

Testing the Effects of Transfection on Mammalian Cytokine RNA Expression

Austin Mitchell Vecchio

Abstract—Abstract outline

This experiment aims to study the effect of genetic expression on polyplex treated cells against β -actin and $\text{INF}\alpha$.

I. INTRODUCTION

Gene therapy is an experimental method for treating disease by introducing a healthy copy of a defective gene into the patient's cells to alter the patients genetic material. The alternative is a non viral gene therapy method which can often have simple and large scale production with low host immunogenicity. However, non viral gene therapy can often yield low transfection efficiencies. New advances in technologies have improved transfection efficiency by leveraging cytokines. Cytokines are small proteins that are involved in various types of cell signaling. Classifications of these signalings include interferons, chemokines and interleukins.

In this experiment, there are two devices used. The first device, the Qubit, is an electronic device that quantifies a substance through fluorescence. The second device is qPCR. Reverse Transcriptase (RT)qPCR is a technique used to describe the level of genetic expression is occurring in vitro by measuring the amount of mRNA within a sample. In this technique, mRNA is reverse transcribed in to complementary DNA by an enzyme labeled reverse transcriptase. For the Real Time PCR experiment, since it is impossible to eliminate all genomic DNA from the cDNA synthesis, two solutions will be prepared. The first solution will be a +RT which will include reverse transcriptase and thus, will have cDNA. The second solution will be a -RT solution which will lack reverse transcriptase. When QPCR is performed on the -RT solution, genomic DNA will be amplified and will be seen in the resulting data.

In this experiment, β -actin and RPL13A will be used for direct controls against the immunological transcripts. $\text{INF}\beta$ will not be used as it will only appear after cycle 40 in the experiment producing undesirable results. The No Template Control will consist of DEPC water. The immunological transcripts that will be used in this experiment are Interleukin-6 (IL6) and Interferon- α ($\text{INF}\alpha$) Beta actin is a type of actin isoform which is highly involved in cell motility, structure and integrity. RPL13A is a gene that codes for the 60S ribosomal L13a protein. Interleukin-6 (IL-6) is a multifunctional cytokine that defends the host in response to immune and hematopoietic activities. Interferon- α is a part of a large subgroup of interferon proteins that help regulate the activity of the immune system.

Advisor: Dipl.-Ing. Michelle Ammerman, Lehrstuhl für Nachrichtentechnik, TUM, WS 2050/2051.

II. METHODS

A. RNA Purification

The cells in trizol were thawed before phase separation. During phase separation, the poly treated cells in trizol were incubated at 42° after which an 0.2mL of chloroform was added per 1mL of Trizol. The homogenized sample was incubated at 42° for 2-3 minutes following a vigorous shake for 15 seconds. The sample was then centrifuged 12,000 g for 15 minutes at 4°C before transferring the aqueous phase to a second tube. (100)Again, the sample was centrifuged 12,000 g for 15 minutes at 4°C . The pellet was not washed in fear of losing RNA. The solution then, underwent a series of centrifuges in between the removal of ethanol to result in an RNA pellet. The pellet was then air dried to remove remaining ethanol. The pellet was incubated at 42° following a resuspension in 20 μL of RNase free water.

B. cDNA Synthesis

For the first step of cDNA Synthesis, two solutions (+RT/-RT) were formed from the following compounds: 10x dsDNase Buffer, dsDNase, Template RNA, polyplex 24hour RNA and nuclease free water. Both solutions were incubated at 42° after being centrifuged. The solutions were then chilled on ice, centrifuged and placed back on ice. For both solutions, 5X Reaction mix and nuclease free water was added. In the +RT solution, Maxima Enzyme mix (reverse transcriptase) was added to the mixture. The -RT solution used DEPC H₂O instead of Maxima Enzyme mix so that the -RT solution can simulate the +RT solution without synthesizing RNA into cDNA. When the -RT solution undergoes QPCR, any contaminating genomic DNA will be amplified. The two solutions will be then mixed gently and centrifuged. An incubation period will take place at 25°C for 10 min followed by a second incubation session at 50°C for 15 min. The reaction will be terminated by heating the solutions at 85°C for 5 min. This process was conducted using Maxima First Strand cDNA Synthesis Kit with dsDNase (Thermo Scientific, Cat: K1671)

C. QPCR

Each well will contain 10 μL of 2X iQaq universal SYBR Green Supermix, 5 μL of a primer (IL6, $\text{INF}\alpha$, β -actin or RPL13A), and 5 μL of the respective diluted cDNA. The cDNA needs to be diluted to 4ng/ μL from their respective concentrations before it is added to the wells. The concentrations of cDNA for both sets of Poly24 and no washed cells were all around 100ng/ μL . Therefore only one set of calculations were needed for determining the proper dilution

of cDNA into water for either $\text{INF}\alpha$ and IL6 or β -actin and RPL13A and the respective -RT wells. For just cells, a different set of calculations were needed since the original cDNA concentrations were around 11 ng/ μ L. All wells contained the Green Supermix. The columns were sorted in the following manner: 1-3 contained IL6, 4-6 contained $\text{INF}\alpha$, 7-9 contained β -actin and 10-12 contained RPL13A. The QrtPCR was ran for 40 cycles.

D. Gel Electrophoresis

In order to further understand the results gained from the QrtPCR, an 1.2and gel electrophoresis was ran on the PCR product.

E. Analysis

The results from the QrtPCR produced Ct values from each well. A Ct value is a numeric inverse correlation to the quantity of nucleic acid detected by the apparatus. The Ct values can be used in combination with the $\Delta\Delta\text{Ct}$ formula to produce fold inductions.

III. RESULTS

A. Qubit

Following the cDNA synthesis, the concentrations were measured by the Qubit. The concentrations are as follows: SD1 (standard 1): 54.27ng/ μ L, SD2 (standard 2): 1057.52ng/ μ L, Tk: 8.75ng/ μ L, DA: 68.0ng/ μ L, AV: 487.0ng/ μ L, TK: 28.0ng/ μ L.

B. QrtPCR

TABLE I
WELLS FOR IMMUNOLOGICAL RESPONSES

	IL6	$\text{INF}\alpha$	β -actin	RPL13A
Poly24[AV]-NoWash	3.85E-05	7.58E-03	5.09E-05	1.05E-02
Poly24[AV]-Cells	3.10E-02	3.54E-02	3.94E-02	4.27E-02
NoWash-Cells	8.01E+02	4.65E+00	7.01E+02	4.08E+00
Poly24[Tk]-NoWash	8.20E-05	4.64E-03	3.77E-05	6.04E-03
Poly24[Tk]-Cells	6.54E-02	5.90E-02	2.57E-02	2.55E-02
Poly6+iCRT-NoWash6	500.25	421.07	25.83	22.29

C. Gel Electrophoresis

An agarose gel was ran to further analyze the amplified cDNA from the QrtPCR reaction.

IV. DISCUSSION

The mRNA was purified and converted to cDNA. The resulting concentration was relatively high compared to peers. This eludes that the treatment for these cells of polycationic DNA for 24 hours could result in higher transcription rates.

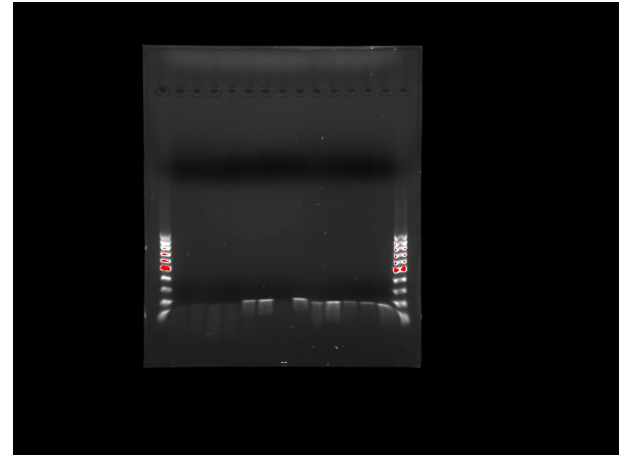


Fig. 1. Agarose Gel

TABLE II
PCR WELLS FOR IMMUNOLOGICAL RESPONSES

	1	2	3	4	5	6
A (Poly24[TK])	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L
B (Just Cells)	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L
C (No Wash)	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L
D (Poly24[AV])	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L
E (-RT)	Ts	Ni	AV	Ts	Ni	AV
E (NTC)	Ts	Ni	AV	Ts	Ni	AV
G	Mi (-RT)	Mi (NTC)		Mi (-RT)	Mi (NTC)	
H						

TABLE III
PCR WELLS FOR CONTROLS

	7	8	9	10	11	12
A (Poly24[TK])	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L
B (Just Cells)	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L
C (No Wash)	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L
D (Poly24[AV])	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L	4ng/ μ L
E (-RT)	Ts	Ni	AV	Ts	Ni	AV
E (NTC)	Ts	Ni	AV	Ts	Ni	AV
G	Mi (-RT)	Mi (NTC)		Mi (-RT)	Mi (NTC)	
H						

V. CONCLUSION

To futher understand the nature polyplex treated cells in response to a specific duration, it would be suggested that the polyplex cells are treated in 3 hour increments from 3 hours to 24 hours.

VI. FIGURES

REFERENCES

- [1] J. Hagenauer, E. Offer, and L. Papke. Iterative decoding of binary block and convolutional codes. *IEEE Trans. Inform. Theory*, vol. 42, no. 2, pp. 429-445, Mar. 1996.
- [2] T. Mayer, H. Jenkac, and J. Hagenauer. Turbo base-station cooperation for intercell interference cancellation. *IEEE Int. Conf. Commun. (ICC)*, Istanbul, Turkey, pp. 356-361, June 2006.
- [3] J. G. Proakis. *Digital Communications*. McGraw-Hill Book Co., New York, USA, 3rd edition, 1995.

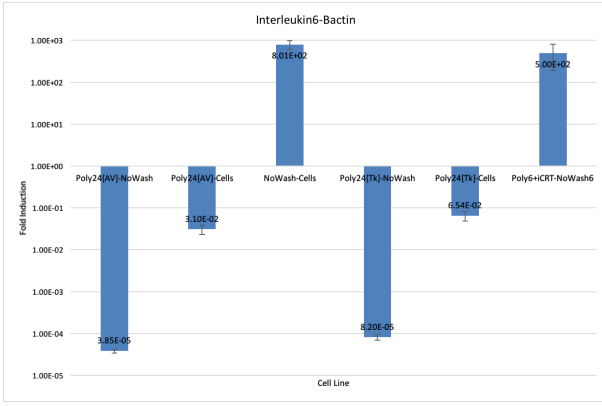


Fig. 2. Logarithmic fold Inductions for Interleukin-6 vs β -actin. From left to right, the comparisons are as follows: Polyplex treated cells for 24 hours version [AV] vs Cells that were not washed, Polyplex treated cells for 24 hours version [AV] vs Just cells, Cells that were not washed vs Just cells, Polyplex treated cells for 24 hours version [Tk] vs Cells that were not washed, Polyplex treated cells for 24 hours version [Tk] vs Just cells, and Polyplex treated cells for 6 hours with iCRT vs Cells that were not washed after 6 hours.

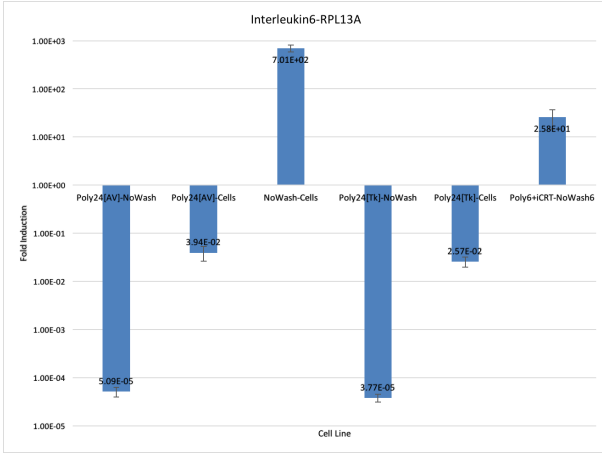


Fig. 3. Logarithmic fold Inductions for Interleukin-6 vs RPL13A. From left to right, the comparisons are as follows: Polyplex treated cells for 24 hours version [AV] vs Cells that were not washed, Polyplex treated cells for 24 hours version [AV] vs Just cells, Cells that were not washed vs Just cells, Polyplex treated cells for 24 hours version [Tk] vs Cells that were not washed, Polyplex treated cells for 24 hours version [Tk] vs Just cells, and Polyplex treated cells for 6 hours with iCRT vs Cells that were not washed after 6 hours.

- [4] F. R. Kschischang. Giving a talk: Guidelines for the Preparation and Presentation of Technical Seminars. <http://www.comm.toronto.edu/frank/guide/guide.pdf>.
- [5] IEEE Transactions \LaTeX and Microsoft Word Style Files. <http://www.ieee.org/web/publications/authors/transjnl/index.html>

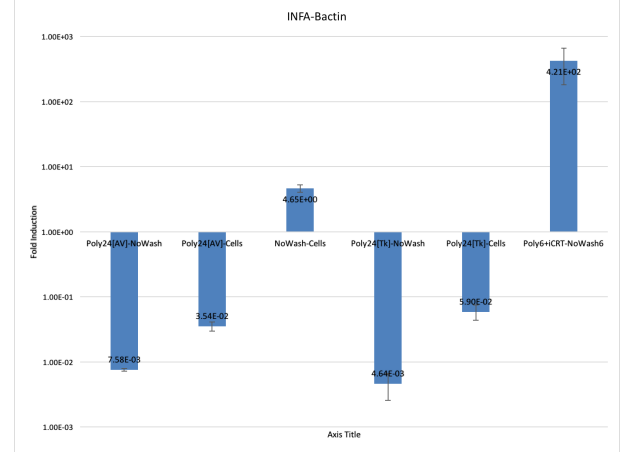


Fig. 4. Logarithmic fold Inductions for Interferon- α vs β -actin. From left to right, the comparisons are as follows: Polyplex treated cells for 24 hours version [AV] vs Cells that were not washed, Polyplex treated cells for 24 hours version [AV] vs Just cells, Cells that were not washed vs Just cells, Polyplex treated cells for 24 hours version [Tk] vs Cells that were not washed, Polyplex treated cells for 24 hours version [Tk] vs Just cells, and Polyplex treated cells for 6 hours with iCRT vs Cells that were not washed after 6 hours.

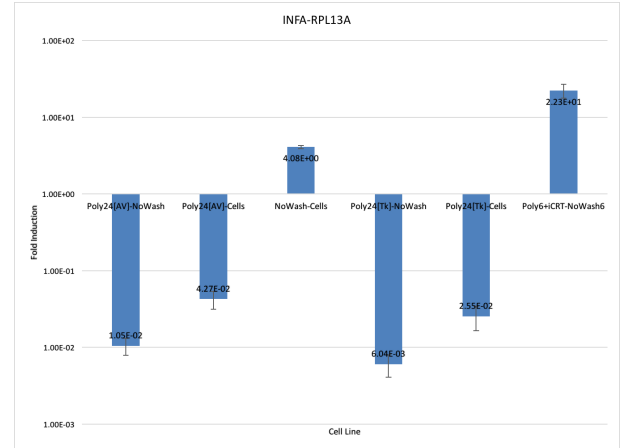


Fig. 5. Logarithmic fold Inductions for Interferon- α vs RPL13A. From left to right, the comparisons are as follows: Polyplex treated cells for 24 hours version [AV] vs Cells that were not washed, Polyplex treated cells for 24 hours version [AV] vs Just cells, Cells that were not washed vs Just cells, Polyplex treated cells for 24 hours version [Tk] vs Cells that were not washed, Polyplex treated cells for 24 hours version [Tk] vs Just cells, and Polyplex treated cells for 6 hours with iCRT vs Cells that were not washed after 6 hours.