

DRAFT 1: Coconut Rhinoceros Beetle FIDL

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Figure 1: Male coconut rhinoceros beetle head and pronotum.

1 Coconut rhinoceros beetle (CRB) (Fig. 1),
2 *Oryctes rhinoceros* (L.) (Coleoptera: Scarabaeidae)
3 is a major pest of coconut palm and oil palm. It is
4 native to southeast Asia, but it has invaded many
5 Pacific islands. American affiliated islands invaded
6 by CRB include American Samoa, the Republic of
7 Palau, Guam, Oahu (Hawaii), and Rota (Common-
8 wealth of the Northern Mariana Islands). With the
9 exception of American Samoa, these islands have re-
10 cently been invaded by a newly discovered variant of
11 CRB referred as the CRB-G biotype. This highly in-
12 vasive biotype is problematic because it is resistant
13 to *Oryctes rhinoceros* nudivirus OrNV, a classical
14 biological control agent which previously controlled
15 CRB on Pacific Islands. Throughout this leaflet,
16 CRB-G refers to the virus-resistant biotype and
17 CRB-S refers to virus-susceptible biotypes which can
18 be readily controlled by OrNV.

1 Biology

1.1 Taxonomy

The coconut rhinoceros beetle (CRB), *Oryctes rhinoceros* L., is a member of the scarab beetle family, Scarabaeidae, and the subfamily Dynastinae. Taxonomic expertise is required to differentiate *O. rhinoceros* from several other similar *Oryctes* species, some of which also attack coconut and other palms.

1.2 Life cycle and reproduction

CRB has four life stages: eggs, grubs, pupae and adults with the grubs having three substages called instars (Fig. 2). The life span depends on environmental conditions, varying between 9 months and 18 months. Generation time varies between 5 months and 9 months. The CRB sex ratio is usually close to 50:50 and females lay about 65 eggs during their lifetime. Under optimal environmental conditions with an unlimited food supply, CRB populations have the potential to grow at a rate of 3,250% per generation.

1.3 Larval feeding behavior

Grubs feed only on decaying vegetation and do no harm. They feed in breeding sites which can be composed of a wide variety decaying vegetation. Preferred sites are standing dead coconut stems and fallen coconut logs and fronds. But piles of anything with a high concentration of decaying vegetation can be used as a breeding site including green-waste, dead trees of any species, saw dust, and manure. CRB breeding sites have even been found in commercially bagged soil purchased from hardware stores. An active breeding site will contain all CRB life stages.

It is interesting to note that CRB individuals are at their heaviest during the prepupal stage (third

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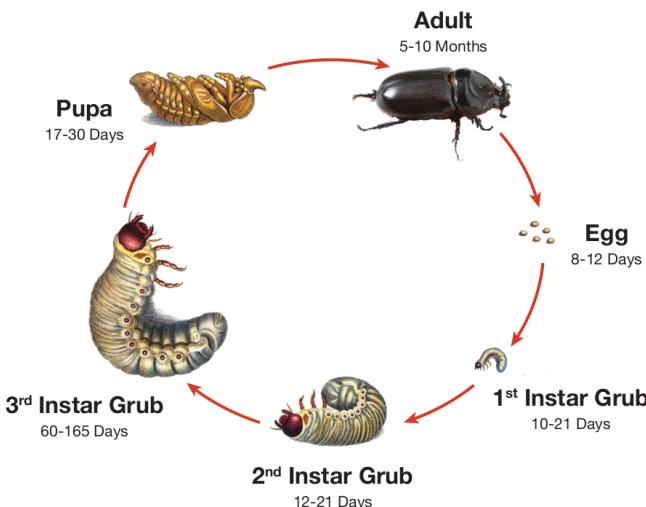


Figure 2: Coconut rhinoceros beetle life cycle.

hole is found (Fig. 5 and 6).

Adults feed for only a few days in the crown before exiting the bore hole to fly off to a breeding site for mating and oviposition. Adults feed every few weeks during their adult lifespan. Thus a single adult can damage several trees.

The impact of v-shaped cuts reduces production of nuts and degrades the aesthetic appeal of ornamental palms. However, the damage is not necessarily permanent. Even severely damaged palms can be nursed back to full health if all further attacks from adults are prevented.

CRB can cause palm mortality if a bore hole damages the single meristem (growing tip) within the base of the crownshaft. Mature palm mortality caused by CRB is rare unless the adult population and the associated attack rate is very high.



Figure 3: Coconut palms severely attacked by coconut rhinoceros beetle.

1.5 Population dynamics

With unlimited food and no control from natural enemies, CRB populations are capable of growing at a rate of about 3,250% per generation. Devastating, uncontrolled outbreaks of CRB are often triggered by a tropical cyclone, massive land clearing

54 instar grubs which have finished feeding). Thus the
55 source of almost all the biomass in a CRB population
56 comes from conversion of decaying vegetation during
57 the larval stage, not from live plant material.

1.4 Adult feeding behavior and damage symptoms

60 CRB causes damage only in the adult stage. Both
61 sexes feed on sap released from macerated soft tis-
62 sues within stems of several tropical monocot plants.
63 Primary host plants are coconut palm and oil palm,
64 but they also attack many other palm species. Oc-
65 casionally, they will attack other monocots such as
66 banana, sago, pandanus, taro, pineapple, sugarcane,
67 papaya and agave. However, CRB is not reported to
68 be a major economic pest of these secondary hosts.

69 In both coconut palm and oil palm, CRB adults fly
70 into the crown and force their way down between the
71 leaf axils until they find a position where they can
72 bore a horizontal hole into the crownshaft towards
73 the white tissue at the center of the stem. When bor-
74 ing this hole, they will pass through one or more de-
75 veloping fronds. When these damaged fronds emerge
76 from the crownshaft several weeks later, the damage
77 will become evident as v-shaped cuts which are a
78 distinctive sign of CRB damage (Fig. 3 and 4). Oc-
79 casionally, similar damage will appear in palms that
80 have been trimmed to prevent contact with power
81 lines or to remove nuts. However, in these cases,
82 there will be no bore holes associated with the dam-
83 age. If CRB caused a v-shaped cut, this can always
84 be confirmed by removing outer petioles until a bore



Figure 4: V-shaped cuts caused by adult CRB.



Figure 6: CRB bore hole in stem made visible by removing petioles.



Figure 5: CRB bore holes through petioles.

1.6 Geographic distribution

Islands in the Pacific and Indian Oceans were invaded by CRB during two waves of movement (Fig. 7). The first wave occurred started in 1909 when CRB was accidentally transported to from Sri Lanka to Samoa with shipment of rubber tree seedlings and it ended during the 1970s. All of the CRB range expansion during this period was south of the equator except for the invasion of the Ryukyu Islands (Japan) starting in 1921 and invasion of the Palau Islands prior to 1943.

The second wave of CRB invasions started in 2007 with discovery of CRB on Guam, followed by invasion of Oahu (Hawaii), Port Morseby (Papua New Guinea), Guadalcanal, Savo and Malaita (Solomon Islands), and Rota (Commonwealth of the Northern Mariana Islands). Beetles in the second wave of invasions are genetically different from those in the first wave and these are being referred to as the Guam biotype or CRB-G for short.

2 Control tactics

2.1 Eradication

In theory, eradication of CRB from a newly invaded area can be attained by blocking invasion pathways coupled with finding and destroying all breeding sites. In practice, eradication has proven to be very difficult after initial establishment of a CRB population, despite early detection and rapid response.

Only one of many eradication attempts has succeeded. This was accomplished on the tiny (36 km^2) Niuatopapu Island (also known as Keppel Island), which lies between Samoa and Tonga. During a period spanning 1922 to 1930 all CRB breeding sites

108 or military activity which generates large amounts
109 of dead and decaying vegetation over a large area.
110 Within a few months, this plentiful food supply gener-
111 ates massive numbers of adults which bore into
112 palm crowns. The high rate of attack results in high
113 palm mortality. These dead palms quickly begin to
114 rot and become ideal CRB breeding sites, generat-
115 ing even higher numbers of adults which bore into
116 and kill even more palms. In the absence of natural
117 enemies or introduced biological control agents, this
118 self-sustaining feedback cycle may result in the loss
119 of most palm trees on an island.

120 An example of this positive feedback cycle oc-
121 curred in Palau as a result of massive destruction of
122 palms and other vegetation during the Second World
123 War. Prior to the war, CRB was very rare in Palau
124 but shortly afterwards about 50% of coconut palms
125 were killed by CRB throughout the archipelago, and
126 some islands lost all of their palms.

127 Uncontrolled outbreaks of CRB-G are currently
128 happening on Guam and some of the Solomon Is-
129 lands.



Figure 7: Screenshot of an online interactive web map [1] showing the geographic distribution of the coconut rhinoceros beetle. Green markers: native range; Orange markers: first detected during the 20th century; Red markers first detected during the 21st century; Open circle: population includes CRB-G biotype; Filled circle: population is exclusively CRB-G biotype.

cut into pieces and burnt or buried to remove potential breeding sites. In some situations, larvae may develop in the crown of live palms. This only occurs where there are large accumulations of organic matter in the frond bases. The organic matter should be removed where possible.

Disposal of dead felled palms Mature palm trees will fall after being weakened by fungal diseases (*Ganoderma*), after strong winds during tropical cyclones or after the felling of senile palms prior to replanting. Dead palms on the ground should be cut up into manageable lengths or chipped prior to disposal by burning or deep burial.

Covering of palm stumps Felled palms leave a stump which is suitable for development of larvae as it rots. In management of palm plantations in Asia, where a zero-burning policy is in operation, ground cover is planted shortly after felling to cover the debris and make it less attractive to the flying beetles. The legumes *Mucuna* spp. and *Pueraria javanica* are ground cover plants that are commonly used, as they will add nitrogen to the soil and cover the decaying trunks.

Management of organic matter and compost Heaps of organic matter, particularly palm debris, provide excellent food material for development of CRB larvae. Any deep piles of organic material will be attractive to the egg laying females. Heaps of fronds or empty fruit bunches are particularly susceptible. Sawdust from sawmills that process palm timber is also a favorable resource for beetle development. General compost, farmyard manure and even organic garbage can provide sites for development of the larvae. The first step in reducing the threat of beetles emerging from composts is management of the organic matter. Palm debris should be spread among the palms to break down rapidly and release nutrients rather than being piled in heaps. Compost or farmyard manure should be turned regularly, and larvae removed, or pigs and chickens can assist by eating exposed larvae. In urban environments, organic material is often gathered during environmental clean-up and composted, but this may provide a centre for re-infestation of the locality. Compost can be sterilized or fumigated to kill larvae; however, this process is energy-demanding and expensive. Sterile compost will also be susceptible to re-invasion.

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163 were located and destroyed.

2.2 Sanitation

165 Sanitation includes detection and destruction of active and potential CRB breeding sites.

Breeding site detection Local searches for breeding sites are usually initiated in response to visible damage to palms or capture of adults in pheromone traps. In Guam and Hawaii, dogs trained to sniff out CRB grubs have been deployed to assist human searchers. Recent research suggests that CRB adults fitted with miniature radio transmitters or harmonic radar tags may be a cost effective way of detecting cryptic breeding sites. The essential idea is that the radio transmitters and tags will accumulate at breeding sites where adults aggregate. These tags can then be detected by ground and/or aerial surveys using radio receivers or harmonic radar transceivers.

Removal of standing dead palms CRB adults are attracted to standing dead palm trees that have begun to rot from the crown. Females will lay their eggs in the rotting palm trunk and the developing larvae will feed on the decaying fibers near the top of the trunk, which starts to decompose in the center forming a protective tube for larval development. As the larvae increase in size and strength of their mandibles, they can penetrate further down the trunk leaving a column of frass and cut fibers for the early instars. Dead standing palms should be felled,

238 Where feasible, compost heaps can be covered with
239 netting to trap emerging beetles.

240 Burning CRB breeding material is the most de-
241 pendable method for removing the food source for
242 CRB grubs. In Hawaii CRB sanitation programs,
243 breeding site material is being burned on-site using
244 air-curtain burners and some is being trucked to a
245 waste-to-energy electrical power generation plant.

246 2.3 Trapping

247 CRB trapping can be used for different purposes in-
248 cluding surveillance for early detection, monitoring
249 growth and spread of a population over time, and
250 for population suppression by mass trapping. In all
251 cases, the trap needs to be attractive enough to draw
252 in beetles from a distance strong and enough to con-
253 tain them once they are captured. Olfactory and
254 visual attractants can be used to increase trap catch.

255 **Artificial breeding sites** One of the first traps to
256 be developed was the Hoyt trap made from a metal
257 can set on top of a coconut trunk or wooden post
258 (Fig. 8). The can was capped with a length of co-
259 conut stem with a hole in the center large enough
260 for a beetle to enter. The trap system was used
261 extensively and functioned because it mimicked a
262 standing, decaying coconut stem which is attractive
263 to CRB adults.

264 An artificial breeding site trap can easily be con-
265 structed simply by laying coconut log sections on
266 the ground (Fig. 9). Trap catch can be enhanced by
267 covering the logs with netting (see section on netting
268 below) and providing a pheromone source.

269 **Pheromone traps** Design and utility of traps
270 changed with the discovery of ethyl chrysanthemate
271 as an attractant. This was rapidly superseded
272 by ethyl-4-methyloctanoate (E4-MO) commonly re-
273 ferred to as oryzcalure, the male-produced aggrega-
274 tion pheromone of CRB, which could be synthesized.
275 E4-MO attracts both sexes, has been used for more
276 than 30 years and is produced commercially by sev-
277 eral companies.

278 Pheromone traps for surveillance need to be ro-
279 bust, inexpensive, attractive to beetles, difficult to
280 exit and simple to service. Simple bucket traps are
281 used extensively for monitoring throughout the Pa-
282 cific Islands and Southeast Asia (Fig. 10). Vaned
283 bucket traps (Fig. 11) and panel traps (Fig. 12)

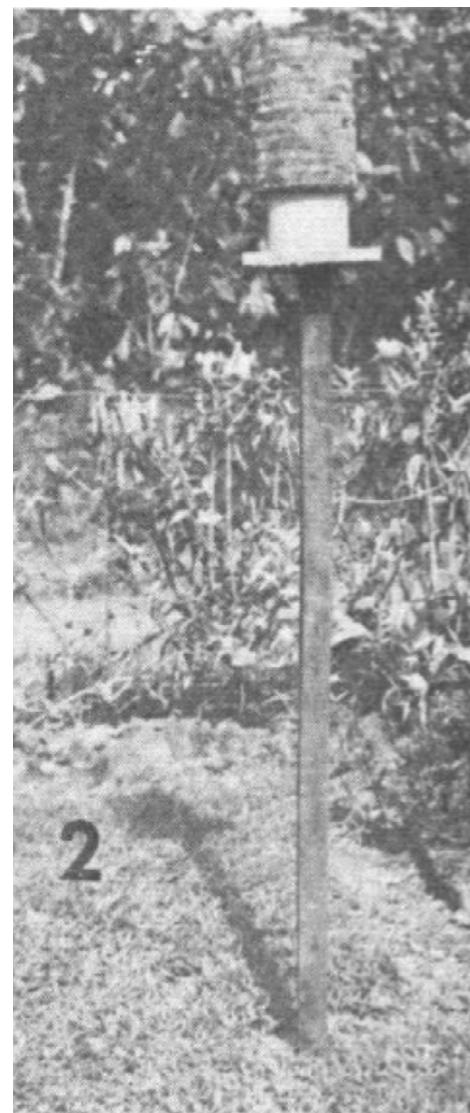


Figure 8: caption

284 have been used in surveillance trapping in Guam and
285 Hawaii where thousands of traps have been deployed
286 to monitor of the spread of CRB populations and
287 success of control activities.

288 **Efficacy of pheromone traps for population sup-
289 pression** Trapping removes adults from the popu-
290 lation and may contribute to pest and damage reduc-
291 tion. Bucket traps baited with pheromone have been
292 reported to reduce CRB populations in Malaysia and
293 the related *O. monoceros* in West Africa.

294 However, efficacy of pheromone traps for popula-
295 tions of CRB-G is in question. Mass trapping was
296 performed on Guam shortly after detection of CRB
297 in the Tumon Bay hotel are in 2007. There was no in-
298 dication of population suppression and trapping did

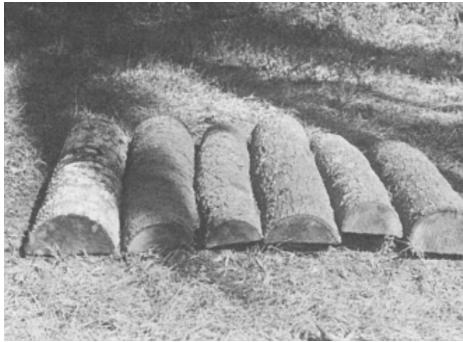


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Figure 11: caption

beetles cannot navigate to pheromone sources. 317

In mark-release-recapture experiments on Guam 318
only 64 of 567 (11%) of marked beetles were re- 319
captured in a grid of traps baited with oryzalure, 320
indicating that oryzalure is not highly attractive to 321
CRB-G biotype. Unfortunately, there are no com- 322
parative data for the CRB-S biotype. 323

Netting 324 Tekken, a gill net used by Chamorro fish- 325
ermen on Guam, has proven to be an effective trap- 326
ping tool for coconut rhinoceros beetles. Beetles are 327
captured when a strand of the netting falls into the 328
gap behind a beetle's pronotum, in the same way 329
that fish are caught when a strand falls into a gill 330
slit. In Guam, heaps of organic waste covered with 331
tekken catch beetles emerging from the pile and also 332
attracted to the heaps for mating and oviposition. 333
Cheap and simple pheromone traps can also be made 334
by attaching pieces of tekken to fences and placing 335
an oryzalure dispenser at the center of each piece.

2.4 Chemical control

Federal and state pesticide regulations should be 337
checked before planning any chemical control activi- 338
ties. 339

Chemical control of CRB is difficult because all life 340
stages live in protected habitats: grubs and adults 341

299 not reduce damage to palms with the mass trapping 300 areas. During 2010, the trap catch rate in Tumon 301 Bay was only 0.006 beetles per trap day, but CRB 302 damage was visible in 100% of coconut palms. In 303 contrast, a similar mass trapping program in Samoa 304 trapped 0.150 per trap-day, 25 times the Guam trap- 305 catch rate, but the proportion of damaged coconut 306 palms was only 30%. Note that the Guam popula- 307 tion is the CRB-G biotype and the Samoan popula- 308 tion is the CRB-S biotype.

309 Three possible explanations have been suggested 310 to account for these observations:

- 311 1. Traps baited with oryzalure are more attractive 312 to CRB-S than CRB-G.
- 313 2. CRB-G individuals do far more damage than 314 CRB-S individuals.
- 315 3. At very high population levels and trap densi- 316 ties there is so much pheromone in the air that



Figure 12: caption



Figure 13: Tekken beetle.

342 may found inside dead logs or buried in or under
 343 heaps of decaying vegetation and adults may be
 344 found boring into palm crowns only briefly, for a few
 345 days during feeding bouts.

346 **Foliar application** Foliar insecticide application is
 347 aimed at preventing damage or mortality of palms
 348 caused by adults. The pyrethroid, cypermethrin has
 349 been used to successfully protect coconut palms and
 350 oil palms. A high enough volume should be applied
 351 so that the pesticide runs down the midrib and pools
 352 at the base of the petiole where it meets the crown-
 353 shaft. This is the location at which CRB adults ini-
 354 tiate bore holes. Foliar sprays may be applied by
 355 hydraulic sprayers or by pesticide applicator drones.



Figure 14: Tekken pile.



Figure 15: Defence trap

Trunk injection Trunk injections of systemic insec-
 356 ticides have been applied to oil palms and coconut
 357 palms with variable success.
 358

Treatment of breeding sites Cypermethrin ap-
 359 plied as a drench controls all life stages heaps of loose
 360 breeding sites material, but may not kill adults and
 361 grubs boring within logs.
 362

2.5 Biological control

Biological control is the use of natural enemies
 364 (predators, pathogens, parasites) to suppress pest
 365 populations. In its native range, CRB is attacked by
 366 a community of co-evolved natural enemies, includ-
 367 ing pathogenic viruses and fungi, predatory carabid
 368 and elaterid beetles and parasitic *Scolia* wasps. The
 369 relative impact of each natural enemy species within
 370 this native community is poorly known, with addi-
 371 tional control strategies often needed to complement
 372 biological control in coconut and oil palm planta-
 373 tions.

When CRB-S invaded the Pacific, it was the focus
 375 of a substantial biological control program. The aim
 376 was to find one or more natural enemies in the na-
 377 tive range that could be introduced to the invaded
 378 range to suppress CRB-S populations. This process
 379 is known as classical biological control. Among many
 380 natural enemies introduced to the Pacific, very few
 381 predators or parasites established. Incidental preda-
 382 tion by pigs and chickens on CRB larvae may as-
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sist with control of this pest and can be useful for control of larvae in household or community waste piles. Local species of generalist arthropod predators (centipedes, beetles, ants) may feed on CRB larvae; however, there is little evidence that this contributes significantly to CRB control.

Only one pathogen provided significant control of CRB-S: the *Oryctes rhinoceros* nudivirus (OrNV) discovered in Malaysia by Alois M. Huger in 1963. This virus infects CRB-S larvae and adults, causing death after 6–30 days. Infected adults are weakened prior to death so that they stop feeding, their mobility is reduced and females stop laying fertile eggs. Once established in countries invaded by CRB-S, the virus significantly reduced CRB populations and the damage they caused.

Another approach to biological control for CRB was to create a biopesticide from a known pathogen. CRB adults and larvae can be infected by the fungus *Metarhizium majus* (formerly *M. anisopliae* var. *majus*). This fungus has been developed into a biopesticide that can be applied to CRB breeding sites in both the native and invaded range.

The recent invasion of CRB-G has changed CRB management wherever it is found. CRB-G is not susceptible to strains of OrNV introduced originally to control CRB-S in the Pacific. A new biological control effort is underway in order to identify strains of OrNV from CRB's native range that are effective against CRB-G. Until an effective OrNV strain is discovered, biopesticides containing *M. majus* are the only option for biological control of CRB-G.

In some jurisdictions, biological control agents are regulated as pesticides. For example, the U.S. Environmental Protection Agency regulates OrNV and *M. majus* as pesticides. Federal and state pesticide regulations should be checked before using biological control agents for CRB.

2.6 Damage surveys

A standardized method has been developed for quantifying CRB damage (for details see Jackson et al 2020). This method uses a five-level scale illustrated in the following figures (Figs 16, 17, 18, 19, 20).

This method can be applied by direct visual observation or by scoring images or video frames. Research is underway to develop automated detection and mapping of CRB damage by applying computer vision techniques to images recorded from the

ground-based and aerial drone surveys.



Figure 16: Index 0; Zero damage; No CRB damage visible



Figure 17: Index 1; Light damage; Notching or tip damage; less than less than 20% foliar loss



Figure 18: Index 2; Medium damage; Multiple fronds damaged; Notching and breakage; 20% to 50% foliar loss



Figure 19: Index 3; High damage; greater than 50% foliar loss



Figure 20: Index 4: Dead or moribund; Meristem destroyed

bination of the tactics described above. 449

For coconut palms planted for subsistence, ornamental purposes, or in commercial plantations, IPM 450
for CRB involves: 451
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1. monitoring of palm damage to detect localized 453
CRB outbreaks and to check that control is suc- 454
cessful 455
2. biological control with OrNV (in the invaded 456
and native range) and other co-evolved natural 457
enemies (in the native range only) 458
3. sanitation to remove organic waste, dead palms 459
and other potential breeding sites 460

Sanitation is an essential component of IPM for 461
CRB that complements biological control. Local- 462
ized CRB outbreaks will occur when breeding sites 463
are left uncontrolled, especially after cyclones and 464
tropical storms, when palms are often toppled by 465
high winds and large amounts of green waste is cre- 466
ated. Historically, outbreaks of CRB often follow 467
cyclone damage or large scale land clearing. Larvae 468
will develop in the decaying fronds, the trunk and 469
even the roots of the felled palms and also in many 470
other forms of decaying vegetation. 471

Some additional options may be incorporated into 472
IPM programmes for coconut, particularly for com- 473
mercial plantations or ornamental palms. These 474
more costly options include pheromone traps to 475
monitor adult beetle activity and complement it with 476
visual surveys of palm damage. Occasionally, trap 477
catches may be high enough to contribute to pop- 478
ulation suppression in coconut plantations, but this 479
strategy is more relevant to oil palm (discussed be- 480
low). If a recent invasion of CRB is targeted for 481
eradication, insecticides may be necessary for suc- 482
cess. 483

For higher value crops, particularly oil palm, the 484
same components are needed as for coconut: moni- 485
toring, biological control; and sanitation. More 486
costly IPM components are recommended for oil 487
palm because the crop's financial value makes 488
greater investment in control worthwhile. Thus, 489
IPM for CRB in oil palm involves: 490

1. monitoring of beetle activity with pheromone 491
traps and palm damage particularly for young 492
palms 493
2. biological control with OrNV (invaded and na- 494
tive range) and other natural enemies (native 495

434 2.7 Integrated pest management

435 Integrated pest management (IPM), is a broad-
436 based approach that integrates practices for eco-
437 nomic control of pests. The Food and Agricul-
438 ture Organization of the United Nations IPM as
439 "The careful consideration of all available pest con-
440 trol techniques and subsequent integration of appro-
441 priate measures that discourage the development of
442 pest populations. It combines biological, chemical,
443 physical and crop specific (cultural) management
444 strategies and practices to grow healthy crops and
445 minimize the use of pesticides, reducing or minimiz-
446 ing risks posed by pesticides to human health and
447 the environment for sustainable pest management."
448 IPM for management of CRB and includes com-

range) plus application of *M. majus* as a biopesticide to breeding sites that cannot be removed

3. sanitation to remove organic waste, particularly during plantation renewal when large amounts of waste is generated

4. insecticide treatments for young palms that are most sensitive to CRB damage.

Note that pollinators of oil palm are vulnerable to insecticides, so applications should be scheduled carefully to avoid flowering. When oil palm plantations are renewed, complete clean-up of the organic waste is challenging. In CRB's native range, an additional strategy is to break up and spread the waste as a thin layer, then plant a fast-growing cover crop, often a legume, over the waste matter.

References

- Pallippambil, Godshen R. 2015.** New Pest Response Guidelines: *Oryctes rhinoceros* (L.) Coleoptera: Scarabaeidae, Coconut rhinoceros beetle. U.S. Department of Agriculture, Animal Plant Health Inspection Service, Plant Protection and Quarantine. Available online at <https://tinyurl.com/mpdmvfpt> 536
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- Bedford, Geoffrey O. 1980.** Biology, ecology, and control of palm rhinoceros beetles. Annual Review of Entomology 25 (1980): 309–39. Available online at <https://tinyurl.com/yh79wmwc> 512
513
514
515
- Gressitt, J. Linsley 1953.** The coconut rhinoceros beetle (*Oryctes rhinoceros*) with particular reference to the Palau Islands. Bernice P. Bishop Museum. Bulletin 212. Available online at <https://tinyurl.com/npsab5d4> 516
517
518
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- Jackson, Trevor, Sean Marshall, Sarah Mansfield and Fereti Atumurirava 2020.** Coconut rhinoceros beetle (*Oryctes rhinoceros*): A manual for control and management of the pest in Pacific Island countries and territories. Pacific Community (SPC). Available online at <https://tinyurl.com/yxk4u27j> 521
522
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- Marshall, Sean D. G., Aubrey Moore, Maclean Vaqalo, Alasdair Noble, and Trevor A. Jackson 2017.** A new haplotype of the coconut rhinoceros beetle, *Oryctes rhinoceros*, has escaped biological control by *Oryctes rhinoceros* nudivirus and is invading Pacific Islands. Journal of Invertebrate Pathology, 149, 127-134. Available online at <https://tinyurl.com/mtpp29da> 528
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