Logistic Regression from Scratch

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Notebook for the Logistic Regression Chapter

First, load the package and plasma data set.

```
library(HSAUR)

## Loading required package: tools
data(plasma)
```

Logistic Regression using R

Use R's glm() function first as our ground truth.

```
set.seed(1234)
i <- sample(1:nrow(plasma), 0.75*nrow(plasma), replace=FALSE)
train <- plasma[i,]
test <- plasma[-i,]
glm1 <- glm(ESR~fibrinogen, data=train, family=binomial)
probs <- predict(glm1, newdata=test, type="response")
pred <- ifelse(probs> 0.5, 2, 1)
acc <- mean(pred == as.integer(test$ESR))
summary(glm1)</pre>
```

```
##
## glm(formula = ESR ~ fibrinogen, family = binomial, data = train)
##
## Deviance Residuals:
      Min 1Q
                   Median
                                 3Q
                                         Max
## -0.9852 -0.7375 -0.5074 -0.1920
                                      2.2554
##
## Coefficients:
              Estimate Std. Error z value Pr(>|z|)
## (Intercept) -5.6141
                          2.6591 -2.111
                                           0.0347 *
               1.5084
                          0.8543
                                  1.766
                                           0.0775 .
## fibrinogen
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for binomial family taken to be 1)
##
      Null deviance: 26.992 on 23 degrees of freedom
## Residual deviance: 22.716 on 22 degrees of freedom
## AIC: 26.716
##
## Number of Fisher Scoring iterations: 4
```

Logistic Regression from Scratch

First we need to define the sigmoid function.

```
# function to return a vector of sigmoid values from an input matrix
sigmoid <- function(z){
   1.0 / (1+exp(-z))
}
# set up weight vector, label vector, and data matrix
weights <- c(1, 1)
data_matrix <- cbind(rep(1, nrow(train)), train$fibrinogen)
labels <- as.integer(train$ESR) - 1</pre>
```

Then we need code for gradient descent. The algorithm used here first starts with all weights = 1, then iterates. Notice we get the same weights (coefficients) as R gave us, but it took a lot longer.

```
weights <- c(1, 1) # repeat this for rerunning the block
learning_rate <- 0.001
for (i in 1:500000){
   prob_vector <- sigmoid(data_matrix %*% weights)
   error <- labels - prob_vector
   weights <- weights + learning_rate * t(data_matrix) %*% error
}
weights</pre>
### [,1]
```

```
## [,1]
## [1,] -5.614088
## [2,] 1.508385
```

Predict with the weights we generated

```
# predict with our weights
test_matrix <- cbind(rep(1, nrow(test)), test$fibrinogen)
test_labels <- as.integer(test$ESR) - 1
predicted <- test_matrix %*% weights
probabilities <- exp(predicted) / (1 + exp(predicted))
predictions <- ifelse(probabilities > 0.5, 1, 0)
mean(predictions == test_labels)
```

```
## [1] 1
```

Visualization that the log odds is a line.

```
plasma_log_odds <- cbind(rep(1, 32), plasma$fibrinogen) %*% weights
plot(plasma$fibrinogen, plasma_log_odds, col=plasma$ESR)
abline(weights[1], weights[2])</pre>
```

