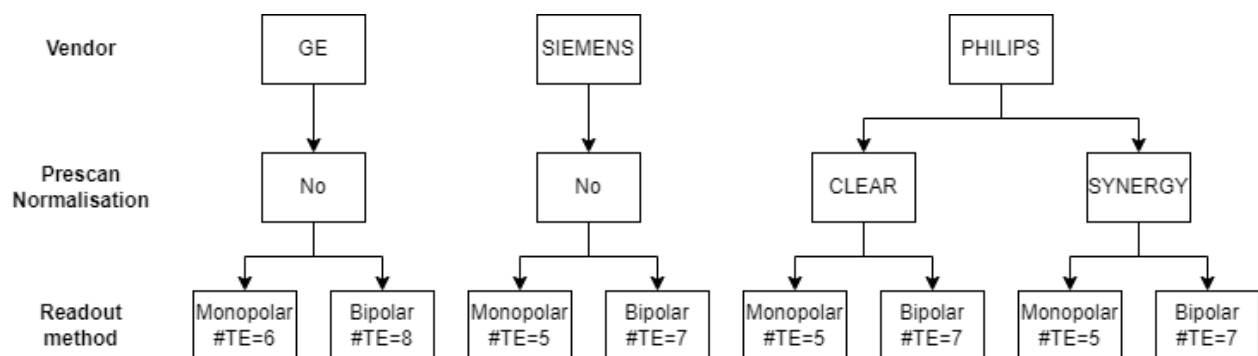


Figure S1: An illustration of the raw data available.

## 1 Data preparation and organization

2 Background: There are two zip files available in Zenodo containing the example data  
3 organised in two different ways:

4 (a) "QSM\_CONSENSUS\_Paper\_Example\_DICOM\_code.zip"

5 This zip file contains example DICOM images exported from the scanners  
6 without any modifications. The code directory accompanied with this file  
7 contains the scripts to (1) convert the DICOM images to NIFTI format, (2)  
8 organise the NIFTI images according to BIDS v1.8.0, and (3) perform QSM  
9 reconstruction.

10  
11 (b) "QSM\_Consensus\_Paper\_example\_Data\_Result\_Code.zip"

12 This zip file contains all data and results that were produced by running all the  
13 scripts provided in the code directory.

14  
15 The following Data Preparation section provides information on all the pre-processing  
16 steps to prepare the unmodified DICOM images to the BIDS format data that is ready  
17 for QSM reconstruction in SEPIA. If the readers are interested in the QSM  
18 reconstruction and work on the  
19 "QSM\_Consensus\_Paper\_example\_Data\_Result\_Code.zip" file only, they may skip  
20 Section "Data Preparation".  
21

## 22 Data preparation

23 The scripts created for this section were tested on a Mac system (macOS 13.2) and a  
24 Linux system (CentOS 7). These were not tested on Windows systems and would  
25 require adaptations to work on that OS.

26  
27 Most imaging software in the field typically deals with images in Analyze or NiftI format.  
28 As such the raw data (imaging data after coil combination) provided has to be converted  
29 to this format in 4 steps:

30 Step 1: Unzip the received data and reformat the directory structure

31 Scripts:

- 32 • Preparation\_01\_rename\_received\_data.sh

33  
34 Step 2: Convert DICOM images into NiftI format

35 Dependency: dcm2nii (version 1.0.20220720)

36 Scripts:

- 37 • Preparation\_02\_convert\_dicom2nii.sh

38  
39 Step 3: Rename the files according to the BIDS format (Brain Imaging Data  
40 structure)

41 Dependency: Matlab R2016b onwards

42 The naming strategy is as follows:

- 43 • Vendors are identified using the session tag: ses-  
44 <GE|PHILIPS|SIEMENS>

- For GE and SIEMENS, different readout methods are identified using the acquisition tag: acq-<Bipolar|Monopolar>;
- For PHILIPS, the normalisation method is also printed on the acquisition tag, i.e., acq-<BipolarCLEAR|BipolarSYNERGY|MonopolarCLEAR|MonopolarSYNERGY>

Script:

1. Preparation\_03\_rename\_to\_bids\_format.m

Step 4: Prepare NIFTI data for SEPIA

Dependency: (1) Matlab R2016b onwards, (2) SEPIA v1.2.2.4

Involves the following operation:

- Combining individual multi-echo 3D volumes into a single 4D volume with TE in the 4th dimension;
- Obtaining header info (e.g., B<sub>0</sub> direction and TE) from NIFTI header and JSON sidecar files and saving as SEPIA's header format;
- (GE only) Correcting inter-slice opposite polarity on real and imaginary images and exporting phase images from the corrected real/imaginary data

Script:

- Preparation\_04\_prepare\_for\_sepia.m

## Data organization

The following tree diagram illustrates the directory structure of how the data are organised after running all the scripts provided in the code directory “QSM\_Consensus\_Paper\_Example\_Code/”. The content of the different directories is mentioned after the comment “%” symbol. Note that similar directories exist under the “/derivatives/SEPIA/SIEMENS/” and “/derivatives/SEPIA/PHILIPS/” as under “/derivatives/SEPIA/GE/”.

```
QSM_Consensus_Paper_Example_DICOM_Code/
|-- QSM_CONSENSUS_DATA.zip           % Zip file containing all unmodified DICOM images
|-- protocols                       % Protocol text/HTML files
|-- QSM_Consensus_Paper_Example_Code % Containing all the scripts
|   |-- doc                         % Containing manual to use the Example data
|   |-- From_DICOM_zip_file_to_SEPIA_ready % Scripts for preparing QSM_CONSENSUS_DATA.zip
|   |-- SEPIA_Pipeline_FANSI         % SEPIA pipeline config files with FANSI recon
|   |-- SEPIA_Pipeline_MEDI          % SEPIA pipeline config files with MEDI recon
|-- raw                             % DICOM images
|-- converted                       % dcm2niix output
|   |-- GE
|   |   |-- Bipolar                 % Bipolar readout acquisition
|   |   |-- Monopolar               % Monopolar readout acquisition
|   |-- PHILIPS
|   |   |-- Bipolar_CLEAR           % with CLEAR normalisation
|   |   |-- Bipolar_SYNERGY        % with SYNERGY normalisation
|   |   |-- Monopolar_CLEAR
|   |   |-- Monopolar_SYNERGY
|   |-- SIEMENS
|   |   |-- Bipolar
|   |   |-- Monopolar
|-- derivatives                     % directory contains all derived output
    |-- SEPIA                      % SEPIA output
        |-- GE
```

```

1 | |-- Bipolar
2 | |   |-- GRE
3 | |   |-- Pipeline_FANSI % Full QSM recon using FANSI for dipole inversion
4 | |   |-- Pipeline_MEDI % Full QSM recon using MEDI for dipole inversion
5 | |   |-- Monopolar
6 | |   |-- GRE
7 | |   |-- Pipeline_FANSI % Full QSM recon using FANSI for dipole inversion
8 | |   |-- Pipeline_MEDI % Full QSM recon using MEDI for dipole inversion
9 | |-- PHILIPS
10 | |-- SIEMENS

```

## QSM reconstruction pipeline

This section describes all the QSM reconstruction processing steps performed in SEPIA. All the processing steps are specified in the SEPIA pipeline configuration files, which are in the sub-directories of the script directory:

“QSM\_Consensus\_Paper\_Example\_Code/SEPIA\_Pipeline\_FANSI/” and “QSM\_Consensus\_Paper\_Example\_Code/SEPIA\_Pipeline\_MEDI/”, corresponding to the two processing pipelines demonstrated as follows.

## Environment and dependencies

The data were processed using the following setup:

### Operating system

- Linux CentOS 7

### Environment

- Matlab R2021a (but the scripts are backwards compatible with earlier Matlab versions from R2016b to R2022a)

### Dependencies

The following QSM toolboxes have to be downloaded and integrated into SEPIA following the instruction provided on the SEPIA documentation website ([https://sepia-documentation.readthedocs.io/en/latest/getting\\_started/Installation.html](https://sepia-documentation.readthedocs.io/en/latest/getting_started/Installation.html)):

- SEPIA v1.2.2.4 (<https://github.com/kschan0214/sepia/releases/tag/v1.2.2.4>)
- MRITools v3.5.6 (<https://github.com/korbinian90/CompileMRI.jl/releases/tag/v3.5.6>)
- MEDI toolbox (release: 15th January 2020) (<http://pre.weill.cornell.edu/mri/pages/qsm.html>)
- FANSI toolbox [v3] (<https://gitlab.com/cmilovic/FANSI-toolbox>)

## QSM reconstruction using Example data

This section describes all the QSM reconstruction settings that were used on the example data. All the methods and algorithm parameters mentioned were already specified in the SEPIA pipeline configuration files (sepia\_<GE|PHILIPS|SIEMENS>\_<Monopolar|Bipolar>\_config.m), which can be found in the sub-directories of the code folder “QSM\_Consensus\_Paper\_Example\_Code/”: “SEPIA\_Pipeline\_FANSI/” and “SEPIA\_Pipeline\_MEDI/”. Here, we provide an overview of the main parameters of each of these pipelines (Tables S1-S4) for the readers’ convenience.

## Step 1: Preparation

- (GE only) Phase data is inverted before QSM recon processing (i.e., phase = - phase) so that paramagnetic susceptibility gives a positive value while diamagnetic susceptibility gives a negative value, same as the data from other vendors. This step was performed with the option provided by SEPIA.
- Brain mask is obtained by using MEDI toolbox implementation of FSL's BET on the 1st echo magnitude image, using default setting -f 0.5 -g 0
- (Bipolar readout data only) Bipolar readout correction based on (Li et al., 2015) using the implementation provided with SEPIA.
- Note that the relevant sequence parameters such as echo time and slice orientation are automatically derived from the data.

## Step 2: Total field estimation and echo combination

Table S1: Algorithm parameters for total field estimation and echo combination.

Parameters	Values	Remark
Echo phase combination	ROMEO total field calculation	(Dymerska et al., 2020)
MCPC-3D-S phase offset correction	On	
Mask for unwrapping	SEPIA mask	FSL's BET mask
Using ROMEO Mask in SEPIA	Off	
Exclude voxel using relative residual with threshold	0.3 (applied on weighting map)	See <a href="https://sepia-documentation.readthedocs.io/en/latest/method/weightings.html">https://sepia-documentation.readthedocs.io/en/latest/method/weightings.html</a>

## Step 3: Background field removal

Table S2: Algorithm parameters for background field removal.

Parameters	Values	Remark
Method	VSHARP	(Li et al., 2011); SEPIA's implementation
Maximum spherical mean value filtering size	12	Unit: voxel
Minimum spherical mean value filtering size	1	Unit: voxel
Remove residual B1 field	No	
Erode brain mask before BFR	1	Unit: voxel
Erode brain mask after BFR	0	

## Step 4: Dipole inversion

We demonstrate the dipole inversion steps with two recommended methods (FANSI and MEDI).

#### Step 4.1 : FANSI dipole inversion

Table S3: Algorithm parameters for dipole field inversion using “SEPIA\_Pipeline\_FANSI” pipeline.

Parameters	Values	Remark
Method	FANSI	(Milovic et al., 2019, 2018)
Iteration tolerance	0.1	
Maximum number of iterations	400	
Gradient L1 penalty, regularisation weight	0.0005	
Gradient consistency weight	0.05	
Fidelity consistency weight	1	
Solver	Non-linear	
Constraint	TV	
Method for regularisation spatially variable weight	Vector field	
Using weak harmonic regularisation	On	
Harmonic constraint weight	150	
Harmonic consistency weight	3	
Reference tissue	Brain mask	

#### Step 4.2: MEDI Dipole inversion

Table S4: Algorithm parameters for dipole field inversion using “SEPIA\_Pipeline\_MEDI” pipeline.

Parameters	Values	Remark
Method	MEDI	(Liu et al., 2011)
Regularisation parameter (lambda)	2000	
Method of data weighting	1	SNR weighting
Percentage of voxels considered to be edges	90	
Array size for zero padding	[0 0 0]	

<b>Performing spherical mean value operator</b>	On	
<b>Radius of the spherical mean value operation</b>	5	Unit: voxel
<b>Performing modal error reduction through iterative tuning (MERIT)</b>	On	
<b>Performing automatic zero reference (MEDI+0)</b>	Off	
<b>Reference tissue</b>	Brain mask	

# Adaptation of the example pipeline to other studies

The provided SEPIA pipeline configuration file (sepia\_<GE|PHILIPS|SIEMENS>\_<Monopolar|Bipolar>\_config.m) can be reused for other studies, assuming the data in these studies have the compatible input directory described in the SEPIA documentation website ([https://sepia-documentation.readthedocs.io/en/latest/getting\\_started/Data-preparation.html](https://sepia-documentation.readthedocs.io/en/latest/getting_started/Data-preparation.html)):

This can be done by updating the “input” variable in the configuration file to the location of the input directory that contains all the essential data in your computer. Alternatively, if a graphical operation is preferred, the SEPIA pipeline configuration files can be imported to the SEPIA’s GUI by using the “Load config” button on the bottom left of the GUI display and then select the configuration .m file. The GUI will then be updated to the specified methods and algorithm parameters according to the text in the configuration file. Readers can then specify the required input and output information on the “I/O” panel on the GUI.

1    Example results

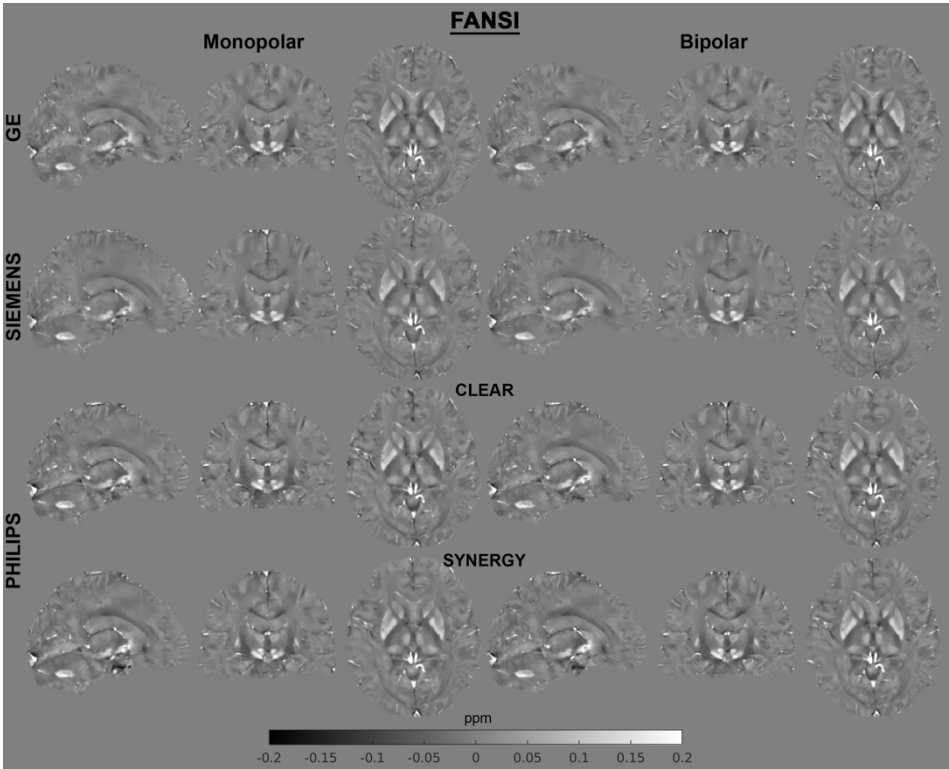
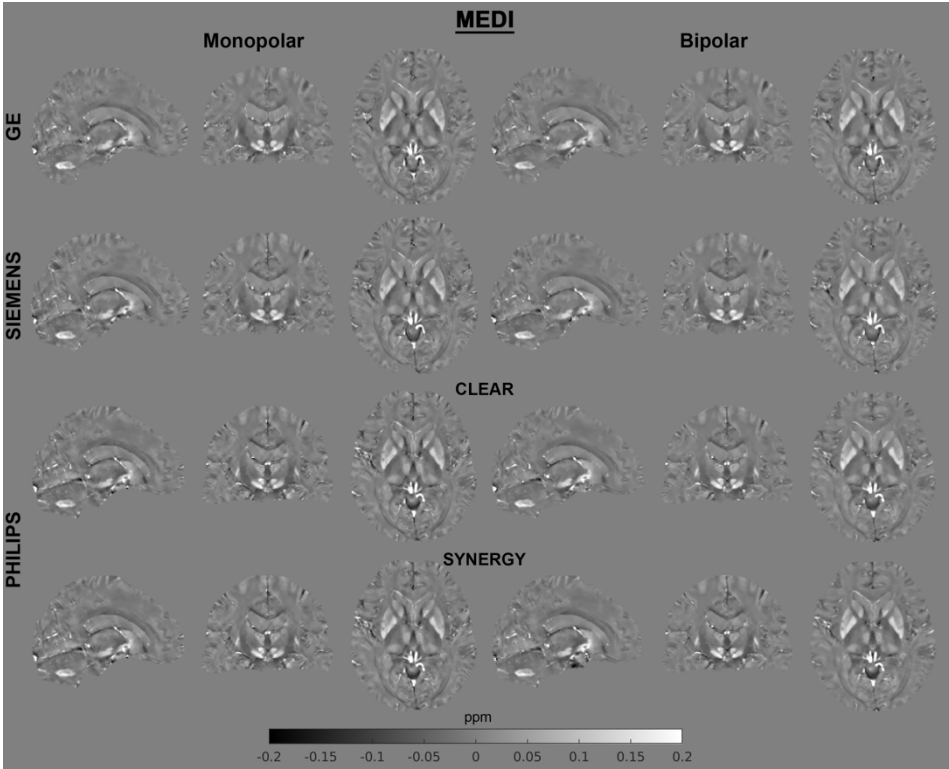


Figure S2: Susceptibility maps derived using the "SEPIA\_Pipeline\_FANSI" processing pipeline.





1 Figure S3: Susceptibility maps derived using the “SEPIA\_Pipeline\_MEDI” processing  
2 pipeline.  
3

## References

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