# Answers to the exam of Linear Models, Generalized Linear Models and Linear Algebra, dd 29 January 2010

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The answers given below are rather elaborate to give you some extra background information. Generally, you can give shorter answers during the examination!

1 (30). We study the relationship between amount of lime applied to a soil and the pH of the soil. We have 3 soils and apply no lime, 1 unit of lime, and 2 units of lime. After application of lime, the pH values are measured: 5.8, 6.2, 6.3. The relationship is modeled by simple linear regression, where variable rate represents the amount of lime (with values 0,1,2):  $pH = \alpha + \beta rate + \epsilon$ ;  $var(\epsilon) = \sigma_{\epsilon}^{2}$  (constant).

a (2) Calculate by hand the least-squares estimates of intercept  $\alpha$  and slope  $\beta$ . Using  $\sum x = 3$ ,  $\sum y = 18.3$ ,  $\sum x^2 = 5$ ,  $\sum xy = 0 \times 5.8 + 1 \times 6.2 + 2 \times 6.3 = 18.8$ , n = 3, we get  $\hat{\beta} = \frac{\sum xy - \sum x \sum y/n}{\sum x^2 - (\sum x)^2/n/} = \frac{18.8 - 3 \times 18.3/3}{5 - 3^2/3} = 0.25$ , and  $\hat{\alpha} = \bar{y} - \hat{\beta}\bar{x} = 6.1 - 0.25 \times 1 = 5.85$ .

In reality a larger experiment was done. In total 25 soils were examined, using 5 levels of lime (0, 1, 2, 4, and 8 units), each applied to 5 randomly selected soils. Below we fit the simple linear regression model, regressing pH on rate.

```
> lime.reg <- lm(pH ~ rate, data=lime)</pre>
> summary(lime.reg)
Call:
lm(formula = pH ~ rate, data = lime)
Residuals:
                 1Q
                     Median
                                       3Q
     Min
                                                Max
-0.14663 -0.03783 -0.00128 0.03217
Coefficients:
             Estimate Std. Error t value Pr(>|t|)
(Intercept) 5.73783
                           0.02119 270.83 < 2e-16
               0.03172
                            0.00514
                                         6.17 2.7e-06
Residual standard error: 0.0727 on 23 degrees of freedom
Multiple R-squared: 0.624,
                                       Adjusted R-squared: 0.607
F-statistic: 38.1 on 1 and 23 DF, p-value: 2.68e-06
b (1) Give the estimates of \alpha, \beta, and \sigma_{\epsilon}.
\hat{\alpha} = 5.738, \, \hat{\beta} = 0.0317, \, \text{and } \hat{\sigma_{\epsilon}} = 0.0727.
```

- c (2) Give practical interpretations of  $\alpha$ ,  $\beta$ , and  $\sigma_{\epsilon}$ .
- $\alpha$  is the expected soil pH if no lime is applied;  $\beta$  is the expected change in soil pH for a unit change in lime amount;  $\sigma_{\epsilon}$  may be described as the expected deviation of the pH of a soil from the population mean at constant lime rate.
- d (2) In the output above, a number of hypothesis tests is performed. For all, give  $H_0$ ,  $H_1$ , test-statistics, P-values, and conclusions.

Three tests:

- 1. "Omnibus" F-test:  $H_0: \beta=0, H_1: \beta\neq 0, F=MSR/MSE$  with outcome F=38.1, P-value  $P(F_{1,23} \ge 38.1) = 2.68e - 06$ ; P-value < 0.05, so reject  $H_0$ , slope is not equal to zero, relationship lime and pH is proven.
- 2. t-test:  $H_0: \alpha=0, H_1: \alpha\neq 0, t=(\hat{\alpha}-0)/se(\hat{\alpha})$  with outcome t=270.83; P-value < 2e-16, so  $H_0$ is rejected, intercept is not zero; this is a test without meaning.
- 3. t-test:  $H_0: \beta=0, H_1: \beta\neq 0, t=(\hat{\beta}-0)/se(\hat{\beta})$  with outcome t=6.17; P-value  $P(|t_{23}|>6.17)=2.7e-6$ , so  $H_0$  is rejected, slope deviates from zero; notice that  $F=t^2$ : in simple linear regression omnibus F-test and t-test for slope are equivalent.
- e1 (2) Write down the model in matrix notation, making clear what is what.

$$\text{Linear model is } y = X\beta + \epsilon, \, \text{here:} \left[ \begin{array}{c} y_1 \\ y_2 \\ y_3 \\ y_4 \\ y_5 \\ y_6 \\ \dots \\ y_{24} \\ y_{25} \end{array} \right] = \left[ \begin{array}{c} 1 & 0 \\ 1 & 0 \\ 1 & 0 \\ 1 & 0 \\ 1 & 0 \\ 1 & 0 \\ 1 & 1 \\ \dots & \dots \\ 1 & 8 \\ 1 & 8 \end{array} \right] + \left[ \begin{array}{c} \epsilon_1 \\ \epsilon_2 \\ \epsilon_3 \\ \epsilon_4 \\ \epsilon_5 \\ \epsilon_6 \\ \dots \\ \epsilon_{24} \\ \epsilon_{25} \end{array} \right].$$

Second column of X contains the  $2\bar{5}$  values of lime

e2 (3) Write down the normal equations, and solve them. You may want to use that  $\sum_{i=1}^{25} X_i = 75$ ,  $\sum_{i=1}^{25} X_i^2 = 425$ ,  $\sum_{i=1}^{25} Y_i = 145.82$ , and  $\sum_{i=1}^{25} X_i Y_i = 443.82$ .

Normal equations are  $X'X\mathbf{b} = X'y$ , here in matrix notation:  $\begin{bmatrix} n & \sum x \\ \sum x & \sum x^2 \end{bmatrix} \begin{bmatrix} \hat{\alpha} \\ \hat{\beta} \end{bmatrix} = \begin{bmatrix} \sum y \\ \sum xy \end{bmatrix}.$  Fill in the numbers:  $\begin{bmatrix} 25 & 75 \\ 75 & 425 \end{bmatrix} \begin{bmatrix} \hat{\alpha} \\ \hat{\beta} \end{bmatrix} = \begin{bmatrix} 145.82 \\ 443.82 \end{bmatrix}.$  Hence two equations with two unknowns:  $\begin{bmatrix} 25\hat{\alpha} + 75\hat{\beta} & = 145.82 \\ 75\hat{\alpha} + 425\hat{\beta} & = 443.82 \end{bmatrix}.$  Solving, accord a great transfer of the first of the fi

Solving, second equation minus 3 times first equation gives  $200\hat{\beta} = 6.36$ , so  $\hat{\beta} = 0.0318$ . Then  $\hat{\alpha} =$  $(145.82 - 75 \times 0.0318)/25 = 5.7374$ . Notice slight differences with R-output due to rounding.

e3 (3) Show how the standard error of  $\hat{\beta}$  can be obtained using matrix calculation.

Let **b** be the least-squares solution  $\mathbf{b} = (X'X)^{-1}X'y$  with X and y as in e2. Then variance-covariance matrix of **b** is  $Var(\mathbf{b}) = \sigma_{\epsilon}^2 (X'X)^{-1}$ .

Now, the inverse  $X'X^{-1} = \begin{bmatrix} 25 & 75 \\ 75 & 425 \end{bmatrix}^{-1} = \frac{1}{25 \times 425 - 75^2} \begin{bmatrix} 425 & -75 \\ -75 & 25 \end{bmatrix}$ . The second element of the diagonal is needed:  $\frac{1}{25 \times 425 - 75^2} \times 25 = 25/5000 = 0.005$ . Variance  $\sigma_{\epsilon}^2$  is estimated as  $\hat{\sigma}_{\epsilon}^2 = MSE = 0.0727^2$ . Then,  $se(\hat{\beta}) = \sqrt{0.0727^2 \times 0.005} = 0.0051407$ , as given by R.

e4 (1) Sketch the geometrical interpretation of least squares estimation. For a geometrical interpretation look at figure 10.2 in Fox, page 222.

The simple linear regression may be too simple. Let's compare it with the model that gives a mean to each level of variable rate.

f (3) Calculate the F-statistic for the test of goodness of fit based on the residual sums of squares of the simple linear regression model and the one-way ANOVA model. How would you judge the significance of the outcome?

> deviance(lime.reg)

[1] 0.121455

```
> lime.anova <- lm(pH ~ as.factor(rate), data=lime)</pre>
```

> deviance(lime.anova)

#### [1] 0.09909

 $F = \frac{(SSE_{RM} - SSE_{FM})/(dfe_{RM} - dfe_{FM})}{SSE_{FM}/dfe_{FM}} \text{ with } RM = \text{reduced model} = \text{simple linear regression model, and } FM = \text{full model} = \text{one-way ANOVA model. Fill in: } F = \frac{(0.121455 - 0.09909)/3}{0.09909/20} = 1.5047.$ 

The test-statistic behaves under  $H_0$  like a random draw from a  $F_{3,20}$  distribution. Because the ratio 1.5 is not far from 1, we can say (with some knowledge of F-distributions, without looking in the F-table) that  $H_0$ : "Extra parameters to go from regression to ANOVA model not needed" will not be rejected. In other words, the simple linear regression is good enough, goodness-of-fit is ok.

g (1) Write down the ANOVA model, used above, in matrix notation.

The linear model  $y = X\beta + \epsilon$  for one-way ANOVA situation with overparameterization will be:

$$\begin{bmatrix} y_1 \\ y_2 \\ y_3 \\ y_4 \\ y_5 \\ y_6 \\ \dots \\ y_{24} \\ y_{25} \end{bmatrix} = \begin{bmatrix} 1 & 1 & 0 & 0 & 0 & 0 & 0 \\ 1 & 1 & 0 & 0 & 0 & 0 & 0 \\ 1 & 1 & 0 & 0 & 0 & 0 & 0 \\ 1 & 1 & 0 & 0 & 0 & 0 & 0 & 0 \\ 1 & 1 & 0 & 0 & 0 & 0 & 0 & 0 \\ \dots & (x_1) & (x_2) & (x_3) & (x_4) & (x_5) \\ 1 & 0 & 0 & 0 & 0 & 0 & 1 \\ 1 & 0 & 0 & 0 & 0 & 0 & 1 \end{bmatrix} \begin{bmatrix} \mu \\ \alpha_1 \\ \alpha_2 \\ \alpha_3 \\ \alpha_4 \\ \alpha_5 \end{bmatrix} + \begin{bmatrix} \epsilon_1 \\ \epsilon_2 \\ \epsilon_3 \\ \epsilon_4 \\ \epsilon_5 \\ \epsilon_6 \\ \dots \\ \epsilon_{24} \\ \epsilon_{25} \end{bmatrix}$$

Variables  $x_1 cdots x_5$  are five dummy variables for 5 levels of lime.

In the experiment two types of lime were used (AL and GL), both applied at rates 0, 1, 2, 4 and 8, in total to 50 soils. The effects of the two types of lime on soil pH may be different. We want to include both types of lime in the linear model, and make comparisons between them.

h1 (1) How are linear models of this type generally called? ANCOVA models = Analysis of covariance models

We fit a linear model that allows two separate regression lines for the two types of lime, see output below.

## > anova(lime.reg2)

## Analysis of Variance Table

```
Response: pH

Df Sum Sq Mean Sq F value Pr(>F)

type 1 0.5243 0.5243 142.94 1.03e-15

rate 1 1.3783 1.3783 375.78 < 2e-16

type:rate 1 0.2911 0.2911 79.36 1.41e-11

Residuals 46 0.1687 0.0037
```

h2 (2) Write down the fitted regression lines for the two types of lime.

```
typeAL: \hat{y} = 5.781 + 0.0857 \times rate
typeGL: \hat{y} = 5.781 - 0.043 + (0.0857 - 0.0540) \times rate = 5.738 + 0.0317 \times rate.
```

h3 (2) Test  $H_0$ : "two regression lines run parallel". Give test-statistic, P-value and conclusion.

Either F-test or t-test (because it concerns a single parameter):

F-test: TS is  $F = ((SSE_{parallel lines} - SSE_{FM})/1)/MSE_{FM}$  with outcome F = 79.4, P-value=  $P(F_{1,46} > 79.4) << 0.0001$ , so reject  $H_0$ : different slopes.

t-test: TS is  $t = ((\hat{\beta}_{\text{typeGL:rate}} - 0)/se_{\text{typeGL:rate}})$  with outcome t = -0.054/0.006 = -8.91, P-value=  $2P(t_{46} < -8.91) << 0.0001$ , so reject  $H_0$ : different slopes.

h4 (2) Compare the P-values for the t-tests for the individual coefficients, with the P-values for the F-tests in the anova-table. Which P-values are different and which are the same? Explain.

The ANOVA table contains sequential sums of squares for type, rate, and type:rate, in that order. In that case, the F-tests make the follow model comparisons, all concerning single degrees of freedom:

for type: intercept -> intercept + type

for rate: intercept + type -> intercept + type + rate

for type:rate: intercept + type + rate -> intercept + type + rate + type:rate

The t-tests for individual coefficients are partial tests, so that the effect of a variable is judged in presence of all other variables. So, for instance the coefficient of type is the difference in intercepts of the two lime types, but in the model with different slopes. So, this concerns the model comparison intercept + rate + type:rate -> intercept + type + rate + type:rate, which differs from the F-test done above. Summarizing, the interaction-test for difference in slopes gives identical P-values for F and t (because it concerns identical model comparisons), but the tests for type and rate do not.

If all regressors were uncorrelated (which is not the case), would the t-tests and F-test produce identical results here.

h5 (3) Show, using the output above, that the intercepts are not significantly different. This suggests that a model with common intercept is sufficient here. Comment on this model in the light of the principle of marginality.

The P-value from the t-test for comparison of intercepts is 0.092. Testing at  $\alpha = 0.05$ , we do not reject the null hypothesis of equal intercepts. If we fit a model without type, hence forcing the intercepts to be equal, we violate the principle of marginality, because the model contains the interaction (type:rate), but only one of the "main effects" (rate). Still, this model would make perfect sense.

2 (11). In a study on sleeping behaviour of animals, the sleep duration is related to a number of explanatory variables: log(body weight), log(brain weight), log(lifespan), log(gestation), predation ((1=least likely to be preyed upon, 5=most likely to be preyed upon), and exposure (1=least exposed, 5=most exposed).

We fit a multiple linear regression model, explaining totalsleep from all 6 quantitative regressors.

```
> sleep.lm <- lm(totalsleep ~ logbodywgt + logbrainwgt + loglifespan + loggest + exposure + predation,
+ data=sleep)
> summary(sleep.lm)
Call:
lm(formula = totalsleep ~ logbodywgt + logbrainwgt + loglifespan +
    loggest + exposure + predation, data = sleep)
Residuals:
     Min
               10
                    Median
                                 30
                                         Max
-6.90186 -1.68891 -0.26267 1.61512 6.40439
Coefficients:
             Estimate Std. Error t value Pr(>|t|)
(Intercept) 22.053211 3.075389 7.1709 6.427e-09
logbodywgt -0.063608
                       0.524492 -0.1213
                                           0.90403
logbrainwgt -0.659306
                       0.781667 -0.8435
                                           0.40353
loglifespan 0.584594
                        0.842920 0.6935
                                           0.49162
loggest
            -1.682804
                        0.738533 -2.2786
                                           0.02760
exposure
            -0.030104
                        0.611942 -0.0492
                                           0.96099
                       0.491140 -2.4604
predation
           -1.208401
                                           0.01787
Residual standard error: 3.0496 on 44 degrees of freedom
Multiple R-squared: 0.62624,
                                    Adjusted R-squared: 0.57528
F-statistic: 12.287 on 6 and 44 DF, p-value: 4.5068e-08
```

a (2) Give the P-value of the "omnibus" F-test and compare it with the P-values of the t-tests for the individual coefficients. Comment on the difference. What could be the reason?

Omnibus F-test: P=4.5e-8. So,  $H_0$ : "all regression coefficients zero" is strongly rejected, there is explanatory power in this model. Individual t-tests: most P-values are large, smallest (0.018) is for predation. Individual t-tests tell that effects are absent or just significant. Reason is multicollinearity: t-test measures effect of a regressor, partial to all other regressors. Due to correlation between regressors, an individual regressor is not contributing a lot extra.

b1 (3) Give the assumptions that underly linear models.

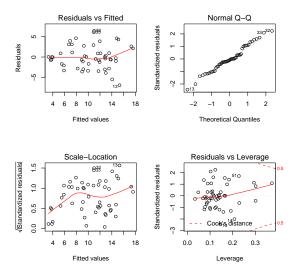
In linear models errors  $\epsilon_i$  are assumed to

- 1) be independent
- 2) have constant variance
- 3) be normally distributed
- 4) have expectation zero

In short:  $\epsilon_i$  are i.i.d.,  $\sim N(0, \sigma_{\epsilon}^2)$ 

b2 (4) Check whether there are problems with those assumptions using the plots and output below. Comment on all residual plots (skip the plot left-below), but also on collinearity (see output next page).

```
> plot(sleep.lm)
```



#### > vif(sleep.lm)

```
logbodywgt logbrainwgt loglifespan loggest exposure predation 14.8986711 20.9921439 3.5710519 3.1901759 5.0180650 2.8723461
```

- \*) Plot residuals vs fitted values is used for checking constant variance. There are slight indications that variance increases with mean: variability (in y-direction) seems to increase for larger fitted values.
- \*) Normal QQ-plot is used for checking normality. Points lay reasonably well on straight line, no indications of deviations from normality.
- \*) Plot of leverage vs standardized residuals can be used to check for potential influence (leverage), and regression outliers. All standardized residuals are roughly in the range (-2,2), so no problems here. There are 4 observations with leverage exceeding the threshold  $2 \times p/n = 2 \times 7/51 = 0.27$ . The standardized residuals are not large for these observations, though, does not look problematic.
- \*) Variables logbodywgt and logbrainwgt have VIF values exceeding the threshold value 10: each of the two is highly related to one or more other regressors. This will cause problems, like inflated standard errors. In subquestion 2a we already saw the effects of multicollinearity.
- c (2) Using stepwise regression a smaller model is obtained. Interpret the first step of the backward elimination procedure below. Which criterion is used for selection and how is it defined? Which variable will be removed from the model after this step?

## > step(sleep.lm)

```
Start: AIC=120.2
totalsleep ~ logbodywgt + logbrainwgt + loglifespan + loggest +
    exposure + predation
              Df Sum of Sq
                                R.S.S
- exposure
               1
                     0.023 409.237 118.206
- logbodywgt
               1
                     0.137 409.351 118.220
- loglifespan
               1
                     4.473 413.688 118.758
- logbrainwgt
                     6.616 415.831 119.021
<none>
                            409.214 120.203
- loggest
               1
                    48.286 457.501 123.892
                    56.300 465.514 124.777
- predation
               1
Step: AIC=118.21
totalsleep ~ logbodywgt + logbrainwgt + loglifespan + loggest +
   predation
```

```
Df Sum of Sq
                               RSS
                     0.188 409.425 116.229
- logbodywgt
              1
- loglifespan 1
                     4.867 414.104 116.809
- logbrainwgt 1
                     6.681 415.918 117.032
<none>
                           409.237 118.206
- loggest
                    54.772 464.008 122.612
                   153.289 562.526 132.431
- predation
Step: AIC=116.23
totalsleep ~ logbrainwgt + loglifespan + loggest + predation
              Df Sum of Sq
                               RSS
                                       AIC
                     5.506 414.931 114.911
- loglifespan 1
<none>
                           409.425 116.229
- logbrainwgt
                    42.383 451.809 119.253
- loggest
               1
                    54.998 464.424 120.658
- predation
                   157.247 566.673 130.806
               1
Step: AIC=114.91
totalsleep ~ logbrainwgt + loggest + predation
              Df Sum of Sq
                               RSS
                                       AIC
<none>
                           414.931 114.911
                    40.679 455.610 117.680
- logbrainwgt 1
- loggest
                    51.611 466.542 118.890
               1
- predation
                   177.374 592.305 131.062
Call:
lm(formula = totalsleep ~ logbrainwgt + loggest + predation,
                                                                  data = sleep)
Coefficients:
                                         predation
(Intercept) logbrainwgt
                              loggest
   23.08575
                -0.58799
                             -1.61560
                                          -1.28089
```

The AIC is used for backward elimination.  $AIC = -2\log(\text{likelihood}) + 2p$  with p the number of parameters in the model, smaller values point to better fitting models. Each variable is removed from the model in turn, and the resulting AIC's are reported. For 2 regressors the AIC deteriorates (becomes larger) by removal, so these variables are important. For 4 regressors removal makes the AIC smaller (better), so these regressors are candidates for removal. The largest improvement of AIC is obtained by the removal of exposure, so this will be the first variable to be removed.

3 (14). In an experiment 48 rats were exposed to a poison (3 types: I, II, III) and a (not further specified) treatment (4 treatments, labeled A, B, C, D). Each combination of poison and treatment was applied to exactly 4 rats, so we have a balanced design. The survival time in tens of hours was measured. Here we analyze the  $y = \log(\text{survival time})$  using a two-way ANOVA model.

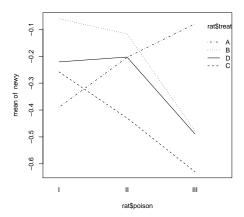
The two-way ANOVA model with interaction in means notation is  $y_{ijk} = \mu_{ij} + \epsilon_{ijk}$ , with i = 1, 2, 3 for poisons, j = 1, ..., 4 for treatments, and k = 1, ..., 4 for replicates; furthermore,  $\operatorname{var}(\epsilon_{ijk}) = \sigma_{\epsilon}^2$ .

a (1) Write down this model in effects notation.

$$y_{ijk} = \mu + \alpha_i + \beta_j + \alpha \beta_{ij} + \epsilon_{ijk}$$

b (3) Explain what it means if factors poison and treatment interact. Sketch a "profile plot" that would indicate a situation with interaction.

Interaction between poison and treatment means that the effect of poison is not the same for all four treatments. An example of a profile plot with interaction is given below, showing poison A with different response pattern over the treatments (does not correspond to data in this question):



c (2) Use the output below to test the null-hypothesis of "no interaction". Give  $H_0$ , test statistic and outcome, P-value and conclusion.

```
> rat.aov <- aov(y ~ poison + treat + poison*treat, data=rat)
> anova(rat.aov)
```

## Analysis of Variance Table

# Response: y

Df Sum Sq Mean Sq F value Pr(>F)
poison 2 0.987849 0.493924 48.43244 6.1953e-11
treat 3 0.670925 0.223642 21.92948 2.9868e-08
poison:treat 6 0.074642 0.012440 1.21986 0.31894
Residuals 36 0.367136 0.010198

> tapply(rat\$y, rat\$poison, mean)

 $H_0$ : "no interaction"  $\Leftrightarrow H_0: \alpha\beta_{11} = \alpha\beta_{12} = \ldots = \alpha\beta_{34}$ Test statistic  $F = \frac{SS_{\text{poison:treat}/6}}{MSE}$  with outcome 1.22, and P-value  $P(F_{6,36} \ge 1.22) = 0.32$ . P-value > 0.05, so do not reject  $H_0$ , we cannot find evidence for interaction between poison and treatment. d1 (1) Estimate the difference in expected log(surval time)  $(\mu_{1.} - \mu_{2.})$  between poison types I and II (averaged over the treatments).

 $y_{1..}-y_{2..}=-0.2324-(-0.3135)=0.081$ , so log(survival time) is on average 0.0815 longer for poison I compared to poison II.

d2 (2) To determine whether this difference is significant, the standard error of difference (sed) is needed. Write the sed $(y_{1..}-y_{2..})$  as a function of  $\sigma_{\epsilon}^2$ .

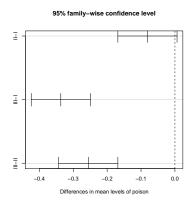
First look at the variance of the difference (sed<sup>2</sup>): sed<sup>2</sup> $(y_{1..}-y_{2..})=\sigma_{\epsilon}^2/16+\sigma_{\epsilon}^2/16=\sigma_{\epsilon}^2/8$ , because 16 rats were used for poisson I and 16 for II. So sed $(y_{1..}-y_{2..})=\sigma_{\epsilon}/\sqrt{8}$ .

d3 (3) Estimate this standard error, and use it get the Least Significant Difference LSD (using 5% significance level; you may use t-value 2.028). Is the difference significant?

$$\widehat{\text{sed}}(y_{1..} - y_{2..}) = \sqrt{MSE}/\sqrt{8} = \sqrt{0.010198}/\sqrt{8} = 0.0357.$$
  $LSD = t_{36}(0.975) \times \text{sed}(y_{1..} - y_{2..}) = 2.028 \times 0.0357 = 0.0724.$  The estimated difference is 0.0815, which is larger than  $LSD = 0.0724$ , so we would call the difference significant.

d4 (2) Tukey's multiple comparison approach is an alternative. Are poisons I and II significantly different according to the graph below? Compare with d3).

> plot(TukeyHSD(rat.aov, "poison"))



I and II are not signficantly different according to the Tukey approach, because the Tukey 95% confidence interval for  $\mu_{1.} - \mu_{2.}$  does contain the value 0.

- 4 (14). In a study on the research productivity of PhD students in biochemistry, the number of articles published by the student during the last three years (art) is related to a number of explanatory variables: number of kids five years or younger (kid5), number of articles published by the mentor (ment), gender (fem). Variable fem is an indicator variable for female. We assume an additive model at this moment.
- a (2) Write down a generalized linear model that is appropriate for the data we have here. Pay attention to the three components of a glm.
- 1) Random part of the model:  $art_i \sim Poisson(\mu_i)$ ,  $art_i$  independent (i = 1, ..., 915)
- 2) Systematic part of the model: linear predictor  $\eta_i = \beta_0 + \beta_1 \operatorname{kid5}_i + \beta_2 \operatorname{ment}_i + \beta_3 \operatorname{fem}_i$
- 3) Link function:  $log(\mu_i) = \eta_i$ .

Number of Fisher Scoring iterations: 5

b (3) Interpret the coefficients given below (assuming that the model fits well), both in terms of significance as in terms of direction of the effect, and effect on the expected number of articles.

#### > head(articles)

```
fem ment kid5 art
    0
         8
              2
                  3
2
    0
         7
              0
                  0
3
    0
        47
              0
                  4
4
    0
        19
              1
                  1
5
    0
         0
              0
                  1
    0
> articles.glm <- glm(art ~ kid5 + ment + fem, family=poisson(link=log), data=articles)</pre>
> summary(articles.glm)
glm(formula = art ~ kid5 + ment + fem, family = poisson(link = log),
    data = articles)
Deviance Residuals:
    Min
                10
                      Median
                                     30
                                              Max
-3.57458 -1.56330
                    -0.36517
                                0.55834
                                          5.55137
Coefficients:
              Estimate Std. Error z value Pr(>|z|)
            0.4367326  0.0468254  9.3268 < 2.2e-16
(Intercept)
kid5
            -0.1437870 0.0365734 -3.9315 8.443e-05
ment
             0.0256452  0.0019523  13.1361 < 2.2e-16
            -0.2431065 0.0542073 -4.4848 7.300e-06
fem
(Dispersion parameter for poisson family taken to be 1)
   Null deviance: 1817.41 on 914 degrees of freedom
Residual deviance: 1640.85 on 911 degrees of freedom
AIC: 3316.59
```

All three regression coefficients are significantly different from zero, with the strongest effect (in terms of P-value) for ment (number of articles published by the mentor).

The higher the number of kids, the lower number of articles. For every extra kid the predicted number of articles has to be multiplied by  $e^{-0.144} = 0.866$ , so a relative decrease of 100-87%=13%.

The higher the output of the mentor, the larger the number of articles. Every extra article of the mentor leads to a multiplication factor of  $e^{0.0256} = 1.026$ , so a relative increase of 2.6%.

Females have lower number of articles than males: females produce on average  $e^{-0.243} = 0.784$  times the number of articles of males, so a relative decrease of 100-78%=22%.

c (3) Explain the differences in "Analysis of deviance" tables obtained by anova and drop1 statements, given on the next page.

```
> anova(articles.glm, test = "Chisq")
Analysis of Deviance Table
Model: poisson, link: log
Response: art
Terms added sequentially (first to last)
      Df Deviance Resid. Df Resid. Dev
                                         P(>|Chi|)
NULL
                         914
                               1817.405
kid5
            4.196
                               1813.210
                                              0.041
                         913
          152.049
                         912
                               1661.161 6.1814e-35
ment
           20.309
                               1640.851 6.5884e-06
fem
                         911
> drop1(articles.glm, test = "Chisq")
Single term deletions
Model:
art ~ kid5 + ment + fem
       Df Deviance
                        AIC
                                LRT
                                       Pr(Chi)
           1640.85 3316.59
<none>
           1657.00 3330.74
kid5
                              16.15 5.8470e-05
           1781.51 3455.25
                             140.66 < 2.22e-16
ment
           1661.16 3334.90
                              20.31 6.5884e-06
fem
```

The anova function sequentially adds variables to the model, starting with an intercept-only model. The column labeled 'Deviance' shows the *change* in residual deviance by entering this variable to the model. The residual deviance itself is given in column 'Resid.Dev'.

The drop1 function drops a single term, always from the full model. Notice that R is not consequent in naming the output, because the column labeled 'Deviance' now shows the residual deviance, corresponding to the column 'Resid. Dev' in the anova output. The column 'LRT' contains the differences in deviance, always starting from the full model.

Comparing column 'Deviance' from anova with column 'LRT' from drop1 shows differences, because the model comparisons are different for sequential ("Type I" in SAS terminology) and partial ("Type II") approaches, as we saw earlier in question 1h4. Only for the last term fem, the model comparisons are identical, and hence are the deviances (20.31).

d (3) Check the goodness of fit of the model by interpretation of the deviance. What value would you expect if all is well, and how "significant" is the deviation here?

Look at the residual deviance in the full model, which should behave like a random draw from a  $\chi^2_{911}$  distribution if model fits well. A  $\chi^2_{911}$  distributed variable has expectation 911, and variance  $2 \times 911 = 1822$ , or standard deviation 42.7. Here, the residual deviance equals 1640.85 on 911 d.f. Notice that the deviance is much larger than would be expected by change (average value would be 911). Its value of 1640.95 is close to 20 s.d.'s above the mean, so the deviation is highly "significant". Conclusion: goodness of fit is not ok.

e (2) Describe how you would handle the problem (if needed at all), and what the consequences would be for the conclusions in b).

There may be different causes for a high deviance value, like non-fitting systematic part of model, outliers, or clustered responses (like the same mentor having different PhD students). It is not clear what the causes are here. A simple and rather standard approach is to allow for overdispersion by inclusion of an extra scale parameter. All standard errors are multiplied with factor  $\sqrt{1640.85/911} = 1.34$  (and instead of  $\chi^2$  tests F-tests are done). As the denominators of the Wald test statistics become larger, the Wald test-statistics themselves are closer to zero, and P-values increase. E.g. for kid5 Wald statistic z=2.93. The results will remain significant here, though.

f (1) Mention another type of application of Poisson glm's.

Poisson glm's can also be used for analysis of contingency tables.

- 5 (10). Generalized Linear Models: general theory.
- a (2) Write the Poisson distribution as a member of the exponential family.

Let  $y \sim Poisson(\lambda)$ , then probability distribution is  $f(y) = \frac{e^{-\lambda}\lambda^y}{y!}$ . The log-likelihood is  $LL(\lambda) = \log(\frac{e^{-\lambda}\lambda^y}{y!}) = -\lambda + y\log(\lambda) - \log(y!) = \frac{y\log(\lambda) - \lambda}{1} + (-\log(y!)) = \frac{y\log(\lambda) - \exp(\log(\lambda))}{1} + (-\log(y!))$ . Compare this with the general formula for the exponential family:  $LL = \frac{y\theta - b(\theta)}{\phi} + c(y, \phi)$ .

- b (4) Specify using this notation: 1) the canonical parameter; 2) the dispersion parameter; 3) the expectation  $\mu$ ; 4) the variance function  $V(\mu)$ ; 5) the canonical link function.
- 1) canonical parameter is  $\theta = \log(\lambda)$
- 2) dispersion parameter  $\phi = 1$
- 3) function  $b(\theta) = \exp(\theta)$ , so that expectation  $\mu = b'(\theta) = \frac{d}{d\theta} \exp(\theta) = \exp(\theta) = \exp(\log(\lambda)) = \lambda$ 4) variance function  $V(\mu) = b''(\theta) = \frac{d^2}{d\theta^2} \exp(\theta) = \exp(\theta) = \lambda$
- 5) canonical link function is link function which brings the mean to the scale of the canonical parameter; it can be obtained by starting with the expectation  $\mu = b'(\theta)$ , so that  $\theta = (b')^{-1}(\mu)$ , and the link function is  $(b')^{-1}$ . Here,  $b'(\theta) = \exp(\theta)$ , with inverse log. So, the canonical link function is the log function (natural logarithm).
- c (1) Give the name of the estimation principle in glm.

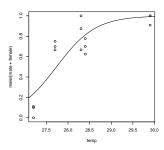
Maximum likelihood estimation.

- d (1) Different types of tests are used in glm. Mention at least two.
- 1) Likelihood ratio tests; 2) Wald tests; 3) score tests.
- e (2) Explain the (residual) deviance.

The (residual) deviance of a model is twice the difference in maximized log-likelihood of the saturated model and the current model. The saturated model is the model with as many parameters as observations (and as such useless), resulting in perfect predictions. The maximized log-likelihood of this model is the highest possible likelihood obtainable for the given set of data. The deviance is twice the difference between this log-likelihood and that of the current model. In easier words, it tells how much can be gained by making the model larger than the current model. It is like a residual sum of squares, which measures how much the model can explain more by extending the current model until predictions are perfect (in which case the residual sum of squares is zero).

6 (11). In a study of the effect of incubation temperature on the sex of turtles, an experiment was conducted with 5 temperatures. The number of male and female turtles born was recorded. We analyze the data using logistic regression.

```
> glmturtle <- glm(cbind(male,female) ~ temp, family=quasibinomial (link=logit), data=turtle)
> summary(glmturtle)
Call:
glm(formula = cbind(male, female) ~ temp, family = quasibinomial(link = logit),
    data = turtle)
Deviance Residuals:
     Min
                      Median
                                    30
                10
                                             Max
-2.07212 -1.02915
                    -0.27137
                               0.80867
                                         2.55496
Coefficients:
             Estimate Std. Error t value Pr(>|t|)
                       17.08133 -3.5898 0.003296
(Intercept) -61.31832
              2.21103
                         0.61216 3.6119 0.003159
(Dispersion parameter for quasibinomial family taken to be 2.0186618)
    Null deviance: 74.5080 on 14 degrees of freedom
Residual deviance: 24.9425 on 13 degrees of freedom
AIC: NA
Number of Fisher Scoring iterations: 5
> plot(male/(male+female) ~ temp, data=turtle)
> ld <- data.frame(temp=seq(26.9,30,by=0.1))</pre>
> phat <- predict(glmturtle, ld, type="response")
> lines(phat ~ ld$temp)
```



a (2) Write down the fitted model.

$$\log(\frac{p}{1-p}) = a + b \text{ temp} = -61.32 + 2.211 \text{ temp}$$
, with p the probability of a male.

b (3) Use the fitted model to find the temperature T75 where 75% of the turtles are males.

For 75% males, logit is  $\log(0.75/0.25) = 1.0986$ . So, we want to find the temperature T75 for which 1.0986 = -61.32 + 2.211 temp. It then follows that  $T75 = (1.0986 - (-61.32))/2.211 = 28.23^{\circ}$ .

c (4) Give an approximate standard error of T75 using the delta method. Below is the estimated variance-covariance matrix V of the parameters, that you may need.

```
> sigma2 <- sum(residuals(glmturtle,type="pearson")^2)/13
> (V <- sigma2 * summary(glmturtle)$cov.unscaled)</pre>
```

(Intercept) 291.769 -10.45456 (Intercept) -10.455 temp

T75 = f(a,b) = (1.0986 - a)/b, and we need an approximate variance of T75 by the delta method:  $V(T75) \approx (\frac{\partial f(a,b)}{\partial a})^2 v_{11} + (\frac{\partial f(a,b)}{\partial b})^2 v_{22} + 2 \times \frac{\partial f(a,b)}{\partial a} \frac{\partial f(a,b)}{\partial b} v_{12}$ , everything evaluated at maximum likelihood estimates.

First the partial derivative w.r.t. a:  $\frac{\partial (1.0986-a)/b}{\partial a} = -1/b$ , evaluated at mle: -1/2.21103 = -0.45228. Next the partial derivative w.r.t. b:  $\frac{\partial (1.0986-a)/b}{\partial b} = -(1.0986-a)/b^2$ , evaluated at mle:  $-(1.0986-a)/b^2$ 

 $(-61.318))/2.211^2 = -12.7677.$ 

Combining:  $V(T75) \approx (-0.45228)^2 \times 291.769 + (-12.7677)^2 \times 0.37473 + 2 \times -0.45228 \times -12.7677 \times 0.37473 + 2 \times -0.45228 \times -0.45228$ -10.45456 = 59.68345 + 61.0863 - 120.7413 = 0.02845 and standard error  $\sqrt{0.02845} = 0.169$ .

d (2) Give an approximate 95% confidence interval for T75, using the approximate standard error from c). (Use standard error 0.15 if you don't have the answer from c ).

Approximate 95 % c.i. for T75 is  $28.23 \pm 1.96 \times 0.169 = (27.90, 28.56)$ .