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**Biological Control of the Cycad
Aulacaspis Scale, *Aulacaspis
yasumatsui***

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1. Abstract - Cave

2. Introduction - Cave

3. Economic impact of CAS - Cave and Wright

4. Ecological impact of CAS - Moore

Ecological impact of CAS invasions varies greatly with location, largely due to differences in characteristics of host plant populations, climate, and presence of natural enemies. When CAS arrived in Florida (1995) and Hawaii (1998), it became an economic pest of ornamental cycads which could be protected using a combination of pesticide applications and biological control. However, when CAS arrived in Guam (2003), it started out as an economic pest of ornamental cycads *Cycas revoluta* and *Cycas micronesica* but rapidly spread to the wild *Cycas micronesica* population, causing an uncontrolled island-wide outbreak. This chapter will be focused on the ecological impact to *C. micronesica* and the forest ecosystem on Guam.

4.1. *C. micronesica* taxonomy and pre-invasion status

Cycas micronesica was described as a species in 1994 (Hill [1994](#)). Prior to description, it was identified as *C. rumphii* or *C. circinalis*. This tree-sized cycad is endemic to Micronesia and it is the only endemic gymnosperm on these islands. It grows in the wild in a wide variety of habitats on Guam, the Northern Mariana Islands, the Yap Islands, and the Palau Islands. Individuals are either male or female.

Based on an island-wide census of Guam's trees performed by the U. S. Forest Service in 2002, a year before arrival of CAS, *C. micronesica* was identified as the most abundant tree in Guam's forests Donnegon et al. 2004. It thrived because it had evolved tolerance to local abiotic threats such as typhoons and drought and for this reason it was often used as a low maintenance ornamental plant. Indigenous people of the Mariana Islands, the Chamorros, named this plant *fadang* and used it as a source of starch. But there is no evidence that the plant was ever planted as a crop.

4.2. First detection and invasive pathway for arrival of *A. yasumatsui* on Guam

Arrival of CAS on Guam was predicted: on February 13 2000 T. E. Marler published an article in the Gardening section of the Pacific Daily News entitled *Looking out for scale insects* (Haynes and Thomas E. Marler 2005). Alarmed by establishment of CAS in Hawaii, Marler warned of pending arrival on Guam and pleaded for a ban cycad imports to the island.

CAS was first detected in the Tumon Beach hotel area of Guam near the end of 2003 on *C. micronesica* and *C. revoluta* growing as ornamental plants at two hotels. In those days, almost every hotel on Guam had cycad displays near their entrances.

The invasion pathway along which CAS travelled to Guam is unknown. It is likely that this pest arrived via importation of infested cycads from Hawaii, Florida or elsewhere. However, there is no evidence of this: there are no records of legal cycad importation to Guam in the two years prior to detection of CAS on the island (R. Campbell, Guam Plant Inspection Facility, personal communication to AM).

An intriguing alternative is that CAS arrived on Guam as crawlers. For several years prior to arrival, there was an active infestation of CAS on *C. revoluta* growing in an outdoor garden at the Honolulu International Airport located within a few hundred meters of where passengers boarded a daily 7.5 hour flight to Guam. Possibly, crawlers were carried on clothing of passengers visiting this garden or airborne scale crawlers may have been blown into cargo holds, wheel wells, or other spaces on Guam-bound aircraft. The location of initial CAS detection sites in Tumon Bay are only about 1 km downwind of the Guam International Airport.

4.3. Impacts of CAS on the Guam population of *C. micronesica*

The Guam CAS outbreak, which was severe and spread rapidly throughout the island (Table 4.3, Figure 1), was well documented by Marler and others. Within 12 years, *C. micronesica* went from being the most abundant tree in Guam's forests to being listed under the U.S. Endangered Species Act. It should be noted that since arrival of CAS, several other recently arrived invasive insect species and even some native species have begun attacking and damaging Guam's cycads (T. Marler and R. Muniappan 2006, Moore, R. H. Miller, T. E. Marler, et al. 2013, (Deloso et al. 2020)). However, CAS is considered to be the prime driver of population decline because it is the only herbivore causing damage severe enough to result in mortality.

Rapid decline of the Guam *C. micronesica* population was documented by Thomas E. Marler and Krishnapillai 2020 (Figure 1). Data are annual cycad stem counts in 120 permanent plots established after infestation of Guam's wild cycads, but prior to any mortality. It can be seen in the figure that cycad stem count declined to only 12.5% of the original within the first 3 years of the survey (2005-2008) and continued to decline to only 4% of the original count in succeeding years (2009-2020). In addition to high plant mortality surviving cycads stopped reproducing in the Guam plots: the last seedling (0-10 cm tall) was seen in 2006 and the last juvenile (10-100 cm tall) was seen in 2014. Between 2014 and 2020 the annual mortality rate for mature plants was relatively stable at about 14 per ha.

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Table 1: Timeline for the Guam CAS infestation.

2000	T. E. Marler predicts of arrival of CAS on Guam in a Pacific Daily News article (Haynes and Thomas E. Marler 2005) in response to detection of this pest in Florida during 1996 (Howard et al. 1999) and Hawaii during 1998 (Heu, Chun, and Nagamine 2003)
2002	An initial US Forest Service survey indicates that <i>C. micronesica</i> is Guam’s most abundant tree (with stem diameter greater than 5 inches) and estimates a population size of 1,571,556 (Donnegon et al. 2004)
2003	CAS first detected on ornamental <i>Cycas revoluta</i> and <i>C. micronesica</i> at hotels in Tumon Bay
2006	<i>C. micronesica</i> added to the IUCN Red List of Endangered and Threatened Species
2013	A second US Forest Service survey ranks <i>C. micronesica</i> , misidentified in the report as <i>C. circinalis</i> , is Guam’s xxrd most abundant tree (with stem diameter greater than 5 inches) and estimates a population size of n,nnn,nnn (Lazaro et al. 2020)
2015	<i>C. micronesica</i> listed as a threatened species under the US Endangered Species Act (United States Government 2015)

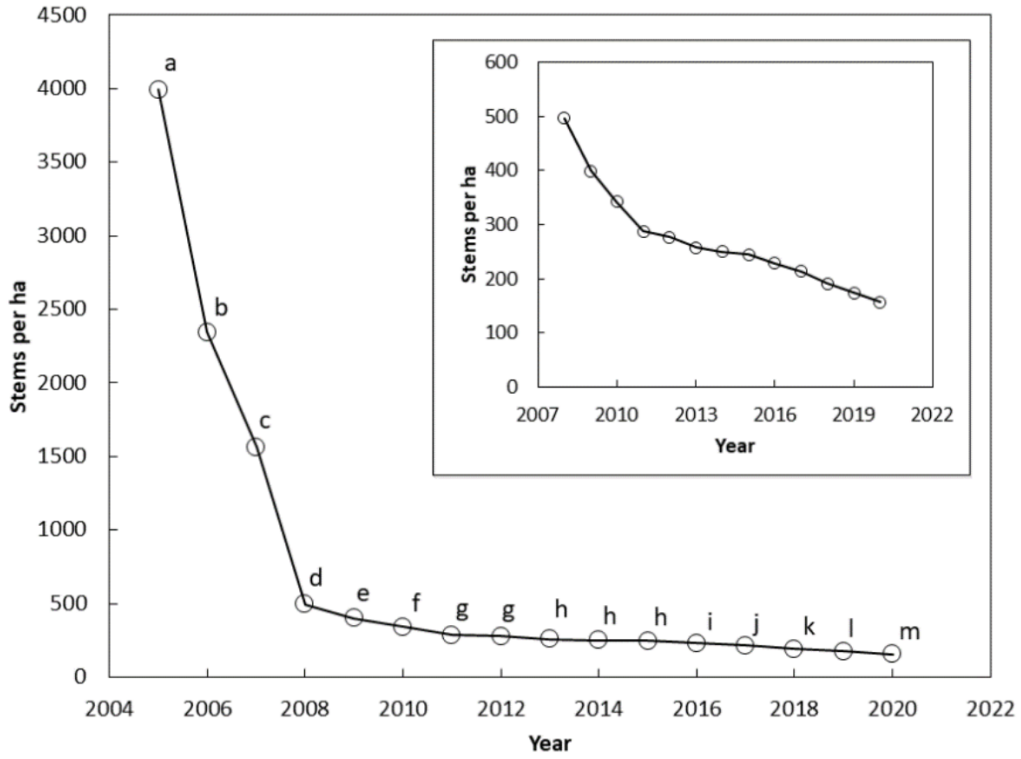


Figure 1: [From Thomas E. Marler and Krishnapillai 2020] The number of *Cycas micronesica* stems per ha (all size categories) among 12 Guam habitats from 2005 until 2020. The inset shows results from 2008 until 2020 with smaller vertical axis range. Ordinates of markers with the same letter are not significantly different.

4.4. Impacts of CAS on *C. micronesica* individuals

Immediate impacts of CAS on health of *C. micronesica* individuals is very obvious. On Guam, unprotected plants typically die within 18 months (REF) after being infested. Several overlapping generations of CAS totally encrust all leaves within a few months of infestation. Plant mortality is caused by a combination of nutrient removal by CAS sap feeding plus blockage of photosynthesis. Scales covers of dead insects are tightly affixed to the plant and these do not easily wash off. So photosynthesis may be blocked even after scale insects are killed. RANGASWAMY Muniappan et al. 2012 suggest that toxic effects of CAS saliva may also be involved in cycad mortality.

Secondary impacts of CAS on health of *C. micronesica* individuals are not obvious.

4.4.1. Mature plants which have survived CAS infestation have reduced reproductive capacity

Perhaps the most important secondary impact is much lower reproductive capability in plants recovering from CAS-infestation.

Thomas E. Marler and Gil N. Cruz 2019 reported that seeds from CAS-infested plants are deficient in nonstructural carbohydrates and germination rates are much lower: 43% of seeds from healthy plants germinated versus 7% of seeds from infested plants.

In addition, Thomas Marler and Terry 2021 reported that mature male plants which survived the initial CAS invasion have significantly smaller cones than prior to infestation. Average cone volume was 24% of pre-infestation levels in 2007 and this increased to 57% in 2021.

4.4.2. Mature plants which have survived CAS infestation are more susceptible to typhoon damage

C. micronesica and other endemic plants on Guam have evolved tolerance for typhoon strength winds (greater than 118 km/h) because tropical cyclones frequently visit this island. (Thomas Marler 2013) reported on results of nondestructive winching stress tests performed on *C. micronesica* stems to simulate effects of typhoon strength winds. Stems of plants which had not been infested by CAS were significantly stiffer than those which had been infested by CAS for either two or five years. Marler hypothesized from this finding that CAS-infested plants were much more susceptible to stem failure during typhoons.

Evidence supporting this hypothesis came two years after the original article was published, from a damage survey after Typhoon Dolphin passed over Guam on May 15, 2015. Thomas E Marler, J. H. Lawrence, and G. Cruz 2016 compare the level of damage from Typhoon Dolphin with that of a previous cyclone, Supertyphoon Paka:

Supertyphoon Paka caused severe damage to Guam's forest resources in 1997 when the *C. micronesica* population was healthy and not threatened by any known invasive insect herbivores. Less than 2% of the healthy *C. micronesica* population exhibited windsnap damage during peak winds of 298 km/h. In contrast, Typhoon Dolphin's peak winds of only 170 km/h. caused windsnap of 6% (mean of our eight sampling sites) of Guam's unhealthy *C. micronesica* population after only 10 years of *A. yasumatsui* infestations.

Windsnap is synonymous with *stem failure*. It is interesting to note that, because energy is a function of the square of wind speed, maximum winds generated by Paka were about 3 times more powerful than those of Dolphin.

4.5. CAS-induced cascading impacts on Guam's forest ecosystems

An *ecological cascade effect* is a series of secondary extinctions that are triggered by the primary extinction of a key species in an ecosystem. *C. micronesica* is considered to be a key species in Guam forest ecosystems because it was the most abundant tree prior to arrival of CAS. Loss of this plant is likely to threaten survival other organisms.

4.5.1. Herbivores which depend on *C. micronesica* for food

Marianus fruit bat, *Pteropus mariannus* Haynes and Thomas E. Marler 2005 reported:

The fleshy, aromatic covering of fadang seeds is a preferred food item for the endangered Mariana fruit bat, *Pteropus marianus marianus*. Fadang is so resistant to most types of disturbance that its seeds are sometimes the only bat food item available in the forest following the destructive winds of a passing cyclone. Fewer than 100 Mariana fruit bats remain on Guam and it is unknown what effect the loss of fadang will have on these endangered bats.

In 2020, the US Fish and Wildlife Service estimated that only 45 fruit bats remain on Guam in a single roost site on Andersen Air Force Base.

***Dihammus marianarum* and *Anatrachyntis* sp.** Thomas Marler and J. Lawrence 2013 reported:

At least two arthropods depend on the *C. micronesica* population; the endemic stem borer *Dihammus marianarum* exploits cycad stem tissue for larval food, and the cone borer *Anatrachyntis* sp requires microstrobili (male cones) for larval food.

Dihammus marianarum (= *Acalolepta marianarum*) is an endemic longhorn beetle (Cerambycidae), which became a secondary pest of *C. micronesica* after plants were infested with CAS. This beetle has many larval host plants (T. Marler and R. Muniappan 2006) so *C. micronesica* is probably not critical for its survival.

On the other hand the cosmopterigid moth, *Anatrachyntis* sp, has been identified as a probable pollinator of *C. micronesica* and it has been suggested that this insect may be an obligate symbiont, meaning that it cannot survive without the cycad.

An abundance of larvae are found in every male cone following pollen shedding. Pupation takes place in a silken cocoon on the surface of the cone (T. Marler and R. Muniappan 2006). Terry et al. 2009 used sticky traps to sample insects and pollen in the vicinity of *Cycas micronesica* cones. They reported:

On female cone sticky traps, 30% of the pollen grains were associated with *Anatrachyntis* moths or moth scales and less than 5% with other insects; however, over 60% of the pollen was not associated with any insect, suggesting some pollen is wind dispersed.

Based on these observations, they hypothesized that *C. micronesica* is pollinated both by wind and insects, with *Anatrachyntis* sp being an important insect pollinator.

No lepidopteran had previously been identified as a cycad pollinator. However, nine years later Hua, Salzman, and Pierce 2018 reported that a moth in the same genus, *Anatrachyntis badi*, was discovered feeding in male cones of a native Florida cycad, *Zamia integrifolia*, and suggested that this species may also be an insect pollinator.

Details of the hypothesized symbiosis between the unidentified *Anatrachyntis* species and *C. micronesica* on Guam are not fully understood. The relationship between partners in this symbiosis differs markedly from than between flowering plants and their insect pollinators:

1. The reward for pollination service is provided to the caterpillar stage by the male plants which provide food in the form of sacrificial tissues within the male cone. The male cone may also provide a site free from competitors and natural enemies. The presumption here is that the caterpillars are protected by cycad toxins to which they have evolved tolerance.
2. There is no known reward provided to adults moths carrying pollen to female cones for pollination to take place. It is not known how *Anatrahynitis* moths are attracted to female cones of *C. micronesica*.

The biology of *Atrachyntis* on Guam is largely unknown apart from its association with *C. micronesica*. It is possible that this species is highly specialized and that its survival is totally dependent on availability of male cones. Alternatively, it is possible that this moth is not an obligate symbiont, but has broad host range.

4.5.2. Ecosystem services provided by *C. micronesica*

Nitrogen fixation and impacts on forest soils *C. micronesica* is the only dominant native tree on Guam which nitrogen-fixing symbionts. Therefore, the native biota of Guam's habitats has developed with this living resource provisioning the ecosystem with nitrogen. (Thomas Marler and Terry 2011). In the case of cycads, the nitrogen fixing symbionts are cyanobacteria growing in specialized structures called coralloid roots.

4.5.3. Prognosis for Guam's Forests

Currently, mature *C. micronesica* in Guam's forests are dying and these are not being replaced by seeds or juvenile plants. It is obvious that without a change, this endemic plant, the most abundant tree in Guam's forests only two decades ago, is headed towards local extinction.

Pending extirpation of Guam's native cycads is not the first or last ecological disaster caused by invasive species in Guam's forests (Moore 2018). The brown tree snake, *Boiga irregularis*, first detected in 1953 caused extinction or extirpation of all Guam's forest birds, also removing the ecosystem services they provided such as seed dispersal, insectivory, and pollination. The coconut rhinoceros beetle, *Oryctes rhinoceros*, first detected in 2007 is killing large numbers of coconut palms, *Cocus nucifera*, and the introduced palma brava, *Heterospatha elata*. These palm species were identified as Guam's second and third most abundant trees in the 2002 forestry survey. (Donnegon et al. 2004).

Restoration of Guam's forests to their pristine state is no longer possible. However, there is an urgent need to control CAS and other key invasive species so that some recovery can take place, without further loss of biodiversity.

5. Natural enemies of CAS - Cave

6. Classical biological control

6.1. Florida - Cave

6.2. Hawaii - Wright

6.3. Guam - Moore

6.3.1. *Rhyzobius lophanthae*

About 100 adults of *Rhyzobius lophanthae* were field collected on Maui and imported to Guam during November 2004. This coccinellid was originally introduced to California from Australia in 1892 and to Hawaii from California in 1894. It was observed feeding voraciously on CAS shortly after arrival of this new pest in Hawaii.

R. lophanthae was previously introduced to Guam on two separate occasions under various synonyms: *R. satelles* Blackburn, *Lindorus lophanthae* (Blaisdell), and *R. pulchellus* Montrouzier (Nafus and Schreiner 1989). In 1925 and 1926 *Rhyzobius satelles* was imported to Guam from California to control the coconut scale, *Aspidiotus destructor* Signoret. However, attempts at field establishment failed.

Nafus and Schreiner 1989 also reported:

In 1971, *Rhyzobius satelles* Blackburn (as *R. pulchellus* Montrouzier) was introduced to Guam from New Caledonia to aid in the control of coconut scales and citrus scales. A single specimen of *R. satelles* was recovered in 1978, indicating establishment. The beetle, however, is very uncommon; an intensive survey of coconut insects in 1984 yielded no specimens.

The beetles from Maui were reared on scale-infested *C. micronesica* cuttings placed in a large screened camping tent set up in a laboratory. Adult offspring were collected for field release by aspirating them from the walls of the tent into plastic vials. Field releases were initiated on February 16 2005 at the Guam National Wildlife Refuge at Ritidian Point. The beetles established readily. By July 7 2005 high densities on adults were observed on cycads anywhere within a 1 km radius of the release site. Establishment and dispersion of the beetles were monitored using yellow sticky traps deployed between June 2005 and May 2006. Unexpectedly, we were also able to monitor CAS crawlers and adult males using these traps (Fig. 2) (Moore 2017). Following establishment of *R. lophanthae* at Ritidian Point, laboratory-reared and field-collected beetles were released at about 30 other sites throughout Guam.

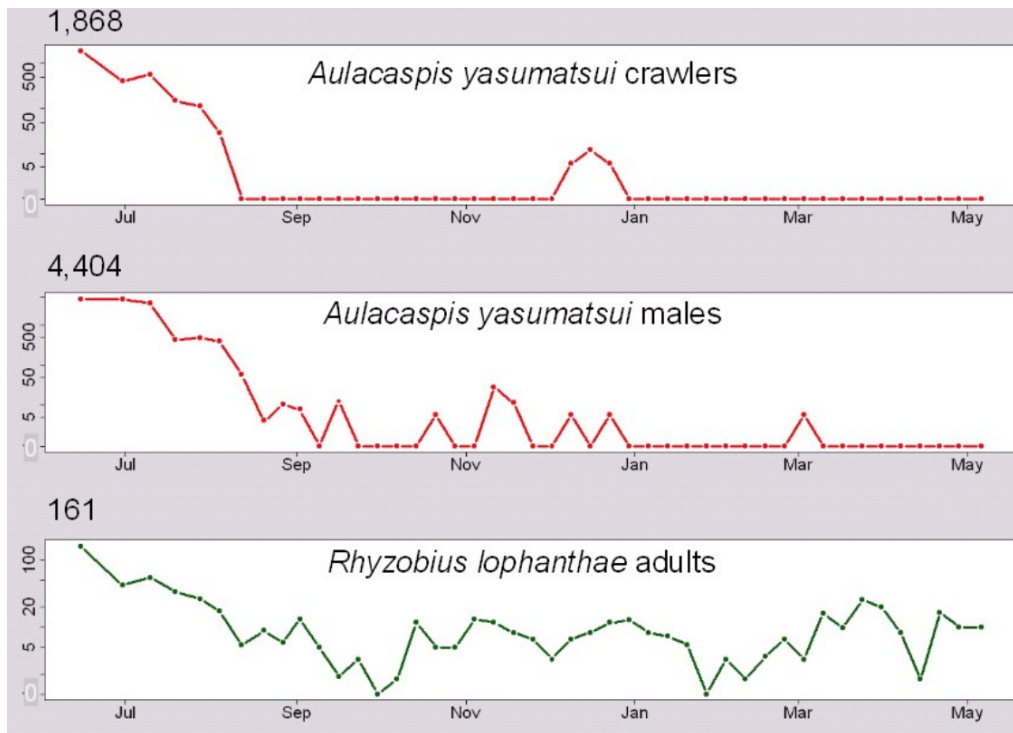


Figure 2: Insects trapped on yellow sticky cards at Ritidian Point, Guam following field release of *Rhyzobius lophanthae* in February, 2005. X axis runs from July 2005 through May 2006; Y-axis, in log scale, is number of insects trapped per square meter per day. [Figure from Moore, R. H. Miller, and Thomas E. Marler 2013]

By about 2010, *R. lophanthae* larvae or adults could be found on almost every CAS-infested cycad on Guam, preventing CAS from killing mature cycads. By 2010, about 90% of wild cycads had been killed on Guam (REF). Unfortunately, the *C. micronesica* population is not recovering because almost all seeds and seedlings are being killed by CAS and other causes (REF). Thomas Marler, R. Miller, and Moore 2013 showed that *R. lophanthae* predation of CAS is significantly reduced close to the ground and suggest that this may partially account for failure of the beetle to protect CAS seedlings. They also suggested:

The causes of reduced scale predation by *R. lophanthae* near the ground are unknown, but a parasitoid biological control agent may not exhibit these same limitations. Furthermore, because a parasitoid would be much smaller than *R. lophanthae*, it would likely be better able to access scale infestations within cracks and crevices on *C. micronesica* and *C. revoluta* trees.

6.3.2. *Coccobius fulvus*

6.3.3. *Aphytis lignanensis*

6.3.4. *Arrhenophagus*

Ask Mark, Janis about Bernarr's report on fortuitous introduction of CAS parasitoids.

Ask Reddy about his report.

Ask Arnold Harra.

6.4. Elsewhere - Cave

7. Prospects for future action - Cave, Wright, and Moore

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Notes from key references.

A. Haynes and Marler 2005

A.1. fadang plants provide crucial food for other organisms

A second reason for funding this project is that fadang plants provide crucial food for other organisms. The fleshy, aromatic covering of fadang seeds is a preferred food item for the endangered Mariana fruit bat, *Pteropus marianus marianus*. Fadang is so resistant to most types of disturbance that its seeds are sometimes the only bat food item available in the forest following the destructive winds of a passing cyclone. Fewer than 100 Mariana fruit bats remain on Guam and it is unknown what effect the loss of fadang will have on these endangered bats. (For more information on Guam's fruit bats, refer to the following website: <http://www.fws.gov/pacific/pacificislands/wesa/marianabatindex.html>.) We have only just begun our herbivory surveys of *Cycas micronesica*. In our preliminary work, we have identified the indigenous stem borer, *Dihammus marianarum* (Coleoptera: Cerambycidae), as a common cycad consumer. When these surveys are completed, there may be other native arthropod cycad consumers. Thus, the loss of this host species may also lead to the loss of several native species. Haynes and Thomas E. Marler 2005

A.2. symbiotic relationships

A third reason this project deserves emergency status is that cycad plants are part of a tripartite symbiotic system of organisms. As with many species of plants, cycad roots are invaded by mycorrhizal fungi. However, cycads also produce coralloid roots that are host to nitrogen-fixing cyanobacteria. Work in ex-situ sites reveals that any available genotype of cyanobacteria can invade coralloid roots. This is also probably true for mycorrhizae. Thus, determining the genetic variation of cycad mycorrhizae and cyanobacteria in the native habitat is needed to shed light on conservation of these important organisms. Loss of *Cycas micronesica* from Guam's various populations could easily result in the permanent loss of local genotypes of these other organisms that make up this complex, interconnected system. Haynes and Thomas E. Marler [2005](#)

A.3. pollinator(s)

A fourth reason for considering this a top priority for emergency funding is the nature of cycad pollination dynamics. We now know that cycads are pollinated almost exclusively by obligate insect pollinators (Norstog & Nicholls, 1997; Whitelock, 2002). We are attempting to identify the insect(s) which co-evolved with Guam's cycad population as the pollinators. At the present time, so many of the habitats are infested with CAS that it is difficult to find individual cycads suitable for pollination studies. The probability of one or more endemic, obligate insect pollinators occurring on Guam is highly likely. These beneficial insects will also be lost along with Guam's cycad population should we continue to stand by and let the CAS epidemic continue unchecked. Haynes and Thomas E. Marler [2005](#)

A.4. fadang is an iconic plant

Our height increment data indicate that many of Guam's coastal cycad plants are hundreds of years old. These plants have survived the Spanish-American War and two world wars; they have survived innumerable tropical cyclones; they have survived the invasion of intentionally introduced feral deer and pig populations and the accidental introduction of various insect species. Some of these individual plants "watched" Ferdinand Magellan and his fleet sail along Guam's coast on 6 March 1521. And they endured the Spanish-Chamorro Wars that decimated the indigenous human population. Truly, the remaining plants comprise a long-lived botanical and cultural treasure, one that is in danger of disappearing forever. We simply cannot elect to continue to watch this tragedy without attempting to intervene. The impending cascade of detrimental effects is looming too large to justify apathy. Haynes and Thomas E. Marler [2005](#)

B. 2005 - Report and recommendations on cycad aulacaspis scale

B.1. list of known CAS biocontrol agents

BIOLOGICAL CONTROL

In practice, the introduction of predators or parasitoids is the most cost- and labor-effective method of controlling scale insect infestations. It is also the standard approach for long-term control of introduced exotic scale pests (Meyerdirk, 2002). The existence of effective natural biocontrol agents for CAS can be assumed, since the scale is usually in low or moderate densities in wild populations of *Cycas* in Thailand, where CAS is native, where it does not cover foliage like it does in cultivated plants (Tang et al., 1997). At least three such predators or parasitoids have been identified and tested in the field as biocontrol agents for CAS. In 1996, Dr. Richard Baranowski of the University of Florida-Homestead (retired), working with Banpot Naponpeth, director of the Natural Biological Control Research Center at Kawetsart University in Bangkok, Thailand, identified two potential biocontrol organisms. This research was conducted in part on the grounds of Nong Nooch Tropical Garden. Both insects were evaluated and then widely released in Florida as biocontrol agents of CAS. *Coccobius fulvus* (Compere & Annecke) (Hymenoptera: Aphelinidae) is a parasitoid 3 wasp no larger than its host, ca. 1 mm long (see Fig. 1). Observations of this organism in south Florida suggest that this wasp, by itself, is not aggressive enough to control CAS on *Cycas* plants, such as *C. revoluta*, that are highly susceptible to CAS (Caldwell, 2005), but it can be effective in large, heavily infested plants of other *Cycas* species and/or plants with dense foliage in which pesticide application is inhibited (Wiese et al., in press). *Coccobius fulvus* has also been released as a biocontrol agent of other diaspid scale insects in the U.S. (Meyerdirk, 2002). *Cybocephalus binotatus* Grouvelle (Coleoptera: Nitidulidae) is a predatory beetle not much longer than CAS (see Fig. 2). The adult punctures the scale cover and chews on the living scale underneath; it will also deposit eggs under the scale cover, where its larvae then feed on CAS eggs. A study of the effects of this predator on a similar species of *Aulacaspis* on mangos suggests that, because it requires a substantial scale population to maintain effective numbers, it must be re-released periodically into infested areas to maintain effective control (Lagadec, 2004). Thus, an active, ongoing release program may be necessary for effective control using this biocontrol agent. Recently the beetle released in Florida has been re-identified as *Cybocephalus nipponicus* (Endrody-Younga) (R. Cave, pers. comm.). The ladybird beetle, *Rhyzobius lophanthae* (Blaisdell) (Coleoptera: Coccinellidae), a native of Australia that is often called the “scale destroyer,” has been successfully used as a control agent for CAS on cultivated *Cycas* plants in Hawaii by the University of Hawaii and the Hawaii Department of Agriculture (Hara et al., undated). It has also been reared and released on Guam since February 2005 to combat the CAS infestation in wild *Cycas* mi-

Mites The mite, *Hemiarcoptes* sp. (prob. *H. coccophagus*), has been found to control a related species of *Aulacaspis* in a lab setting (Meyerdirk, 2002). Fungi An unidentified fungus is known to grow on masses of the scale, *Aulacaspis tegalensis* (Zehntner) (Meyerdirk, 2002). Another unidentified fungus has been observed growing on CAS in Florida (Caldwell, 2005). Tang et al. 2005

B.2. CAS is fatal

CAS will kill an infected *Cycas* host plant within a year if no control measures are taken. Tang et al. 2005

B.3. Primary Recommendations to the CSG

To reiterate, the activities of major urgency that the CSG should pursue are the following:

1. A major priority must be to promote research on identifying new biocontrol agents for CAS and determining how to improve the effectiveness and accelerate the establishment of biocontrol organisms in newly infested areas.
2. Work together with the IUCN/SSC Invasive Species Specialist Group and the Global Invasive Species Programme to alert plant protection organizations of countries throughout the tropics and subtropics— especially those that possess wild species of *Cycas*—about the threat of CAS. Provide them with information and techniques for effective exclusion of CAS. This will require tapping into the IUCN and/or other high profile media outlets.
3. Assist with locating funding for current control efforts in Guam and Taiwan. Aid in collating and documenting such efforts, so as to identify the most effective techniques and avoid repeated duplication of ineffective control measures. Tang et al. 2005