

p53 in human neural development

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Abstract

Here my abstract will come!

1 Laymen summary

My laymen summary will put here.

2 Introduction

TP53 is a tumor suppressor gene that is mutated or inactivated in around 50% of all human cancers [Bouaoun et al., 2016]. It is called the guardian of the genome [Ananiev et al., 2011]. This is because p53 is rapidly upregulated by stress responses like DNA damage [Zilfou and Lowe, 2009]. Other stressors are hypoxia, oncogene activation and differentiation cues. Upon cellular stress, p53 elicits responses such as DNA repair, apoptosis or cell cycle arrest. P53 is a part of the p53 family that also contains p63 and p73. They work together to evoke the cellular responses mentioned.

The tumor suppressor functions of TP53 have been studied extensively to find therapeutic targets for cancer. However, its role during human brain development is less known. P53 is expressed throughout the whole mouse brain during early embryogenesis (Gottlieb, 1997). Additionally, p53 is highly expressed in human embryonic stem cells and gradually decreases expression during cortical differentiation (van de Leemput et al., 2014). The same process holds for murine cortical development with sporadic p53 activity in the adult mouse brain (Schmid, Lorenz, Hameister, & Montenarh, 1991).

Studies with the p53 knock out (KO) mice surprisingly reported the mice to develop normally, despite being susceptible to tumors (Donehower

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et al., 1992). However, further analyses found that between 8 and 23% of the p53 KO females, developed exencephaly which is an overgrowth of the neural tissue hindering the closure of the neural tube (Armstrong, Kaufman, Harrison, & Clarke, 1995; Sah et al., 1995). Additionally, one case of a KO male with exencephaly has been reported. When neural stem cells (NSC) are isolated at E13.5 from these mice it has been found that p53 influences NSC proliferation and differentiation (Liu et al., 2013). Loss of p53 stimulates NSC proliferation and directs differentiation of NSC towards the neuronal fate and away from becoming astrocytes.

In addition, several studies have shown the importance of maintaining correct levels of p53 protein in the central nervous system (CNS) to avoid inducing aberrant apoptosis and cell cycle arrest [9, 10].

Some proteins in the p53 pathway also influence neural development. P53 response gene MDM2 acts as negative feedback by binding p53 for degradation (Haupt, Maya, Kazanietz, & Oren, 1997). Mdm2 KO mice display extensive p53-dependent apoptosis in the ventricular zone of the cerebral cortex, which induces neuroepithelium degeneration (Xiong, Van Pelt, Elizondo-Fraire, Liu, & Lozano, 2006). This indicates that regulation of p53 during neural development is crucial.

So far, insight in the role of p53 during human neural development is lacking. Studying early human brain development provides a challenge. Human fetal material is rare and often of low quality. Researchers therefore have relied on mouse models. Indeed, the mouse and human brain have apparent differences: mice have a decreased cortical size as well as a general absence of outer radial glia cells, which are important during human cortical development (Hansen, Lui, Parker, & Kriegstein, 2010).

Recent developments in the stem cell biology have opened new windows of studying human brain development. The discovery of cellular reprogramming gives the possibility to derive induced pluripotent stem (iPS) cells derived from human various somatic cells (Takahashi et al., 2007; Takahashi & Yamanaka, 2006). Neural stem cells and neurons can subsequently be derived from these stem cells. Importantly, a protocol has been developed to human iPS cells for the generation of 3D brain organoids (Lancaster et al., 2013; Lancaster & Knoblich, 2014). Brain organoids have been shown to almost perfectly match early human brain development and to express key gene pathways operating during human neurodevelopment (Camp et al., 2015; Kelava & Lancaster, 2016a; Luo et al., 2016).

In this study, the human iPS cell-derived brain organoid system will be used to study the role of p53 during human brain development. Additionally, we will use a 2D system of neural development by using human neuroepithelial-like stem (NES) cells derived from iPS cells. Since there is a trade-off between complexity of a system and homogeneity of results (Kelava & Lancaster, 2016b), a combination of a complex 3D and a more controlled 2D in vitro system will allow for more comprehensive analysis of the p53

knock-down (KD) phenotype. With this system, we believe we can aid the understanding of p53's role during early human brain development.

We found that loss of p53 results in disorganization of the stem cell layer and a delay in generation of TBR1 positive neurons. To further understand p53's role in neural stem cell self-renewal and differentiation we used NES cells. Upon knocking down p53 in NES cells we observed centrosome amplification leading to G2/M-phase arrest, reduced proliferation rate, and chromosomal rearrangements. Gene set enrichment analysis (GSEA) of p53 KD NES cells showed downregulation of DNA damage repair pathways as a possible mechanism for the increase in genomic instability. Interestingly, p53 loss did not impede differentiation of TUJ1 positive neurons. Metabolism result.

3 Materials and Methods

3.1 Samples

cells etc.

3.2 sections

Articles are usually structured into sections, and books into chapters and chapter sections. Sections can be cross referenced, if you `\label` them. For example the Introduction is section 3 in this document. Sub-sections are possible

3.2.1 sub-sub-sections are also possible

subsection without number

4 Documentation

5 Tables

Here are some examples of tables.

Table 1: Orbits of S_n in Ω^3 (with $n > 3$) and in Ω^4 (with $n > 4$)

Orbits in Ω^3		Orbits in Ω^4	
Orbit type	multiplicity	Orbit type	multiplicity
$\{(i, i, i)\}$	1	$\{(i, i, i, i)\}$	1
$\{(i, i, j)\}$	3	$\{(i, i, i, j)\}$	4
$\{(i, j, k)\}$	1	$\{(i, i, j, j)\}$	3
		$\{(i, i, j, k)\}$	6
		$\{(i, j, k, l)\}$	1
Total	5	Total	15

subrepresentation		dim
<i>Diagonal</i>		
1_D	$= \{f \mid f_{ij}=0, f_{ii}=c\}$	1
1_D^\perp	$= \{f \mid f_{ij}=0, \sum_{i=1}^n f_{ii}=0\}$	$n-1$
<i>Off-diagonal</i>		
1_O	$= \{f \mid f_{ii}=0, f_{ij}=c\}$	1
sym^+	$= \{f \mid f_{ij}=\alpha_i+\alpha_j, \sum \alpha_i=0\}$	$n-1$
sym	$= \{f \mid f_{ij}=f_{ji}, \sum_i f_{ij}=\sum_j f_{ij}=0\}$	$\frac{n(n-3)}{2}$
alt^+	$= \{f \mid f_{ij}=\alpha_i-\alpha_j, \sum \alpha_i=0\}$	$n-1$
alt	$= \{f \mid f_{ij}=-f_{ji}, \sum_i f_{ij}=\sum_j f_{ij}=0\}$	$\frac{(n-1)(n-2)}{2}$
Name	Model formula	Conditions
diagonal		
1	$\mu_{ii} = \mu$	
\mathbb{R}^Ω	$\mu_{ii} = \mu + \alpha_i$	
off-diagonal		
Symmetric		
1	$\mu_{ij} = \mu$	
sym^+	$\mu_{ij} = \alpha_i + \alpha_j$	
sym	$\mu_{ij} = \gamma_{ij}$	$\gamma_{ij} = \gamma_{ji}$
Alternating		
alt^+	$\mu_{ij} = \alpha_i - \alpha_j$	
alt	$\mu_{ij} = \gamma_{ij}$	$\gamma_{ij} = -\gamma_{ji}$
Differential Effects		
DE_θ	$\mu_{ij} = \alpha_i \cos \theta + \alpha_j \sin \theta$	$\theta \in [0, \pi)$
DE	$\mu_{ij} = \alpha_i \cos \theta + \alpha_j \sin \theta$	

Nicely placed tables with captions, numbers and labels like table 1, can be produced with `\begin{table}` and `\end{table}`.

6 Lists

Devotees of the bullet point:

- Should use the `\begin{itemize}` command to start a bulleted list.
- Should use the `\item` command to add items to the list.
- Should use the `\end{itemize}` command to end a bulleted list.

Numbered lists can be useful

1. because I say so.
2. because they break up the visual flatness of a page.
3. because they can be nested which is useful for
 - (a) examination questions.
 - (b) algorithms.
 - (c) er.

7 Bibliography and references

Quite nice bibliography creating facilities are available using the BibTeX program. Basically you create a separate file containing all your references, cite them in the document using standard commands and then place a command at the end of you document to create the reference list — only references that you actually cited will appear on this list.

Here are some example citations: [Billon, 2004].

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References

- [Ananiev et al., 2011] Ananiev, J., Tchernev, G., Patterson, J. W., Gulubova, M., and Ganchev, G. (2011). p53 - "The Guardian of Genome". *Acta Medica Bulgarica*, 38(2):72–82.
- [Armesilla-Diaz et al., 2009] Armesilla-Diaz, A., Bragado, P., del Valle, I., Cuevas, E., Lazaro, I., Martin, C., Cigudosa, J. C., and Silva, A. (2009). P53 Regulates the Self-Renewal and Differentiation of Neural Precursors. *Neuroscience*, 158(4):1378–1389.
- [Armstrong et al., 1995] Armstrong, J. F., Kaufman, M. H., Harrison, D. J., and Clarke, A. R. (1995). High-frequency developmental abnormalities in p53-deficient mice. *Current Biology*, 5(8):931–936.
- [Bian et al., 2018] Bian, S., Repic, M., Guo, Z., Kavirayani, A., Burkard, T., Bagley, J. A., Krauditsch, C., and Knoblich, J. A. (2018). Genetically engineered cerebral organoids model brain tumor formation. *Nature Methods*, 15(8):631–639.
- [Billon, 2004] Billon, N. (2004). Roles for p53 and p73 during oligodendrocyte development. *Development*, 131(6):1211–1220.
- [Bouaoun et al., 2016] Bouaoun, L., Sonkin, D., Ardin, M., Hollstein, M., Byrnes, G., Zavadil, J., and Olivier, M. (2016). TP53 Variations in Human Cancers: New Lessons from the IARC TP53 Database and Genomics Data. *Human Mutation*, 37(9):865–876.
- [Bourdon, 2007] Bourdon, J. C. (2007). P53 and Its Isoforms in Cancer. *British Journal of Cancer*, 97(3):277–282.
- [Camp et al., 2015] Camp, J. G., Badsha, F., Florio, M., Kanton, S., Gerber, T., Wilsch-Bräuninger, M., Lewitus, E., Sykes, A., Hevers, W., Lancaster, M., Knoblich, J. A., Lachmann, R., Pääbo, S., Huttner, W. B., and Treutlein, B. (2015). Human cerebral organoids recapitulate gene expression programs of fetal neocortex development. *Proceedings of the National Academy of Sciences*, 112(51):201520760.
- [Checler and Alves Da Costa, 2014] Checler, F. and Alves Da Costa, C. (2014). P53 in neurodegenerative diseases and brain cancers. *Pharmacology and Therapeutics*, 142(1):99–113.

- [Claes et al., 2018] Claes, C., Van den Daele, J., and Verfaillie, C. M. (2018). Generating tissue-resident macrophages from pluripotent stem cells: Lessons learned from microglia. *Cellular Immunology*, 330(January):60–67.
- [Cruz-Acuña et al., 2018] Cruz-Acuña, R., Quirós, M., Huang, S., Siuda, D., Spence, J. R., Nusrat, A., and García, A. J. (2018). PEG-4MAL hydrogels for human organoid generation, culture, and in vivo delivery. *Nature Protocols*, 13(September).
- [da Silva et al., 2018] da Silva, B., Mathew, R. K., Polson, E. S., Williams, J., and Wurdak, H. (2018). Spontaneous Glioblastoma Spheroid Infiltration of Early-Stage Cerebral Organoids Models Brain Tumor Invasion. *SLAS Discovery*.
- [Dell’Anno et al., 2018] Dell’Anno, M. T., Wang, X., Onorati, M., Li, M., Talpo, F., Sekine, Y., Ma, S., Liu, F., Cafferty, W. B. J., Sestan, N., and Strittmatter, S. M. (2018). Human neuroepithelial stem cell regional specificity enables spinal cord repair through a relay circuit. *Nature Communications*, 9(1):3419.
- [Dezonne et al., 2017] Dezonne, R. S., Sartore, R. C., Nascimento, J. M., Saia-Cereda, V. M., Romaão, L. F., Alves-Leon, S. V., De Souza, J. M., Martins-De-Souza, D., Rehen, S. K., and Gomes, F. C. A. (2017). Derivation of Functional Human Astrocytes from Cerebral Organoids. *Scientific Reports*, 7:1–14.
- [Di Lullo and Kriegstein, 2017] Di Lullo, E. and Kriegstein, A. R. (2017). The use of brain organoids to investigate neural development and disease. *Nature Reviews Neuroscience*, 18(10):573–584.
- [Donehower et al., 1992a] Donehower, L. A., Harvey, M., Slagle, B. L., McArthur, M. J., Montgomery, C. A., Butel, J. S., and Bradley, A. (1992a). Mice deficient for p53 are developmentally normal but susceptible to spontaneous tumours. *Nature*, 356(6366):215–221.
- [Donehower et al., 1992b] Donehower, L. A., Harvey, M., Slagle, B. L., McArthur, M. J., Montgomery, C. A., Butel, J. S., and Bradley, A. (1992b). Mice deficient for p53 are developmentally normal but susceptible to spontaneous tumours. *Nature*, 356(6366):215–221.
- [Donthamsetty et al., 2014] Donthamsetty, S., Brahmbhatt, M., Pannu, V., Rida, P. C., Ramarathinam, S., Ogden, A., Cheng, A., Singh, K. K., and Aneja, R. (2014). Mitochondrial genome regulates mitotic fidelity by maintaining centrosomal homeostasis. *Cell Cycle*, 13(13):2056–2063.

- [Duffy et al., 2017a] Duffy, M. J., Synnott, N. C., and Crown, J. (2017a). Mutant p53 as a target for cancer treatment. *European Journal of Cancer*, 83:258–265.
- [Duffy et al., 2017b] Duffy, M. J., Synnott, N. C., and Crown, J. (2017b). Mutant p53 as a target for cancer treatment. *European Journal of Cancer*, 83:258–265.
- [Falk et al., 2012] Falk, A., Koch, P., Kesavan, J., Takashima, Y., Ladewig, J., Alexander, M., Wiskow, O., Taylor, J., Trotter, M., Pollard, S., Smith, A., and Brüstle, O. (2012). Capture of neuroepithelial-like stem cells from pluripotent stem cells provides a versatile system for in vitro production of human neurons. *PLoS ONE*, 7(1):1–13.
- [Floriddia et al., 2012] Floriddia, E. M., Rathore, K. I., Tedeschi, A., Quadrato, G., Wuttke, A., Lueckmann, J.-M., Kigerl, K. A., Popovich, P. G., and Di Giovanni, S. (2012). p53 Regulates the Neuronal Intrinsic and Extrinsic Responses Affecting the Recovery of Motor Function following Spinal Cord Injury. *Journal of Neuroscience*, 32(40):13956–13970.
- [Frese et al., 2017] Frese, C. K., Mikhaylova, M., Stucchi, R., Gautier, V., Liu, Q., Mohammed, S., Heck, A. J., Altelaar, A. F., and Hoogenraad, C. C. (2017). Quantitative Map of Proteome Dynamics during Neuronal Differentiation. *Cell Reports*, 18(6):1527–1542.
- [Fuentes et al., 2016] Fuentes, L., Lebenkoff, S., White, K., Gerdt, C., Hopkins, K., Potter, J. E., Grossman, D., Project, P. E., and Sciences, R. (2016). HHS Public Access. 93(4):292–297.
- [Giandomenico et al., 2018] Giandomenico, S. L., Mierau, S. B., Gibbons, G. M., Wenger, L. M., Masullo, L., Sit, T., Sutcliffe, M., Boulanger, J., Tripodi, M., Derivery, E., Paulsen, O., Lakatos, A., and Lancaster, M. (2018). Cerebral organoids at the air-liquid interface generate diverse nerve tracts with functional output. *bioRxiv*, page 353151.
- [Gil-Perotin, 2006] Gil-Perotin, S. (2006). Loss of p53 Induces Changes in the Behavior of Subventricular Zone Cells: Implication for the Genesis of Glial Tumors. *Journal of Neuroscience*, 26(4):1107–1116.
- [Gil-Perotin et al., 2011] Gil-Perotin, S., Haines, J. D., Kaur, J., Marin-Husstege, M., Spinetta, M. J., Kim, K. H., Duran-Moreno, M., Schallert, T., Zindy, F., Roussel, M. F., Garcia-Verdugo, J. M., and Casaccia, P. (2011). Roles of p53 and p27Kip1 in the regulation of neurogenesis in the murine adult subventricular zone. *European Journal of Neuroscience*, 34(7):1040–1052.

- [Gong et al., 2016a] Gong, L., Pan, X., Chen, H., Rao, L., Zeng, Y., Hang, H., Peng, J., Xiao, L., and Chen, J. (2016a). P53 isoform $\Delta 133p53$ promotes efficiency of induced pluripotent stem cells and ensures genomic integrity during reprogramming. *Scientific Reports*, 6(July):1–8.
- [Gong et al., 2016b] Gong, L., Pan, X., Chen, H., Rao, L., Zeng, Y., Hang, H., Peng, J., Xiao, L., and Chen, J. (2016b). P53 isoform $\Delta 133p53$ promotes efficiency of induced pluripotent stem cells and ensures genomic integrity during reprogramming. *Scientific Reports*, 6(July):1–8.
- [Gottlieb, 1997] Gottlieb, E. (1997). Transgenic mouse model for studying the transcriptional activity of the p53 protein: age- and tissue dependent changes in radiation-induced activation during embryogenesis. *Embo J.*, 16(6):1381–1390.
- [Hackland et al., 2017] Hackland, J. O., Frith, T. J., Thompson, O., Marin Navarro, A., Garcia-Castro, M. I., Unger, C., and Andrews, P. W. (2017). Top-Down Inhibition of BMP Signaling Enables Robust Induction of hPSCs Into Neural Crest in Fully Defined, Xeno-free Conditions. *Stem Cell Reports*, 9(4):1043–1052.
- [Hansen et al., 2010] Hansen, D. V., Lui, J. H., Parker, P. R., and Kriegstein, A. R. (2010). Neurogenic radial glia in the outer subventricular zone of human neocortex. *Nature*, 464(7288):554–561.
- [Haupt et al., 1997] Haupt, Y., Maya, R., Kazaz, A., and Oren, M. (1997). Mdm2 promotes the rapid degradation of p53. *Nature*, 387(6630):296–299.
- [Higuchi et al., 2006] Higuchi, H., Wakita, H., Nagasawa, H., Matsui, Y., Weill, D. F., Wood, B. J., Blundy, J. D., Hess, P. C., Virgo, D., Keppler, H., Dubinsky, E. V., Dardon, a., Chazot, G., Vannucci, R., Prowatke, S., Hametner, K., Gu, D., Petibon, C., Jenner, G. a., Dorfman, a. M., Dingwell, D. B., Vangroos, a. F. K., and Carmicheal, I. S. E. (2006). (30, 31) for basalts analogous to the gabbroic melt of experiment Z10 and combine D. *Handbook of Chemistry and Physics*, 312(June):1650–1653.
- [Hoffman et al., 2017] Hoffman, G. E., Hartley, B. J., Flaherty, E., Ladran, I., Gochman, P., Ruderfer, D. M., Stahl, E. A., Rapoport, J., Sklar, P., and Brennand, K. J. (2017). Transcriptional signatures of schizophrenia in hiPSC-derived NPCs and neurons are concordant with post-mortem adult brains. *Nature Communications*, 8(1).
- [Hollstein et al., 1991] Hollstein, M., Sidransky, D., Vogelstein, B., and Harris, C. C. (1991). P53 Mutations in Human Cancers. 253(5015):49–53.
- [Huang et al., 2012] Huang, T. T., Zou, Y., and Corniola, R. (2012). Oxidative stress and adult neurogenesis-Effects of radiation and superox-

- ide dismutase deficiency. *Seminars in Cell and Developmental Biology*, 23(7):738–744.
- [Hwang et al., 2018] Hwang, L. A., Phang, B. H., Liew, O. W., Iqbal, J., Koh, X. H., Koh, X. Y., Othman, R., Xue, Y., Richards, A. M., Lane, D. P., and Sabapathy, K. (2018). Monoclonal Antibodies against Specific p53 Hotspot Mutants as Potential Tools for Precision Medicine. *Cell Reports*, 22(1):299–312.
- [Karzbrun et al., 2018] Karzbrun, E., Kshirsagar, A., Cohen, S. R., Hanna, J. H., and Reiner, O. (2018). Human brain organoids on a chip reveal the physics of folding. *Nature Physics*, 14(5):515–522.
- [Kelava and Lancaster, 2016a] Kelava, I. and Lancaster, M. A. (2016a). Dishing out mini-brains: Current progress and future prospects in brain organoid research. *Developmental Biology*, 420(2):199–209.
- [Kelava and Lancaster, 2016b] Kelava, I. and Lancaster, M. A. (2016b). Stem Cell Models of Human Brain Development. *Cell Stem Cell*, 18(6):736–748.
- [Khoury and Bourdon, 2010a] Khoury, M. P. and Bourdon, J. C. (2010a). The isoforms of the p53 protein. *Cold Spring Harbor perspectives in biology*, 2(3):1–11.
- [Khoury and Bourdon, 2010b] Khoury, M. P. and Bourdon, J. C. (2010b). The isoforms of the p53 protein. *Cold Spring Harbor perspectives in biology*, 2(3):1–11.
- [Kim et al., 2017] Kim, J., Choi, I., and Lee, Y. (2017). Involvement of Atm and Trp53 in neural cell loss due to Terf2 inactivation during mouse brain development. *Histochemistry and Cell Biology*, 148(5):489–501.
- [Koch et al., 2009] Koch, P., Opitz, T., Steinbeck, J. A., Ladewig, J., and Brustle, O. (2009). A rosette-type, self-renewing human ES cell-derived neural stem cell with potential for in vitro instruction and synaptic integration. *Proceedings of the National Academy of Sciences*, 106(9):3225–3230.
- [Kuijpers and Hoogenraad, 2011] Kuijpers, M. and Hoogenraad, C. C. (2011). Centrosomes, microtubules and neuronal development. *Molecular and Cellular Neuroscience*, 48(4):349–358.
- [Lancaster, 2018a] Lancaster, M. A. (2018a). Crinkle-Cut Brain Organoids. *Cell Stem Cell*, 22(5):616–618.
- [Lancaster, 2018b] Lancaster, M. A. (2018b). Crinkle-Cut Brain Organoids. *Cell Stem Cell*, 22(5):616–618.

- [Lancaster and Knoblich, 2014] Lancaster, M. A. and Knoblich, J. A. (2014). Generation of cerebral organoids from human pluripotent stem cells. *Nature Protocols*, 9(10):2329–2340.
- [Lancaster et al., 2013] Lancaster, M. A., Renner, M., Martin, C. A., Wenzel, D., Bicknell, L. S., Hurles, M. E., Homfray, T., Penninger, J. M., Jackson, A. P., and Knoblich, J. A. (2013). Cerebral organoids model human brain development and microcephaly. *Nature*, 501(7467):373–379.
- [Liu et al., 2013] Liu, H., Jia, D., Li, A., Chau, J., He, D., Ruan, X., Liu, F., Li, J., He, L., and Li, B. (2013). p53 Regulates Neural Stem Cell Proliferation and Differentiation via BMP-Smad1 Signaling and Id1. *Stem Cells and Development*, 22(6):913–927.
- [Liu et al., 2017] Liu, Z., Zhang, C., Skamagki, M., Khodadadi-Jamayran, A., Zhang, W., Kong, D., Chang, C. W., Feng, J., Han, X., Townes, T. M., Li, H., Kim, K., and Zhao, R. (2017). Elevated p53 Activities Restrict Differentiation Potential of MicroRNA-Deficient Pluripotent Stem Cells. *Stem Cell Reports*, 9(5):1604–1617.
- [Lundin et al., 2018] Lundin, A., Delsing, L., Clausen, M., Ricchiuto, P., Sanchez, J., Sabirsh, A., Ding, M., Synnergren, J., Zetterberg, H., Brolén, G., Hicks, R., Herland, A., and Falk, A. (2018). Human iPS-Derived Astroglia from a Stable Neural Precursor State Show Improved Functionality Compared with Conventional Astrocytic Models. *Stem Cell Reports*, 10(3):1030–1045.
- [Luo et al., 2016] Luo, C., Lancaster, M. A., Castanon, R., Nery, J. R., Knoblich, J. A., and Ecker, J. R. (2016). Cerebral Organoids Recapitulate Epigenomic Signatures of the Human Fetal Brain. *Cell Reports*, 17(12):3369–3384.
- [Madhavan et al., 2018] Madhavan, M., Nevin, Z. S., Shick, H. E., Garrison, E., Clarkson-Paredes, C., Karl, M., Clayton, B. L., Factor, D. C., Allan, K. C., Barbar, L., Jain, T., Douvaras, P., Fossati, V., Miller, R. H., and Tesar, P. J. (2018). Induction of myelinating oligodendrocytes in human cortical spheroids. *Nature Methods*.
- [Mansour et al., 2018] Mansour, A. A., Gonçalves, J. T., Bloyd, C. W., Li, H., Fernandes, S., Quang, D., Johnston, S., Parylak, S. L., Jin, X., and Gage, F. H. (2018). An in vivo model of functional and vascularized human brain organoids. *Nature Biotechnology*, 36(5):432–441.
- [Marin Navarro et al., 2018] Marin Navarro, A., Susanto, E., Falk, A., and Wilhelm, M. (2018). Modeling cancer using patient-derived induced pluripotent stem cells to understand development of childhood malignancies. *Cell Death Discovery*, 4(1):7.

- [Medelnik et al., 2018] Medelnik, J. P., Roensch, K., Okawa, S., del Sol, A., Chara, O., Mchedlishvili, L., and Tanaka, E. M. (2018). Signaling-Dependent Control of Apical Membrane Size and Self-Renewal in Rosette-Stage Human Neuroepithelial Stem Cells. *Stem Cell Reports*, 10(6):1751–1765.
- [Meiliana and Wijaya, 2011] Meiliana, A. and Wijaya, A. (2011). Epigenetic Reprogramming Induced Pluripotency. *The Indonesian Biomedical Journal*, 3(2):93.
- [Monzel et al., 2017] Monzel, A. S., Smits, L. M., Hemmer, K., Hachi, S., Moreno, E. L., van Wuellen, T., Jarazo, J., Walter, J., Brüggemann, I., Boussaad, I., Berger, E., Fleming, R. M., Bolognin, S., and Schwamborn, J. C. (2017). Derivation of Human Midbrain-Specific Organoids from Neuroepithelial Stem Cells. *Stem Cell Reports*, 8(5):1144–1154.
- [Muguruma et al., 2015] Muguruma, K., Nishiyama, A., Kawakami, H., Hashimoto, K., and Sasai, Y. (2015). Self-organization of polarized cerebellar tissue in 3D culture of human pluripotent stem cells. *Cell Reports*, 10(4):537–550.
- [Ogawa et al., 2018] Ogawa, J., Pao, G. M., Shokhirev, M. N., and Verma, I. M. (2018). Glioblastoma Model Using Human Cerebral Organoids. *Cell Reports*, pages 1220–1229.
- [Olivier et al., 2010] Olivier, M., Hollstein, M., and Hainaut, P. (2010). TP53 Mutations in Human Cancers: Origins, Consequences, and Clinical Use. *Cold Spring Harbor Perspect Biol*, pages 1–17.
- [Qian et al., 2018] Qian, X., Jacob, F., Song, M. M., Nguyen, H. N., Song, H., and Ming, G. L. (2018). Generation of human brain regionspecific organoids using a miniaturized spinning bioreactor. *Nature Protocols*, 13(3):565–580.
- [Quadrato et al., 2017] Quadrato, G., Nguyen, T., Macosko, E. Z., Sherwood, J. L., Yang, S. M., Berger, D. R., Maria, N., Scholvin, J., Goldman, M., Kinney, J. P., Boyden, E. S., Lichtman, J. W., Williams, Z. M., McCarroll, S. A., and Arlotta, P. (2017). Cell diversity and network dynamics in photosensitive human brain organoids. *Nature*, 545(7652):48–53.
- [Renner et al., 2017] Renner, M., Lancaster, M. A., Bian, S., Choi, H., Ku, T., Peer, A., Chung, K., and Knoblich, J. A. (2017). Selforganized developmental patterning and differentiation in cerebral organoids. *The EMBO Journal*, 36(10):1316–1329.
- [Robles and Harris, 2010] Robles, A. I. and Harris, C. C. (2010). Clinical outcomes and correlates of TP53 mutations and cancer. *Cold Spring Harbor perspectives in biology*, 2(3):a001016–a001016.

- [Sabapathy, 2015] Sabapathy, K. (2015). The Contrived Mutant p53 Oncogene Beyond Loss of Functions. *Frontiers in Oncology*, 5(December):1–8.
- [Sabapathy, 2016] Sabapathy, K. (2016). p73: a Positive or Negative Regulator of Angiogenesis, or Both? *Molecular and Cellular Biology*, 36(6):848–854.
- [Sabapathy and Lane, 2018] Sabapathy, K. and Lane, D. P. (2018). Therapeutic targeting of p53: All mutants are equal, but some mutants are more equal than others. *Nature Reviews Clinical Oncology*, 15(1):13–30.
- [Sah et al., 1995] Sah, V. P., Attardi, L. D., Mulligan, G. J., Williams, B. O., Bronson, R. T., and Jacks, T. (1995). A subset of p53-deficient embryos exhibit exencephaly. *Nature Genetics*, 10(2):175–180.
- [Schmid et al., 1991] Schmid, P., Lorenz, A., Hameister, H., and Montenarh, M. (1991). Expression of p53 during mouse embryogenesis. *Development (Cambridge, England)*, 113(3):857–65.
- [Shahsavani et al., 2017a] Shahsavani, M., Pronk, R. J., Falk, R., Lam, M., Moslem, M., Linker, S. B., Salma, J., Day, K., Schuster, J., Anderlid, B.-M., Dahl, N., Gage, F. H., and Falk, A. (2017a). An in vitro model of lissencephaly: expanding the role of DCX during neurogenesis. *Molecular Psychiatry*, (July):1–11.
- [Shahsavani et al., 2017b] Shahsavani, M., Pronk, R. J., Falk, R., Lam, M., Moslem, M., Linker, S. B., Salma, J., Day, K., Schuster, J., Anderlid, B.-M., Dahl, N., Gage, F. H., and Falk, A. (2017b). An in vitro model of lissencephaly: expanding the role of DCX during neurogenesis. *Molecular Psychiatry*, (July):1–11.
- [Shin et al., 2013] Shin, M. H., He, Y., and Huang, J. (2013). Embryonic stem cells shed new light on the developmental roles of p53. *Cell and Bioscience*, 3(1):1.
- [Skene et al., 2018] Skene, N. G., Bryois, J., Bakken, T. E., Breen, G., Crowley, J. J., Gaspar, H. A., Giusti-Rodriguez, P., Hodge, R. D., Miller, J. A., Muñoz-Manchado, A. B., O’Donovan, M. C., Owen, M. J., Pardiñas, A. F., Ryge, J., Walters, J. T., Linnarsson, S., Lein, E. S., Sullivan, P. F., and Hjerling-Leffler, J. (2018). Genetic identification of brain cell types underlying schizophrenia. *Nature Genetics*, 50(6):825–833.
- [Steinemann et al., 2013] Steinemann, D., Göhring, G., and Schlegelberger, B. (2013). Genetic instability of modified stem cells - a first step towards malignant transformation? *American journal of stem cells*, 2(1):39–51.

- [Tailor et al., 2013a] Tailor, J., Kittappa, R., Leto, K., Gates, M., Borel, M., Paulsen, O., Spitzer, S., Karadottir, R. T., Rossi, F., Falk, A., and Smith, A. (2013a). Stem Cells Expanded from the Human Embryonic Hindbrain Stably Retain Regional Specification and High Neurogenic Potency. *Journal of Neuroscience*, 33(30):12407–12422.
- [Tailor et al., 2013b] Tailor, J., Kittappa, R., Leto, K., Gates, M., Borel, M., Paulsen, O., Spitzer, S., Karadottir, R. T., Rossi, F., Falk, A., and Smith, A. (2013b). Stem Cells Expanded from the Human Embryonic Hindbrain Stably Retain Regional Specification and High Neurogenic Potency. *Journal of Neuroscience*, 33(30):12407–12422.
- [Takahashi and Yamanaka, 2006] Takahashi, K. and Yamanaka, S. (2006). Induction of Pluripotent Stem Cells from Mouse Embryonic and Adult Fibroblast Cultures by Defined Factors. *Cell*, 126(4):663–676.
- [Tanabe et al., 2013a] Tanabe, K., Nakamura, M., Narita, M., Takahashi, K., and Yamanaka, S. (2013a). Maturation, not initiation, is the major roadblock during reprogramming toward pluripotency from human fibroblasts. *Proceedings of the National Academy of Sciences*, 110(30):12172–12179.
- [Tanabe et al., 2013b] Tanabe, K., Nakamura, M., Narita, M., Takahashi, K., and Yamanaka, S. (2013b). Maturation, not initiation, is the major roadblock during reprogramming toward pluripotency from human fibroblasts. *Proceedings of the National Academy of Sciences*, 110(30):12172–12179.
- [Tomasini et al., 2008] Tomasini, R., Tsuchihara, K., Wilhelm, M., Fujitani, M., Rufini, A., Cheung, C. C., Khan, F., Itie-Youten, A., Wakeham, A., Tsao, M. S., Iovanna, J. L., Squire, J., Jurisica, I., Kaplan, D., Melino, G., Jurisicova, A., and Mak, T. W. (2008). TAp73 knockout shows genomic instability with infertility and tumor suppressor functions. *Genes and Development*, 22(19):2677–2691.
- [Tremblay et al., 2016] Tremblay, R., Lee, S., and Rudy, B. (2016). GABAergic Interneurons in the Neocortex: From Cellular Properties to Circuits. *Neuron*, 91(2):260–292.
- [Uhlin et al., 2017] Uhlin, E., Rönnholm, H., Day, K., Kele, M., Tamimies, K., Bölte, S., and Falk, A. (2017). Derivation of human iPS cell lines from monozygotic twins in defined and xeno free conditions. *Stem Cell Research*, 18:22–25.
- [van de Leemput et al., 2014] van de Leemput, J., Boles, N. C., Kiehl, T. R., Corneo, B., Lederman, P., Menon, V., Lee, C., Martinez, R. A., Levi, B. P., Thompson, C. L., Yao, S., Kaykas, A., Temple, S., and Fasano,

- C. A. (2014). CORTECON: A temporal transcriptome analysis of in vitro human cerebral cortex development from human embryonic stem cells. *Neuron*, 83(1):51–68.
- [Vigilante et al., 2018] Vigilante, A., Laddach, A., Moens, N., Meleckyte, R., Vickers, A., Wiseman, E., Tewary, M., and Zandstra, P. (2018). 1,2,4,7,8
- [Wazen et al., 2014] Wazen, R. M., Kuroda, S., Nishio, C., Sellin, K., Brunski, J. B., and Nanci, A. (2014). NIH Public Access. 8(9):1385–1395.
- [Xiong et al., 2006] Xiong, S., Van Pelt, C. S., Elizondo-Fraire, A. C., Liu, G., and Lozano, G. (2006). Synergistic roles of Mdm2 and Mdm4 for p53 inhibition in central nervous system development. *Proceedings of the National Academy of Sciences*, 103(9):3226–3231.
- [Zappone et al., 2000] Zappone, M. V., Galli, R., Catena, R., Meani, N., De Biasi, S., Mattei, E., Tiveron, C., Vescovi, a. L., Lovell-Badge, R., Ottolenghi, S., and Nicolis, S. K. (2000). Sox2 regulatory sequences direct expression of a (beta)-geo transgene to telencephalic neural stem cells and precursors of the mouse embryo, revealing regionalization of gene expression in CNS stem cells. *Development*, 127(11):2367–2382.
- [Zheng et al., 2016] Zheng, X., Boyer, L., Jin, M., Mertens, J., Kim, Y., Ma, L., Ma, L., Hamm, M., Gage, F. H., and Hunter, T. (2016). Metabolic reprogramming during neuronal differentiation from aerobic glycolysis to neuronal oxidative phosphorylation. *eLife*, 5(JUN2016).
- [Zhou et al., 2017a] Zhou, R., Xu, A., Gingold, J., Strong, L. C., Zhao, R., and Lee, D. F. (2017a). LiFraumeni Syndrome Disease Model: A Platform to Develop Precision Cancer Therapy Targeting Oncogenic p53. *Trends in Pharmacological Sciences*, 38(10):908–927.
- [Zhou et al., 2017b] Zhou, R., Xu, A., Gingold, J., Strong, L. C., Zhao, R., and Lee, D. F. (2017b). LiFraumeni Syndrome Disease Model: A Platform to Develop Precision Cancer Therapy Targeting Oncogenic p53. *Trends in Pharmacological Sciences*, 38(10):908–927.
- [Zilfou and Lowe, 2009] Zilfou, J. T. and Lowe, S. W. (2009). Tumor suppressive functions of p53. *Cold Spring Harbor perspectives in biology*, 1(5):1–13.
- [Zinin et al., 2014] Zinin, N., Adameyko, I., Wilhelm, M., Fritz, N., Uhlén, P., Ernfors, P., and Henriksson, M. A. (2014). MYC proteins promote neuronal differentiation by controlling the mode of progenitor cell division. *EMBO Reports*, 15(4):383–391.