While you are waiting for the workshop to begin, please launch Spyder.

All code and slides covered today are on my Github: https://github.com/atchen/python_workshop_part2

Introduction to Python for Biological Data Analysis: Part 2

Ann Chen PhD student in BME August 8, 2018

Acknowledgements



Justin Bois
Lecturer, Division of Biology
and Biological Engineering at Caltech

This workshop is an extremely condensed version of the various Python courses he teaches.

I highly recommend visiting his website (bois.caltech.edu) and checking out BE/Bi/NB 203 and BE/Bi 103.

Most lessons are open-access.

Review of part I

Data type	Definition	Called by	Mutable?
integer	any whole number	an integer	no
float	any real, non-whole number	a float	no
string	a sequence of characters	single, double, or triple quotes	no
list	a sequence of objects	[]	yes
tuple	a sequence of objects	()	no



Review of part I

```
my list = [1, 3.49, 'apple', (1, 'orange')]
print('the type of my list is', type(my list))
print('the type of my list[0] is', type(my list[0]))
print('the type of my list[1] is', type(my list[1]))
print('the type of my_list[2] is', type(my_list[2]))
print('the type of my list[3] is', type(my list[3]))
#lists are mutable
my list[0] = 5
print(my_list)
#tuples are immutable
my tuple = (1, 2)
my tuple[0] = 2 #this will result in an error
my list[3][0] = 2 #this will also result in an error
```

Review of part I

```
def append_a(test_list):
    '''
    Adds the str 'a' after each element in a user-defined list.
    ex: input = [1, 2]. output = [1, 'a', 2, 'a']
    new_list = []
    for i in test_list:
        new_list.append(i)
        new_list.append('a')
    return new_list

print(append_a(my_list))
```

Today's agenda

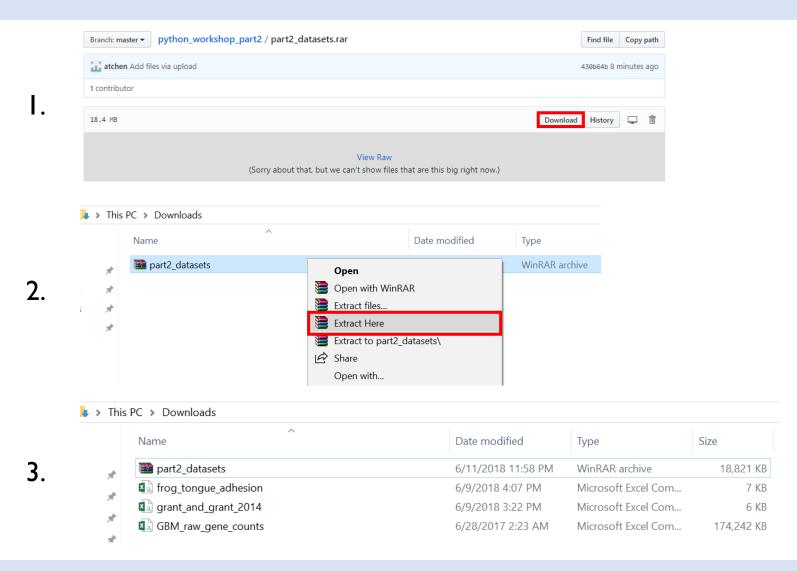
Today, we will use what we learned to analyze datasets from published articles using the Pandas package.

In addition, we will go over how to plot our data using two different packages: seaborn and matplotlib.

We will be covering three datasets from Grant PR, et al, Kleinteich T, et al., and Darmanis S, et al., respectively.



Download and extract files from Github





Loading a CSV file

Download the file grant_and_grant_2014.csv and load it

```
import pandas as pd
import numpy as np
import matplotlib.pyplot as plt

#use pd.read_csv() to read in the data and store in a DataFrame
df = pd.read_csv('C:/Users/Ann/Downloads/grant_and_grant_2014.csv', comment='#')

df.head()
```

The data is stored in a **dataframe**, a data type that Panda uses for convenient data analysis.

This dataset investigates the heritability of beak depth. It includes the maternal beak depth, paternal beak depth, and mean offspring beak depth.

Dataframe columns

To access a column in a dataframe, use the following syntax

```
#slicing a column out of a dataframe by using the column name
df['Average offspring beak depth (mm)']
```

```
In [13]: df.head()
Out[13]:
   Average offspring beak depth (mm) Paternal beak depth (mm) \
                                10.70
                                                          10.90
                                 9.78
                                                          10.70
                                 9.48
                                                          10.70
3
                                 9.60
                                                          10.70
                                10.27
                                                           9.85
   Maternal beak depth (mm)
0
                         9.3
1
                        8.4
                         8.1
                        9.8
4
                       10.4
```

```
In [14]: df['Average offspring beak depth (mm)']
Out[14]:
0     10.70
1     9.78
2     9.48
3     9.60
4     10.27
```

Slicing dataframes with Booleans

Let's say we only want rows with average offspring beak depth ≥ 11 mm. Pandas dataframes can be conveniently sliced with Booleans.

```
#only want indices with offspring beak depth >= to 11 mm
inds = df['Average offspring beak depth (mm)'] >= 11
inds
```

inds is an array of Booleans that is the same length of the dataframe. It is True when the condition is met and False if it is not.

```
#slice out rows we want
df_big_offspring_bd = df.loc[inds]
df_big_offspring_bd
```

Indexing of rows is preserved.



Indexing with loc

Using .loc allows us to index by row. We put an array of Booleans and get back a dataframe containing rows associated with True in the arrays of Booleans.

Notice, the original indexing of rows is preserved. For example, there is no index 2.

```
#this will result in an error
df_big_offspring_bd.loc[2]
```

To get around this, use .iloc which gives indexing with sequential integers.

```
#this will return the third row
df_big_offspring_bd.iloc[2]
```



Renaming columns

Pandas has a nice method for renaming column headers. Let's rename the columns to

```
'Average offspring beak depth (mm)' \rightarrow 'avg_offspring_bd' 
 'Paternal beak depth (mm) \rightarrow 'paternal_bd' 
 'Maternal beak depth (mm) \rightarrow 'maternal_bd'
```

NumPy arrays from dataframe columns

```
offspring_bd = df['avg_offspring_bd'].values
paternal_bd = df['paternal_bd'].values
maternal_bd = df['maternal_bd'].values
```

Now the values of the column are stored in a NumPy array (uses same notation for indexing as a list)

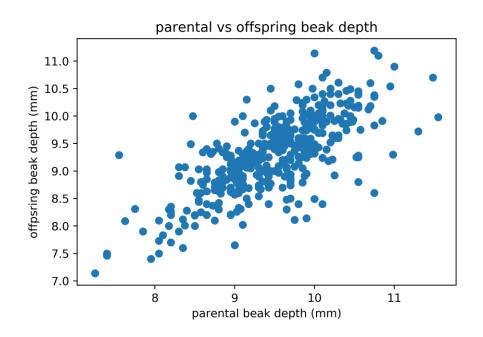
Let's calculate the mean parental beak depth. Conveniently, we don't need to use a for loop for each parent pair.

```
parental_bd = (paternal_bd + maternal_bd) / 2
```



Scatterplots

```
#plot mean parental beak depth vs mean offspring beak depth
plt.scatter(parental_bd, offspring_bd)
plt.xlabel('parental beak depth (mm)')
plt.ylabel('offpsring beak depth (mm)')
plt.title('parental vs offspring beak depth')
plt.savefig('scatter.png', dpi=500)
```



Linear regression with SciPy

Let's perform a linear regression to get the slope and intercept of the data. scipy.optimize.curve_fit() takes a set of data and returns the parameters of the model function that best describes the data.

scipy.optimize.curve_fit(model function, x-data, y-data, guess for parameters). Read the documentation for more information about this function.



Linear regression with SciPy

scipy.optimize.curve_fit() returns two arrays. The first contains the optimal set of parameters for the curve fit. The second is a 2D array with the covariance of the parameters.

```
#parse the results
slope, intercept = popt

#print the results
print('slope =', slope)
print('intercept = ', intercept, 'mm')
```

```
slope = 0.722905188545
intercept = 2.44841830979 mm
```

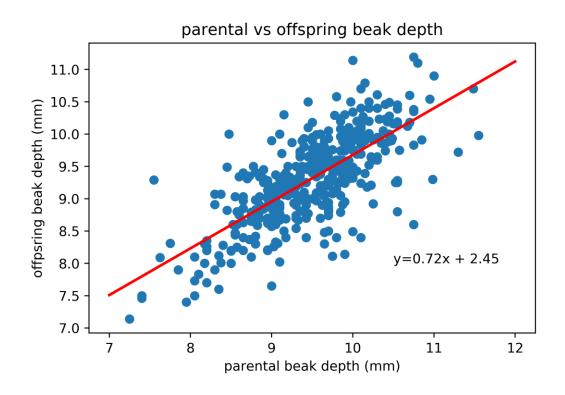


Scatterplot with line of best fit

```
#define range for line
x = np.array([7, 12])
y = linear_fun(x, slope, intercept)

plt.scatter(parental_bd, offspring_bd)
plt.plot(x, y, color='red', linewidth=2)
plt.xlabel('parental beak depth (mm)')
plt.ylabel('offpsring beak depth (mm)')
plt.title('parental vs offspring beak depth')
plt.text(10.5, 8, 'y=%.2fx + %.2f' %(slope, intercept), fontsize=10)
plt.savefig('scatter_w_line.png', dpi=500)
```

Scatterplot with line of best fit



Making a dataframe

```
#use a dictionary to make a dataframe
data_dictionary = {'A': [1, 2, 3], 'B': [4, 5, 6], 'C': [7, 8, 9]}
df = pd.DataFrame(data=data_dictionary, index=[49, 48, 1])
```

More practice with loc and iloc:

```
#returns the second row
df.iloc[1]

#returns the row with the index 1 (third row in this dataframe)
df.loc[1]
```

Frog tongue adhesion

In this dataset, the authors looked at the adhesive strength of the tongue for South American horned frogs. Each row is an experiment where they recorded different metrics.

```
#read dataset and store in dataframe
df = pd.read_csv('C:/Users/Ann/Downloads/frog_tongue_adhesion.csv', comment='#')
df.head()
```

	date	ID	trial number	impact force (m N)	impact time (ms)	impact force / body weight	adhesive force (mN)	time frog pulls on target (ms)	adhesive force / body weight	adhesive impulse (N-s)	total contact area (mm2)	contact area without mucus (mm2)	contact area with mucus / contact area without mucus	contact pressure (Pa)	adhesive strength (Pa)
0	2013_ 02_26	I	3	1205	46	1.95	-785	884	1.27	-0.290	387	70	0.82	3117	-2030
ı	2013_ 02_26	I	4	2527	44	4.08	-983	248	1.59	-0.181	101	94	0.07	24923	-9695
2	2013_ 03_01	-	1	1745	34	2.82	-850	211	1.37	-0.157	83	79	0.05	21020	-10239
3	2013_ 03_01	I	2	1556	41	2.51	-455	1025	0.74	-0.170	330	158	0.52	4718	-1381
4	2013_ 03_01	I	3	493	36	0.80	-974	499	1.57	-0.423	245	216	0.12	2012	-3975

Data extraction

Let's extraction the experiment with index 42.

```
#extract experiment with index 42
df.loc[42]
```

We can also use Boolean slicing to extract data.

Let's say we want to look at trial number 3 on May 27, 2013 of frog III. This is a more common and meaningful case.

```
#set up boolean slicing
date = df['date'] == '2013_05_27'
trial = df['trial number'] == 3
ID = df['ID'] == 'III'

#slice out the row
df_slice = df.loc[date & trial & ID]
```

Computing with dataframes and inserting columns

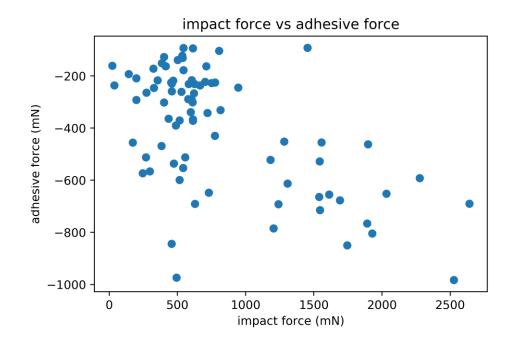
Let's make a new column with impact force in Newtons. We can divide the impact force (mN) column elementwise by 1000, just as with arrays.

```
#add a new columns with impact force in units of newtons
df['impact force (N)'] = df['impact force (mN)'] / 1000
df.head()
```



Plotting how impact force correlates with other metrics

```
plt.scatter(df['impact force (mN)'], df['adhesive force (mN)'])
plt.xlabel('impact force (mN)')
plt.ylabel('adhesive force (mN)')
plt.title('impact force vs adhesive force')
plt.savefig('frog_scatter.png', dpi=500)
```



Making subplots

Let's say we want to plot the adhesive force, total contact area, impact time, and contact pressure against impact force.

```
# set figure = fig and subplots = ax
fig, ax = plt.subplots(4, sharex=True)
plt.xlabel('impact force (mN)')
ax[0].scatter(df['impact force (mN)'], df['impact time (ms)'])
ax[1].scatter(df['impact force (mN)'], df['adhesive force (mN)'])
ax[2].scatter(df['impact force (mN)'], df['total contact area (mm2)'])
ax[3].scatter(df['impact force (mN)'], df['contact pressure (Pa)'])
fig.savefig('frog_subplots.png',dpi=500)
```

Making subplots

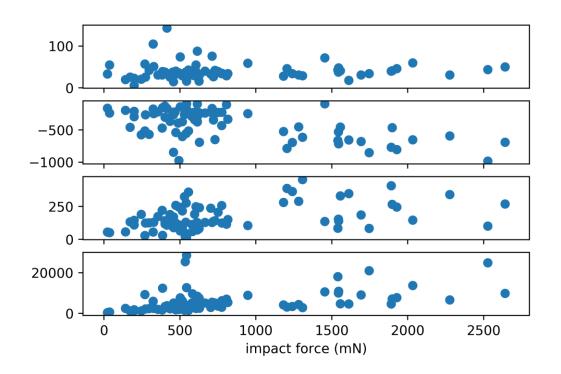


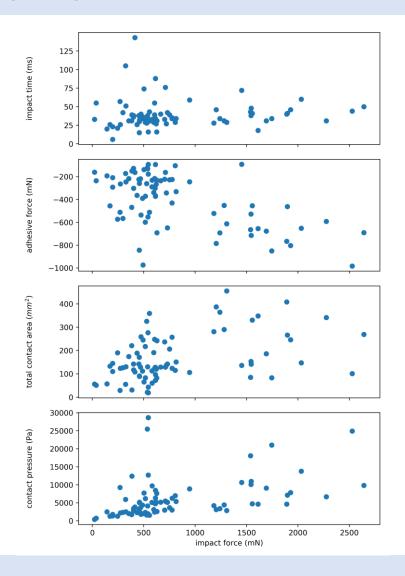
Figure seems a bit stretched out, and there are no y-axis labels.



Subplot parameters

```
fig, ax = plt.subplots(4, sharex=True, figsize=(7,10))
plt.xlabel('impact force (mN)')
ax[0].scatter(df['impact force (mN)'], df['impact time (ms)'])
ax[0].set ylabel('impact time (ms)')
ax[0].get yaxis().set label coords(-0.15, 0.5)
ax[1].scatter(df['impact force (mN)'], df['adhesive force (mN)'])
ax[1].set ylabel('adhesive force (mN)')
ax[1].get yaxis().set label coords(-0.15, 0.5)
ax[2].scatter(df['impact force (mN)'], df['total contact area (mm2)'])
ax[2].set ylabel('total contact area ($mm^2$)')
ax[2].get yaxis().set label coords(-0.15, 0.5)
ax[3].scatter(df['impact force (mN)'], df['contact pressure (Pa)'])
ax[3].set ylabel('contact pressure (Pa)')
ax[3].get yaxis().set label coords(-0.15, 0.5)
plt.tight layout()
fig.savefig('frog subplots.png',dpi=500)
```

Subplot parameters



Summary of what was changed:

- I. Set y-axis labels for subplots
- 2. Change figure size
- 3. Align y-axis labels for subplots
- 4. Improve spacing with plt.tightlayout()



enumerate() as a tool for plotting

It allows us to loop over something and have an automatic counter.

Subplots with enumerate()

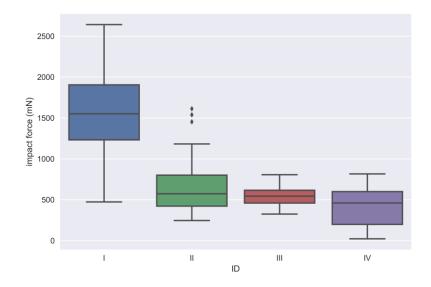
```
#same as before
fig, ax = plt.subplots(4, sharex=True, figsize=(7,10))
plt.xlabel('impact force (mN)')
#instead of typing out each subplot individually, use a for loop
for i,e in enumerate(subplot list):
      ax[i].scatter(df['impact force (mN)'], df[e])
      ax[i].set ylabel(e)
      ax[i].get yaxis().set label coords(-0.15, 0.5)
plt.tight layout()
plt.xlabel('impact force (mN)')
fig.savefig('frog subplots looped.png',dpi=500)
ax[0].scatter(df['impact force (mN)'], df['impact time (ms)'])
ax[0].set ylabel('impact time (ms)')
ax[0].get yaxis().set label coords(-0.15, 0.5)
ax[1].scatter(df['impact force (mN)'], df['adhesive force (mN)'])
                                                                                for i,e in enumerate(subplot_list):
ax[1].set ylabel('adhesive force (mN)')
                                                                                   ax[i].scatter(df['impact force (mN)'], df[e])
ax[1].get yaxis().set label coords(-0.15, 0.5)
                                                                                   ax[i].set ylabel(e)
                                                                                   ax[i].get yaxis().set label coords(-0.15, 0.5)
ax[2].scatter(df['impact force (mN)'], df['total contact area (mm2)'])
ax[2].set ylabel('total contact area ($mm^2$)')
ax[2].get_yaxis().set_label_coords(-0.15, 0.5)
ax[3].scatter(df['impact force (mN)'], df['contact pressure (Pa)'])
ax[3].set ylabel('contact pressure (Pa)')
ax[3].get_yaxis().set_label_coords(-0.15, 0.5)
```

Seaborn: a high level plotting package for Matplotlib

Let's make a box plot of the impact force sorted by frog ID.

```
import seaborn as sns

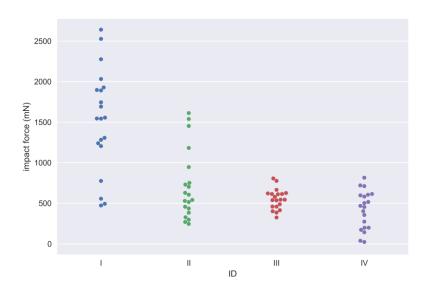
#box plot
sns.boxplot(data=df, x='ID', y='impact force (mN)')
plt.savefig('boxplot.png', dpi=500)
```



Seaborn: a high level plotting package for Matplotlib

Now, let's make a beeswarm plot of the impact force sorted by frog ID.

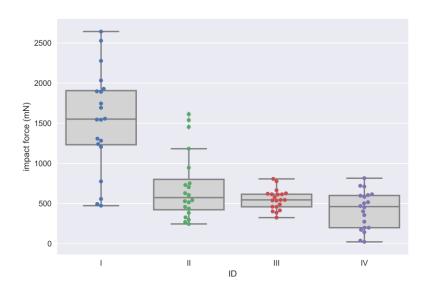
```
#beeswarm plot
sns.swarmplot(data=df, x='ID', y='impact force (mN)')
plt.savefig('beeswarm.png', dpi=500)
```



Seaborn: a high level plotting package for Matplotlib

Lastly, let's overlay the boxplot and the beeswarm plot.

```
#overlay box plot and beeswarm plot
sns.boxplot(data=df, x='ID', y='impact force (mN)', color='lightgray')
sns.swarmplot(data=df, x='ID', y='impact force (mN)')
plt.savefig('boxbeeplot.png', dpi=500)
```



Working with gene expression datasets

Pandas is useful for analyzing extremely large datasets — like single-cell RNA-seq data.

Load in the gene expression matrix. In this case, it is space-delimited. Be patient. It will take a few seconds to load.

```
#read in the scrna-seq dataset. this will take a few seconds
df = pd.read_csv('C:/Users/Ann/Downloads/GBM_raw_gene_counts.csv', sep=' ')
```

This dataset is 25,000x larger than the other datasets we've been using!

It is a 23386 \times 3589 dataframe. Each row is a gene, and each column is a cell. so this dataset contains information for 23386 genes in 3589 cells.

Non-integer index column

```
#let's look at the index column
df.index
```

For this dataset, the index column contains the gene names

```
#df.loc must contain a gene name
df.loc['A2MP1']
```

If you want an integer index column, can use the function df.reset_index()
or set index_col=False in read_csv().



Checking if a gene is in the dataset

```
#can do this manually
'TP53' in df.index
```

Will return True if in the gene is in the dataset and False if it is not.

What if we have a list of genes we want to check? We can write a function.

```
def gene_in_df(gene_list):
    """
    This function takes in a list of gene names and
    returns the names of genes that are not in the dataset.
    """
    genes_in_df = []
    for i in gene_list:
        if i in df.index:
            genes_in_df.append(i)

    print('Gene(s) not in dataset', set(gene_list) - set(genes_in_df))
    return set(gene_list) - set(genes_in_df)
```

Checking if a gene is in the dataset

Let's test the function out.

Gene expression correlation using Pearsonr

```
scipy.stats.pearsonr(x,y)
```

Calculate a Pearson correlation coefficient and its corresponding p-value

The Pearson correlation coefficient measures the linear relationship between two datasets. Coefficient will vary between -1 and +1, with 0 implying no correlation and -1/+1 implying an exact linear relationship.

```
#measure the correlation between TP53 and DERL1
coef, pval = pearsonr(df.loc['TP53'], df.loc['DERL1'])
print('the coefficient is %0.3f' %coef)
print('the p-value is %0.3f' %pval)
```



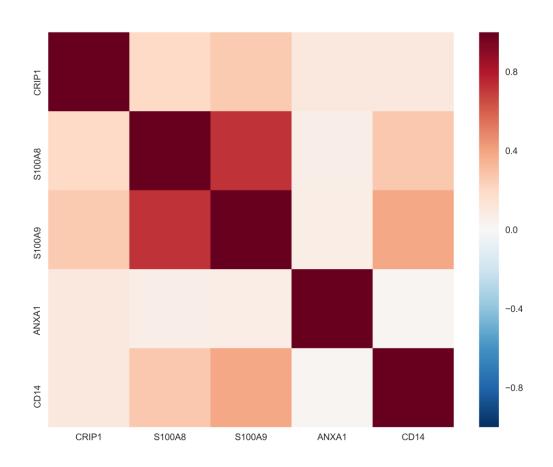
Gene correlation matrix

pandas.DataFrame.corr

Computes pairwise correlation of columns. Default is Pearsonr correlation.

```
gene list = ['CRIP1', 'S100A8', 'S100A9', 'ANXA1', 'CD14']
#make a new dataframe with your genes of interest
coef df = df.loc[gene list]
coef df.head()
#transpose the dataframe first so genes are in columns
#then calculate pairwise correlation
corr = coef df.transpose().corr()
#set color palette where low coefs are blue and high coefs are red
#center the color bar at 0
plt.subplots(figsize=(10, 8))
sns.heatmap(corr, cmap='RdBu r', center=0)
plt.savefig('correlation_heatmap.png', dpi=500)
```

Gene correlation matrix





Fraction of cells expressing a gene

```
gene list = ['TMEM119', 'P2RY12', 'GPR34', 'OLFML3', 'SLC2A5', 'SALL1',
'ADORA3']
def gene_frac_df(gene list):
   This function takes in a list of gene names and returns the fraction of
    cells that express each gene.
   #list to store the fraction for each gene
   gene frac list = []
   for i in gene list:
        if i in df.index:
            #if the gene is in the dataset
            #sum the number of cells that have 0 expression
            gene 0 = (df.loc[i]==0).sum()
            #(total number of cells - cells with 0 expression) / total
            gene frac = (len(df.columns) - gene 0) / len(df.columns)
            #add the fraction to the gene frac list
            gene frac list.append(gene frac)
        else:
            #if the gene is not in the dataset, append 0 to the list
            gene frac list.append(∅)
    return gene frac list
```

ELLE COLLINGE

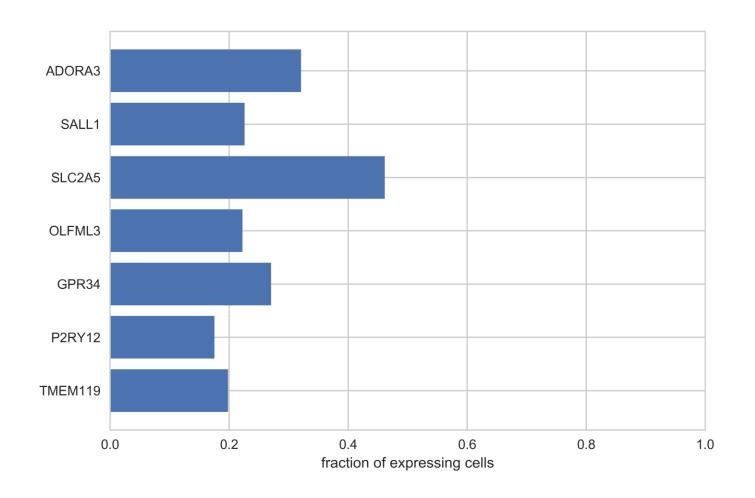
Fraction of expressing genes

```
gene_frac_list = gene_frac_df(gene_list)

#array containing the number of genes
y_pos = np.arange(len(gene_list))

#barh = horizontal bar graph
plt.barh(y_pos, gene_frac_list)
plt.xlim(0,1)
plt.yticks(y_pos, tuple(gene_list))
plt.yticks(y_pos, tuple(gene_list))
plt.xlabel('fraction of expressing cells')
plt.savefig('frac_gene.png', facecolor='white', dpi=500)
```

Fraction of expressing genes





4 general rules to follow

- You can never over-comment your code.
- 2. Write functions that do simple tasks.
- 3. There might be a package for it. It doesn't hurt to check.
- 4. If you have a question, someone has probably asked it online.

Thank you for attending!