

```
> library(PLIER)
```

## 1 Vaccination Dataset

Load data.

```
> data(bloodCellMarkersIRISDMP)
> data(svmMarkers)
> data(canonicalPathways)
> data(vacData)
```

Construct a joint pathway matrix by merging canonicalPathways, bloodCellMarkersIRISDMP and svmMarkers and select genes appearing in both gene expression profile and the joint pathway matrix.

```
> allPaths=combinePaths(bloodCellMarkersIRISDMP, svmMarkers,canonicalPathways)
> cm.genes=commonRows(allPaths, vacData)
```

Normalize the data and count the number of latent variables in the data by `num.pc()`. The result is 24. Then set `max.iter = 250`, `k = 24` and all other parameters to be default.

```
> vacDataN=rowNorm(vacData)
> num.pc(vacDataN[cm.genes,])
```

[illegible]

```
> plierResult=PLIER(vacDataN[cm.genes,], allPaths[cm.genes,],k=24, trace=T, max.iter=150)
```

```
[1] "L2 is set to 82.0573187891353"
[1] "L1 is set to 41.0286593945677"
[1] "L3 is set to 0.0111182399988254"
```

We correlate the decomposition result with SPVs from CellCODE. We have nice one-to-one correspondence, though the "DendriticCell" signature from CellCODE is more closely related to the Type-I interferon transcriptional response so it is probably not cell-type induced variation.

```
> data(SPVs)
> plotMat(cor(t(plierResult$B), SPVs))
```

### Visualize the cross-validation results

```
> plotMat(plierResult$Uauc)
```

Plot all of U and visualize the top genes

```
> plotU(plierResult, auc.cutoff = 0.5, pval.cutoff = 1)
> plotTopZ(plierResult, vacDataN, allPaths, top = 10)
```

The "PID<sub>ATF2</sub> PATHWAY" looks a little tenuous and we can check its statistics.

```
> plierResult$summary[which(plierResult$summary$`LV index`==3),]
\end{Sinput}
\begin{Soutput}
```

	pathway	LV index	AUC	p-value
5	PID_ATF2_PATHWAY	3	0.5718064	0.0536596

```
\end{Soutput}
```

```
\end{Schunk}
```

The association with "PID<sub>ATF2</sub> PATHWAY" is not significant: this pathway has only 42 genes t

```
\begin{Schunk}
```

```
\begin{Sinput}
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```
> plotTopZ(plierResult, vacDataN, allPaths, index=c(3), top=50)
```

```
\end{Sinput}
```

```
\end{Schunk}
```

```
\section{HCC Dataset}
```

Load data

```
\begin{Schunk}
```

```
\begin{Sinput}
```

```
> data(HCCdataTumor)
```

```
> data(canonicalPathways)
```

```
> data(chemgenPathways)
```

```
> data(oncogenicPathways)
```

```
\end{Sinput}
```

```
\end{Schunk}
```

Construct a joint pathway matrix by merging canonicalPathways, chemgenPathways and oncogenic

```
\begin{Schunk}
```

```
\begin{Sinput}
```

```
> CancerPath=combinePaths(canonicalPathways, chemgenPathways, oncogenicPathways)
```

```
> cmHCC=commonRows(HCCdataTumor, CancerPath)
```

```
>
```

```
\end{Sinput}
```

```
\end{Schunk}
```

Remove small pathways, not strictly necessary but saves computation time by making the pathw

```
\begin{Schunk}
```

```
\begin{Sinput}
```

```
> ii=which(colSums(CancerPath[cmHCC,])<20)
```

```
> HCCpath=CancerPath[, -ii]
```

```
\end{Sinput}
```

```
\end{Schunk}
```

Prescale the data

```
\begin{Schunk}
```

```

\begin{Sinput}
> HCCdataN=rowNorm(HCCdataTumor[cmHCC,])
\end{Sinput}
\end{Schunk}
Precompute Chat, which is used to define active pathways and is expensive for large pathway
\begin{Schunk}
\begin{Sinput}
> HCCchat=computeChat(CancerPath[cmHCC,])
\end{Sinput}
\end{Schunk}
Compute the number of latent variables by num.pc(HCCdataUse) and the result is 52. Then set
\begin{Schunk}
\begin{Sinput}
> plierResultHCC=PLIER(HCCdataN, CancerPath[cmHCC,], k = 52, Chat = HCCchat, trace=T)
\end{Sinput}
\begin{Soutput}
[1] "L2 is set to 132.670764279278"
[1] "L1 is set to 66.3353821396389"
[1] "L3 is set to 0.00807273868235351"
\end{Soutput}
\end{Schunk}
Plot the result with a high AUC cutoff so it is not too busy
\begin{Schunk}
\begin{Sinput}
> plotU(plierResultHCC, auc.cutoff = 0.9)
\end{Sinput}
\end{Schunk}
We found two immune components, interferon alpha and genes related to interferon gamma/CD8/T
\begin{Schunk}
\begin{Sinput}
> plotTopZ(plierResultHCC, HCCdataN, CancerPath, index=c(26, 40), top = 20)
\end{Sinput}

```