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Determinants of Nile tilapia's (*Oreochromis niloticus*) growth in aquaculture pond in Batu, Indonesia

EVELLIN DEWI LUSIANA^{1,2,*}, MUHAMMAD MUSA^{1,2}, SYAHRIL RAMADHAN¹

¹Department of Aquatic Resource Management, Faculty of Fisheries and Marine Science, University of Brawijaya. Jl. Veteran, Malang 65145, East Java, Indonesia. Tel./fax.: +62-341-553512, *email: evellinlusiana@ub.ac.id

²AquaRES Research Group, Faculty of Fisheries and Marine Science, University of Brawijaya. Jl. Veteran, Malang 65145, East Java, Indonesia

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Abstract. Lusiana ED, Musa M, Ramadhan S. 2021. Determinants of Nile tilapia's (*Oreochromis niloticus*) growth in aquaculture pond in Batu, Indonesia. *Biodiversitas* 22: 999-1005. Nile tilapia (*Oreochromis niloticus*) has been commonly cultured all over the world, especially in Indonesia, due to high market demand. The success of Nile tilapia culture production depends on the fish growth during cultivation as well as pond environment. Many studies have revealed that quality of water strongly affects fish growth in an aquaculture pond. This study aims to investigate the relationship among water quality like Physico-chemical (temperature, transparency, pH, dissolved oxygen, carbon dioxide, nitrate, and phosphate) and biological (phytoplankton abundance) factors and the specific growth rate of Nile tilapia in a freshwater cultivation pond in Batu City, Indonesia. The data was analyzed using the path model. Results indicated that the most significant factor for fish growth was the phytoplankton abundance. However, since nitrate and phosphate also played roles as determinants of phytoplankton abundance, they can be considered indirect water quality factors of Nile tilapia growth.

Keywords: Nitrate, path model, phosphate, phytoplankton abundance, specific growth rate

INTRODUCTION

Nile tilapia (*Oreochromis niloticus*) has been widely cultured all over the world in response to high market demand; it is easy to breed, has the ability to receive additional feed well, and possesses a high survival rate and economic value (KKP 2010). However, most of the farmers involved in tilapia production, especially those from developing countries, still lack critical information about the cultural process. This in turn limits their ability to optimize production levels (El-Sayed 2006). Such optimization requires an understanding of the various changes in environmental factors that can impact managerial practices important to maximizing fish yield (Bajaj 2017).

There are many variables that determine the quality of a water body, such as temperature, dissolved oxygen, turbidity, transparency, carbon dioxide, pH, alkalinity, nitrate and phosphate levels, primary productivity, biological oxygen demand (BOD), and plankton population (Bhatnagar and Devi 2013; Makori et al. 2017). Generally, factors influencing the growth of fish in aquaculture ponds can be categorized into two groups: abiotic and biotic. While light, water temperature, and chemical constitution are considered to be the most important factors in the first group, the second group consists solely of the single factor of food availability for fish, which would appear to have the greatest significance (Wingfield 1940).

Phytoplankton is one of the food sources for fish in aquaculture ponds. Because, it has the ability to perform photosynthesis, and plays an important role as a natural feed or primary producer for other organisms in the aquatic ecosystems (Vrede et al. 2009). The abundance of

phytoplankton is highly associated with fish production in aquaculture activities, and it is influenced by environmental factors, especially the existence of nutrients in the form of nitrate and phosphate in water (Donald et al. 2013; Lusiana et al. 2019).

One of the production centers of Nile tilapia in Indonesia is found in Freshwater Cultivation, located in Batu city. There, the aquaculture ponds are supplied with water from the Brantas river basin (Lusiana et al. 2018). However, the Brantas river has suffered from water pollution as the result of many human activities that exist along with the river flow (Suroso et al. 2007; Yetti 2011). Domestic and industrial activities have created wastes, and various substances have been discharged into the water, which in turn can affect water quality, both physicochemically and biologically (Syandri et al. 2016). Therefore, further investigation of the influence of water quality on fish growth in ponds is needed. The purpose of this study is to identify the main environmental factors that determine the growth of tilapia fish in the aquaculture ponds in Batu. Many studies stated that environmental factors have direct impact on fish growth (Bhatnagar and Devi 2013; Makori et al. 2017). Thus, the relationship can be simply analyzed using linear regression and correlation approaches (Dauda and Akinwole 2014; Aho et al. 2018). However, phytoplankton, as the biotic factor in the water environment, are also strongly affected by other abiotic factors. Hence, a simple model (regression and correlation) is not sufficient to determine the main water quality factor influencing fish growth. We thus consider a path model to accommodate the complexity of relationships between water quality factors and fish growth. Moreover, this analysis enables to

choose the most contributing factor to the growth of fish. Such information is important for the fish farmer to decide prioritization in water quality management.

MATERIALS AND METHODS

Study area

The study was conducted from February to April 2018 in the freshwater cultivation area in UPBAT (*Unit Pengelola Budidaya Air Tawar*) Punten, Batu City, East Java Province, Indonesia (Figure 1). It is a freshwater cultivation management unit under marine and fisheries service of East Java Province. The cultivation area receives water supply from Brantas River.

Collection and analysis of samples

In this study, water quality factors that comprised both physicochemical (temperature, transparency, pH, dissolved oxygen, carbon dioxide, nitrate, and phosphate) and biological (phytoplankton abundance) variables were considered. Fish growth was measured in weight (gm). Temperature was measured in-situ using thermometer, while transparency using secchi disk. Furthermore, pH paper was used in-situ to obtain the level of pH and DO meter Lutron DO-5510 for DO of the cultivated pond waters. The rest parameters were observed ex-situ at laboratory. Nitrate and phosphate were measured using spectrophotometer UV-Vis Spectroquant Pharo 300. The sampling procedure was conducted every two weeks with five replication during near midday (Morsy 2011) after the fish given feed. According to National Standardization Agency or BSN, the frequency of feeding was three times a day as much as 3% of body weight (Sari et al. 2017).

Specific growth rate

We randomly collected 15 fish at each sampling period by using scoope net (Riswandha et al. 2015). The specific growth rate of the fish can be calculated by applying the formula (Hopkins 1992)

$$SGR = \frac{\ln W_t - \ln W_0}{t} \cdot 100\%$$

Where, SGR = specific growth rate; W_t = final weight (gram); W_0 = initial weight (gram); t = duration of the observation (day).

Path analysis

Path analysis is the extension of linear regression into a model comprising multiple relationships in order to form a simultaneous equation system (Sarwono 2011). The analysis can only be used for variables with a minimum interval scale and that fulfill normality and linearity assumptions. Before performing path analysis, a path diagram must be built that illustrates the overall relationship of the analyzed variables, based on theoretical justification (Hair et al. 2019). Moreover, the data should be standardized based on normal standardization method (Lusiana and Mahmudi 2020). The steps in path analysis are, (i) building a path diagram, (ii) performing a test of normality (Kolmogorov-Smirnov test) and linearity assumption, (iii) performing a test of significance, (iv) applying trimming theory, and (v) identifying the most influential predictors to the response (Kurniawan et al. 2016). The initial construction of the path diagram in this study is depicted in Figure 2.

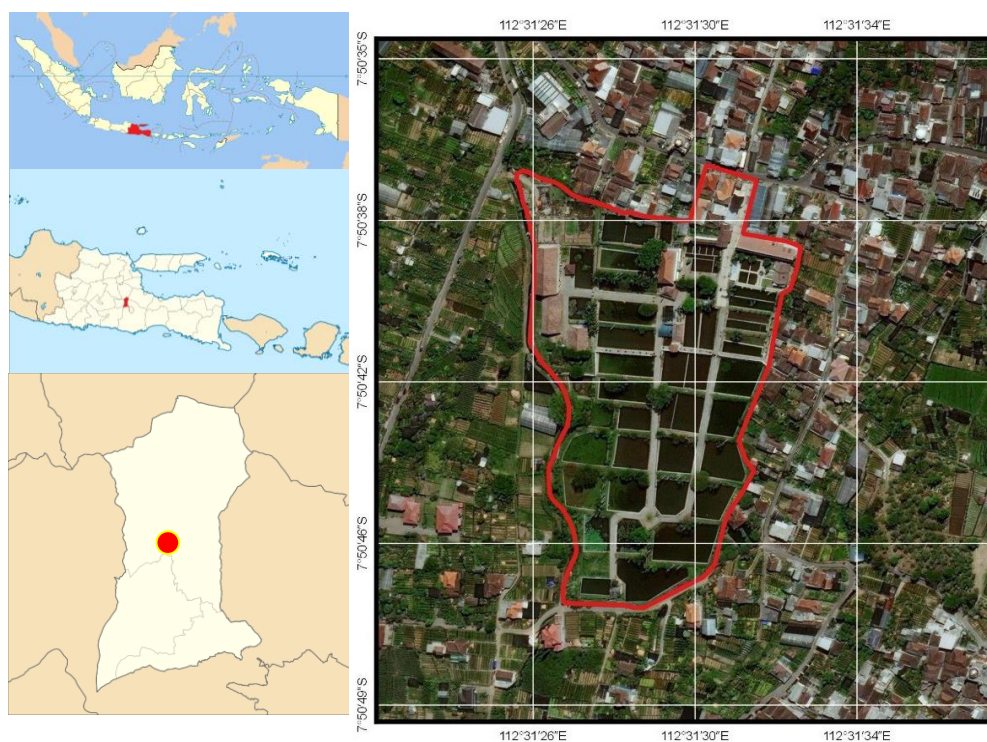


Figure 1. Study area location showing the aquaculture ponds in UPBAT Punten, Batu City, Indonesia (●)

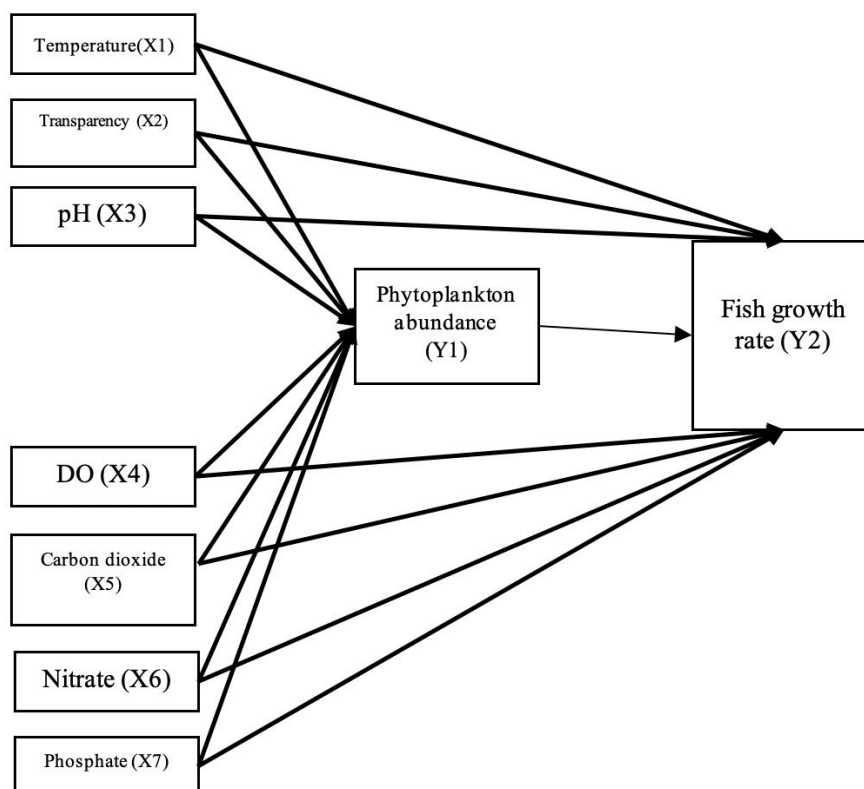


Figure 2. Complete path diagram for initial analysis for this study

RESULTS AND DISCUSSION

Physico-chemical parameters of water

Water temperature was recorded to be around 24 to 25°C during the study period (Table 1). This is classified as the optimum heat for tilapia culture, since the ideal range is 24-32°C (El-Sayed and Kawanna 2008). Temperatures that extend over the boundaries of the range are most probably used to maintain internal physiology rather than growth. Transparency was observed to be between 26.3 cm and 34.3 cm, which is desirable for aquaculture production (Bhatnagar and Devi 2013), and transparency of around 30-60 cm indicates optimum productivity of the pond (Singh and Ngachan 2007). Therefore, it can be said that the water clarity in the study site is optimum for tilapia growth. Notably, measuring transparency is an alternate step for roughly estimating the trophic state of a water body (Pavluk and bij de Vaate 2008). In addition, a high content of fine sediment and organic particles will decrease the water transparency and affect the primary production of the ecosystem (Peck Yen and Rohasliney 2013).

As measured, pH was shown to be under 7; thus, the water was categorized as acidic. A suitable pH for fish culture is 6.7 and 9.5, and the ideal level for fish growth is 7.5 and 8.5 (Singh and Ngachan 2007). On the other hand, the concentration of DO in this study was observed to be around 6 mg/L. As stated in a previous study, adequate DO for a fish pond is greater than 6 mg/L (Banerjee 1967); hence the pond in this study has a suitable DO rate for good fish growth.

Carbon dioxide (CO₂) concentration varied from 7.26 to 12.11 mg/L, which is not favorable for fish growth, since the desirable limit for carbon dioxide in aquaculture is lower than 10 mg/L (Ekubo and Abowei 2011). The increase of CO₂ in aquatic ecosystem did not affect the primary productivity, but it may shift the composition of phytoplankton to be dominated by harmful algae (Keys et al. 2018). The other important factor of water quality is the availability of nutrients, represented by nitrate and phosphate concentrations. Both the compounds in this study were favorable for fish culture with the good ranges of nitrate and phosphate from 0.1 to 4.0 mg/L and 0.05 to 0.07 mg/L, respectively (Singh and Ngachan 2007; Bhatnagar and Devi 2013).

Abundance and community structure of phytoplankton

Total phytoplankton abundance in the Nile tilapia pond in Batu varied from around 8000 cells/mL to more than 10000 cells/mL (Figure 3). The largest biomass was seen in the first and ninth weeks. Four divisions of phytoplankton within this abundance were identified: Chrysophyta, Chlorophyta, Cyanophyta, and Euglenophyta. Chlorophyta was the most dominant division, accounting for more than 40% of total algae, with a total of 12 genera (*Cladophora*, *Chlorella*, *Scenedesmus*, *Spirogyra*, *Platymonas*, *Schroderia*, *Asterococcus*, *Chlorococcum*, *Pediastrum*, and *Straurastum*). The abundance of Chrysophyta was ranging from 2678 to 3224 cell/mL and consisted of seven genera (*Coscinodiscus*, *Cyclotella*, *Cymbella*, *Synedra*, *Gyrosigma*, *Nitzschia*, and *Navicula*). Cyanophyta and Euglenophyta were the least

abundant divisions, comprising five (*Lyngbya*, *Chorococcus*, *Oscillatoria*, *Phormidium*, and *Coelosphaerium*) and two (*Trachelomonas* and *Euglena*) genera, respectively.

These results were inlined with the previous studies which suggested that Nile tilapia fish is effective in filtering Chlorophyta and Cyanophyta (cyanobacteria) from any water sources (Mohamed et al. 2019). It is also reported that the fish can feed on all types of Cyanophyta without any exception for toxic species (Xie et al. 2001). In addition, the long-term exposure of Nile tilapia to these species could accumulate the toxin in the fish tissue and potentially transfer it to upper trophic organisms (Mohamed et al. 2019). Therefore, the control of phytoplankton blooms especially Cyanophyta needs to be performed in the fish pond (Xie et al. 2001).

Specific growth rate of Nile tilapia

The specific growth rate (SGR) of Nile tilapia in the aquaculture pond displayed a fluctuation trend (Figure 4). The highest rate (4.51%) appeared in the first week, followed by a constant decrease until reaching the bottom at the seventh week (1.40%). After this, the SGR increased by 0.75 %. The dynamics in phytoplankton abundance over time as shown in Figure 3 have similar trend to SGR

fluctuation in Figure 4. The maximum rate of SGR occurred along with the highest biomass of phytoplankton in the first week. Identically, the small number of phytoplankton in the seventh week seemed to be associated with the low SGR of tilapia. Hence, descriptively, those results depicted a tight relationship between phytoplankton abundance and growth rate.

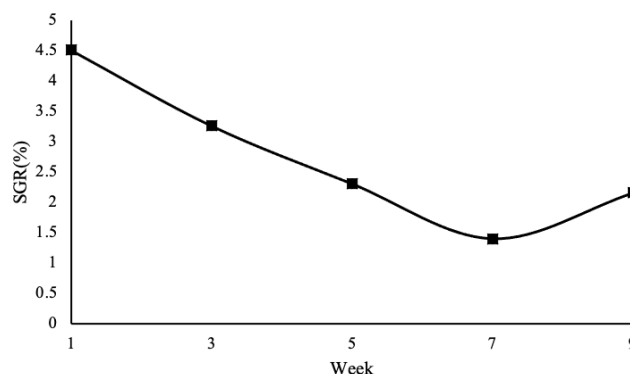


Figure 4. The SGR of Nile tilapia at aquaculture pond in Batu city, Indonesia

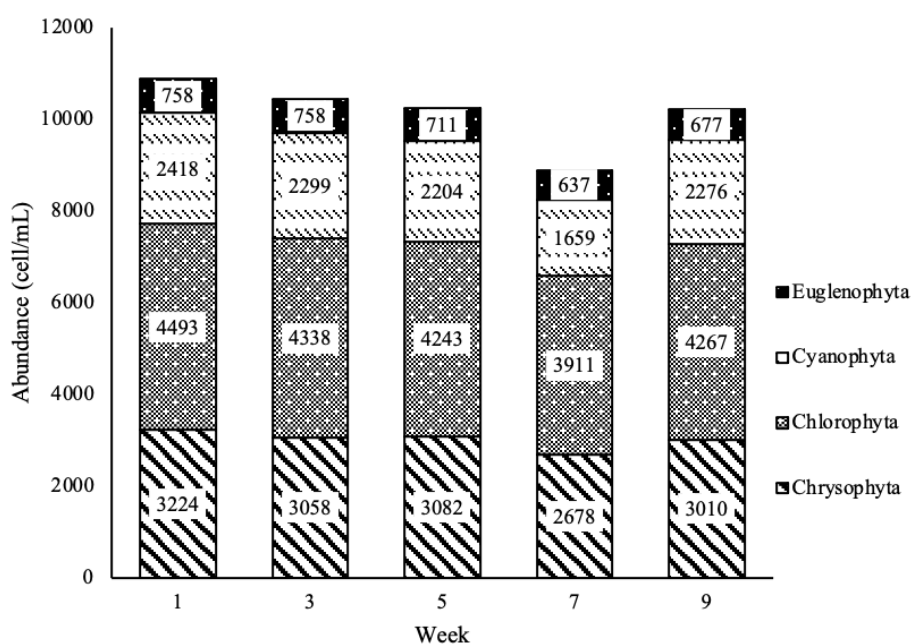


Figure 3. Phytoplankton abundance and composition during the research

Table 1. Physico-chemical parameters of water in Nile tilapia culture pond in Batu city, Indonesia

Variables	Week				
	1	3	5	7	9
Temperature ($^{\circ}\text{C}$)	23.8 \pm 0.764	24.2 \pm 0.764	23.7 \pm 0.577	24 \pm 1.000	24.5 \pm 0.5
Transparency (cm)	29.2 \pm 1.607	30.8 \pm 0.764	32.2 \pm 2.466	26.3 \pm 0.577	34.3 \pm 3.512
pH	6.33 \pm 0.058	6.63 \pm 0.115	6.77 \pm 0.115	6.5 \pm 0.000	6.63 \pm 0.115
DO (mg/L)	6.17 \pm 0.116	6.56 \pm 0.307	6.57 \pm 0.26	5.98 \pm 0.502	6.81 \pm 0.165
CO ₂ (mg/L)	9.08 \pm 1.820	9.69 \pm 2.102	10.29 \pm 2.096	7.26 \pm 1.815	12.11 \pm 1.045
Nitrate (mg/L)	0.787 \pm 0.091	0.853 \pm 0.05	0.896 \pm 0.043	0.478 \pm 0.014	1.166 \pm 0.222
Phosphate (mg/L)	0.023 \pm 0.001	0.024 \pm 0.001	0.025 \pm 0.001	0.013 \pm 0.001	0.032 \pm 0.001

Determinant of growth rate: path analysis results

There are two assumptions underlying path analysis: normality and linearity. The normality assumption should be fulfilled by the response variables. This study engaged with two response variables: phytoplankton abundance (Y1) and fish growth (Y2). The Kolmogorov-Smirnov test showed that both the response variables were normally distributed ($p > 0.05$). Meanwhile, not all variable relationships in the path model have a linear association; only the relationship between transparency, nitrate, and phosphate to phytoplankton abundance, and phytoplankton abundance to fish growth complete the assumption. Consequently, those relationships that were not linear were deleted from the path diagram. Hence, a modified path diagram is shown in Figure 5A, along with the parameter estimation. As the effect of transparency is non-significant ($p < 0.05$), it was trimmed from the path model, thus creating the final path diagram in this study, as shown in Figure 5B.

The concentrations of nitrate and phosphate have a significant ($p < 0.05$) positive effect on phytoplankton abundance (Figures 5A and 5B). Generally, nutrient availability (nitrate and phosphate) is related to expanded biomass, and can prompt changes in the phytoplankton community. The abundance of phytoplankton frequently rises in number yet decreases in quality. Even when phytoplankton cells increase, remarkably, the quantity of species does not increase, which shows that there is a transcendence of specific species (Sidabutar et al. 2016; Wisha et al. 2018). On the other hand, Figure 5A showed that the influence of transparency on phytoplankton abundance was negative. The transparency of surface waters is often related to the amount of plant nutrients in the water. The greater the number of nutrients, the greater

the number of phytoplankton, and, as a result, the less transparent water. In other words, as water transparency increases, algal abundance rapidly declines (Sternier 2009). However, unfortunately, it turned out that this relationship was insignificant ($p < 0.05$) in this study and was deleted from the final model. In addition, only phytoplankton abundance proved to have a direct significant effect on fish growth rate ($p < 0.05$).

In final path model, nitrate was the most dominant predictor of phytoplankton abundance in this study, since its total effect is greater than that of phosphate (Table 2). Meanwhile, the greatest determinant for fish growth is phytoplankton abundance. There are several reasons why phytoplankton is important for aquaculture. They are the foundational component of the natural web food and also used as a supplement for manufactured feed. This is especially significant for small, post-larval crustaceans and small fish soon after stocking; at noontime, these algae produce oxygen through photosynthesis at a faster rate than oxygen diffused from the atmosphere in the culture pond. Lastly, phytoplankton creates turbidity that restricts sunlight penetration to the bottom of the pond, and its bloom is a reliable monitor for aquatic macrophytes that grow under the water surface (Boyd 2016).

In conclusion, Nile tilapia's growth in an aquaculture pond is highly determined by environmental factors, both abiotic and biotic. This study showed that abiotic factors (nitrate and phosphate) have an indirect effect on the SGR of the fish, which was mediated by the biotic factor (phytoplankton abundance). This factor also became the greatest determinant of tilapia growth in the aquaculture pond in Batu. In addition, phytoplankton biomass itself was more strongly affected by nitrate concentration in the water body than by phosphate concentration.

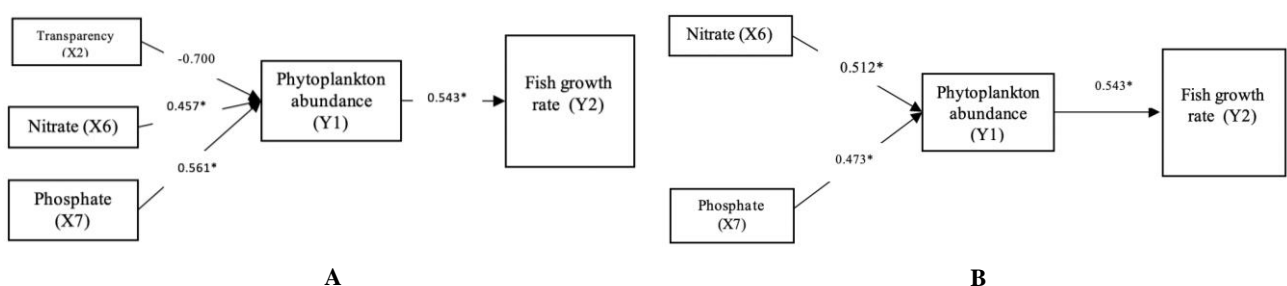


Figure 5. A. Modified path diagram after assumption test; B. final path diagram

Table 2. Direct, indirect effect, and total effect of final path model

Predictor	Response					
	Phytoplankton abundance			Fish growth rate		
	Direct effect	Indirect effect	Total effect	Direct effect	Indirect effect	Total effect
Nitrate	0.512	-	0.512	-	0.278	0.278
Phosphate	0.473	-	0.473	-	0.257	0.257
Phytoplankton abundance	-	-	-	0.543	-	0.543

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