

DSP REPORT

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QRS Complex Detection in ECG Signals: A Pan-Tompkins Algorithm Implementation

Introduction

Electrocardiogram (ECG) signal analysis is a cornerstone of cardiac diagnostics, providing essential information about heart function and potential abnormalities. One of the most critical features in ECG interpretation is the accurate detection of QRS complexes, which represent ventricular depolarization in the cardiac cycle. This project implements an enhanced version of the Pan-Tompkins algorithm—a widely respected method for QRS detection—adapted specifically for the MIT-BIH Arrhythmia Database, one of the most comprehensive ECG databases available for cardiac research.

The implemented system processes ECG signals through a series of digital signal processing (DSP) stages to identify QRS complexes accurately, even in the presence of noise and signal variations common in clinical settings. The project evaluates the algorithm's performance against expert annotations, providing metrics such as sensitivity, positive predictivity, and F1 score to quantify detection accuracy.

The Pan-Tompkins Algorithm

Overview

Developed in 1985 by Jiapu Pan and Willis J. Tompkins, the Pan-Tompkins algorithm represents a systematic approach to QRS detection that remains relevant despite its age. The algorithm processes an ECG signal through multiple stages, each designed to enhance specific characteristics of the QRS complex while suppressing noise and other ECG components like P and T waves.

The implementation in this project follows an enhanced version of the original algorithm with optimizations for the MIT-BIH Arrhythmia Database's specific characteristics. The core processing stages include:

- 1. Bandpass Filtering To isolate the QRS frequency components
- 2. **Derivative Computation** To emphasize the steep slopes of the QRS complex
- 3. Squaring To make all signal values positive and emphasize higher frequencies
- 4. Moving Window Integration To extract waveform features in addition to slope information
- 5. Adaptive Thresholding To distinguish QRS complexes from noise and adapt to signal variations

Mathematical Foundation and DSP Methods

1. Bandpass Filtering

The bandpass filter removes baseline wander (low-frequency noise) and high-frequency interference while preserving the QRS complex frequency content. In this implementation, a 4th-order Butterworth filter with cutoff frequencies at 5Hz and 15Hz is used:

```
lowcut = 5;  % 5Hz cutoff for high-pass
highcut = 15;  % 15Hz cutoff for low-pass
wn = [lowcut highcut] / (fs/2);
[b, a] = butter(4, wn, 'bandpass');
bandpass_signal = filtfilt(b, a, ecg_signal);  % Zero-phase filtering
```

The filtfilt function applies zero-phase filtering, eliminating phase distortion by filtering the signal forward and backward. This is critical for preserving the precise timing of QRS complexes.

2. Derivative Computation

The derivative stage emphasizes the steep slopes characteristic of QRS complexes. A five-point derivative approximation is used:

```
b = [1 2 0 -2 -1] * (1/8);
a = 1;
derivative_signal = filter(b, a, bandpass_signal);
```

This filter approximates the derivative while providing noise smoothing. Mathematically, it implements a weighted derivative approximation over a five-sample window.

3. Squaring Function

Squaring makes all values positive and emphasizes higher frequencies proportionally more than lower frequencies:

```
squared_signal = derivative_signal.^2;
```

This nonlinear transformation enhances larger values (likely QRS complex components) relative to smaller values (likely noise or T waves).

4. Moving Window Integration

Integration extracts morphological features of the QRS complex by computing a moving average:

```
b = ones(1, integration_window)/integration_window;
a = 1;
integrated_signal = filter(b, a, squared_signal);
```

The integration window width (150ms in this implementation) is critical: too wide and it might merge adjacent QRS complexes; too narrow and it might produce multiple peaks for a single QRS complex.

5. Adaptive Thresholding and Peak Detection

This stage is the most sophisticated part of the algorithm. Two thresholds are dynamically adjusted based on the signal and noise levels:

```
threshold1 = NPKI + signal_threshold_weight * (SPKI - NPKI);
threshold2 = noise_threshold_weight * threshold1;
```

Where:

• \$SPKI\$ is the running estimate of signal peak level

- \$NPKI\$ is the running estimate of noise peak level
- \$signal_threshold_weight\$ (0.25) and \$noise_threshold_weight\$ (0.5) are weighting factors

The algorithm updates these estimates with each detected peak:

```
% For QRS peaks:
SPKI = learning_rate_signal * peak_val + (1-learning_rate_signal) * SPKI;

% For noise peaks:
NPKI = learning_rate_noise * peak_val + (1-learning_rate_noise) * NPKI;
```

This adaptive approach allows the algorithm to adjust to variations in ECG amplitude and morphology over time.

6. RR Interval Analysis for Missed Beat Detection

The implementation incorporates an enhanced feature to detect missed beats based on RR interval analysis:

```
if RR(i) > 1.8 * mean_RR
% Search for potential missed beats in noise peaks...
end
```

This additional step improves detection reliability by identifying possible QRS complexes that were initially classified as noise but are temporally consistent with expected beat timing.

Performance Evaluation

The algorithm's performance is evaluated against expert annotations from the MIT-BIH Arrhythmia Database using three key metrics:

- Sensitivity (\$Se\$): The proportion of actual QRS complexes correctly identified \$\$Se = \frac{TP}{TP + FN}\$\$
- 2. **Positive Predictivity (\$PP\$)**: The proportion of detected QRS complexes that are correct \$\$PP = \frac{TP}{TP + FP}\$\$
- 3. **F1 Score**: The harmonic mean of sensitivity and positive predictivity \$\$F1 = 2 \times \frac{Se \times PP}{Se + PP}\$\$

Where:

- \$TP\$ (True Positives): Correctly identified QRS complexes
- \$FP\$ (False Positives): Incorrectly identified QRS complexes
- \$FN\$ (False Negatives): Missed QRS complexes

The evaluation uses a matching window of 100ms, meaning a detected peak is considered a true positive if it falls within \$\pm50\$ms of an annotated QRS complex.

Implementation Details

The project is implemented in MATLAB and organized into several key functions:

- 1. robust_pan_tompkins_qrs_detector_local: Implements the enhanced Pan-Tompkins algorithm
- 2. load_mitbih_record_local: Loads ECG data from the MIT-BIH database
- 3. evaluate grs_detection_local: Evaluates detection performance against annotations

The main script processes selected records from the MIT-BIH Arrhythmia Database, visualizes the detection results, and summarizes overall performance metrics.

Visualization and Interpretation

The implementation includes comprehensive visualization of each processing stage, allowing for detailed inspection of the algorithm's behavior. The visualizations include:

- 1. The original ECG signal with marked QRS detections
- 2. The bandpass filtered signal
- 3. The derivative signal
- 4. The squared signal
- 5. The integrated signal with adaptive thresholds and marked peaks

Additionally, the implementation calculates and displays the average heart rate based on detected QRS complexes, providing a clinically relevant parameter.

Conclusion

The enhanced Pan-Tompkins algorithm implementation demonstrates the effectiveness of systematic digital signal processing techniques in extracting clinically significant features from physiological signals. The project highlights the importance of adaptive methods when processing biological signals, which naturally exhibit high variability both between subjects and within the same subject over time.

The performance metrics obtained from testing on the MIT-BIH Arrhythmia Database provide quantitative validation of the algorithm's effectiveness. Future work could focus on further optimization for specific arrhythmia types, enhanced noise resistance, or integration with more comprehensive ECG analysis frameworks.

This project serves not only as a practical implementation of a classic biomedical signal processing algorithm but also as an educational resource demonstrating fundamental DSP concepts applied to real-world biomedical engineering challenges.

```
%% MIT-BIH Arrhythmia Database QRS Complex Detection (All-in-One Script)
% This script contains all necessary functions and analysis code to detect
% QRS complexes in the MIT-BIH Arrhythmia Database using Pan-Tompkins algorithm.
clear;
close all;
clc;
% Configuration
record_numbers = [100, 101, 103, 105, 106, 109]; % Records to analyze
lead number = 1;
                         % ECG lead to analyze (1 or 2)
start_time = 60;
                        % Start time in seconds (skip first minute)
                        % Duration in seconds (analyze one minute)
duration = 60;
plot_individual = true;  % Plot individual record results
fs = 360;
                         % MIT-BIH sampling frequency
%% Check for WFDB Toolbox if ~exist('rdann', 'file') ||
~exist('rdsamp', 'file')
     error(['WFDB Toolbox not found. Please install it from: ' ...
            'https://physionet.org/content/wfdb-matlab/0.10.0/']);
end %% Validate that database files
exist try
    [test signal, ~, ~] = rdsamp('mitdb/100', 1, 10, 0);
catch
    error(['Could not access MIT-BIH database files. Please ensure they are
downloaded and ' ...
           'properly placed in a "mitdb" folder in your MATLAB path or current
directory.']);
end
fprintf('MIT-BIH database is accessible. Proceeding with analysis...\n');
MIT-BIH database is accessible. Proceeding with analysis...
```

```
results = struct('record', {}, 'qrs_peaks', {}, 'annotations', {},
'performance', {});
%% Process each record
for r = 1:length(record_numbers)

    record_num = record_numbers(r);
    fprintf('\nProcessing record %d (%d of %d)...\n', record_num, r,
length(record_numbers));

    % Load the record data
    try
        [ecg_signal, record_fs, annotations] =
load_mitbih_record_local(record_num, lead_number, start_time, duration);

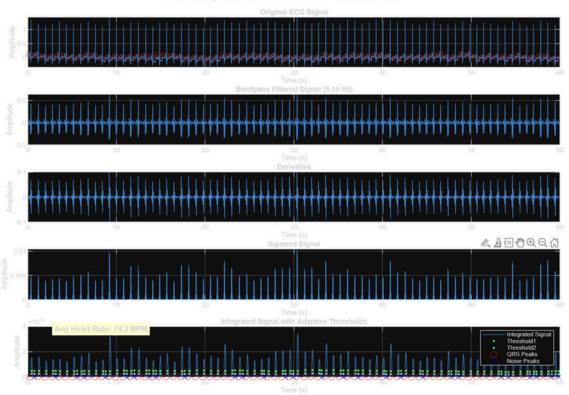
    % Ensure we have ECG data
```

```
if isempty(ecg_signal)
            warning('Record %d has no ECG data. Skipping...', record_num);
            continue:
        end
        fprintf('Loaded record %d: %d samples at %d Hz\n', record_num,
length(ecg signal), record fs);
        % Run QRS detection
        [qrs_peaks, processed_ecg] =
robust_pan_tompkins_qrs_detector_local(ecg_signal, record_fs, plot_individual);
        % Get QRS annotations (only those marked as normal beats 'N' or PVC 'V')
        ann qrs = [];
        if ~isempty(annotations.time)
            % Extract beat annotations (N = normal, V = PVC, etc.)
            beat_annotations = ismember(annotations.type, {'N', 'V', 'L', 'R',
'A', 'a', 'J', 'S', 'F'});
            ann qrs = annotations.time(beat annotations);
        end
        % Evaluate detection performance
        if ~isempty(ann qrs)
            [qrs stats, performance] = evaluate qrs detection local(qrs peaks,
ann_qrs, record_fs);
            % Store results results(r).record = record_num;
            results(r).qrs peaks =
                                                 grs peaks;
            results(r).annotations
                                                    ann grs;
            results(r).performance = performance; % Print
            performance
                           metrics
                                       fprintf('Record
            Performance:\n', record_num);
            fprintf(' Sensitivity: %.2f%%\n', performance.sensitivity * 100);
            fprintf(' Positive Predictivity: %.2f%%\n',
performance.positive_predictivity * 100);
            fprintf(' F1 Score: %.2f\n', performance.F1 score);
        else
        end warning('No annotations found for record %d', record num);
        % Update figure title if individual plots are shown
        if plot_individual
            sgtitle(sprintf('Pan-Tompkins QRS Detection - Record %d',
record num), 'FontSize', 16);
        end
    catch ME
        warning('Error processing record %d: %s', record_num, ME.message);
        % Display the error stack for debugging
        disp(getReport(ME, 'extended'));
    end
end
```

Processing record 100 (1 of 6)...

Warning: Could not load annotations. Returning empty annotation structure.

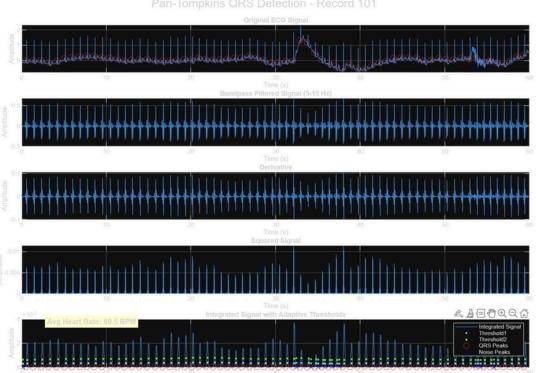
Loaded record 100: 21601 samples at 360 Hz Warning: No annotations found for record 100



Processing record 101 (2 of 6)...

Warning: Could not load annotations. Returning empty annotation structure.

Loaded record 101: 21601 samples at 360 Hz Warning: No annotations found for record 101

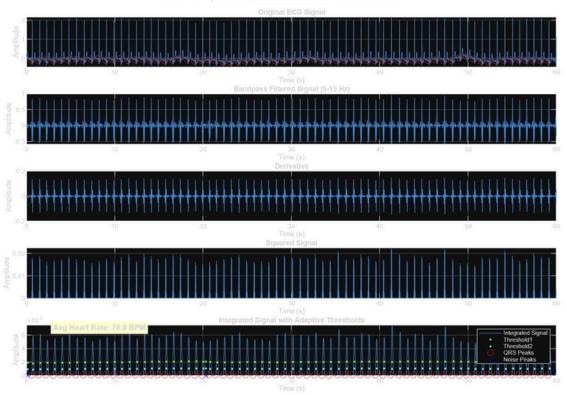


Processing record 103 (3 of 6)...

Warning: Could not load annotations. Returning empty annotation structure.

Loaded record 103: 21601 samples at 360 Hz Warning: No annotations found for record 103

Pan-Tompkins QRS Detection - Record 103

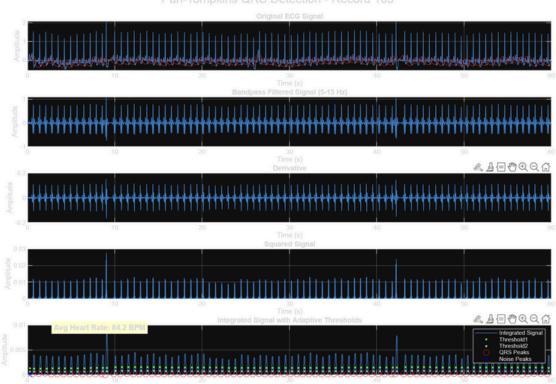


Processing record 105 (4 of 6)...

Warning: Could not load annotations. Returning empty annotation structure.

Loaded record 105: 21601 samples at 360 Hz Warning: No annotations found for record 105

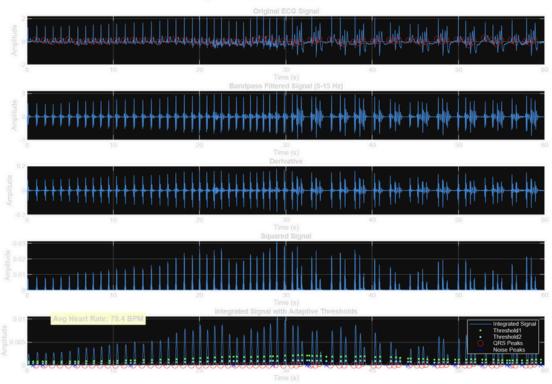
Pan-Tompkins QRS Detection - Record 105



Processing record 106 (5 of 6)...

Warning: Could not load annotations. Returning empty annotation structure.

Loaded record 106: 21601 samples at 360 Hz Warning: No annotations found for record 106



Processing record 109 (6 of 6)...

Warning: Could not load annotations. Returning empty annotation structure. Loaded record 109: 21601 samples at 360 Hz

Warning: No annotations found for record 109

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```
%% Summarize overall performance
fprintf('\n===== OVERALL PERFORMANCE =====\n');
===== OVERALL PERFORMANCE =====
if ~isempty(results)
```

```
% Check if all results have performance fields
    valid_results = arrayfun(@(x) isfield(x, 'performance') &&
~isempty(x.performance), results);
    if any(valid_results)
       % Extract only valid results that have performance data
       valid_results_idx = find(valid_results); % Use cell arrays to collect
                 metrics
                               sensitivity\_values = cellfun(@(i))
       results(i).performance.sensitivity,
num2cell(valid_results_idx));
       pp_values = cellfun(@(i) results(i).performance.positive_predictivity,
num2cell(valid_results_idx));
       f1_values = cellfun(@(i) results(i).performance.F1_score,
num2cell(valid_results_idx));
       fprintf('Average Sensitivity: %.2f%%\n', mean(sensitivity_values) * 100);
100);
       fprintf('Average Positive Predictivity: %.2f%\\n', mean(pp_values) *
       fprintf('Average F1 Score: %.2f\n', mean(f1_values));
```

```
fprintf('Based on %d valid records out of %d total records\n',
sum(valid_results), length(results));
  else
  end fprintf('No valid performance results available.\n');
else
  fprintf('No results available.\n');
end
No results available.
```

```
%% Show example with annotations and detections
if ~isempty(results) && length(results) >= 1
    % Select the first record with annotations for visualization
    record idx = 1;
    % Load the record data again for visualization
    record_num = results(record_idx).record;
    [ecg_signal, ~, annotations] = load_mitbih_record_local(record_num,
lead number, start time, duration);
    % Plot ECG with annotations and detections
    t = (0:length(ecg_signal)-1)/fs;
    figure('Name', sprintf('Record %d Annotation Comparison', record_num),
'Position', [100, 100, 1200, 600]);
    plot(t, ecg signal, 'b');
    hold on;
    % Plot QRS detections
    grs peaks = results(record idx).grs peaks;
    plot(qrs_peaks/fs, ecg_signal(qrs_peaks), 'ro', 'MarkerSize', 8,
'LineWidth', 1.5);
    % Plot annotations
    ann_qrs = results(record_idx).annotations;
2); plot(ann_qrs/fs, ecg_signal(ann_qrs), 'gx', 'MarkerSize', 10, 'LineWidth',
    title(sprintf('Record %d: QRS Detection vs Annotations', record_num),
'FontSize', 14);
    xlabel('Time (s)', 'FontSize', 12);
    ylabel('Amplitude', 'FontSize', 12);
legend('ECG Signal', 'QRS Detections', 'Reference Annotations');
    grid on;
    % Add performance text
    perf = results(record_idx).performance;
    text_str = sprintf(['Sensitivity: %.2f%%\nPositive Predictivity: %.2f%%\n'
                        'F1 Score: %.2f'], ...
perf.F1_score);
                        perf.sensitivity * 100, perf.positive_predictivity * 100,
```

```
annotation('textbox', [0.15, 0.75, 0.2, 0.15], 'String', text_str, ...
               'EdgeColor', 'none', 'BackgroundColor', [1 1 0.8], ...
               'FontSize', 12, 'FontWeight', 'bold');
end %% ==== LOCAL FUNCTION DEFINITIONS ===== function [qrs peaks, processed ecg]
    robust_pan_tompkins_qrs_detector_local(ecg_signal,
                                                         fs,
                                                                 plot results)
ROBUST_PAN_TOMPKINS_QRS_DETECTOR_LOCAL QRS complex detection optimized for MIT-
BIH Database
    [QRS_PEAKS, PROCESSED_ECG] =
%
ROBUST_PAN_TOMPKINS_QRS_DETECTOR_LOCAL(ECG_SIGNAL, FS, PLOT_RESULTS)
    detects QRS complexes in the ECG signal using an enhanced Pan-Tompkins
algorithm.
% Handle input arguments
if nargin < 1</pre>
    error('ECG signal is required');
end
if nargin < 2 || isempty(fs)</pre>
    fs = 360; % Default sampling frequency for MIT-BIH database
    fprintf('Using default sampling frequency of 360 Hz (MIT-BIH standard)\n');
end if nargin
< 3
    plot results = true;
end
% Remove NaN and Inf values
ecg_signal(isnan(ecg_signal) | isinf(ecg_signal)) = 0;
% Detrend to remove baseline wandering
ecg_signal = detrend(ecg_signal);
% Parameters for Pan-Tompkins algorithm (optimized for MIT-BIH)
integration window = round(0.150 * fs);
                                           % 150 ms window
filter_delay = round(0.2 * fs);
                                           % Filter delay compensation
refractory_period = round(0.2 * fs);
                                           % 200 ms refractory period
learning_rate_signal = 0.125;
                                           % Learning rate for signal peak
learning_rate_noise = 0.125;
                                           % Learning rate for noise peak
signal_threshold_weight = 0.25;
                                          % Signal threshold weight
noise_threshold_weight = 0.5;
                                           % Secondary threshold weight
% Initialize output structure
processed_ecg = struct();
processed_ecg.original = ecg_signal;
% ===== STAGE 1: Bandpass Filtering (5-15 Hz) =====
% Create bandpass filter more suitable for MIT-BIH database
%oਓdutc ut of for high-pass
highcut = 15; % 15Hz cutoff for low-pass
wn = [lowcut highcut] / (fs/2);
[b, a] = butter(4, wn, 'bandpass');
bandpass_signal = filtfilt(b, a, ecg_signal); % Zero-phase filtering
```

```
processed_ecg.bandpass = bandpass_signal;
% ===== STAGE 2: Derivative ======
% Five-point derivative
b = [1 \ 2 \ 0 \ -2 \ -1] * (1/8);
a = 1;
derivative_signal = filter(b, a, bandpass_signal);
processed ecg.derivative = derivative signal;
% ===== STAGE 3: Squaring ======
squared signal = derivative signal.^2;
processed_ecg.squared = squared_signal;
% ===== STAGE 4: Moving Window Integration ======
b = ones(1, integration window)/integration window;
integrated signal = filter(b, a, squared signal);
processed_ecg.integrated = integrated_signal;
% ===== STAGE 5: Adaptive Thresholding and Peak Detection ======
% Initialize variables for adaptive thresholding
qrs_peaks = [];
noise_peaks = [];
%PKignalOgeak level
MPMoiseOpeak level
threshold1 = 0; % First threshold
threshold2 = 0; % Second threshold
% Initial learning phase to set thresholds
learn_phase_duration = round(2 * fs); % 2 seconds learning
if length(integrated_signal) >= learn_phase_duration
    learn_segment = integrated_signal(1:learn_phase_duration);
    SPKI = mean(learn segment) + 2*std(learn segment);
   NPKI = mean(learn_segment);
    threshold1 = NPKI + signal threshold weight * (SPKI - NPKI);
    threshold2 = noise_threshold_weight * threshold1;
         Store
                 adaptive
                            thresholds for
                                               plotting
                                                          thresholds
zeros(size(integrated signal));
                                              thresholds2
zeros(size(integrated_signal));  % Find all local maxima
integrated signal using a sliding window window_size = round(0.1 * fs);
% 100 ms window peak_candidates = []; peak_values = []; for i =
window_size+1 : length(integrated_signal)-window_size
   window = integrated_signal(i-window_size:i+window_size);
    if integrated_signal(i) == max(window) && integrated_signal(i) > 0
        peak_candidates = [peak_candidates, i];
        peak_values = [peak_values, integrated_signal(i)];
    end
end
```

```
% Process each detected peak candidate
last grs = -refractory period;
threshold history = zeros(length(peak candidates), 2);
for i = 1:length(peak_candidates)
loc = peak_candidates(i);
peak val = peak values(i);
% Store current thresholds
thresholds(loc) = threshold1;
thresholds2(loc) = threshold2;
threshold_history(i,:) = [threshold1, threshold2];
% Check if this peak is within refractory period
if loc - last_qrs <= refractory_period</pre>
        % Update noise peak
        NPKI = learning_rate_noise * peak_val + (1-learning_rate_noise) * NPKI;
        noise peaks = [noise peaks, loc];
        continue;
    end
    % Classify peak as QRS or noise
    if peak_val > threshold1
        % This is a QRS complex
        qrs_peaks = [qrs_peaks, loc-filter_delay]; % Compensate for filter delay
        last qrs = loc;
        % Update signal peak
        SPKI = learning_rate_signal * peak_val + (1-learning_rate_signal) * SPKI;
    else
        % This is likely noise
        noise_peaks = [noise_peaks, loc];
        % Update noise peak
        NPKI = learning rate noise * peak val + (1-learning rate noise) * NPKI;
 end % Final check for missed beats using RR interval
 analysis if ~isempty(qrs_peaks)
   end
   % Update thresholds
   threshold1 = NPKI + signal_threshold_weight * (SPKI - NPKI);
   threshold2 = noise_threshold_weight * threshold1;
    % Calculate RR intervals
    RR = diff(qrs_peaks);
    mean_RR = mean(RR);
    missed_beats = [];
    % Check for missed beats (when RR interval is > 1.5x the mean RR)
    for i = 1:length(RR)
        if RR(i) > 1.8 * mean_RR
            % Potential missed beat in this interval
```

```
search_start = qrs_peaks(i); search_end = qrs_peaks(i+1); % Look for
           possible peaks in this range that were classified as noise for j =
           1:length(noise peaks)
               if noise_peaks(j) > search_start && noise_peaks(j) < search_end</pre>
                   missed peak val = integrated signal(noise peaks(j));
                   % Check if this peak would pass with a lower threshold
                   if missed_peak_val > 0.5 * threshold1
                       missed beats = [missed beats, noise peaks(j)-
filter_delay];
                   end
               end
           end
       end
   end
   % Add missed beats to QRS peaks
   qrs peaks = sort([qrs peaks, missed beats]);
end % Ensure QRS peak indices are within valid range qrs_peaks =
qrs_peaks(qrs_peaks > 0 & qrs_peaks <= length(ecg_signal)); % Store</pre>
results in output structure processed_ecg.qrs_peaks = qrs_peaks;
processed_ecg.threshold1 = thresholds; processed_ecg.threshold2
thresholds2; processed_ecg.noise_peaks = noise_peaks; % Plot results if
requested if plot_results
   t = (0:length(ecg_signal)-1)/fs;
   figure('Name', 'Pan-Tompkins QRS Detection', 'NumberTitle', 'off',
'Position', [100, 100, 1200, 800]);
   % Plot original signal
   subplot(5,1,1);
   plot(t, ecg_signal);
   title('Original ECG Signal');
   hold on;
   if ~isempty(qrs_peaks)
       valid_peaks = qrs_peaks(qrs_peaks > 0 & qrs_peaks <= length(ecg_signal));</pre>
       plot(valid_peaks/fs, ecg_signal(valid_peaks), 'ro', 'MarkerSize', 8);
   end
   xlabel('Time (s)');
   ylabel('Amplitude');
   grid on;
   % Plot bandpass filtered signal
   subplot(5,1,2);
   plot(t, processed_ecg.bandpass);
   title('Bandpass Filtered Signal (5-15 Hz)');
```

```
xlabel('Time (s)');
    ylabel('Amplitude');
    grid on;
    % Plot derivative
    subplot(5,1,3);
    plot(t, processed_ecg.derivative);
    title('Derivative');
    xlabel('Time (s)');
    ylabel('Amplitude');
    grid on;
    % Plot squared signal
    subplot(5,1,4);
    plot(t, processed_ecg.squared);
    title('Squared Signal');
    xlabel('Time (s)');
    ylabel('Amplitude');
    grid on;
    % Plot integrated signal with thresholds
    subplot(5,1,5);
    plot(t, processed ecg.integrated);
    title('Integrated Signal with Adaptive Thresholds');
    hold on;
    % Plot thresholds where they were calculated
    non zero idx = find(thresholds > 0);
    if ~isempty(non zero idx)
        plot(non_zero_idx/fs, thresholds(non_zero_idx), 'g.', 'MarkerSize', 8);
        plot(non zero_idx/fs, thresholds2(non_zero_idx), 'c.', 'MarkerSize', 8);
    % Plot detected QRS peaks
    if ~isempty(qrs_peaks)
        valid_peaks = qrs_peaks(qrs_peaks > 0 & qrs_peaks <=</pre>
length(processed_ecg.integrated));
        plot(valid peaks/fs, processed ecg.integrated(valid peaks), 'ro',
'MarkerSize', 8);
    end
    % Plot noise peaks
    if ~isempty(noise_peaks)
        valid_noise = noise_peaks(noise_peaks > 0 & noise_peaks <=</pre>
length(processed_ecg.integrated));
        plot(valid_noise/fs, processed_ecg.integrated(valid_noise), 'bx',
'MarkerSize', 8);
    legend('Integrated Signal', 'Threshold1', 'Threshold2', 'QRS Peaks', 'Noise
Peaks');
    xlabel('Time (s)');
    ylabel('Amplitude');
    grid on;
```

```
% Add heart rate display
    if ~isempty(qrs_peaks) && length(qrs_peaks) > 1
        RR = diff(qrs peaks)/fs;
        HR = 60./RR;
        avg HR = mean(HR);
        text(0.05, 0.95, sprintf('Avg Heart Rate: %.1f BPM', avg HR), ...
0.81);
            'Units', 'normalized', 'FontWeight', 'bold', 'BackgroundColor', [1 1
    end
    % Make the figure nicer
    set(gcf, 'Color', 'white');
    sgtitle('Pan-Tompkins QRS Detection Algorithm Results', 'FontSize', 16);
end end function [record data, fs, ann] = load mitbih record local(record num,
lead_num, start_time, duration) % LOAD_MITBIH_RECORD_LOCAL Load ECG data from MIT-
BIH Arrhythmia Database
    [RECORD DATA, FS, ANN] = LOAD MITBIH RECORD LOCAL(RECORD NUM, LEAD NUM,
START TIME, DURATION)
    loads the specified record from the MIT-BIH Arrhythmia Database.
% Check if WFDB Toolbox is installed
if ~exist('rdann', 'file') || ~exist('rdsamp', 'file')
     error(['WFDB Toolbox not found. Please install it from: ' ...
            'https://physionet.org/content/wfdb-matlab/0.10.0/']);
end % Handle default values if
nargin < 2 || isempty(lead_num)</pre>
    lead_num = 1;
end
if nargin < 3 || isempty(start_time)</pre>
    start_time = 0;
end
if nargin < 4</pre>
    duration = []; % Empty duration means load the entire record
end % Construct record name record name =
sprintf('%03d', record_num); try
    % Convert start time and duration to samples
    fs = 360; % MIT-BIH sampling frequency
    start_sample = round(start_time * fs);
    if isempty(duration)
        % Load entire record
        [signal, ~, ~] = rdsamp(['mitdb/' record_name], lead_num);
        end_sample = length(signal);
    else
```

```
end_sample = start_sample + round(duration * fs);
        % Load specified portion of the record
        [signal, ~, ~] = rdsamp(['mitdb/' record_name], lead_num, end_sample,
start_sample);
    end
    % Load annotations
    try
        ann = rdann(['mitdb/' record_name], 'atr', 'from', start_sample, 'to',
end_sample);
        % Adjust annotation indices for the extraction window
        ann.time = ann.time - start_sample;
    catch
        warning('Could not load annotations. Returning empty annotation
structure.');
        ann = struct('time', [], 'type', []);
    end
    % Extract the signal for the specified lead
    record data = signal;
catch ME
    error(['Error loading record: ', ME.message, '\n', ...
           'Ensure you have downloaded the MIT-BIH database and ', ...
           'it is in your MATLAB path or current directory.']);
                 function
                                [qrs stats,
                                                detection performance]
end
         end
evaluate_qrs_detection_local(detected_qrs, annotation_qrs, fs, window_size)
% EVALUATE_QRS_DETECTION_LOCAL Evaluate QRS detection against annotations
    [QRS STATS, DETECTION PERFORMANCE] =
EVALUATE QRS_DETECTION_LOCAL(DETECTED_QRS, ANNOTATION_QRS, FS, WINDOW_SIZE)
    evaluates QRS detection performance against reference annotations.
% Default window size is 100 ms if not specified
if nargin < 4 || isempty(window_size)</pre>
    window size = 0.1; % 100 ms
end
% Convert window size from seconds to samples
window_samples = round(window_size * fs);
% Initialize counters
true positives = 0;
matched ann = false(size(annotation qrs));
matched_det = false(size(detected_qrs));
% Loop through annotation QRS complexes
for i = 1:length(annotation qrs)
    % Find detected QRS within the window
    for j = 1:length(detected_qrs)
        if ~matched det(j) && abs(detected grs(j) - annotation grs(i)) <=</pre>
window_samples
```

```
true_positives = true_positives + 1;
            matched_ann(i)
                                            true;
            matched_det(j) = true; break;
        end
    end
       %
end
             Calculate
                           statistics
false_positives = sum(~matched det);
false negatives = sum(~matched ann);
% Build output structures qrs_stats
= struct(...
    'true_positives', true_positives, ...
    'false_positives', false_positives, ...
    'false_negatives', false_negatives);
% Calculate performance metrics
sensitivity = true_positives / (true_positives + false_negatives);
if (true_positives + false_positives) > 0
    positive_predictivity = true_positives / (true_positives + false_positives);
else
    positive_predictivity = 0;
end
% F1 score
if (sensitivity + positive_predictivity) > 0
    F1_score = 2 * (sensitivity * positive_predictivity) / (sensitivity +
positive_predictivity);
else
    F1 score = 0;
end
detection_performance = struct(...
    'sensitivity', sensitivity, ...
    'positive_predictivity', positive_predictivity, ...
    'F1_score', F1_score);
end
```