BIOSTATS

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Introduction

This textbook aims to introduce fundamental statistical techniques and their applications to biological data. A unique aspect of this book is the "flipped-order" introduction. Many statistics courses start with theory; yet, I found it difficult for those unfamiliar with statistics. I will start with a real example of the method, followed by the explanation for an underlying theory/concept. The author is an ecologist, so some methods in this book might not be popular in other fields.

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Chapter 1

Why statistics?

1.1 Rationale

Why do I need statistics in the first place? - This is the first question I got when I stepped into the field of ecology. I thought this would be an easy question that someone could give me the answer to right away, but actually, it is a deep one. The short answer is "we need statistics because we only have partial information about what we want to know." The long answer? See below.

1.2 Example – fish density in a lake

Let's say we are interested in fish density in a lake (total area $15 \times 15 = 225 \text{ m}^2$). We try to quantify it by catching fish in a certain area (1 m² for example). Since there should be some variation in fish density among sampled points, we decided to repeat this 10 times to take an average. Figure 1.1 shows a graphical representation of this example.

Through this sampling, we got the following dataset:

df_count

```
## # A tibble: 10 x 2
       plot count
##
##
      <int> <int>
##
           1
    1
##
    2
           2
                 1
    3
           3
                 2
##
                 2
##
    4
           4
##
    5
           5
                 0
    6
           6
                 3
##
   7
                 3
```

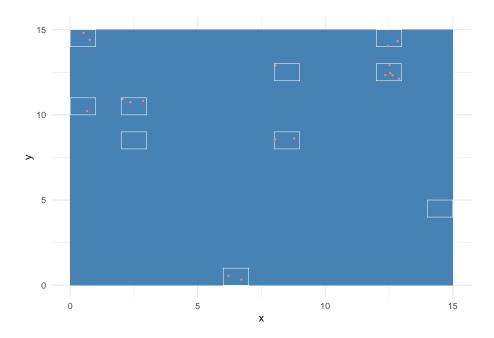


Figure 1.1: Lake fish density example. Points are fish, and white boxes are sampling sites with 1 sq-m $\,$

```
## 8 8 1
## 9 9 5
## 10 10 0
```

Exercise: create a tibble object containing above data

Based on this dataset, we may conclude the average fish density in this lake is 1.9. However, how confident are you in this conclusion? The lake area is $15 \times 15 = 225 \text{ m}^2$ but we sampled only $1 \text{ m}^2 \times 10 = 10 \text{ m}^2$! Indeed, we missed a lot of fish in the lake (Figure 1.2).

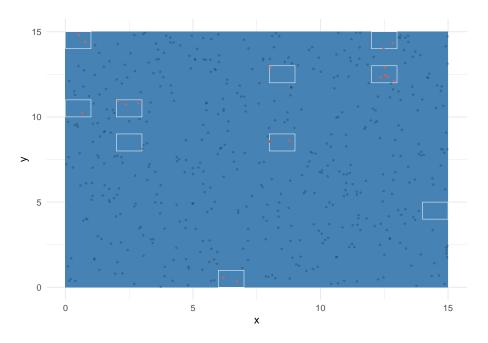


Figure 1.2: True fish distribution in the lake

In ecological research (and almost all research fields), we are rarely able to obtain the perfect dataset with no uncertainty – the data are partial information of what we want to know (true fish density in the entire lake = 2.22). This is the reason why we refer to a data point as a **sample**, and the average fish density in a plot (1.9) is the **estimate** of the true fish density – it is the inference from the incomplete information. Since the average fish density is the estimate, it does not perfectly match the true density – how do we account for this uncertainty?

1.3 Mean & Variance

In the above section, I talked only about the **mean** (total fish counts/number of plots = mean density per plot area $[m^2]$). Let me re-write this in a little more

[1] 1.9

mathematical form – I write a single data point (fish count in a single white box) as y_i with i representing an ID of a plot (therefore, $y_1=1,\ y_2=1,\ y_3=1,\ y_4=2,...y_{10}=3)$, and refer to the number of plots as N. The mean fish counts μ is:

$$\mu = \frac{\sum_{i=1}^{N} y_i}{N}$$

However, the data contain another important aspect of information: variability in fish counts among samples. This variability – called **variance** – expresses the degree of uncertainty in your data. The variance σ^2 can be calculated using the deviation of each data point x_i from the mean μ :

$$\sigma^2 = \frac{\sum_{i=1}^N (y_i - \mu)^2}{N-1}$$

In this equation, $(y_i - \mu)^2$ is the deviation from the mean, but it is squared to make the deviation positive regardless of the value of y_i . Since all the deviations are positive, the summation should represent the overall deviation from the mean. We divide it by N-1 to express it as the "average" of deviations¹. Thus, chiefly speaking, variance can be interpreted as the average squared deviation of each data point.

Let's estimate mean and variance in R. Create tibble() containing the data presented above, and perform the following analysis. Although R has built-in functions for mean (mean()) and variance (var()), I show here how to express the equations above in R codes:

```
# define variables
y <- df_count$count # fish counts
n <- nrow(df_count) # number of plots

print(y)

## [1] 2 1 2 2 0 3 3 1 5 0

print(n)

## [1] 10
# mean
mu <- sum(y) / n
print(mu)</pre>
```

 $^{^{1}}$ The reason for dividing by N-1, not by N, is a statistically deep topic; in essence, this is required to obtain an unbiased estimate of variance.

```
# variance
sigma2 <- sum((y - mu)^2) / (n - 1)
print(sigma2)</pre>
```

[1] 2.32222

Exercise: Make sure these values are identical to what you would obtain from mean() and var() functions in R

1.4 Probability distribution

1.4.1 Choose right one

Mean and variance are unknown constants² – these are what we refer to as **parameters**. Parameters are the focus of statistical inference because it allows the speculations for the *overall* structure of the population under investigation.

To clarify this argument, let's think more about a mean – more precisely speaking, an expected value of a random variable. A mean is a value around which data points are concentrated. This suggests that the probability of observing data points close to this value is very high. This is the reason why we use a mean as a "representative" summary statistic (e.g., mean height, mean weight, etc...), although many of us may use it without recognizing it. Then, what variance does for us? As you move away from the mean, the probability of observing that value will decrease. Variance tells you how sharply the probability decreases as a value deviates from the mean (smaller variance means sharper decline). Thus, these two parameters shape the distribution of probability across a range of possible values – this is what we refer to as a probability distribution. Figure 1.3 is the probability distribution drawn based on the parameters we estimated in the fish data. The distribution has the peak at the mean value (1.9) with curvy declines towards both tails. The y-axis is labeled as "probability density" to allow for the continuous expression of probability for continuous values on the x-axis³.

However, some of you might notice weird things in Figure 1.3. First, the positive probability density extends below the "zero" fish count, which should not happen in nature (we cannot observe negative counts). Second, the fish count is treated as if it is a continuous variable, even though fish counts are always discrete values (there is no way to count 1.1 individuals for an individual survey plot, for example). When comparing this probability distribution with the fish count histogram, they do not look similar at all (Figure 1.4))...even though they should look very similar if the probability distribution represents the data properly.

²At least for frequentist analysis.

³You can define a probability for a range (e.g., 1 < x < 2); otherwise, it must be probability "density" because you cannot define a probability for x = 1.1 due to the continuous nature of the variable x (we don't know whether x = 1.1 to be x = 1.101 or 1.102 or...)

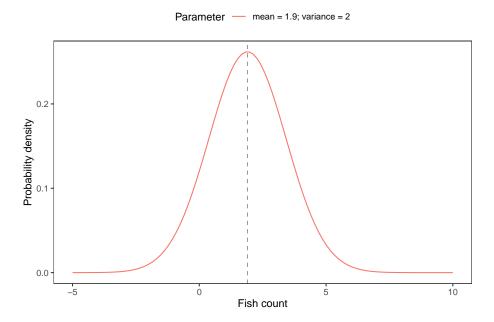


Figure 1.3: Example of probability distribution

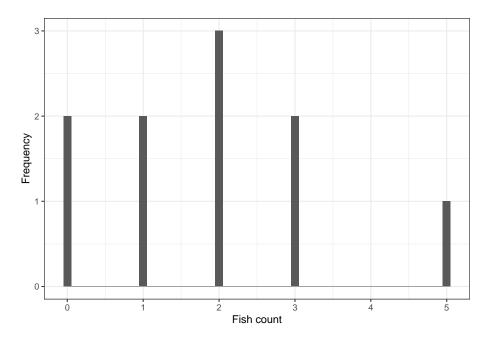


Figure 1.4: Frequency distribution of fish count data

Exercise: Produce a histogram of fish count

Why does this happen? – this is because this particular kind of probability distribution is inappropriate to express count data. The above example is drawn from a **Normal distribution**, whose outcome (values drawn from this distribution) ranges from $-\infty$ to ∞ . To properly represent our fish count data, we must choose a probability distribution that can express discrete values of ≥ 0 (0,1,2,3,...). One possible choice for the fish count data is a **Poisson distribution**, whose outcome is confined to zero and positive discrete values. Unlike a Normal distribution, a Poisson distribution assumes $\mu = \sigma^2$; thus, knowing either μ or σ^2 is sufficient to draw a distribution (we use μ because it's easier) if the Poisson distribution is appropriate to represent the data. When a Poisson distribution is used, the probability distribution differs from that of a Normal distribution. Note that the y-axis is "probability" because the Poisson distribution allows only discrete values.

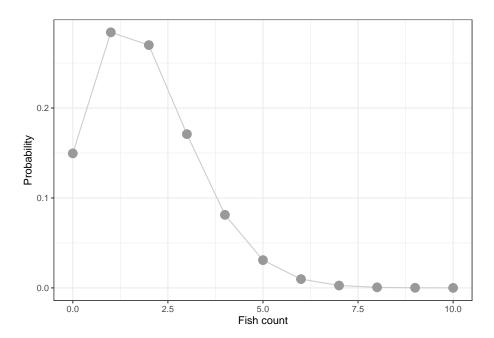


Figure 1.5: Poisson distribution with the mean fish count

To overlay this "probability" distribution on the "frequency" distribution for comparison, we need to calculate a product of the number of samples (in this case, the number of plots = 10) and the probability of fish count Y taking value k, $\Pr(Y=k)$ – this multiplication results in the "expected" frequency of a value in the data. Figure 1.6 shows the comparison – this seems better than the Normal distribution. Yet, there are some deviations from the expectation (e.g., the frequency of fish count Y=1).

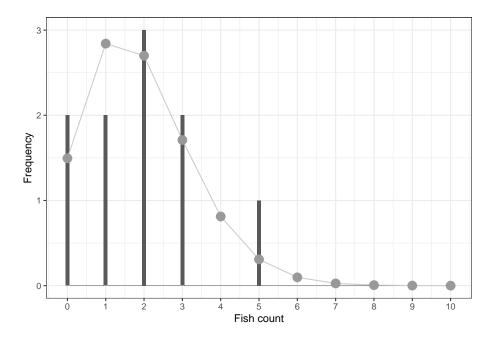


Figure 1.6: Comparison of observed and expected frequencies of fish count

The deviation is due largely to the small sample size – we have only 10 plots. In theory, we can obtain more accurate parameter estimates as the sample size increases. To see this effect, let's increase the sample size to 100 plots – Figure 1.7 shows the updated histogram and the expected distribution with the new mean estimate ($\mu=2.19$). This greatly improved the matching between the expected and observed distributions.

1.4.2 How to choose appropriate one?

We chose a Poisson distribution to express fish counts based on the characteristics of the variable. I used a comparison of a Normal and a Poisson distribution, but there are many more choices when expressing random variables. In ecology, several probability distributions are more frequently used than others, and the following dendrogram describes how we choose an acceptable distribution among them. Answer the questions and find your node.

Once you find your node⁴:

⁴Although this covers common distributions in ecology, following this dendrogram does not guarantee that your choice is appropriate.

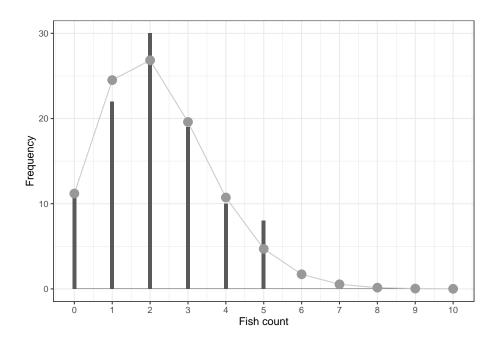


Figure 1.7: Histogram obtained from 100 plots

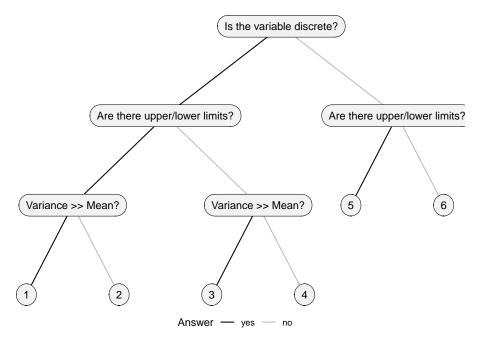


Figure 1.8: Brief guide for how to choose an appropriate probability distribution.

Node	Possible distribu- tion	Possible values	Example in ecological data
1	Beta- binomial	0 to N	Number of fertilized eggs out of N eggs examined. Useful when N is large (tend to be over-dispersed with binomial distribution)
2	Binomial	0 to N	Number of fertilized eggs out of N eggs examined.
3	Negative- binomial	≥ 0	Number of individuals captured. Useful when variance is very large relative to mean (over-dispersed with a Poisson distribution).
4	Poisson	≥ 0	Species richness.
5	Beta	0 to 1	Survival probability.
5	Log- Normal	> 0	Biomass.
6	Normal	$-\infty$ to ∞	Temperature.

Chapter 2

Linear model

Many of you might hate equation – I hated too because no one told me how to read it. However, you will like it once you understand how to interpret; it is actually the clearest explanation. In this section, I will continue to use the example of fish counts in a lake to explain the use of linear models.

2.1 Mean estimate

2.1.1 Normal distribution

In the example of fish counts, we estimated mean and variance to infer how fish counts distribute over a range of possible values (probability distribution). Implicitly, I assumed that the "true" mean – i.e., the expected number of fish counts in a plot – is uniform within the lake. Let's express this assumption as a "model." Let y_i be fish count in plot i:

$$y_i = \mu + \varepsilon_i$$

The above equation reads fish count y_i can be expressed as the sum of mean density μ and residual ε_i (raw deviation from the mean; $\varepsilon_i = y_i - \mu$). Notice that the parameter μ does not have subscript i (i.e., μ is constant across plots) since we assume that the mean fish count is uniform across plots.

We can build this model pretty easily in R using function lm() (Linear Model, LM) with caution that this function assumes a Normal distribution, which we identified as an inappropriate distribution for our fish data. However, let me use this function for the sake of simplicity. In the function lm(), we write a **response** variable (i.e., the variable we want to explain = fish count) on the left side of tilde \sim , and **explanatory** variables (or predictors) on the right. In our simplest model, we do not have explanatory variables, so put 1 on the right.

```
fit <- lm(count ~ 1, data = df_count)</pre>
summary(fit)
##
## Call:
## lm(formula = count ~ 1, data = df_count)
##
## Residuals:
##
              1Q Median
     Min
                            3Q
                                  Max
##
    -1.90 -0.90
                   0.10
                          0.85
                                 3.10
##
## Coefficients:
##
               Estimate Std. Error t value Pr(>|t|)
                 1.9000
                            0.4819
                                      3.943 0.00339 **
## (Intercept)
##
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 1.524 on 9 degrees of freedom
```

This is perhaps the simplest model we can make – the variation in fish count y_i can be expressed as the mean μ plus random variation ε_i . However, where is the mean in the output? In the model output, the estimated mean is shown in the column Estimate under the area Coefficients:. We can prove it by comparing the estimate and the mean calculated with function mean().

Exercise: calculate mean fish count with mean()

Now we see how this model works. however, where is ε_i then? You can get values of ε_i using function resid().

```
eps <- resid(fit)
print(eps)</pre>
```

```
## 1 2 3 4 5 6 7 8 9 10
## 0.1 -0.9 0.1 0.1 -1.9 1.1 1.1 -0.9 3.1 -1.9
```

It is unclear how this is derived, so let's compare it with $y_i - \mu$. Note that SD of the residual (SD = $\sqrt{\text{Variance}}$) is the degree of variation in the data, and the value Residual standard error: should match eps's SD.

Exercise: calculate $y_i - \mu$ and compare it with eps. Also, calculate the SD of eps and compare it with the reported value Residual standard error: in the lm() output.

The model can be written in many ways. It varies by person, but I strongly encourage you to know the following expression of a statistical model:

```
y_i \sim \text{Normal}(\mu, \sigma^2)
```

The interpretation of this equation is that y_i follows a normal distribution with a mean μ and variance σ^2 . When the left and right hands of the equation are related with tilde, it indicates the relationship is stochastic – the outcome y_i always deviates from the expectation μ , unlike equal sign $y_i = \mu$. This expression is identical to the following:

$$y_i = \mu + \varepsilon_i \varepsilon_i \sim \text{Normal}(0, \sigma^2)$$

This expression clarifies that the deviation ε_i is stochastic, but the expectation μ is deterministic. This is the basic structure of statistical model – **observation** is a mixture of deterministic and stochastic components.

2.1.2 Poisson distribution

In the previous section, we have identified that fish count is better described by a Poisson distribution. How can fit a model with a Poisson distribution? R has functionality to perform this type of analysis, and Generalized Linear Model (GLM) (glm()) is the natural extension of lm() function for non-normal distributions. It's pretty easy to use:

```
fit_pois <- glm(count ~ 1, data = df_count, family = "poisson")</pre>
```

The only difference is that we used glm() and specified a Poisson distribution as family = "poisson". Let's see what's in there.

```
summary(fit_pois)
```

```
##
## Call:
## glm(formula = count ~ 1, family = "poisson", data = df_count)
## Deviance Residuals:
##
                   1Q
                        Median
                                                Max
  -1.94936 -0.71853
                       0.07192
                                 0.56940
                                            1.86436
##
## Coefficients:
              Estimate Std. Error z value Pr(>|z|)
                                     2.798 0.00515 **
## (Intercept)
                0.6419
                           0.2294
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for poisson family taken to be 1)
##
       Null deviance: 13.205
                             on 9
                                   degrees of freedom
## Residual deviance: 13.205 on 9 degrees of freedom
## AIC: 36.51
##
```

Number of Fisher Scoring iterations: 5

Wait...it says Estimate 0.64, which is quite different from what we see in our fish count data (mean = 1.9). What's going on? This is because there is internal variable transformation when using a Poisson distribution. Since a Poisson distribution deals with positive integer (including zero), its mean cannot be negative. To avoid this potential problem in some data sets, the function glm() estimates the mean after log-transformation, and then back-transform to an ordinary scale. The reported value is in a natural log-scale – we can retrieve the mean estimate in an ordinary scale by transforming the estimate by exponent exp().

```
# get intercept value in a log-scale
fit_pois$coefficients

## (Intercept)
## 0.6418539
# transform to an ordinary scale
exp(fit_pois$coefficients)

## (Intercept)
## 1.9
```

2.2 Effect estimate

2.2.1

Chapter 3

Appendix: Project Management

3.1 R Project

For this entire book, I will use R Project as a fundamental unit of work-space, in which all the relevant materials (e.g., R scripts .R and data files) are assembled together. There are many ways to organize your project, but I usually make a single R Project for a collection of scripts and data that will lead to a single publication (see example here). To setup an R Project you will need RStudio in addition to base R. While R is a stand-alone software, I strongly recommend to use it with RStudio. RStudio has many functions that help your data analysis. R and RStudio can be installed from the following websites:

- R (you can choose any CRAN mirror to download)
- RStudio

Once you open *RStudio*, you will see the following interface (Figure 3.1). There are three major panels in its first appearance — Console, Environment, and Files. Console is the place where you write your codes and execute calculation/data manipulation/analysis. Environment lists items you saved as an object. Files list any files in a designated location in your computer.

Let's type something in console to see what happens. Copy and paste the following script to Console. You should see x in the environment.

$$x < -c(1, 2)$$

x is an **object** where information is saved - in this case, we stored a sequence of 1 and 2 in object x. Once you store the information into x, you can call it just typing x.

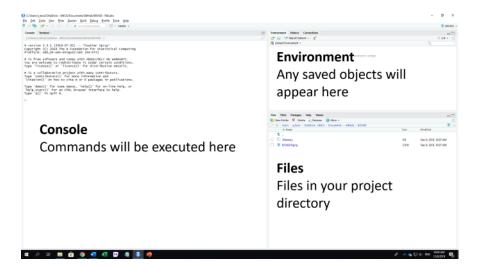


Figure 3.1: RStudio interface.

Х

[1] 1 2

Sweet! You can work on your data like this, however, **it is actually a BAD idea**. As you work on your project, numerous materials will be generated (e.g., I would write > 2000 lines of codes for one project). How do you manage your codes?

3.1.1 Script Editor

It is highly recommended to mange your script in Editor instead. Editor is where you draft and tweak your codes before executing them in Console. To create space for Editor, hit Ctr + Shift + N. A new panel pops up on the top left. Let's type the following script in Editor.

Then, hit Ctr + S to save the Editor file. RStudio will prompt you to enter the file name of the Editor¹.

3.1.2 File Name

It is also critical to have **consistent naming rules** for your files. As you make progress on your project, the number of files in each sub-directory will increase, perhaps exponentially. You will find it difficult navigating yourself unless you

 $^{^{1}}$ In R, an editor file has an extension of .R.

3.1. R PROJECT 23

have clear naming rules for files (and even worse for others). Here are some recommendations:

- NO SPACE. Use underscore.
 - Do: script_week1.R
 - Don't: script week1.R
- NO UPPERCASE. Use lowercase for file names.
 - Do: script_week1.R
 - Don't: Script_week1.R
- BE CONSISTENT. Apply consistent naming rules within a project.
 - Do: R scripts for figures always start with a common prefix, e.g.,
 figure_XXX.R figure_YYY.R(XXX and YYY specifies further details).
 - Don't: R scripts for figures start with random text, e.g., XXX_fig.R
 Figure_Y2.R , plotB.R.

3.1.3 Structure Your Project

If you do not save or randomly locate your codes within your computer, you will lose necessary items sooner or later. For this reason, I assemble all the relevant materials in a single R Project. You can create a new R Project with the following procedure.

- a. Go to File > New Project on the top menu
- b. Select New Directory
- c. Select New Project

A new window pops up and prompts you to name a directory with a location in your computer. Click Browse to select a location for the directory².

The internal structure of an R Project is extremely important to navigate yourself (others once it's published). R Project will be composed of multiple types of files, typically .R, .csv, .rds, .Rmd among others. Unless those files are arranged in an organized manner, it is VERY LIKELY to make severe errors in coding. So I take this seriously. Table 3.1 is my suggested subdirectory structure.

Table 3.1: Suggested internal structure of R Project

Name Content

README. Indiarkdown file explaining contents in the R Project. Can be derived from README. Rmd.

/code Sub-directory for R scripts (.R).

/data_rawb-directory for raw data before data manipulation (.csv or other formats). Files in this sub-directory MUST NOT be modified unless there are changes to raw data entries.

²When you locate your project directories in your computer, I would strongly recommend to create a designated space. For example, in my computer, I have a folder named /r_project in which all R Project directories are located.

Name Content

3.2 Robust coding

While this is not mandatory, I strongly recommend to use RStudio with Git & GitHub (see Chapter ?? Appendix for guidance). Coding is innately error-prone³; every programmers, even bright ones, make mistakes - no exception. However, the critical difference between beginner and advanced programmers is whether they develop robust coding rules with a self-error-detection system. Git is the key to this. I will touch on this occasionally throughout this book.

 $^{^3\}mathrm{well}$ WAY BETTER than manual data manipulation!

Chapter 4

Appendix: Git & GitHub

4.1 Git & GitHub

In this section, I will cover how to integrate *Git* and *GitHub* into *R Studio*. *R Studio* is excellent as is, but it becomes even better when combined with *Git* and *GitHub*. *Git* is a free and open source distributed version control system. *Git* tracks changes in your codes codes while you work on your project so you are aware of any changes you made to your script (and other) files. Tracking changes is extremely important to avoid unintended errors in your codes. This feature also helps avoid creating redundant files. While *Git* is a local system, it has an online counterpart called *GitHub*.

To make this system work, you'll need to go through some processes. The first step is to install *Git* onto your computer:

- Windows: Install *Git* from here. You will be asked about "Adjusting your PATH environment". Choose "*Git* from the command line and also from 3rd-party software" if it is not selected.
- Mac: Follow the instruction here.

Then open R Studio and do Create Project > New Directory > New Project. If you see a check box Create a git repository, check and create a new project (Figure 4.1). You will see a *Git* pane on the upper right panel.

If you can't find the above, do the following: Tools in the menu bar > Terminal > New Terminal, and type where git in the terminal. This will tell you where git executable is located in your computer. Then, go to Tools in the menu bar > Global Options > Git/SVN. You will see *Git executable* in the box, where you can specify the location of git executable.

Next, go to *GitHub* and create your account (free!). But, give some thoughts on your user name. My advice is the following. First, use lowercase only.

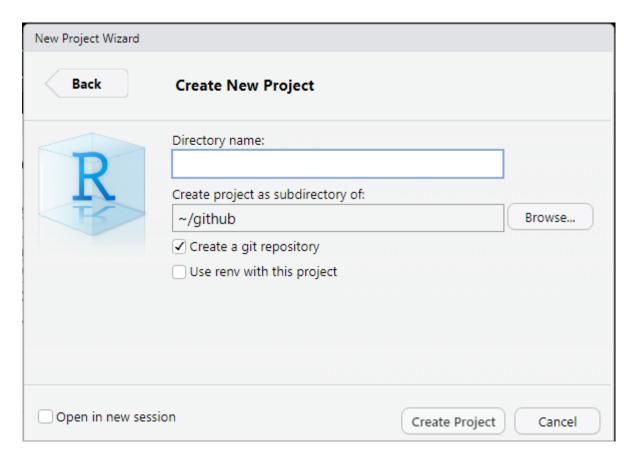


Figure 4.1: After installing Git, you should see 'Create a git repository'.

Second, **include your name** to make it easy to find you on *GitHub*.

R Studio works seamlessly with Git or GitHub, but it is helpful to use a Git client as it provides visual aids. There are choices for a Git client (see options here) but I will use GitHub Desktop (available here) for our exercise. Install GitHub Desktop onto you computer.

4.2 Commit & Push

4.2.1 Register Your Git repo

Open the R Project you've just created as a git repository. Let's make a sample .R file (Ctr + Shift + N) and save it (name as sample.R). For example:

```
## produce 100 random numbers that follows a normal distribution
x <- rnorm(100, mean = 0, sd = 1)

## estimate mean
mean(x)

## estimate SD
sd(x)</pre>
```

Open GitHub Desktop App. You will see the following GUI (Figure 4.2):

Hit current repository (top left) and Add > Add existing repository (Figure 4.3):

4.2.2 Commit

GitHub Desktop will prompt you to enter a local path to your *Git* repository. Browse and select your directory of the R Project - the local Git repository will show up in the list of repositories in *GitHub Desktop* (left side bar in Figure 4.3). Now, you are ready to Commit your files to *Git*! Commit is the procedure to record your file change history in *Git*. To make this happen, click your *Git* repository on GitHub Desktop, and you will see the following:

At this stage, your file (.R or else) is saved onto your local directory, but has not been recorded in *Git* as a change history. To make a Commit, the first thing to do is to choose a file(s). There are checkboxes next to each of the new files. If this box is checked, you are going to commit changes to *Git*. Once you selected the files you want to make a commit, go to the bottom left of the window. There is a small box saying summary (required) or Create sample.R.This is the place where you can put any title that describes what you did in this commit, and you cannot Commit unless you enter this information! For example, I would write initial commit for this exercise from the second commit, you should put a more informative commit message so you can track changes when needed. You can google recommendations for

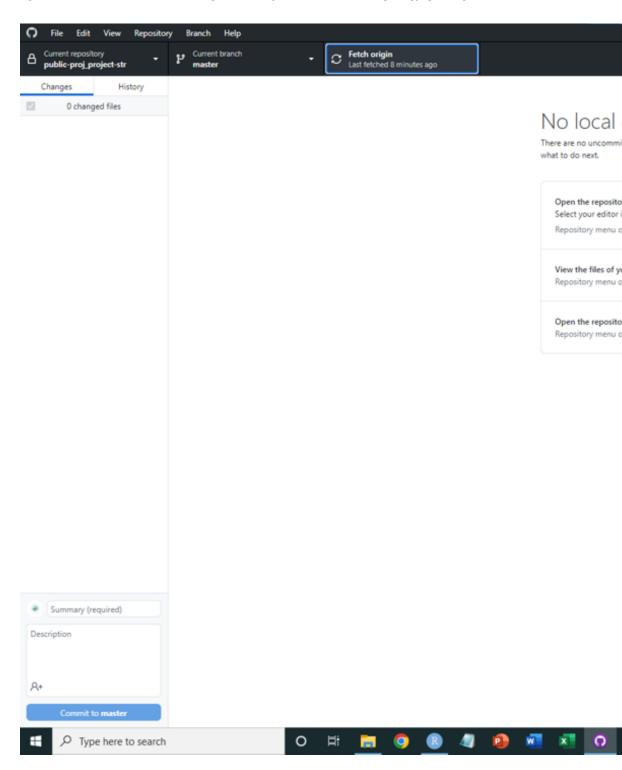


Figure 4.2: GUI for GitHub Desktop

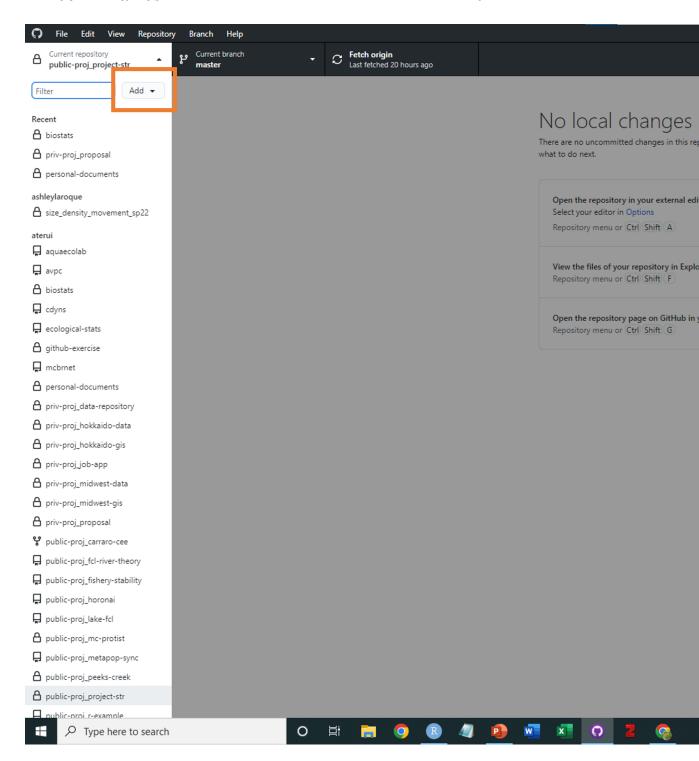


Figure 4.3: Add dropdown on the top left

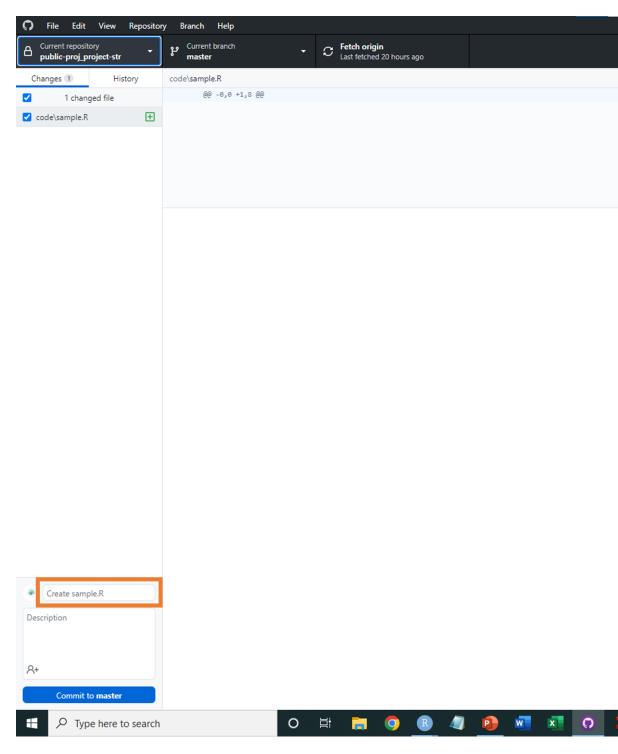


Figure 4.4: You will see 'Create sample.R' or 'summary (required)' on the bottom left

how to write commit titles/descriptions. Then hit commit to master. Now, changes to the selected files have been recorded in *Git*!

4.2.3 Push

Remember, your changes are recorded in your local computer **but not published in your online repository!** To send local changes to the online *GitHub* repository, you will need to **Push** commits via *GitHub Desktop*. Push is the procedure to send your Commit(s) to *GitHub*. Once you do a Commit, *GitHub Desktop* will ask you whether you want to Push it to an online repository (Figure 4.5). If this is the first push, there is no corresponding repository on *GitHub* tied to your local repository, so *GitHub Desktop* will ask you if you want to publish it on *GitHub* (NOTE: although it says 'publish', your repository will remain private unless you explicitly tell *Git Desktop* to make it public). If you are comfortable with the changes you made, **Push** it!

4.2.4 Edit

We went through how we get things uploaded onto *GitHub*, but what happens if we make changes to existing files? To see this, let's make a minor change to your R script. We have created a file sample.R:

```
## produce 100 random numbers that follows a normal distribution
x <- rnorm(100, mean = 0, sd = 1)

## estimate mean
mean(x)

## estimate SD
sd(x)</pre>
```

Edit this as follows:

```
## produce 100 random numbers that follows a normal distribution
x <- rnorm(100, mean = 0, sd = 1)

## estimate mean
median(x)

## estimate SD
var(x)</pre>
```

After making this change, go to *GitHub Desktop* again. *GitHub Desktop* automatically detects the difference between the new and old files and shows which part of the script has been edited! This helps coding quite a bit:

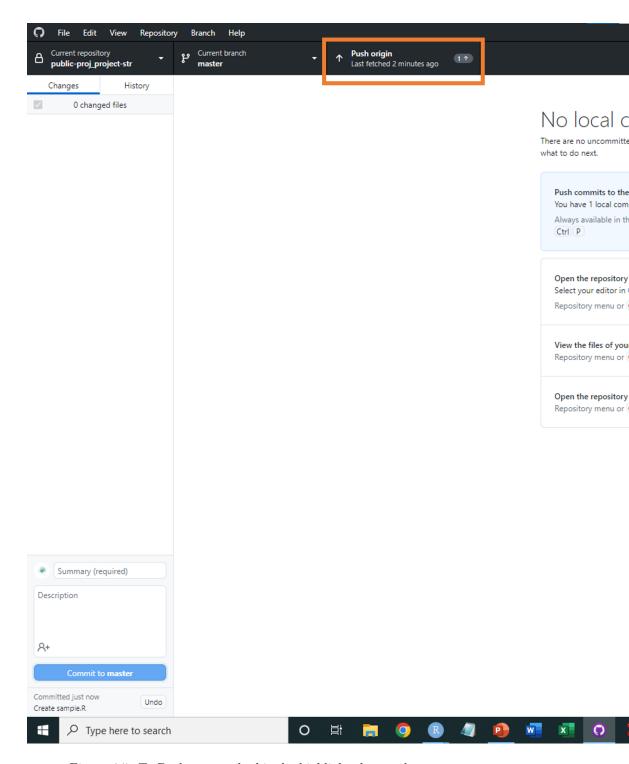


Figure 4.5: To Push your code, hit the highlighted menu button

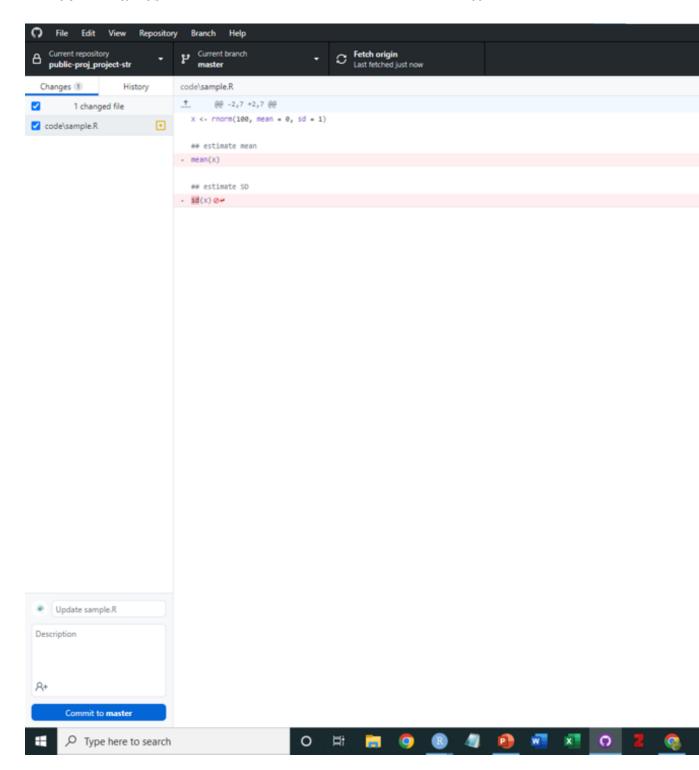


Figure 4.6: Git detects edits to your codes

Chapter 5

Appendix: Data Structure

5.1 Overview

R has 6 basic data types.

- \bullet factor: similar to character, but has levels (alphabetically ordered by default)
- numeric: 20.0, 15.5
- integer: 3, 7
- logical: TRUE , FALSE
- complex: 1+2i (complex numbers with real and imaginary parts)

These elements form one of the following data structures.

- vector: a series of elements. A single data type is allowed in a single vector
- matrix: elements organized into rows and columns. A single data type is allowed in a single matrix
- data frame: looks similar to a matrix, but allows different data types in different columns

5.2 Vector

5.2.1 Create Vector

Below are examples of atomic character vectors, numeric vectors, integer vectors, etc. There are many ways to create vector data. The following examples use c(), :, seq(), rep():

```
#ex.1a manually create a vector using c()
x \leftarrow c(1,3,4,8)
## [1] 1 3 4 8
#ex.1b character
x <- c("a", "b", "c")
## [1] "a" "b" "c"
#ex.1c logical
x <- c(TRUE, FALSE, FALSE)
## [1] TRUE FALSE FALSE
#ex.2 sequence of numbers
x <- 1:5
## [1] 1 2 3 4 5
#ex.3a replicate same numbers or characters
x <- rep(2, 5) # replicate 2 five times
## [1] 2 2 2 2 2
#ex.3b replicate same numbers or characters
x <- rep("a", 5) # replicate "a" five times
## [1] "a" "a" "a" "a" "a"
#ex.4a use seq() function
x \leftarrow seq(1, 5, by = 1)
X
## [1] 1 2 3 4 5
#ex.4b use seq() function
x < - seq(1, 5, by = 0.1)
X
## [1] 1.0 1.1 1.2 1.3 1.4 1.5 1.6 1.7 1.8 1.9 2.0 2.1 2.2 2.3 2.4 2.5 2.6 2.7 2.8
## [20] 2.9 3.0 3.1 3.2 3.3 3.4 3.5 3.6 3.7 3.8 3.9 4.0 4.1 4.2 4.3 4.4 4.5 4.6 4.7
## [39] 4.8 4.9 5.0
```

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5.2.2 Check Features

R provides many functions to examine features of vectors and other objects, for example:

- class() return high-level data structure of the object
- typeof() return low-level data structure of the object
- attributes() metadata of the object
- length() number of elements in the object
- sum() sum of object's elements
- mean() mean of object's elements

Numeric Vector

```
x <- c(1.2, 3.1, 4.0, 8.2)
x

## [1] 1.2 3.1 4.0 8.2

class(x)

## [1] "numeric"

typeof(x)

## [1] "double"

length(x)

## [1] 4

sum(x)

## [1] 16.5

mean(x)

## [1] 4.125

Character Vector

## [1] "character"

## [1] 3</pre>
```

5.2.3 Access

Element ID

Use brackets [] when accessing specific elements in an object. For example, if you want to access element #2 in the vector \mathbf{x} , you may specify as \mathbf{x} [2]:

```
## [1] 2
## [1] 2 2
```

```
## [1] 2 3 2
```

Equation

R provides many ways to access elements that suffice specific conditions. You can use mathematical symbols to specify what you need, for example:

- == equal
- \bullet > larger than
- >= equal & larger than
- \bullet < smaller than
- \leq equal & smaller than
- which() a function that returns element # that suffices the specified condition

The following examples return a logical vector indicating whether each element in **x** suffices the specified condition:

```
# creating a vector
x <- c(2,2,3,2,5)

# ex.1a equal
x == 2

## [1] TRUE TRUE FALSE TRUE FALSE
# ex.1b larger than
x > 2
```

[1] FALSE FALSE TRUE FALSE TRUE

You can access elements that suffice the specified condition using brackets, for example:

```
# ex.2a equal
x[x == 2]

## [1] 2 2 2

# ex.2b larger than
x[x > 2]

## [1] 3 5

Using which(), you can see which elements (i.e., #) matches what you need:
# ex.3a equal
which(x == 2) # returns which elements are equal to 2

## [1] 1 2 4

# ex.3b larger than
which(x > 2)
```

```
## [1] 3 5
```

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5.3 Matrix

5.3.1 Create Matrix

Matrix is a set of elements (*single data type*) that are organized into rows and columns:

```
#ex.1 cbind: combine objects by column
x \leftarrow cbind(c(1,2,3), c(4,5,6))
##
         [,1] [,2]
## [1,]
            1
## [2,]
            2
## [3,]
            3
                 6
#ex.2 rbind: combine objects by row
x \leftarrow rbind(c(1,2,3), c(4,5,6))
Х
         [,1] [,2] [,3]
## [1,]
                 2
                       3
            1
## [2,]
                 5
                       6
#ex.3 matrix: specify elements and the number of rows (nrow) and columns (ncol)
x \leftarrow matrix(1:9, nrow = 3, ncol = 3)
         [,1] [,2] [,3]
## [1,]
                 4
                      7
            1
## [2,]
                 5
                       8
## [3,]
            3
                 6
                       9
```

5.3.2 Check Features

R provides many functions to examine features of matrix data, for example:

- dim() number of rows and columns
- rowSums() row sums

3

6

9

• colSums() column sums

Integer Matrix

[3,]

```
x <- matrix(1:9, nrow = 3, ncol = 3)
x

## [,1] [,2] [,3]
## [1,] 1 4 7
## [2,] 2 5 8</pre>
```

```
class(x)
## [1] "matrix" "array"
typeof(x)
## [1] "integer"
dim(x)
## [1] 3 3
Character Matrix
y <- matrix(c("a", "b", "c", "d", "e", "f"), nrow = 3, ncol = 2)
##
        [,1] [,2]
## [1,] "a" "d"
## [2,] "b" "e"
## [3,] "c" "f"
class(y)
## [1] "matrix" "array"
typeof(y)
## [1] "character"
dim(y)
## [1] 3 2
5.3.3 Access
When accessing matrix elements, you need to pick row(s) and/or column(s), for
example:
x <- matrix(1:9, nrow = 3, ncol = 3)
##
        [,1] [,2] [,3]
## [1,]
           1
                4
## [2,]
          2
                5
                     8
## [3,]
           3
x[2,3] # access an element in row #2 and colum #3
## [1] 8
```

x[2,] # access elements in row #2

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```
## [1] 2 5 8
x[c(2,3),] # access elements in rows #2 and 3
        [,1] [,2] [,3]
##
## [1,]
           2
                 5
## [2,]
                 6
                      9
           3
x[,c(2,3)] # access elements in columns #2 and 3
        [,1] [,2]
## [1,]
                 7
           4
## [2,]
           5
                 8
## [3,]
           6
                 9
You can assess each element with mathematical expressions just like vectors:
x == 2 \# equal
         [,1] [,2] [,3]
## [1,] FALSE FALSE FALSE
## [2,] TRUE FALSE FALSE
## [3,] FALSE FALSE FALSE
x > 2 # larger than
##
         [,1] [,2] [,3]
## [1,] FALSE TRUE TRUE
## [2,] FALSE TRUE TRUE
## [3,] TRUE TRUE TRUE
However, care must be taken when accessing elements, as it will be automatically
converted to vector data:
## [1] 2
## [1] 3 4 5 6 7 8 9
which() needs an additional argument to return both row and column #:
        row col
##
## [1,]
          2
               1
##
        row col
## [1,]
          3
               1
## [2,]
               2
          1
## [3,]
               2
          2
## [4,]
          3
               2
## [5,]
               3
          1
## [6,]
          2
               3
## [7,]
          3
               3
```

5.4 Data Frame

Data frame is a set of elements that are organized into rows and columns, but differ from matrix in several ways.

- it allows multiple data types in different columns
- each column has its name
- you can access columns by name (using \$)

Data frame is the most common data structure when manipulating ecological data. A data set loaded from a spread sheet (we will address this later) will be automatically recognized as a data frame. Here is an example:

Create a data frame

In the following example, variables x and y are organized into a single data frame dat. Variable are renamed when creating a data frame composed of x and y.

```
# Create data frame
x <- c("Pristine", "Pristine", "Disturbed", "Disturbed", "Pristine") # Lake type
y <- c(1.2, 2.2, 10.9, 50.0, 3.0) # TSS: total suspended solids (mg/L)
dat <- data.frame(LakeType = x, TSS = y) # x is named as "LakeType" while y is named a
dat
##
      LakeType TSS
## 1 Pristine 1.2
## 2 Pristine 2.2
## 3 Disturbed 10.9
## 4 Disturbed 50.0
## 5 Pristine 3.0
Call column names
colnames(dat) # call column names
## [1] "LakeType" "TSS"
Access by columns
dat$LakeType # access LakeType
## [1] "Pristine"
                   "Pristine"
                               "Disturbed" "Disturbed" "Pristine"
dat$TSS # access TSS
## [1] 1.2 2.2 10.9 50.0 3.0
You can access elements like a matrix as well:
dat[,1] # access column #1
## [1] "Pristine" "Pristine" "Disturbed" "Disturbed" "Pristine"
```

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```
dat[1,] # access row #1

## LakeType TSS
## 1 Pristine 1.2

dat[c(2,4),] # access row #2 and 4

## LakeType TSS
## 2 Pristine 2.2
## 4 Disturbed 50.0
```

5.5 Exercise

5.5.1 Vector

- a. Create three numeric vectors with length 3, 6 and 20, respectively. Each vector must be created using different functions in R.
- b. Create three character vectors with length 3, 6 and 20, respectively. Each vector must be created using different functions in R.
- c. Copy the following script to your R script and perform the following analysis:
- Identify element IDs of x that are greater than 2.0
- Identify element values of x that are greater than 2.0

5.5.2 Matrix

- a. Create a numeric matrix with 4 rows and 4 columns. Each column must contain identical elements.
- b. Create a numeric matrix with 4 rows and 4 columns. Each row must contain identical elements.
- c. Create a character matrix with 4 rows and 4 columns. Each column must contain identical elements.
- d. Create a character matrix with 4 rows and 4 columns. Each row must contain identical elements.
- e. Copy the following script to your R script and perform the following analysis:
- Identify element IDs of x that are greater than 2.0 (specify row and column IDs)
- Identify element values of x that are greater than 2.0 and calculate the mean.

5.5.3 Data Frame

a. Create a data frame of 3 variables with 10 elements (name variables as x, y and z. x must be character while y and z must be numeric.

- b. Check the data structure (higher-level) of $\mathtt{x},\,\mathtt{y}$ and \mathtt{z}
- c. Copy the following script to your R script and perform the following analysis:
- \bullet Calculate the means of temperature and abundance for states VA and NC separately.