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1 Introduction

1.1 Background

1.2 Experiment

Virus expressing GCaMP6f was injected into the V1 of mice. Approximately 3 weeks post infection, mice were imaged under a 2-photon microscope while sinusoidal drifting gratings were presented on a computer screen placed 3 inches from the mouse (1 degree of visual space ~ 21.3 pixels on the screen). The stimulus was varied as:

1. Sinusoidal drifting gratings at 16 different directions (0:22.5:337.5). Spatial frequency was fixed at 0.03 cycles per degree.
2. Each direction was repeated 10 times (i.e. 10 trials per direction). Directions are presented in a randomized order.
3. Each direction was presented for 2s and was always preceded by a 4s gray screen. Therefore the total duration of the stimulus is 6s.
4. Calcium signals (GCaMP6f in awake mice) were acquired from awake mice at 20 frames per seconds. Thus, sampling rate is 20Hz.

1.3 Aim

- To map orientation tuning responses of excitatory pyramidal neurons in V1.
- To determine response reliability at preferred orientation.
- To determine signal and noise correlations between neuron as a function of orientation tuning.

2 Dataset

2.1 Restructuring the data

There are 19200 Ca readings for each mouse. The experiment is done using 16 different stimuli, Let us think of it as 16 classes. The angle of the drifting pattern changes between the classes. It is implicit that 0° and 180° will have same orientation, but the direction of drifting will be opposite.

First, the data is split into 16 parts corresponding to each class. There are 10 trials for each class. The data is split into 160 subsequences of $19200/160 = 120$ length. Now this is a time-series of 120 points for each trial, and there are ten sets of data for each class.

3 Analysis

3.1 Quantifying the neuron activity

The 120 point time series provide the activity of single cell in response to a particular stimulus in a trial. The activity of cell has to be studied. The first 80 samples are expected to have spikes due to only background activity, as the stimulus was gray during that time. Peaks in Ca concentration denotes the spikes in the neuron. From neuroanatomy, spike rate is a good quantify to measure neuron activity. In Calcium imaging, the amplitudes corresponds to Ca ion concentration. From the neuron models, more the Ca concentration, more is the spike rate. Following methods are used for quantifying spike rate.

3.1.1 Thresholding

3.1.2 Moving Average model

3.2 Estimating orientation selectivity of neuron

3.2.1 Circular variance of direction and orientation

As discussed in the paper, Circular variance is a more robust quantifier of orientation and direction selectivity. A scalar value representing the neuron response is extracted from the 6s long time series for each of the orientations and directions (There are 8 orientations and 16 directions). As we have 10 trials for each class, we will have 160 such values for each neuron. The values are first expressed as vectors in the orientation space. The vector sum of the values provide the ‘preferred orientation’; Magnitude should indicate how much orientation selective the neuron is.

Let $R(\theta)$ be the response of neuron to angle θ and we denote the ‘preferred direction’ as θ_{pref} . Normalized vector sum in orientation and direction space is found as.

$$L_{ori} = \frac{\sum_k R(\theta_k) \exp(2i\theta_k)}{\sum_k R(\theta_k)} \quad (1)$$

$$L_{dir} = \frac{\sum_k R(\theta_k) \exp(i\theta_k)}{\sum_k R(\theta_k)} \quad (2)$$

- Neurons with orientation selectivity are expected to have high L_{ori} .
- Simple cells are orientation selective, but less direction selective. They are expected to have large L_{ori} and small L_{dir} .
- Complex cells are orientation selective as well as direction selective. They are expected to have both large L_{ori} and large L_{dir} .

Circular orientation variance and direction variance is given as

$$CirVar = 1 - |L_{ori}| \quad (3)$$

$$DirCirVar = 1 - |L_{dir}| \quad (4)$$