NATURE NOTES



Check for updates

Plant-arthropod interactions of an endangered California lupine

Carina I. Motta^{1,2} | Justin C. Luong^{2,3} | Katja C. Seltmann²

¹Departamento de Biodiversidade, Universidade Estadual Paulista Júlio de Mesquita Filho, Rio Claro, São Paulo, Brazil

²Vernon and Mary Cheadle Center for Biodiversity and Ecological Restoration, University of California, Santa Barbara, California, USA

³Environmental Studies Department, University of California, Santa Cruz, California. USA

Correspondence

Carina I. Motta and Katja C. Seltmann, Vernon and Mary Cheadle Center for Biodiversity and Ecological Restoration, University of California, Santa Barbara, CA, USA.

Emails: carinaisabellamotta@gmail.com (CIM) and seltmann@ccber.ucsb.edu (KCS)

Funding information

United States Fish and Wildlife Service; Vernon and Mary Cheadle Center for Biodiversity and Ecological Restoration at UC Santa Barbara

Abstract

The reintroduction of endangered plant species is an essential conservation tool. Reintroductions can fail to create resilient, self-sustaining populations due to a poor understanding of environmental factors that limit or promote plant success. Biotic factors, specifically plant–arthropod interactions, have been shown to affect the establishment of endangered plant populations. *Lupinus nipomensis* (Nipomo Mesa lupine) is a state of California (California Rare Plant Rank: 1B.1) and federally (65 FR 14888) endangered endemic plant with only one extant population located along the central California coast. How arthropods positively or negatively interact with *L. nipomensis* is not well known and more information could aid conservation efforts. We conducted arthropod surveys of the entire *L. nipomensis* extant population in spring 2017. Observed arthropods present on *L. nipomensis* included 17 families, with a majority of individuals belonging to Thripidae. We did not detect any obvious pollinators of *L. nipomensis*, providing support for previous studies suggesting this lupine is capable of self-pollinating, and observed several arthropod genera that could potentially impact the reproductive success of *L. nipomensis* via incidental pollination or plant predation.

KEVWODDS

coastal dune, endemic, Fabaceae, Lupinus nipomensis, pollination, restoration

TAXONOMY CLASSIFICATION

Restoration ecology

1 | INTRODUCTION

Plant-arthropod interactions, through processes such as pollination, herbivory, and frugivory, play an important role in the reproductive success of plant species (Nemec & Bragg, 2008; Schweizer et al., 2013; Strong et al., 1995). Low pollinator visitation diversity or abundance can explain reduced distribution, low reproductive output, or failure to establish novel plant populations in habitats that are otherwise abiotically suitable (Karron, 1987; Kearns et al., 1998).

Herbivorous arthropods meanwhile have been shown to negatively impact establishment and seed production of plants (Bevill et al., 1999; Münzbergová & Herben, 2005). Both the mutualistic and antagonistic interactions between plants and arthropods are important drivers in determining the survival of a plant (Stahl et al., 2018).

Despite the importance of arthropod interactions to plant species, current conservation efforts to restore endangered plants often prioritize the presence of appropriate abiotic conditions to select reintroduction sites (Falk et al., 1996; Godefroid et al., 2011; Guerrant

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2022 The Authors. Ecology and Evolution published by John Wiley & Sons Ltd.

& Kaye, 2007). Knowledge of herbivory and pollination has been shown to be important in maximizing reproduction and increasing establishment success of rare plant species (Archer & Pyke, 1991; Kay, 2008). Due to the often small population size of endangered plants, arthropod herbivores can have strong impacts through a reduction in plant size, growth, and recruitment (Ancheta & Heard, 2011; Myers & Sarfraz, 2017). Similarly, pollination is especially important to endangered plant species in order to maintain the population size and increase outcrossing of individuals (Horth, 2019; Reiter et al., 2017; Steffan-Dewenter et al., 2001). Maintaining genetic diversity and reducing the likelihood of inbreeding depression is essential to the conservation of rare plant species (Falk, 1990; Lee et al., 2018).

Lupinus nipomensis Eastw. (Fabaceae, Nipomo Mesa lupine) is a state and federally endangered annual forb endemic to Nipomo, California, USA (USFWS, 2009; Figure 1). Historically, *L. nipomensis* has occurred at low densities in back dunes and inter-dune habitat. The loss of coastal back dune habitat due to land conversion, fragmentation, and competition with the invasive perennial veldt grass (*Ehrharta calycina* Sm., Poaceae) limits the range and potential for natural regeneration of the *L. nipomensis* population (Skinner & Pavlik, 1994). The entire extant population is geographically isolated within a 5 km² area along the central California coast in the Guadalupe-Nipomo Dune Complex and comprises seven dispersed colonies. Total population size is dependent on winter and spring climatic conditions and ranges between 139 and 771 individuals per year (USFWS, 2019).

The reintroduction of Nipomo Mesa lupine, and other rare plants with a limited initial population size, requires specific abiotic conditions to maximize reproductive output from reintroduction efforts.



FIGURE 1 Flower peduncle of *Lupinus nipomensis* Eastw. (Nipomo mesa lupine). Image provided by JC Luong

Luong et al. (2019) provide an overview of abiotic microhabitat characteristics (i.e., landscape slope and aspect) and seed treatment relevant to *L. nipomensis* fecundity during reintroduction efforts. A foundational study of *L. nipomensis* by Walters and Walters (1989) primarily focuses on abiotic drivers of reproduction, specifically changes observed in flowering and fruit set as a factor of rainfall received. Walters and Walters (1989) also include a record of herbivorous arthropod interactions, but do not specifically include pollinator observations. Because other annual lupines (e.g., *Lupinus bicolor* Lindl.) are known to be pollinated by bees and flies, the absence of pollinator observations has led practitioners and researchers to hypothesize that *L. nipomensis* is capable of self-pollinating (Luong et al., 2019; Moldenke, 1976; USFWS, 2019).

In this study, we sought to establish a baseline of plant–arthropod interactions of in situ *L. nipomensis*, especially targeted pollinator observations, which have not previously been studied. We surveyed arthropod use of *L. nipomensis* and classified plant visitors as potential pollinators when observed on or in the flowers. The primary goal of this study was to create an inventory of observed arthropod interactions with *L. nipomensis* to inform future research and conservation efforts.

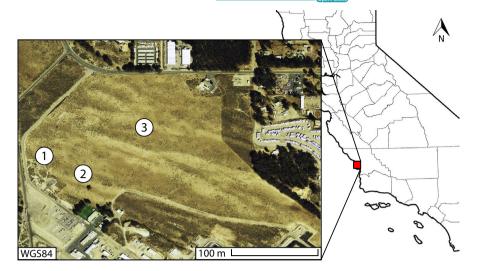
2 | METHODS

2.1 | Study area

This study was conducted in the spring of 2017 at the Phillips 66 Oil Refinery (35.0388889, -120.5894444) in San Luis Obispo County, California, USA (Figure 2). The region is characterized by a Mediterranean-type climate with cool, wet winters and hot, dry summers, while also receiving occasional inputs of water from coastal fog (Baguskas et al., 2016). San Luis Obispo County receives an average precipitation of 33.1 cm annually, has an average wind speed of 10.83 km/h (November–May), and average low and high temperatures ranging from 5.92 to 18.9°C during the growing season (Western Regional Climate Center, 2020). We classified cloud cover on a 4-point scale and recorded ambient air temperature and wind speed during each visit (see complete dataset Luong et al., 2021).

Phillips 66 is required to mitigate for their oil development by establishing protected areas where extant populations are regularly monitored and restricting management actions that could negatively affect *L. nipomensis* populations (USFWS, 2009). This protected area in which we conducted our study is a coastal back dune ecosystem and the oldest part of a dune complex. These less disturbed areas often have later successional plants with increased soil stability as well as higher plant and insect diversity (Buckler, 1979; Ferrier et al., 2012; Miller et al., 2010). The study site is dominated by non-native, invasive perennial veldt grass (*Ehrharta calycina*) with scattered native annual forbs and perennial shrubs. The area is actively grazed by cattle during *L. nipomensis* dormant season (June–November) to suppress the invasive veldt grass.

FIGURE 2 Map of sampled *Lupinus* nipomensis colonies (labeled 1–3) and geographic placement of study site in San Luis Obispo County, California, USA. The study area is indicated by the red box and county lines are displayed in gray on the map of California, USA. The inset box displaying the study area is approximately $400 \text{ m} \times 300 \text{ m}$



The entire population of *L. nipomensis* is restricted to seven colonies within the Phillips 66 protected area. Two of the colonies were excluded due to their wide and sparse distribution, making these colonies ineffective for targeted sampling. One colony was newly rediscovered from historic occurrence data and not found until partway through the course of the study. A fourth population was located along a roadside with different ambient disturbance characteristics compared to the other colonies. Plant visitor observations and vegetation monitoring were conducted at the remaining three colonies within the protected area in accordance with our California Fish and Wildlife permit (Permit No. 2081(a)-16–010) (Figure 2). The three colonies sampled are consistent colonies containing a majority of *L. nipomensis* individuals and are representative of the population as a whole.

2.2 | Plant visitor observations

One monitoring plot was established per colony (n = 3). Each plot was 8m × 8m and contained 22–96 L. nipomensis individuals. We conducted arthropod visitor surveys using a standardized observationbased protocol useful for recording flower and foliage visitations (Herrera, 1990; Thompson, 2001). Observers conducted a 40-min observation session of L. nipomensis arthropod visitors at each of the three plots per visit. A total of 48 h of L. nipomensis arthropod visitor surveys were conducted across all plots (16 h per plot) over the course of 24 visits between March 2017 and May 2017. Short observation times and single field season were due to limited accessibility, small population size, and the permitting protocol to access private property. Observations occurred between 12:00 and 15:00 (Pacific Standard Time), during the warmest part of the day when diurnal insects are most active to observe the greatest potential suite of insects visiting L. nipomensis (Willimer, 1983). One set of early and late sampling (beginning at 9:00 and 16:00, respectively) was conducted to increase the likelihood of sampling temporal niche visitors.

During each arthropod visitor survey, *L. nipomensis* individuals within the monitoring plot were closely observed and any visiting arthropods (floral and otherwise) were captured using aspirations and

hand or net collections (Chacoff & Aizen, 2006; Kleijn & Langevelde, 2006). It is important to note that not all flower visitors observed were pollinators, but they are considered potential or "incidental pollinators." Incidental pollinators move pollen from flower to flower while foraging for other resources (Anandhan et al., 2020; Kearns et al., 1998). Nets were only used when necessary to minimize damage as dictated by the California Department of Fish and Wildlife collection permit (Scientific Collecting Permit SC-13574). Incidental observations of nearby arthropods visiting other plant species within the plot were also recorded, and when possible, captured. At the end of each 40-minute survey, beat sampling was conducted on, and an inflorescence was collected from, each L. nipomensis individual in the plot. Inflorescence samples were stored in air-tight bags with an ethyl acetate cotton ball and kept cool until returned to the lab. At the lab, inflorescences were dissected and any arthropods found were placed in 75% ethanol vials. Specimen collection complied with California State and Federal laws and samples were vouchered at the Invertebrate Zoology Collection at the University of California, Santa Barbara (specimen catalog numbers are included in Appendix S1).

Collected insects were identified to family and received lower classifications if possible (Carvalho, 1955; Daniel & Franz, 2012; Gibson et al., 1997; Herring, 1976; Hoddle et al., 2012; Iowa State University Department of Entomology, 2017; Marshall, 2012; Ross et al., 2002; Schuh & Slater, 1995; Slater & Baranowski, 1978). Specimens identified to family were sorted into putative species based on morphology or morphospecies (Samways et al., 2010). New host records were determined and recorded via the Global Biotic Interactions (GloBI) database and through literature searches (Poelen et al., 2014). Families classified as flower visitors are considered potential pollinators and were observed on or in flower parts of *L. nipomensis*.

2.3 | Data visualization

Interaction data between *L. nipomensis* and visiting arthropods was visualized in R Studio (version 1.4.1106). An interaction web was

created using the *bipartite* package (Dormann, 2021). Arthropod icons were created or retrieved from http://phylopic.org/. Icons for Asilidae, Coccinellidae, Curculionidae, and Miridae are under a creative common license (http://creativecommons.org/licenses/by/3.0/) by G Monger, M Broussard, JC Giron, and K Garcia, respectively. The map of the study area was created using WGS84 imagery from ArcGIS (version 10.4, Environmental Systems Research Institute 2012) and edited in Adobe Illustrator (version 24.1.3).

3 | RESULTS

A total of 351 arthropod individuals were observed interacting with *L. nipomensis* during our surveys (Table 1). Records of the 157 vouchered specimens are available on the UCSB Invertebrate Zoology Collection, Global Biodiversity Information Facility (GBIF), and Global Biotic Interactions (GloBI) databases (Appendix S1). Twenty-two unique morphospecies from 8 orders and 17 families were classified. At least 1 individual from 11 unique families were found in or on a *L. nipomensis* flower (Figure 3).

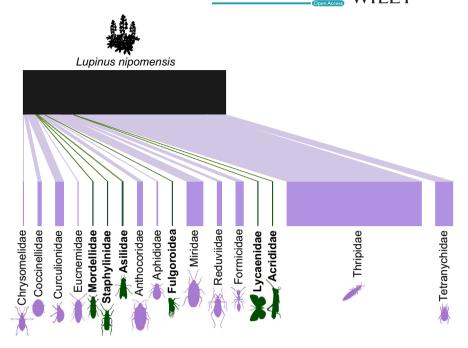
Individuals observed visiting L. nipomensis that were identified to genus and species include members of orders Diptera, Coleoptera, Hemiptera, Hymenoptera, and Lepidoptera (Appendix S1). The one Dipteran observed was Delia Iupini (Coquillett, 1901) (Anthomyiidae). The Coleoptera species were Diabrotica undecimpunctata Mannerheim, 1843 (Chyrsomelidae), Apleurus sp., Scaphomorphus sp., Trigonoscuta sp., and Rhigopsis effracta LeConte, 1874 (Curculionidae). Hemipterans identified to lower classifications include Orius sp. (Anthocordidae), Closterocoris amoenus (Provancher, 1887) (Miridiae), Lygus sp. (Miridae), and Apiomerus californicus Berniker & Szerlip, 2011 (Reduviidae). Collected hymenopterans were all Formicidae, including Crematogaster sp. and Linepithema humile (Mayr, 1868). A lepidoptera individual, Plebejus Iupini (Boisduval, 1869) (Lycaenidae), was observed landing on a vegetative part of L. nipomensis. Thysanoptera individuals were identified to genus (Thrips sp.) and accounted for 234 out of the total 320 individuals observed (Figure 3).

Incidental observations were made of arthropods visiting other plant species within the plot, including *Acmispon glaber* (Vogel) Brouillet (Deerweed, Fabaceae), *Amsinckia spectabilis* Fisch. & C.A. Mey. (seaside fiddlehead, Boraginaceae), *Collinsia heterophylla* Graham (purple

Order	Family	Flower visitor	No. of morphospecies	No. of individuals
Coleoptera				
	Chrysomelidae	Yes	1	1
	Coccinellidae	Yes	1	2
	Curculionidae	Yes	4	14
	Eucnemidae	Yes	1	1
	Mordellidae	No	1	1
	Staphylinidae	No	1	1
Diptera				
	Anthomyiidae	No	1	1
Hemiptera				
	Anthocoridae	Yes	1	9
	Aphididae	Yes	1	2
	Fulgoroidea	No	1	1
	Miridae	Yes	1	28
	Reduviidae	Yes	2	10
Hymenoptera				
	Formicidae	Yes	2	13
Lepidoptera				
	Lycaenidae	No	1	1
Orthoptera				
	Acrididae	No	1	1
Thysanoptera				
	Thripidae	Yes	1	234
Trombidiformes				
	Tetranychidae	Yes	1	31
Total			22	351

TABLE 1 Order, Family, flower visitation, number of morphospecies, and number of individuals of arthropods that were observed interacting with *Lupinus nipomensis*

FIGURE 3 Interaction network of arthropod families containing individuals found on *Lupinus nipomensis*. Families containing at least one individual found in or on flowers are colored in purple while families only containing individuals found on other, vegetative parts of the plant are colored in green with bolded text. The figure is organized alphabetically by Order and Family within Order, corresponding to Table 1



Chinese houses, Plantaginaceae), *Ehrharta calycina* Sm. (perennial veldt grass), *Lupinus chamissonis* Eschsch. (dune bush lupine, Fabaceae), and *Nemophila menziesii* Hook. & Arn. (baby blue eyes, Boraginaceae). We observed primarily pollinating insects, such as *Apis mellifera* Linnaeus, 1758 (Apidae), *Bombus vosnesenskii* Radoszkowski, 1862 (Apidae), Syrphidae, and Lepidopterans, visiting neighboring flowers (within 0–5 m of *L. nipomensis*), but never visiting *L. nipomensis* itself. A complete list of observed individuals is available in Appendix S1.

4 | DISCUSSION

Plant-arthropod interaction studies associated with restoration efforts are becoming more common due to a greater appreciation of the role arthropods play in the success of plants through positive (e.g., pollination) and antagonistic (e.g., herbivory) interactions (Bucharova et al., 2021; Cariveau et al., 2021; Sabatino et al., 2021). While our study did not detect obvious pollinators, we recorded a high number of arthropod individuals that could be incidental pollinators and inadvertently pollinate L. nipomensis while feeding on pollen or other plant resources (Gill, 1991). We observed over 200 Thysanoptera individuals present in L. nipomensis flowers (Table 1). Thysanoptera have been known to pollinate members of several angiosperm families, including fabaceous plants (Varatharajan et al., 2016; Velayudhan & Annadurai, 1986). Individuals of Thysanoptera we found within L. nipomensis inflorescences were observed to have pollen attached to their body (Appendix S2). However, Thysanoptera are well-known flower pests that consume pollen, potentially causing withering of flowers and lowering plant reproductivity (Reitz, 2009). We also observed arthropods known to be important pollinators (i.e., Apis mellifera) visiting neighboring plants, including another lupine species, Lupinus chamissonis Eschsch. (Aslan et al., 2016; Hung et al., 2018). While Apis mellifera and other members of Apidae are known

to pollinate lupine species and were detected in our survey plots, we did not observe these arthropods interacting with *L. nipomensis* (Luong et al., 2019). Our study supports previous findings that *L. nipomensis* is capable of self-pollinating due to an apparent lack of visitation by arthropods known to pollinate, such as bees and flies (Cullen et al., 2021; Nye & Anderson, 1974; USFWS, 2019). If pollination is occurring among *L. nipomensis* individuals, this service is likely being performed by incidental pollinators, such as Thysanoptera.

Arthropod genera were observed that could affect the reproductive success of L. nipomensis due to herbivory. The Dipteran found, Delia Iupini (Anthomyiidae), was collected from a gall present on a L. nipomensis individual and has been observationally implicated to reduce fecundity in a previous study of L. nipomensis (Walters & Walters, 1988). While some galls can be benign, most are detrimental to plant health and, in some cases, have been shown to threaten endangered plant species (Harris & Pitzschke, 2020; Kolesik et al., 2019). Members of Curculionidae, which are known plant pests, were observed within the stem and interacting with D. lupini galls (Johnson-Cicalese et al., 1990; Petrova et al., 2006). The impact of D. lupini gall presence on L. nipomensis reproductivity was not quantified in this study; however, it is possible there were additional individuals we did not observe that are affecting the fecundity of this lupine. Formicidae species we observed included Linepithena humile, the invasive Argentine ant (Holway, 1999). Argentine ants have been shown to impact floral visitation patterns and nesting success of other arthropods with the potential to create cascading, negative effects, and reduce pollinator visitation (Plentovich et al., 2021; Sahli et al., 2016; Underwood & Fisher, 2006). The presence of a gall-inducing and invasive arthropods may further inhibit recruitment within this singular extant population of L. nipomensis.

Plants face both biotic and abiotic barriers to reproductive success, and for rare, endangered species, these barriers can ultimately result in extirpation or extinction (Rejmánek, 2018). Small

populations, like that of *L. nipomensis*, can face pollination limitation, as small plant populations often do not attract pollinators due to low pollen rewards (Shi et al., 2005). A reduction in pollination leads to reduced outcrossing, ultimately resulting in a lower genetic diversity that further threatens already small, rare plant populations (Gray, 2019). Simultaneously, herbivorous arthropods can impact seed production and recruitment of plant species (Lucas-Barbosa, 2016). Active intervention may be necessary to promote outcrossing via hand pollination, as well as protect *L. nipomensis* from herbivorous arthropods to ensure the genetic diversity and successful establishment of new individuals (Serrano et al., 2021; Walsh et al., 2019).

Our results suggest that if pollination is occurring among *L. nipomensis* individuals, it is likely being performed by incidental pollinators. Additionally, we observed herbivorous arthropods that may threaten fecundity of this lupine. Further work is necessary to determine the frequency, as well as mode, of pollination and whether the potential threats this lupine faces from herbivorous arthropods will affect the establishment of novel populations during restoration efforts.

ACKNOWLEDGMENTS

We thank the Land Conservancy of San Luis Obispo and the Phillip-66 refinery in Arroyo Grande for their cooperation and support. We greatly appreciate the data collection assistance of Marli Heininger, Barry Lao, Rachel Behm, Chelsea Chung, and Lindsey Roddick. We thank Lisa Stratton and Connie Rutherford for their logistical support. The data analysis and visualization advice provided by Ana Miller-ter Kuile was greatly appreciated. We are also grateful for Matt Guilliams at the Santa Barbara Botanic Garden for providing us the use of their scanning electron microscope and Leica imaging software. Matthew L. Gimmel assisted in identification of beetles observed in this study. We also thank Patrick Turner, Adam Lambert, and Azucena Lucatero. This study was funded by the United States Fish and Wildlife Service, Ventura Field office under a USFWS cooperative agreement award with the CFDA program 15.650 agreement number F12AC00683 for "Recovery Activities for the endangered plant Nipomo Lupine." Additional funding support was provided by the Vernon and Mary Cheadle Center for Biodiversity and Ecological Restoration at UC Santa Barbara.

CONFLICT OF INTEREST

Any findings or recommendations expressed in this material do not necessarily reflect the views of funding agencies. The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

AUTHOR CONTRIBUTION

Carina Isabella Motta: Formal analysis (lead); Visualization (lead); Writing – original draft (equal); Writing – review & editing (lead). Justin C. Luong: Conceptualization (equal); Data curation (equal); Funding acquisition (equal); Methodology (equal); Project administration (lead); Writing – original draft (equal); Writing – review &

editing (equal). **Katja C Seltmann:** Conceptualization (equal); Data curation (equal); Formal analysis (supporting); Funding acquisition (equal); Methodology (equal); Project administration (equal); Visualization (supporting); Writing – original draft (supporting); Writing – review & editing (equal).

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are openly available in "Plant-arthropod interactions of an endangered California lupine" at https://doi.pangaea.de/10.1594/PANGAEA.938184. Most of the data are freely available and can be used under the terms of the license mentioned on the data set description. Records of vouchered specimens can be found on the Global Biotic Interactions (GloBI) database, available at: https://github.com/globalbioticinteractions/lupinus-nipomensis-interactions-2017.

ORCID

Carina I. Motta https://orcid.org/0000-0001-7127-7638

Justin C. Luong https://orcid.org/0000-0003-2118-4788

Katja C. Seltmann https://orcid.org/0000-0001-5354-6048

REFERENCES

- Anandhan, R., Kazmi, S. I., & Dey, S. (2020). Large carpenter bees as key pollinators of gliricidia sepium. *Indian Journal of Entomology*, 82(4), 691–693. https://doi.org/10.5958/0974-8172.2020.00188.1
- Ancheta, J., & Heard, S. B. (2011). Impacts of insect herbivores on rare plant populations. *Biological Conservation*, 144(10), 2395–2402. https://doi.org/10.1016/j.biocon.2011.06.019
- Archer, S., & Pyke, D. A. (1991). Plant-animal interactions affecting plant establishment and persistence on revegetated rangeland. *Journal of Range Management*, 44(6), 558–565. https://doi.org/10.2307/4003036
- Aslan, C. E., Liang, C. T., Galindo, B., Kimberly, H., & Topete, W. (2016). The role of honey bees as pollinators in natural areas. *Natural Areas Journal*, 36(4), 478–488. https://doi.org/10.3375/043.036.0413
- Baguskas, S. A., Still, C. J., Fischer, D. T., D'Antonio, C. M., & King, J. Y. (2016). Coastal fog during summer drought improves the water status of sapling trees more than adult trees in a California pine forest. *Oecologia*, 181(1), 137–148. https://doi.org/10.1007/s0044 2-016-3556-y
- Bevill, R. L., Louda, S. M., & Stanforth, L. M. (1999). Protection from natural enemies in manning rare plant species. *Conservation Biology*, 13(6), 1323–1331. https://doi.org/10.1046/j.1523-1739.1999.98450.x
- Bucharova, A., Lampei, C., Conrady, M., May, E., Matheja, J., Meyer, M., & Ott, D. (2021). Plant provenance affects pollinator network: Implications for ecological restoration. *Journal of Applied Ecology*, 1–11. https://doi.org/10.1111/1365-2664.13866
- Buckler, W. R. (1979). Dune type inventory and barrier dune classification study of Michigan's Lake Michigan shore (p. 23). Geological Survey Division, Michigan Dept. of Natural Resources.
- Cariveau, D. P., Bruninga-socolar, B., & Pardee, G. L. (2021). A review of the challenges and opportunities for restoring animal-mediated pollination of native plants. *Emerging Topics in Life Sciences*, 4, 99–109. https://doi.org/10.1042/ETLS20190073
- Carvalho, J. C. (1955). Keys to the genera of Miridae of the world (Hemiptera). Boletim do Museu Paraense Emilio Goeldi, 11(2), 5–151.
- Chacoff, N. P., & Aizen, M. A. (2006). Edge effects on flower-visiting insects in grapefruit plantations bordering premontane subtropical forest. *Journal of Applied Ecology*, 43, 18–27. https://doi.org/10.1111/j.1365-2664.2005.01116.x

- Cullen, N., Xia, J., Wei, N. A., Kaczorowski, R., Arceo-Gómez, G., O'Neill, E., Hayes, R., & Ashman, T.-L. (2021). Diversity and composition of pollen loads carried by pollinators are primarily driven by insect traits, not floral community. *Oecologia*, 196(1), 131–143. https://doi.org/10.1007/s00442-021-04911-0
- Daniel, D. H., & Franz, N. M. (2012). An interactive key to the genera of Entiminae weevils of Arizona (Curculionidae: Entiminae). Available at: http://mx.speciesfile.org/projects/85/public/site/asu/home Accessed: 30 June 2017
- Dormann, C. F., Fruend, J., Gruber, B., Dormann, M. C. F., LazyData, T. R. U. E., & ByteCompile, T. R. U. E. (2021). *Package 'bipartite'*. CRAN.
- Falk, D. A. (1990). Endangered forest resources in the U. S.: Integrated strategies for conservation of rare species and genetic diversity. Forest Ecology and Management, 35, 91–107. https://doi. org/10.1016/0378-1127(90)90234-3
- Falk, D. A., Millar, C. I., & Olwell, M. (1996). Restoring diversity: strategies for reintroduction of endangered plants. Island Press.
- Ferrier, S. M., Bangert, R. K., Hersch-Green, E. I., Bailey, J. K., Allan, G. J., & Whitham, T. G. (2012). Unique arthropod communities on different host-plant genotypes results in greater arthropod diversity. Arthropod-Plant Interactions, 6(2), 187–195. https://doi.org/10.1007/s11829-011-9177-9
- Gibson, G. A. P., Huber, J. T., & Wooley, J. B. (1997). Annotated Key to the Genera of Nearctic Chalcidoidea (Hymenoptera). NRC Research Press.
- Gill, R. A. (1991). The value of honeybee pollination to society. *Acta Horticulturae*, 288, 62–68.
- Godefroid, S., Piazza, C., Rossi, G., Buord, S., Stevens, A.-D., Aguraiuja, R., Cowell, C., Weekley, C. W., Vogg, G., Iriondo, J. M., Johnson, I., Dixon, B., Gordon, D., Magnanon, S., Valentin, B., Bjureke, K., Koopman, R., Vicens, M., Virevaire, M., & Vanderborght, T. (2011). How successful are plant species reintroductions? *Biological Conservation*, 144(2), 672–682. https://doi.org/10.1016/j.biocon.2010.10.003
- Gray, A. (2019). The ecology of plant extinction: Rates, traits and island comparisons. *Oryx*, 53(3), 424–428. https://doi.org/10.1017/S0030605318000315
- Guerrant, E. O., & Kaye, T. N. (2007). Reintroduction of rare and endangered plants: Common factors, questions and approaches. Australian Journal of Botany, 55(3), 362–370. https://doi.org/10.1071/BT06033
- Harris, M. O., & Pitzschke, A. (2020). Plants make galls to accommodate foreigners: some are friends, most are foes. *New Phytologist*, *225*(5), 1852–1872. https://doi.org/10.1111/nph.16340
- Herrera, C. M. (1990). Daily patterns of pollinator activity, differential pollinating effectiveness, and floral resource availability, in a summer-flowering Mediterranean shrub. *Oikos*, 58(3), 277–288. https://doi.org/10.2307/3545218
- Herring, J. L. (1976). Keys to Genera of Anthocoridae of America North of Mexico, with Description of a New Genus (Hemiptera: Heteroptera). *Florida Entomological Society*, *59*(2), 143–150.
- Hoddle, M. S., Mound, L. A., & Paris, D. L. (2012). *Thrips of California*. CBIT Publishing. Available at: http://keys.lucidcentral.org/keys/v3/thrips_of_california/Thrips_of_California.html
- Holway, D. A. (1999). Competitive mechanisms underlying the displacement of native ants by the invasive argentine ant. *Ecology*, 80(1), 238–251.
- Horth, L. (2019). 'Understanding the impact of plant arthropod interactions, pollination, and canopy light on the rare orchid, small whorled pogonia (*Isotria medeoloides*). *Plant Ecology*, 220(6), 563–576. https://doi.org/10.1007/s11258-019-00936-x
- Hung, K. J., Kingston, J. M., Albrecht, M., Holway, D. A., & Kohn, J. R. (2018). The worldwide importance of honey bees as pollinators in natural habitats. *Proceedings of the Royal Society B: Biological Sciences*, 285(1870), 20172140.
- Iowa State University Department of Entomology. (2017). Bugguide.net Identification. Available at: http://bugguide.net/node/view/15740

- Johnson-Cicalese, J. M., Wolfe, G. W., & Funk, C. R. (1990). Biology, distribution, and taxonomy of billbug turf pests (Coleoptera: Curculionidae). Environmental Entomology, 19(4), 1037–1046.
- Karron, J. D. (1987). The pollination ecology of co-occurring geographically restricted and widespread species of Astragalus (Fabaceae). *Biological Conservation*, 39(3), 179–193. https://doi. org/10.1016/0006-3207(87)90033-4
- Kay, T. N. (2008). Vital steps toward success of endangered plant reintroductions. *Native Plants Journal*, 9(3), 313–322. https://doi. org/10.1353/npj.0.0025
- Kearns, C. A., Inouye, D. W., & Waser, N. M. (1998). Endangered mutualisms: The conservation of plant-pollinator interactions. *Annual Review of Ecology and Systematics*, 29(1998), 83–112. https://doi.org/10.1146/annurev.ecolsys.29.1.83
- Kleijn, D., & Langevelde, F. V. (2006). Interacting effects of landscape context and habitat quality on flower visiting insects in agricultural landscapes. *Basic and Applied Ecology*, 7, 201–214. https://doi. org/10.1016/j.baae.2005.07.011
- Kolesik, P., Butterworth, N., Lemmon, J., Gibson, T., & Wallman, J. F. (2019). First gall midge (Diptera: Cecidomyiidae) known to feed on plant family Atherospermataceae: a new species of Asphondylia damaging the endangered Australian tree Daphnandra johnsonii. Austral Entomology, 58(2), 317–323. https://doi.org/10.1111/aen.12387
- Lee, S.-R., Choi, J.-E., Lee, B.-Y., Yu, J.-N., & Lim, C. E. (2018). Genetic diversity and structure of an endangered medicinal herb: Implications for conservation. *AoB PLANTS*, 10(2), ply021. https://doi.org/10.1093/aobpla/ply021
- Lucas-Barbosa, D. (2016). Integrating studies on plant-pollinator and plant-herbivore interactions. *Trends in Plant Science*, 21(2), 125–133. https://doi.org/10.1016/j.tplants.2015.10.013
- Luong, J. C., Motta, C. I., & Seltmann, K. C. (2021). Plant-arthropod interactions of an endangered California lupine. Retrieved from https://doi.pangaea.de/https://doi.org/10.1594/PANGAEA. 938184
- Luong, J. C., Nolan, M. P., & Stratton, L. C. (2019). The importance of pilot studies and understanding microhabitat requirements when reintroducing endemic plants during coastal dune restoration. *Journal* of Coastal Conservation, 23(3), 553–562. https://doi.org/10.1007/ s11852-019-00684-0
- Luong, J. C., Turner, P. L., Phillipson, C. N., & Seltmann, K. C. (2019). Local grassland restoration affects insect communities. *Ecological Entomology*, 44(4), 471–479. https://doi.org/10.1111/een.12721
- Marshall, S. (2012). Flies: The Natural History & Diversity of Diptera, 1st ed. Firefly Books.
- Miller, T. E., Gornish, E. S., & Buckley, H. L. (2010). Climate and coastal dune vegetation: Disturbance, recovery, and succession. *Plant Ecology*, 206(1), 97–104. https://doi.org/10.1007/s11258-009-9626-z
- Moldenke, A. R. (1976). California pollination ecology and vegetation types. *Phytologia*, 34(4), 305–361.
- Münzbergová, Z., & Herben, T. (2005). Seed, dispersal, microsite, habitat and recruitment limitation: Identification of terms and concepts in studies of limitations. *Oecologia*, 145(1), 1–8. https://doi.org/10.1007/s00442-005-0052-1
- Myers, J. H., & Sarfraz, R. M. (2017). Impacts of insect herbivores on plant populations. *Annual Review of Entomology*, 62, 207–230. https://doi.org/10.1146/annurev-ento-010715-023826
- Nemec, K. T., & Bragg, T. B. (2008). Plant-Feeding hemiptera and orthoptera communities in native and restored mesic tallgrass prairies. *Restoration Ecology*, 16(2), 324–335. https://doi. org/10.1111/j.1526-100X.2007.00306.x
- Nye, W. P., & Anderson, J. L. (1974). Insect pollinators frequenting strawberry blossoms and the effect of honey bees on yield and fruit quality. Journal of the American Society for Horticultural Science, 99(1), 40–44.

- Petrova, V., Čudare, Z., & Cibuļskis, R. (2006). Predators and herbivores beetles (Coleoptera) naturally occurring on strawberry (Latvia). *Acta Biologica Universitatis Daugavpiliensis*, 6(1–2), 155–159.
- Plentovich, S., Graham, J. R., Haines, W. P., & King, C. B. A. (2021). 'Invasive ants reduce nesting success of an endangered Hawaiian yellow-faced bee. *Hylaeus Anthracinus*', *NeoBiota*, 154, 137–154. https://doi.org/10.3897/neobiota.64.58670
- Poelen, J. H., Simons, J. D., & Mungall, C. J. (2014). Global biotic interactions: An open infrastructure to share and analyze species-interaction datasets. *Ecological Informatics*, 24, 148–159. https://doi.org/10.1016/j.ecoinf.2014.08.005
- Reiter, N., Vlcek, K., O'Brien, N., Gibson, M., Pitts, D., Brown, G. R., Bower, C. C., & Phillips, R. D. (2017). Pollinator rarity limits reintroduction sites in an endangered sexually deceptive orchid (Caladenia hastata): Implications for plants with specialized pollination systems. Botanical Journal of the Linnean Society, 184, 122–136. https://doi.org/10.1093/botlinnean/box017
- Reitz, S. R. (2009). Biology and ecology of the western flower thrips (Thysanoptera: Thripidae): The making of a pest. Florida Entomologist, 92(1), 7–13. https://doi.org/10.1653/024.092.0102
- Rejmánek, M. (2018). Vascular plant extinctions in California: A critical assessment. Diversity and Distributions, 24(1), 129–136. https://doi. org/10.1111/ddi.12665
- Ross, A. H., Thomas, M. C., & Skelley, P. E. (2002). American Beetles, Volume II Polyphaga: Scarabaeoidea through Curculionidea. CRC Press.
- Sabatino, M., Rovere, A., & Meli, P. (2021). Restoring pollination is not only about pollinators: Combining ecological and practical information to identify priority plant species for restoration of the Pampa grasslands of Argentina. *Journal for Nature Conservation*, 61, 126002.
- Sahli, H. F., Krushelnycky, P. D., Drake, D. R., & Taylor, A. D. (2016). Patterns of floral visitation to native Hawaiian plants in presence and absence of invasive argentine ants 1. Pacific Science, 70(3), 309-322. https://doi.org/10.2984/70.3.3
- Samways, M. J., McGeoch, M. A., & New, T. R. (2010). *Insect conservation:* a handbook of approaches and methods. Oxford University Press.
- Schuh, R. T., & Slater, J. A. (1995). True Bugs of the World (Hemiptera:Heteroptera): Classification and Natural History. Cornell University Press.
- Schweizer, D., Gilbert, G. S., & Holl, K. D. (2013). Phylogenetic ecology applied to enrichment planting of tropical native tree species. *Forest Ecology and Management*, 297, 57–66. https://doi.org/10.1016/j. foreco.2013.02.012
- Serrano, H. C., Pinto, M. J., Branquinho, C., & Martins-Loução, M. A. (2021). Ecology as a tool to assist conservation of a rare and endemic Mediterranean plantago species. *Frontiers in Ecology and Evolution*, 9, https://doi.org/10.3389/fevo.2021.614700
- Shi, X. J., Michaels, H. J., & Mitchell, R. J. (2005). Effects of self-pollination and maternal resources on reproduction and offspring performance in the wild lupine, *Lupinus perennis* (Fabaceae). Sexual Plant Reproduction, 18, 55-64. https://doi.org/10.1007/s00497-005-0250-3
- Skinner, M. W., & Pavlik, B. M. (1994). CNPS inventory of rare and endangered vascular plants of California. The California Native Plant Society.
- Slater, J., & Baranowski, R. (1978). How to know the true bugs (Hemiptera-Heteroptera). William C. Brown Publishing.
- Stahl, E., Hilfiker, O., & Reymond, P. (2018). Plant-arthropod interactions: Who is the winner? *Plant Journal*, *93*(4), 703–728. https://doi.org/10.1111/tpj.13773

- Strong, D. R., Maron, J. L., Connors, P. G., Whipple, A., Harrison, S., & Jefferies, R. L. (1995). High mortality, fluctuation in numbers, and heavy subterranean insect herbivory in bush lupine, Lupinus arboreus. *Oecologia*, 104(1), 85–92.
- Steffan-Dewenter, I., Mïnzenberg, U., & Tscharntke, T. (2001).

 Pollination, seed set and seed predation on a landscape scale.

 The Royal Society, 268, 1685–1690. https://doi.org/10.1098/rspb.
 2001.1737
- Thompson, J. D. (2001). How do visitation patterns vary among pollinators in relation to floral display and floral design in a generalist pollination system? *Oecologia*, 126, 386–394. https://doi.org/10.1007/s004420000531
- Underwood, E. C., & Fisher, B. L. (2006). The role of ants in conservation monitoring: If, when, and how. *Biological Conservation*, 132, 166–182. https://doi.org/10.1016/j.biocon.2006.03.022
- USFWS (2009). Lupinus nipomensis (Nipomo Lupine) 5-Year Review: Summary and Evaluation. USFWS.
- USFWS (2019). Nipomo Mesa Lupine (Lupinus nipomensis) 5-Year Review: Summary and Evaluation. USFWS.
- Varatharajan, R., Maisnam, S., Shimray, C. V., & Rachana, R. R. (2016). Pollination potential of thrips (Insecta: Thysanoptera) – an overview. Zoo's Print, 31(4), 6–12.
- Velayudhan, R., & Annadurai, R. S. (1986). Pollination potential of thrips (Insecta: Thysanoptera) in some fabaceous plants. *Proceedings: Animal Sciences*, 95(1), 109–116. https://doi.org/10.1007/BF031 79364
- Walsh, S. K., Pender, R. J., Junker, R. R., Daehler, C. C., Morden, C. W., & Lorence, D. H. (2019). Pollination biology reveals challenges to restoring populations of Brighamia insignis (Campanulaceae), a critically endangered plant species from Hawai'i. Flora, 259, 151448. https://doi.org/10.1016/j.flora.2019.151448
- Walters, D. R., & Walters, B. K. (1988). Taxonomy, demography, and ecology of Lupinus nipomensis Eastwood. California Polytechnic State University.
- Walters, D. R., & Walters, B. K. (1989). The Natural history of the nipomo mesa lupine (Lupinus nipomensis Eastwood) PART ONE. Crossosoma, 15(3), 1-14.
- Western Regional Climate Center. (2020). Web. 22 September 2020. CEMP historical weather data. https://wrcc.dri.edu/Climate/summaries.php
- Willimer, P. G. (1983). Thermal constraints on activity patterns in nectar-feeding insects. *Ecological Entomology*, 8(4), 455–469. https://doi.org/10.1111/j.1365-2311.1983.tb00524.x

SUPPORTING INFORMATION

Additional supporting information may be found in the online version of the article at the publisher's website.

How to cite this article: Motta, C. I., Luong, J. C., & Seltmann, K. C. (2022). Plant–arthropod interactions of an endangered California lupine. *Ecology and Evolution*, 12, e8688. https://doi.org/10.1002/ece3.8688