REVIEW



Endothelin-1 axes in the framework of predictive, preventive and personalised (3P) medicine

Adriana Torres Crigna¹ · Barbara Link¹ · Marek Samec² · Frank A. Giordano¹ · Peter Kubatka³ · Olga Golubnitschaja⁴

Received: 5 June 2021 / Accepted: 11 June 2021 / Published online: 4 August 2021 © The Author(s) 2021

Abstract

Endothelin-1 (ET-1) is involved in the regulation of a myriad of processes highly relevant for physical and mental well-being; female and male health; in the modulation of senses, pain, stress reactions and drug sensitivity as well as healing processes, amongst others. Shifted ET-1 homeostasis may influence and predict the development and progression of suboptimal health conditions, metabolic impairments with cascading complications, ageing and related pathologies, cardiovascular diseases, neurodegenerative pathologies, aggressive malignancies, modulating, therefore, individual outcomes of both non-communicable and infectious diseases such as COVID-19. This article provides an in-depth analysis of the involvement of ET-1 and related regulatory pathways in physiological and pathophysiological processes and estimates its capacity as

- a predictor of ageing and related pathologies,
- a sensor of lifestyle quality and progression of suboptimal health conditions to diseases for their targeted prevention
- and as a potent target for cost-effective treatments tailored to the person.

 $\label{lem:keywords} \begin{tabular}{ll} Keywords & Predictive preventive personalised medicine (PPPM/3PM) \cdot Endothelin \cdot ET-1 \cdot Endothelin axis \cdot \\ & Vasoconstriction \cdot Vasodilation \cdot Vasospasm \cdot Nitric oxide \cdot Stroke \cdot Female and male health \cdot Mental health \cdot \\ & Lifestyle \cdot Stress reaction \cdot Sense regulation \cdot Wound healing \cdot Pregnancy \cdot Embryonic development \cdot Pain sensitivity \cdot \\ & Drug sensitivity \cdot Treatment target \cdot Suboptimal health \cdot Ageing \cdot Metabolic impairments \cdot Cardiovascular disease \cdot \\ & Neurodegeneration \cdot Cancer \cdot COVID-19 \cdot Individual outcomes \\ \end{tabular}$

Abbreviations		AD	Alzheimer's disease
5-HAT	Serotonin	AMD	Age-related macular degeneration
ACS	Acute coronary syndrome (ACS)	AMI	Acute myocardial infarction
		AngII	Angiotensin II
Olga Golubnitschaja olga.golubnitschaja@ukbonn.de Department of Radiation Oncology, University Hospital Bonn, Rheinische Friedrich-Wilhelms-Universität Bonn, Bonn, Germany		BP	Blood pressure
		CHF	Congestive heart failure
		CNS	Central nervous system
		COX	Cyclooxygenase
		CRAE	Combined resistance and aerobic exercise
Clinic of Obstetrics and Gynecology, Jessenius Faculty of Medicine, Comenius University in Bratislava, 036 01 Martin, Slovakia		CVD CVS	Cardiovascular disease Cerebrovascular
Department of Medical Biology, Jessenius Faculty of Medicine, Comenius University in Bratislava,		DOCA	Deoxycorticosterone acetate
		E2	Oestradiol
036 01 Ma	rtin, Slovakia	EAH	Essential arterial hypertension
Predictive, Preventive and Personalised (3P) Medicine, Department of Radiation Oncology, University Hospital Bonn, Rheinische Friedrich-Wilhelms-Universität Bonn, Bonn, Germany		EC	Endothelial cells
		ECE	Endothelin-converting enzyme
		EDD	Endothelium-dependent dilation



edn1 Gene encoding ET-1 ERAs Endothelin receptor antagonists

ET-1 Endothelin-1

ET-2 Endothelin-2
ET-3 Endothelin-3

ETA Endothelin A receptor ETB Endothelin B receptor

HELLP syndrome Haemolysis, elevated liver enzymes

and low platelets syndrome

i.c.v Intracerebroventricularly

IL Interleukin LV Left ventricle

MABs Monoclonal antibodies MMP Metalloproteinase MMP-2 Metalloproteinase-2

NO Nitric oxide
NOS NO synthase
OC Ovarian cancer
OM Olfactory mucosa

ORF Oligomerisation of open reading

frame

PAH Pulmonary arterial hypertension

PDE Phosphodiesterase

PDGF Platelet-derived growth factor

PE Preeclampsia
PG Prostaglandin
PGE-2 Prostaglandin E2
PG12 Prostacyclin

PH Paedriatic hypertension

PHACTR1 Phosphatase and actin regulator 1

PPH Paediatric pre-hypertension

Prepro-ET-2 Peproendothelin-2
Prepro-ET-3 Peproendothelin-3
RA63 Rendomab-A63
RB1 Rendomab-B1
RB4 Rendomab-B4

ROS Reactive oxygen species SAH Subarachnoid haemorrhage

SARS-CoV-2 Severe acute respiratory syndrome

coronavirus 2

SBP Systolic blood pressure SCD Sickle cell Disease

 $\begin{array}{ll} SNP & Single-nucleotide polymorphism \\ TGF-\beta & Transforming growth factor-beta \\ TNF-\alpha & Tumour necrosis factor-alpha \end{array}$

TS Takotsubo syndrome

VCAM-1 Vascular adhesion molecule-1 VSMCs Vascular smooth muscle cells VSMCs Vascular smooth muscle cells vWF Von Willebrand factor (vWF)

WH Wound healing
WKY rats Wistar–Kyoto rats
YAP Yes-associated protein



Preamble

Endothelin (ET) is involved in the regulation and performance of a myriad of processes, which physiologically occur in a healthy human body. To them belong:

- Maintaining versus diminishing physical well-being [1,
 2]
- Regulation of stress reactions and mental health: chronic as well as episodic psychosocial factors provoke social, environmental and emotional stress reactions; elevated plasma endothelin-1 (ET-1) influences individual differences in autonomic and hemodynamic responses to stress [3, 4]
- Female and male health [5–8]
- Pregnancy and embryonic development [9–12]
- Wound healing [13–16]
- Regulation of senses [17–20]

amongst others.

On the other hand, shifted ET homeostasis may influence and predict the development and progression of:

- Suboptimal health conditions [1, 18–24]
- Ageing and related pathologies [5, 25–28]
- Vascular stiffness and ageing, cardiovascular diseases and "young" ischemic stroke [1, 29–35]
- Metabolic impairments with cascading complications
 [1, 36–38]
- Neurodegenerative disorders [39–42]
- Particularly aggressive subtypes of cancer such as metastasing breast and prostate malignancies [7, 20, 43-50]
- ET-1 axes are involved in thermoregulation and attenuate the heat loss, modulate pain and drug sensitivity [20, 51–55], therapy response [56–58] and individual outcomes of both non-communicable [59] and infectious diseases, such as during the current COVID-19 pandemic [60–63].

This article provides an in-depth analysis of the involvement of ET-1 and related regulatory pathways in physiological and pathophysiological processes and estimates its potential as the diagnostic, prognostic and treatment target in the framework of 3P medicine.

Historical notes

Endothelins were first described in 1985 by Hickey et al. as factors with vasoconstrictor actions acquired from a culture of bovine aortic endothelial cells [64]. They were suggested to have a chemical composition similar to peptides,

due to the abolishing activity of trypsin [65]. Thereafter, Yanagisawa et al. defined the structure of these constricting factors as a 21-amino acid peptide named endothelin (designated endothelin-1 or ET-1), from a porcine aortic endothelial cell culture [66].

Shortly after, a similar peptide family named sarafotoxins was discovered in the cardiotonic venom of snakes and was seen to have a similar sequence to ET-1 [67, 68]. In humans, two further endogenous isoforms of endothelins were described while analysing the gene encoding ET-1 (edn1): endothelin-2 (ET-2) and endothelin-3 (ET-3). All isoforms consist of 21 amino acid residues, yet possess differential expression subject to their tissue or cell of origin [69]. In fact, due to differences in affinities for the three isoforms, in the upcoming year, two G protein-coupled receptors were identified, namely endothelin A receptor (ETA) [70] and endothelin B receptor (ETB) [71], where ET-1 and ET-2 signal through ETA in a more potent manner than ET-3, while all three isoforms are equally efficient in receptor ETB [72]. In fact, endothelins are secreted by numerous different cells, such as endothelial cells (EC), vascular smooth muscle cells (VSMCs), fibroblasts, renal medulla, leukocytes and macrophages [72]. The objective of this review is to summarise endothelin mediation in normal physiology and focus on its role in the pathogenesis of a number of affections and diseases.

Endothelin-1 axes: function, physiology and measurement

After the discovery of ET-1, permanently increasing interest for its potent and sustained vasoconstrictor action in the pharmaceutical industry and academia sectors has been monitored. Endothelin isoforms are synthesised by respective cell types and tissues in the human being and encoded by responsible genes located in different chromosomes (ET-1, chromosome 6 and ET-2 and ET-3, chromosomes 1 and 20 respectively) [69]. ET-1, the main and parent component of the endothelin family, is secreted by most cell types with an increased expression in vascular EC, smooth muscle cells, cardiac myocytes fibroblasts, podocytes, macrophages and fibroblasts [73–75]. ET-1 has been described to be a multifunctional peptide involved in many physiological (cell differentiation and growth), pathological processes (cancer development and inflammatory events) and cell functions [76]. It plays a crucial role in pulmonary physiology, autoimmune disorders, neurological function and fluid and electrolyte transport [66, 69, 77, 78].

Synthesis of the ET-1 bioactive 21-aa peptide is carried out in multiple stages by the encoding gene edn1 via a proteolytic pathway [79]. Human gene edn1 transcription generates an mRNA that consequently encodes a 212-aa

prepro-ET-1 [66]. Once prepro-ET-1 enters the endoplasmic reticulum furin-like peptidase split prepro-ET-1 into a 38-aa inactive peptide intermediate named big ET-1 [80]. Lastly, big ET-1 is converted by the endothelin-converting enzyme (ECE) to form the active form of ET-1 [81, 82]. ET-2 and ET-3 are also constituted from their inactive Big ET by ECE. Various ECE human isoforms have been described at different subcellular locations, ECE-1a, ECE-1b, ECE-1c and ECE-1d [83]. ET-1 plasma and circulating typical ET levels in numerous species is ~ 1 pM suggesting that upon standard physiological conditions, endothelins are not circulating hormones but truly operate as paracrine and autocrine factors [84, 85].

267

ET-1 bioavailability regulation is described to happen mainly at a transcriptional level [86–88]. Proper reactivity of edn1 to stimuli is required for a correct regulation of ET-1 expression within the systems of the human body. In fact, modifications in edn1 expression or genetic polymorphisms have described alteration and pathogenesis of numerous diseases such as diabetic retinopathy, cancer, heart failure, cardiomyopathy or asthma [89–93]. ET elevated circulating levels are often related to cardiovascular diseases (CVD) such as ischaemic heart disease, hypertension, chronic heart failure, ischaemic heart disease or pulmonary hypertension. Nevertheless, increased ET levels have also been reported in non-CVD [85]. An enhancement of ET-1 mRNA expression has been reported in numerous cells by transforming growth factor-beta (TGF-β), interleukins, insulin, angiotensin II (AngII), tumour necrosis factor-alpha (TNF- α) and norepinephrine [66, 94, 95]. In EC, an upregulation of ET-1 mRNA is exerted by hypocapnia and downregulated by hypoxia [96].

As aforementioned, ET isoforms and ET receptors are distributed within a spectrum of tissues and by a variety of cells; hence, their interaction induces signal cascades promoting multifunctional outcomes. In fact, endotheliumreleased ET-1 (mainly secreted to the basal face) interacts via ETA with the smooth muscle, prompting vasoconstriction. In contrast, ET-1 coupling with ETB receptor regulates the secretion of relaxing factors such as prostacyclin (PGl2) and nitric oxide (NO) on neighbouring EC, inducing relaxation of smooth muscles [85]. Data suggests that endogenic ET-1 regulates peripheral blood flow via smooth muscle ETA, while diminution in blood pressure (BP) is regulated by endothelial cell ETB receptor [85, 97–100]. ET-1-ETA system has been described to be of importance in the cardiac and cranial neural crest, whereas ET-3-ETB in enteric neuron and melanocyte development [101, 102].



Characterisation of ET subtypes

ET-2 differs from ET-1 in humans and other mammals namely cats, dogs, cattle and monkeys by two amino acids, Trp⁶ and Leu⁷. Synthesis of ET-2 has been described to be comparable to that of ET-1. Edn2 gene is transcripted into peproendothelin-2 (Prepro-ET-2) which is then cleaved by a furin into Big ET-2. Then, ECE-1 and ECE-2 convert Big ET-2 into mature peptide ET-2 [82]. Gardiner et al. reported in an in vivo study how Big ET-2 was transformed to exert cardiovascular effects similarly to Big ET-1 [103]. However, there is still some controversy regarding the efficiency of conversion of Big ET-2 by ECE, despite sharing an identical cleavable bond with ET-1; Big ET-2 conversion rate by ECE-1 and ECE-2 was respectively 5-7 and 7–10% as rapid as Big ET-1 transition [82]. A quantitative RT-PCR study on rats revealed ET-1 mRNA expression in all 16 analysed organs, though only detected ET-2 expression in lung, ovary, heart, stomach and intestine, being ET-2 distribution more organ restricted [104]. Increased levels were reported in ovary and in all intestine regions (duodenum, jejunum, ileum, colon and rectum) [105, 106]. Medulla oblongata and pituitary glands presented higher ET-2 mRNA levels than ET-1, though most brain areas (cerebellum and cerebrum) had low or undetectable levels of ET-2 [107].

ET-2 is synthesised in spectra of human tissues: their mRNA and/or peptide have been identified in the heart, lung, kidney, vasculature, intestine and ovaries, promoting intestinal contraction, ovulation, thermoregulation and lung alveolarisation [108–112]. Big ET-2 presented greater levels (~ 2 pmol/l) than Big ET-1 in human plasma [113, 114]. Nevertheless, ET-2 concentration (~ 0.9 pmol/l) is lower than ET-1 levels [115]. Both in mice and rats, ET-2 differs from ET-1 by three amino acids (Asn⁴, Trp⁶ and Leu⁷) and is termed vasoactive intestinal contractor due to its original identification in contracting mice ileum [116].

ET-2 presents distinct physiological and pathophysiological properties than ET-1. In fact, removal of the ET-2 gene in mice translates into a phenotype with severe hypothermia, growth deceleration, hypoglycemia and ketonemia [117]. ET-2 gene selective deletion in epithelial cells prompted great changes in mice lung morphology leading to low blood oxygen and elevated carbon dioxide levels. ET-2 function is exerted in a paracrine manner, being ET-2 mRNA present in epithelial cells and receptor mRNA in the mesenchyme. Furthermore, ET-2 appears to have a significant role in ovarian physiology [117].

Many studies have emphasised an important role of ET-1 as a follicular development and luteal phase regulator [118–120]; however, ET-2 expression in ovaries is in fact higher than ET-1 [121]. Both ET-2 and ECE-1 were

transitorily expressed during ovulation in rat ovaries [122]. Palanisamy et al. reported ET-2 expression in the granulosa cells in pre-ovulatory follicles. After superovulation induction in mice, they observed a drastic enhancement of ET-2 mRNA expression 11 h after, coinciding with follicular breach [108].

ET-2 also acts as a chemokine, being a chemoattractant for neutrophils even at low levels [123]. Furthermore, ET-2 stimulated macrophage chemotaxis via ETB receptor through the MAPK pathway, displaying similar activity as CCL2 inflammatory chemokine [124].

ET-3 differs in 6 amino acids within the N-terminal loop when compared to ET-1 and ET-2, which confers ET-3 with selectivity for ETB receptor [125]. The ET-3 synthetic pathway is primarily similar to that of ET-1 and ET-2. The transcription product of the edn3 gene, peproendothelin-3 (Prepro-ET-3), is cleaved by the furin enzyme into the inactive Big ET-3. The capacity of ECE-1 and ECE-2 to be able to cleave Big ET-3 has been argued. Data report a reduced conversion rate of Big ET-3 (1–3 and 4–9% respectively) when compared to Big ET-1 [82]. Nevertheless, Big ET-3 induces vasoconstriction in vivo, suggesting the existence of cleaving enzymes [103, 126]. Indeed, a zinc-dependent endopeptidase with a similar ECE-1 sequence, Kell, effectively cleaved Big ET-3, proposing an alternative synthetic enzyme for ET-3 [127, 128].

ET-3 is expressed in melanocytes, intestinal epithelial cells, renal tubular epithelial cells, placenta, melanocytes and brain neurons promoting the secretion of anti-inflammatory and vasodilating agents (particularly PG12 and NO) [74, 129]. Furthermore, ET-3 has been detected in the heart, endometrium, brain and pituitary glands, where its levels were greater than ET-1 [130]. In human plasma, ET-3 was detectable at low ratios of ~ 0.3 pmol/l, as well as Big ET-3 at ~ 6 pmol/l [113]. Human EC are unable to synthesise ET-3, thereby inactive circulating Big ET-3 may be originated in the adrenal glands [131]. Elevated concentration of immunoreactive ET-3 has been found in the lungs, brain, intestine and pituitary gland of rats [132]. Prepro-ET-3 mRNA was detected in the submandibular gland, kidney, eye, stomach and spleen rat tissues [133], while Big ET-3 was seen in mast cells and macrophages within the gastrointestinal tract of rats [134]. Significant increases in Big ET-3 concentrations have been reported in haemodialysis patients (along with Big ET-1 and -2), even if their active peptide levels were moderately elevated [135]. Nevertheless, concentration changes related to disease have not been broadly researched.



Balanced release of ET-1 and nitric oxide is crucial for health protection

NO is produced in the arterial wall and is one of the most effective vasodilator molecules, which arbitrate endothelium-dependent relaxation [136]. NADPH-dependent NO synthase (NOS) generates NO through an enzymatic conversion of L-arginine to L-citrulline [137]. NOS3 constitutive expression promotes NO production by the endothelium. NO functionality is extensive, due to which it acts as the main endothelium-derived relaxing factor, in order to maintain vascular homeostasis [136]. Endothelial cellproduced factors, ET-1 and NO, exhibit opposing actions on smooth muscle cell contraction; however, when balanced, they regulate local vascular tone [136]. Studies have revealed that NO and ET-1 can be mutually regulated in order to reach vascular tone homeostasis. In fact, stimulation of NO production in EC exerted a reduction of ET-1 expression and secretion [138]. Another study showed ET-1 induction of eNOS uncoupling [139] and blockage of ETA receptor mends NO-dependent vascular function in mice with atherosclerosis [140].

Endothelial dysfunction is characterised by a transition of the endothelium to a pro-inflammatory and reduced vasodilation state, in which there is an alteration of vasoactive factor (NO, ET-1) equilibrium [141]. Several studies have suggested that decreased NO concentrations and elevated vasoconstrictive ET-1 and serotonin (5-HT) might have an impact on high BP onset. In fact, imbalanced expression of ET-1 versus NO, together with the impaired 5-HT release, has been reported in essential arterial hypertension (EAH) [142]. EAH pathogenesis is associated with the endothelium and constitutes a social burden [142]. Aflyatumova et al. studied the association between the plasma ET-1, serum NO, serum 5-HT and platelet 5-HT and BP in adolescent males, along with their use as preclinical biomarkers for endothelial dysfunction and EAH [142]. Plasma ET-1 and serum 5-HT concentrations were increased regarding the controls in both paediatric pre-hypertension (PPH) and hypertension (PH) children, although PH presented significantly higher levels. NO serum levels on the other hand were higher in PPH than in PH patients, correlating negatively with BP values [142]. Thus, results suggest that ET-1, NO and 5-HT may be related to BP in adolescents and could potentially function as preclinical biomarkers of EAH. These results are in line with another study which described increased ET-1 plasma concentrations in hypertensive adolescents when compared to healthy subjects, combined with correlating ET-1 levels with systolic blood pressure (SBP) levels [143].

Gender differences in ET-1 functionality

Numerous gender differences have been observed in the endothelin system; in fact, gonadal hormones play a crucial role in modulating gender-related disparities in the ET system [144]. Sex steroids are crucial for vascular homeostasis regulation [145]. In both experimental and human hypertension models, plasma ET-1 levels were significantly higher elevated in males than females [146–148]. Female plasma ET-1 levels oscillate throughout the menstrual cycle, being lowest during the luteal and follicular phase and higher in the menstrual phase [149]. Female sexual hormones inhibit ECE action along with ET-1 mRNA expression and diminish ETB receptor expression, thus restraining ET-1 levels [144]. In a bilateral ovariectomy study on female Sprague–Dawley rats, oestrogen replacement treatment decreased lung expression of ETA receptor and renal inner medullar expression of ETB receptor [150]. Another study with oestrogen administration in transexual male patients reported a decrease in ET-1 levels [146]. Furthermore, in pregnancy, ET-1 levels are also decreased, concordant with elevated oestrogen levels [151, 152]. The other significant female hormone, progesterone, was seen to inhibit in vitro ET-1 secretion both in resting and stimulated EC [153, 154].

Conversely, testosterone appears to increase ET-1 synthesis both in vitro and in vivo [144, 154, 155]. Testosterone also modulates vascular reactions to exogenous ET-1 [156]. However, a rat orchidectomy analysis revealed elevated ET-1, ETA and ETB receptors in the portal veins, suggesting that testosterone might act suppressing the ET system [157]. Erectile dysfunction may be an early indication of hypertension, being endothelial dysfunction often the link between these two conditions. Hypertension is strongly correlated with an increased release of procontracticle factors such as AngII, aldosterone and ET-1, which impacts vascular and erectile structures [6]. In the vasculature, ET-1 elicits reactive oxygen species (ROS), production by NADPH oxidase enzyme, which in turn releases ET-1, favouring a prohypertensive [158, 159]. Animal model studies reveal ET-1 to be essential in salt-sensitive hypertension-related ED [160, 161]. Furthermore, ET-1 generated Ca²⁺ influx alteration, prompting smooth muscle contraction in isolated penile tissue [162]. An in vitro study showed increased ET-1 Ca²⁺ influence in human smooth muscle cells derived from corpus cavernosum from patients with ED when matched with healthy subjects [163]. These data propose gonadal hormones to have tissue-specified effects on the ET system; hence, an altered equilibrium in menopausal women between estrogens and testosterone might worsen ET-1 vascularrelated pathologies [5]. Regarding ET-1 sensitivity, males have been demonstrated to exert increased vasoconstriction than females in response to ET-1, which has been associated



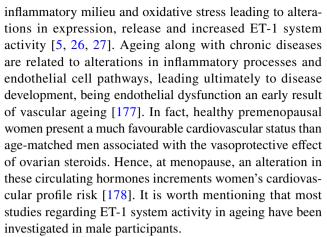
with the increased expression of ETA and ETB receptor in the saphenous veins and in the renal medulla in males compared to females [144, 164, 165]. Similarly, cultivated cerebral arteries from women presented a decreased vascular sensitivity to ET-1 in comparison to those from men [166]. Clinical and basic research studies have demonstrated that females have reduced ETA-mediated vasoconstriction than men [164, 167–169]. Females also display great vascular endothelial smooth muscle ETB receptor-mediated vessel enlargement, which counteracts the global constrictor vascular tone [169, 170]. In fact, a study in which an ETB receptor antagonist (BQ-788) was administered to healthy subjects revealed ETB receptor-mediated vasoconstriction in men and vasodilation in women [170]. This may indicate a leading role of ETB receptor function in women that in turn provide better outcomes in vascular-related disorders than in men. In a similar study, Stauffer et al. showed increased blood flow in men after administration with ETA receptor antagonist (BQ-123) when compared to women, indicating greater ETA receptor-related tonic constriction [169]. Indeed, specific patterns of ET-1 receptor subtype expression and localisation lead to gender disparities in vascular responsiveness to ET-1.

Gender difference within the ET-1 system has been also associated with vascular mediators in downstream cascades activated via ET receptor activation. Zimmerman et al. have reported gender differences concerning ET-1 mediation in oxidative stress generation, modulation of NO levels, inflammation induction and Ang II hypertension [171]. It has been hypothesised that the activation state of the ETB receptor limits ET-1-induced increase in plasma within oxidative stress in females compared to males [172]. Furthermore, female rats that lack ETB receptors have been demonstrated to have diminished renal NOS activity relative to males [173]. Also, intramedullar ET-1 infusion increased diuretic and natriuretic responses in female but not male rats, proposing larger ETA- and ETB-mediated increases in NO in females [174]. Further studies have described ET-1 effect on increasing calcium release from the inner medullar collecting duct cells in males as compared to females [175].

ET-1, age and ageing: is differential diagnostic approach feasible?

In developed countries, ageing entails an increased prevalence of a variety of non-communicable and chronic diseases [176]. Amongst them are vascular diseases, myocardial infarction, stroke, heart failure, diabetes, obesity and cancer, with an increased incidence in the course of ageing [25, 176].

Ageing is related to particular changes in both the innate and adaptive immune systems, promoting an increased



In human brachial arteries, an increase in ET-1 expression was related to ageing along with alterations in ETA-ETB ratios [28]. Ageing men displayed an enhancement in ET-1-dependent vasoconstrictor tone, being potentially alleviated with aerobic exercise [179]. Donato et al. revealed an increased ET-1 expression in vascular EC of healthy elderly men, related to a decreased endothelium-dependent dilation (EDD). In old mice, ET-1 receptor A signalling suppressed EDD [28]. Indeed, elevated endogenous ET-1 mRNA and protein levels in plasma and vascular tissue have been associated with ageing in both female and male rats [180, 181]. In studies with aged mice and rats, they observed ET-1 dampened vasoconstriction (albeit increased ET-1 concentrations) potentially due to functional inability of endothelin receptor signalling pathways [179, 182, 183]. Furthermore, in aged male rats, ET-1 may prompt renal injury, which suggests an action of ET across vascular barriers into the kidney [184]. A study researching aged female mice hearts noted upregulation of ET-1 expression in in vitro cultures of late passage cardiac fibroblasts, along with downregulation of oestrogen receptor-α (ER-α), important negative ET-1. Comparable results with in vitro cultures of senescent cardiac fibroblasts were reported, observing also an increased expression of fibronectin and collagens, which suggests that ageing-related cardiac fibrosis is in part subject to ET-1 upregulation [185].

Other age-associated diseases such as glaucoma or agerelated macular degeneration (AMD) presented increased ET-1 which suggested its role in the diagnosis and pathogenesis of AMD [186, 187]. Enhanced levels of ET-1 were found in AMD patients treated with angiogenesis inhibitors (bevacizumab), yet its implication has not yet been determined [188]. Glaucoma patients have revealed increased ET-1 circulating and intraocular levels and anterograde-retrograde axonal transport [189–193]. ET-1 levels were able to predict postsurgery intraocular pressure in patients undertaking primary open-angle glaucoma eye surgery [194].



ET-1 relevance for endocrine status and multi-faceted hormonal regulation

Diseases related to physiological ageing leading to endocrine changes, such as menopause, are enhanced with age, presenting constraints of the arterial vascular bed, arterial hypertension, insulin resistance and diabetes [195–197].

In fact, as previously described, premenopausal women are more advantageous than age-matched men regarding the cardiovascular phenotype, largely on account of the vasoprotective role of female sex hormones [198]. However, significant hormonal alterations are associated with menopause such as decreased levels of progesterone and oestrogen [199, 200]. Furthermore, estradiol (E2) also declines while follicle-stimulating hormone elevates, increasing the androgen:oestrogen ratio in postmenopause women [201]. Together with the ageing process, this may have implications on vascular and endothelial function, transitioning to a high-risk cardiac profile [199, 200]. It is known that E2 affects NO production and secretion in a direct manner via ERα, which might promote endothelial dysfunction due to a decrease in NO accessibility [202]. In a study with postmenopausal women, they found lower expression of ERα receptors and eNOS in peripheral vein-derived endothelial cells in comparison to young subjects, which suggest flowmediated vasodilation to be dependent on their expression [203]. This indicates that endothelial function deterioration is partially dependent on reduction of ER α expression and eNOS, in turn affecting NO secretion.

The endothelin system and ET-1 are crucial in vascular dysfunction pathogenesis and have been seen to be affected by ageing. In fact, studies have shown an increase of endothelin in plasma in postmenopausal women [204, 205] along with elderly men [28] in comparison with young adults. In a menopause rat model, they observed greater mesenteric artery reactivity to ET-1 precursor, Big ET-1, compared to young females, which improved with oestrogen treatment [206]. Moreover, in studies with ovariectomised rats, the treatment with oestrogen neutralises ET-1 expression, suggesting a vital role of oestrogen on ET-1 regulation [147, 168, 207]. In contrast, testosterone is known to elevate ET-1 expression in both in vivo and in vitro conditions, thereby modulating responses to external ET-1 [154, 156]. In a Brazilian study on postmenopausal women, they revealed a direct relation between ET-1 and testosterone levels in serum [208].

Indeed, in postmenopausal women, ET-1 promotes the deterioration of endothelial function. However, a study with ETB antagonist BQ-788 has demonstrated the involvement of ETB receptor in restoring vasodilation in ageing women [204]. Contrarily, in the same study, the ETA antagonist showed no changes in vasodilator capacity, suggesting the

loss of endothelial function to be associated with a decline in ETB receptor-mediated dilation [204].

Furthermore, in other endocrine defects such as thyroid diseases (Hashimotos's thyroditis, Graves' disease), they have also detected increased ET-1 plasma levels when compared to healthy controls [209]. Other studies have also described elevated ET-1 plasma levels in Graves' disease [210, 211]. Nevertheless, patients suffering from endemic goitre did not present increased ET-1 levels [209]. They failed to observe a relation between ET-1 and thyroid disease parameters (thyroxine, thyroid volume, thyroid-stimulating hormone) [209]. Letizia et al. also reported a lack of correlation between the ET-1 levels and thyroid hormones [211]. Another study has observed that thyrocytes from Graves' disease patients were stimulated by ET-1 to proliferate to a higher extent than healthy subjects [210]. Molet et al. demonstrated that after 24-h treatment with beta chemokines (macrophage inflammatory protein 1-alpha and monocyte chemotactic protein 1), EC elevated ET-1 mRNA in vitro secretion [212]. Thus, beta chemokines are involved in inducing ET-1 release in autoimmune and inflammatory diseases namely, Graves' disease and Hashimotos's thyroiditis [209]. In a study with hyperthyroidism patients, they observed increased plasma ET-1 concentration as opposed to control subjects [213]. They further discovered serum triiodothyronine and free thyroxine to be positively correlated to plasma ET-1 levels, along with a decrease in ET-1 levels after hyperthyroidism treatment compared to pretreatment values [213].

ET-1 has also been described to regulate the development and secretion of the adrenal gland [214]. The involvement of the ET system was detected both in human and rat adrenal cortex and in vitro carcinoma cells [215, 216]. Preproendothelin-1, ECE-1 and ETA and ETB mRNA were found in normal rat and human adrenal cortex [214]. In vivo studies with ET-1 administration to rats and dogs revealed an increase in blood pressure and plasma aldosterone levels [217], speculating the influence of ET-1 on the adrenal cortex [217]. Furthermore, this was supported by increased plasma aldosterone levels due to subcutaneous ET-1 infusion in rats [218]. Altogether, the ET system has an important function in the regulation of adrenal cortex function and potential contribution in adrenal gland pathogenesis like Conn's adenoma [214].

ET-1 association with adverse health effects related to abnormal body weight

Studies investigating endothelin mediation in low BMI subjects are currently limited. However, an epidemiologic study aimed to determine the correlation between BMI (comprising low BMI) and endothelial dysfunction



[38]. Physiological control of vascular tone needs for an equilibrium between vasoconstricting and vasodilating factors [219]. Indeed, endothelin-1 and AngII circulating levels were similarly increased in low BMI, normal, obese and extremely obese groups. Levels of oxidative stress were analogous in low BMI, normal and obese subjects, although significantly greater when compared to normal BMI. Thus, results suggest that besides obesity, low BMI may also present a risk factor for diminished endothelium-dependent vasodilation in subjects, as a result of reduced NO bioavailability, with potential CVD prevalence [38]. Moreover, subjects with abnormally low BMI (underweight) have reported an increased risk of breast cancer with a poor prognosis [220]. Ferri et al. have reported increased plasma ET-1 levels in lean patients with essential hypertension, hyperlipemia and glucose intolerance compared to normotensive or subjects lacking these metabolic disorders [221]. Fasting insulin levels in plasma were also correlated to elevated ET-1 plasma levels, referred to as a risk factor for hypertension [221]. Another study tested leptin vascular actions on ET-1 system and NO pathway balance in both lean and obese individuals and tested the effects of ETA receptor antagonist, BQ123 and NOS inhibitor, NG-monomethyl L-arginine (L-NMMA) [222]. Leptin circulating levels were significantly greater in obese patients than in lean subjects, as previously observed [223, 224]. After infusion of BQ123 and L-NMMA, hyperleptinemia lean subjects revealed enhanced vasodilator responsiveness and greater vasoconstrictor response to L-NMMA during ETA receptor antagonism [222]. These data suggest that in healthy subjects, leptin supports vascular homeostasis by balancing both the ET-1 system and NO pathway, whereas exogenous leptin actions are absent in hyperleptinemic patients with obesity-related metabolic syndrome (MetS) [222]. Besides, increased production of ET-1 vasoconstrictor protein, impaired NO-dependent endothelial function and oxidative stress have been related to increased abdominal fat and body mass index [225].

Subjects with elevated body mass index (BMI) (both overweight and obese) present a chronic metabolic disorder with a distinct detriment of their cardiovascular health along with increased morbidity and mortality [226, 227]. Characterised by an increased BP and an incidence of hypertension [228], this threat may result from endocrine and paracrine dysregulation and chronic inflammatory state that derive from irregularities in adipose tissue function [229]. This consequently elicits an instability amidst the NO pathway and ET-1 system resulting from a disruption in vascular homeostasis [229].

Endothelial dysfunction is marked by diminished NO levels in obesity as presented by Steinberg et al. [230]. In patients with elevated BMI, they observed a dampened

increase in leg blood flow in reaction to intra-arterial delivery of muscarinic receptor agonist methacholine, compared to lean subjects [230]. These results were reproduced by several groups [231, 232]. A further study revealed an impairment of brachial artery flow-mediated dilation (FMD) in obese patients. This suggests obesity-related metabolic abnormalities to prompt brachial artery endothelial dysfunction [233]. Furthermore, Woo et al. reported an association between brachial artery EDD impairment and mild to moderate obesity [234, 235]. Nutrition adjustments have been seen to partially restore vascular abnormalities specifically when combined with physical activity [234].

Moreover, increased ET-1 vascular action in pathologic conditions such as obesity, hypertension, diabetes mellitus and insulin resistance lead to BP abnormalities [36, 37, 236, 237]. Levin et al. revealed a key role of ET-1 in the pathophysiology of vasomotor morbidities linked to the formation of atherosclerotic plaque and endothelial dysfunction [238]. This knowledge was supported by a study investigating ET-1 and NO system interaction in obese patients, where the addition of selective ETA blocker (BQ-123) compensated the limitation in endothelium-dependent vasodilation, confirming ET-1 contribution to abnormal vascular homeostasis in such patients [239]. Indeed, increased ET-1-mediated vasoconstriction in insulin-resistant conditions was identified in obesity and diabetes subjects [240]. Yoon et al. performed a study with obese patients, observing increased peripheral vasodilation in this group when compared to lean subjects. This revealed endothelin as a potent vasoconstrictor affecting vascular tone and diastolic blood pressure [228]. Another study investigated the relationship between elevated ET-1 activity in hypertensive subjects and increased body index. Increased vasodilator responses to ETA blockage in overweight and obese patients were seen when compared to lean hypertensive subjects [241]. This ETA-dependent vascular reaction in a hypertensive and increased BMI setting may suggest an increased vasoconstrictive response to ET-1 or an enhanced ET-1 synthesis at the receptor site [241]. In a study researching ET-1-mediated vasoconstrictive tone, they revealed a dampened forearm constrictor response to external ET-1 in obese and overweight patients in relation to lean subjects [242]. Moreover, selective ETA receptor blockage elicited an increased forearm vasodilator response in subjects with higher BMI [242]. Shortly, ET-1 vascular action increased in overweight and obesity along with diminished endothelium-dependent vasodilation which may lead to an enhanced risk of hypertension and atherosclerosis disease. Physical activity and lifestyle changes can indeed improve endothelial function and reduce selective markers of endothelial activation [243]. In fact, a study carried out with obese men revealed a



direct correlation between the percentage of weight loss and decline in plasma ET-1 concentration upon the conclusion of a low-caloric diet, which may improve obesity-induced endothelial dysfunction [244].

ET-1 as the trigger of pro-inflammatory pathways

Numerous studies have proven the involvement of ET-1 in vascular inflammatory processes. ET-1 has been described to be related to inflammatory responses through activation of transcription factors (i.e. NF-κB) and expression of pro-inflammatory cytokines such as IL-1, IL-6 and TNF- α [245], which in turn stimulate secretion of ET-1 [246]. In particular, TNF- α is known to enhance inflammatory responses by means of cytokine cascades and is involved in the pathogenesis of several diseases such as rheumathoid arthritis, sepsis, Crohn's disease, diabetes and obesity [247]. TNF-α induces ET-1 secretion in vascular EC [248, 249]. Besides, activated T cells release of TNF-α and IFN-γ induce monocytes to produce ET-1 in human peripheral blood mononuclear cells [250]. Other studies claim T cell-secreted TNF-α, IFN-γ, IL-4 and IL-10 to generate distinct macrophage types [251, 252]. Studies have proven that ETA receptor antagonists, namely BQ1232, have a favourable impact on TNF- α levels [253], lowering their concentration in patients after bypass grafting [254]. TNF- α and IL-1 β expressions were also inhibited in an oxidative stress lung rat model by BQ123 (Chen et al. 2010). ETB receptor antagonist studies display some conflicting results. Tonari et al. revealed inhibition of TNF-α expression by BQ788 antagonist in patients with optic nerve damage [255]. Nevertheless, another group stated no substantial reduction in TNF-α levels in rat hearts following BQ788 infusion [256].

Indeed, increased production of pro-inflammatory cytokines can trigger the production of prostaglandin (PG) in vascular endothelial and smooth muscle cells [257]. A number of pathways (NF-κB, cyclooxygenase (COX) and NADPH oxidase-dependent) have been seen to promote ROS production in different cell types [258–260]. COX is a key enzyme in PG synthesis [178], and studies have indicated that its expression can be induced by ET-1, together with prostaglandin E2 (PGE-2) release by NF-κB and MAPKs [257]. ET-1 stimulated surface expression of vascular adhesion molecule-1 (VCAM-1) in TNFα-stimulated vascular EC. This ET-1-promoted increase may be due to TNF-α concomitant inhibitory action on endothelin-induced NO production [261]. A further study with hypertensive patients revealed stimulation of arterial VCAM-1 by ET-1 [262]. VCAM-1 along with intercellular adhesion molecules prompt the adhesion of inflammatory cells to the vascular surface, involved in atherosclerosis evolution [263]. Anggrahini et al. investigated the effects of ET-1 on vascular inflammation and neointima formation in an ET-1-knockout mice model. They observed a decrease of ET-1 adverse effects, suggesting its participation in vasoconstriction and inflammatory cell recruitment to the vessels [264]. Moreover, ROS are critical physiological molecules in vascular cells which overproduction leads to the development of atherosclerosis and endothelial dysfunction [265]. The increase in oxidative stress, monocyte/macrophage infiltration and decrease in high-density lipoproteins are effects exerted by ET-1, leading to the progression of atherosclerosis and aneurysms [266]. Furthermore, overexpression of ET-1 in the endothelium of atherosclerotic mice was marked by a reduction of endothelial signalling pathways in charge of endothelium-dependent relaxation [267]. ET-1 receptor antagonist might be beneficial in preventing numerous vascular-related diseases [268, 269].

ET-1 functionality is linked to both physiologic and impaired healing

Chronic wounds (non-healing)/ulcers are wounds that have not completed the physiologic wound healing (WH) process in an effective manner, arresting usually at early inflammatory stages [13]. Some chronic wound features include neutrophil infiltration, extended inflammatory phase and consistent infections [270, 271]. Risk factors related to the chronic wound-deficient healing process are both modifiable (smoking, malnutrition, obesity, diabetes, alcohol consumption, CVD) and non-modifiable (genetic traits and ageing) [13]. Wound-associated pain comprises both physiologic and psychologic factors, involving also stress and anxiety [15]. The immune system might be compromised by cortisol overproduction, and catecholamine-induced poor tissue oxygenation affecting WH in a direct manner [15]. In fact, individuals suffering from Flammer syndrome present elevated ET-1 levels causing systemic hypoxia as a result of improper vasoconstriction or deficient vasodilation [18, 272]. Wound healing in the liver is an integrated process that involves remodelling, fibrogenesis and disruption of liver structure, and accumulation of extracellular proteins leads to the conversion of the liver to fibrotic and even cirrhotic state [14]. ET is of great importance for WH in the liver; in fact, ET-1 and -3 circulating levels were seen elevated both in cirrhosis patients and in preclinical hepatic WH models [273, 274]. Immunoreactive ET-1 levels in the liver might be related to liver disease severity [275]. Sinusoidal EC usually produce ET in a healthy liver, although following an injury, ET-1 synthesis transitions to hepatic stellate cells [276–279]. Furthermore, ET-1's primary aims in the liver are stellate



cells, which possess a greater number of ET receptors and enhance their binding sites upon activation [280–282]. Activation of stellate cells is exerted by numerous factors such as cytokines, chemokines, growth factors, oxidative stress and ET-1 [283, 284]. ET-1 also interacts with TGF- β in modulating hepatic stellate cell activation and increases TGF- β 1 mRNA besides stimulating the release of TGF- β 1 in these cells. Inhibition of ET signalling by the ET receptor antagonists has decreased hepatic fibrogenic response [284–287]. In a chronic liver injury model, bosentan was able to inhibit ET-1-induced fibrogenesis [284].

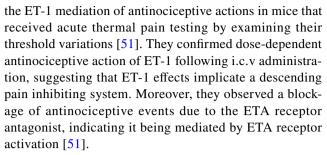
ET-1 is involved in the regulation of senses

Olfactory-sensitive neurons and their axons are enveloped by glia-like cells called sustentacular cells, which help to sustain the olfactory mucosa (OM) structure and ionic integrity [17]. A group demonstrated that sustentacular cells were sensitive to ET-1 and thereby able to uncouple gap junctions [288, 289]. Moreover, endothelin has been described to inhibit gap junctional communication effectively in astrocytes [290]. ET receptors were also greatly expressed in the OM, being ETB receptor mostly expressed in the olfactory sensory neurons, while most non-neuronal cells mainly express ETA receptors [288, 289]. Another investigation found ET-1 acted as a neuroprotective for olfactory cells [291]). In a recent study, they evaluated if OM-produced ET-1 could have an effect on olfactory processing in young rats [17]. OM responses to odorant stimulation following local ET-1 application revealed no modification of response amplitude but slowed treatment recovery. The maternal odour recognition orientation test was decreased after ET-1 treatment, which overall suggests ET-1 olfactory response regulation only partly via gap junction uncoupling [17].

Relation between increased levels of ET-1 levels in blood serum in retinitis pigmentosa and FS has also been described [292]. In accordance, a case report described how a female patient suffering from FS presented significantly elevated serum endothelin levels [19]. ET-1 effect on PGE-2 increased production could potentially suppress the feeling of thirst in such individuals [19].

ET-1 modulates pain and drug sensitivity

Endothelins contribute to numerous pain-related processes, namely, pain caused by cancer, inflammation or Sickle cell disease (SCD). In the central nervous system (CNS), administration of ET-1 intracerebroventricularly (i.c.v) was formerly reported to present antinociceptive actions, which suggests ET-1 involvement in pain transmission [293, 294]. In fact, in a study, they investigated



In addition, ET-1 has been described to be overexpressed in breast carcinoma patients and is associated with poor prognosis, as well as displaying a strong link between both preoperative and postoperative pain sensitivity [20]. The younger age breast cancer patient cohort might be at increased risk for elevated pain sensitivity [53]. In another study, ET-1 administration to study participants elicited dose-dependent effects regarding spontaneous pain and temperature perception [54]. Furthermore, in SCD patients and mouse models, ET-1 blood plasma levels have been described to be elevated, resulting in acute and chronic pain episodes. It is thought that ETA receptors support SCDderived pain by primary sensory neuron NF-κB-triggered upregulation of Nav1.8 [55]. In a study, they investigated the mechanism by which ET-1/ETA receptors participate in SCD-associated pain. They showed that ET-1 and ETA receptor levels were elevated in the dorsal root ganglia of humanised mouse SCD models, but pharmacologic inhibition of ETA receptors (in primary sensory neurons) by ABT-627 [295] mitigated basal and post-hypoxia pain hypersensitivities [55]. These findings suggest the ETA receptor as a potential target for SCD pain management, although further clinical research must be performed.

ET-1/ET-1R axis activation gives cells the potential to exert changes in cell fate and accomplish deleterious features [296]. ET-1 expression has been detected in numerous malignancies such as in advanced tumoral contexts [297, 298], where elevated ET-1R indicated worsened prognosis [296]. Within tumours, ET-1 generates signals which induce pro-survival transcriptional answers, securing tumoral cells from cancer therapy-induced apoptosis [297, 299, 300]. Indeed, ovarian cancer (OC) patients have poor survival rates due to late diagnosis at clinical stages along with recurrence of the disease due to failure of platinum-based chemotherapy [301]. Chemotherapy resistance consists of adaptive signalling pathways which develop specific transcriptional profiles [302]. Platinum-resistant OC tumours have been seen to express greater levels of ETA receptor, being linked to worse disease prognosis [300]. It was revealed that the ET-1/ETA receptor axis hinders the ves-associated protein (YAP) pathway in platinum-resistant OC cells, which crosstalk hinders chemotherapy-induced apoptosis [52]. Tocci et al. effectively described co-therapy of ET-1 receptor antagonist and cisplatin to achieve sensibilisation of



platinum-based therapy-resistant cells, reducing in turn their metastatic potential [52]. Conclusively, blocking the ET-1 receptor enhances platinum-induced apoptosis and hampers adaptive systems, constituting a favourable therapeutic approach to enhance OC patient drug sensitivity.

ET-1 modulates stress reactions and mental health

Psychological social risk factors such as social environment (job or family stress, low socioeconomic status, detrimental life events) and emotional factors (depression, anxiety, exhaustion) have been known to strongly relate to a higher risk of CVD [3, 303]. In the INTERHEART study, they evaluated crucial standard CVD risk factors and a series of psychosocial factors (depression, financial stress) related to acute MI. Results revealed an odds ratio in women of 3.49 and 2.58 in men, independent of ethnicity and geographic origin [304]. Notably, these odd ratios were comparable to CVD risk factors such as diabetes, hypertension or smoking [305]. This suggests an overall contribution of psychosocial factors to an increased prevalence of CVD, being ET-1 balance disruption directly linked to these psychosocial-induced mechanisms [3]. Yammime et al. thoroughly reviewed studies dealing with the relation between these factors and found studies proving heterogeneous findings [3]. Studies on young men and women with CVD family history revealed ET-1 higher levels in African Americans compared to European Americans, following psychological challenges [306]. Sex differences in ET-1 stress reactivity were evaluated, observing significantly increased ET-1 plasma levels in males compared to females [307]. However, there was great variability in protocols used for ET-1 assessment, time of sample collection, laboratory detection procedures and the type and duration of mental/psychological challenges, making it challenging to draw clear conclusions [308–311]. Evidently, ET-1 measurements have to be obtained at regular intervals during a post-stress recovery period in order to elucidate ET-1-specific kinetics [308]. Vascular dementia follows Alzheimer's disease (AD) as the most common type of dementia worldwide, caused mainly by ischaemic or hemorrhagic cerebrovascular (CVS) disease; they cause cognitive detriment and neurodegeneration [312–314]. A recent study focused on examining whether a combination of Shenmayizhi formula and Ginkgo extract positively impacts mild to moderate VaD [4]. Serum indexes of vascular endothelial function, namely, ET-1, NO, VEGF and von Willebrand factor (vWF) were measured. Posttreatment serum ET-1 and vWF levels had decreased, whereas VEGF and NO levels increased in the SMYZF group, presenting more significant changes than the Ginkgo group [4]. This suggests that SMYZF administered with Ginkgo tablets may potentially improve vascular endothelial function, assisting with cognitive improvement and neurological functions in VaD patients. Other studies also described an association between vascular cognitive impairment severity and ET-1 levels [315, 316]. Furthermore, ET-1-induced vasoconstriction prompts cerebral ischaemia and hypoxia which in turn promotes dementia development [317].

275

ET-1 and cardiovascular diseases

ET-1 levels have been reported to be elevated in a number of CVD such as acute myocardial infarction (AMI) [318], coronary artery disease, hypertension, atherosclerosis and congestive heart failure (CHF), amongst others [29]. On account of its vascular tone and contractive potential, ET-1 has been suggested to be involved with hypertension development in humans [29]. In murine models of hypertension, ET-1 levels were increased only if accelerated hypertension is displayed [319, 320]. Studies comparing ET-1 expression in blood vessels from deoxycorticosterone acetate (DOCA)-salt rats and normotensive Wistar-Kyoto (WKY) rats revealed larger amounts in the former, which indicated strain-related differences in ET-1 production, responsiveness or proportion of ETA and ETB vessel receptors [321]. Human studies also revealed an increase in BP after infusion of ET-1 to healthy subjects [322]. ET-1 blocking with bosentan led to a decrease in BP in hypertensive patients [323]; nevertheless, it also diminished BP in essential hypertension subjects with normal ET-1 levels [324], potentially indicating that plasma ET-1 levels do not show the real state of endothelin action.

Hypercholesterolemia is linked with elevated ET-1 levels in human tissues and plasma [325]. It has been hypothesised that ET-1 may be relevant for atherosclerosis formation at all stages, even at the outset [326, 327] seen that ET receptor blockade decreased premature atherosclerosis [328]. More recent animal studies have described how oxidised low-density lipoprotein prompt mRNA expression and release of ET-1 from EC, proposing a key role of ET-1 in atherosclerosis progression [329]. ET receptor antagonist was also able to hinder early atherosclerosis formation in hyperlipidemic hamsters [328]. In humans, there are many factors that can influence atherosclerosis such as hypertension, obesity, diabetes mellitus or hypercholesterolemia [330], and even cytokine release and inflammation may promote atherosclerosis [331]. Furthermore, ET-1 increased levels induce platelet-derived growth factor (PDGF), fibroblast growth factor, TGF- β 1 and vascular adhesion molecules synthesis [332].

Ischaemic heart disease is the primary mortality cause worldwide [333]. In experimental models, coronary vessel constriction was induced by ET-1 [333], demonstrating that



coronary vasosparm is related to ET-1 hyperactivity. ET-1 activation impeded NO-mediated dilation, which might promote vasospam in coronary arteries [334]. ET receptor antagonist extended rat long-term survival following AMI [335]. Coronary artery disease patients might eventually develop acute coronary syndrome (ACS) or AMI. In fact, AMI patients presented elevated plasma ET-1 levels [336] correlating with 1-year mortality [337]. Further studies have shown how endogenous ET-1 exerted vasoconstrictor on arteries, demonstrated by the increased coronary flow following ET-1 receptor blockade in both coronary artery diseased [338] and common coronary patients [339]. Patients with ACS presented enhanced myocardium and left ventricular tissue perfusion following selective ETA blocker therapy [340]. In accordance with these results, ETA and ETB receptor blockage evidenced vasodilation in coronary atherosclerosis patients [338].

The endothelin system is also involved in chronic heart failure pathophysiology, being ET-1 plasma levels higher in patients with CHF, and resting values almost twofold/threefold greater than in healthy subjects [341]. This raise is thought to be primarily due to an increased big ET-1 and ET-1 production [342], supported by the release by the lungs and myocardial cells [343]. CHF patients' symptoms and hemodynamics have been correlated to circulating ET-1 levels [31], being big ET and ET-1 independent survival predictors [344, 345]. Other investigators also showed how plasma ET-1 in CHF was related to more severe disease [346].

Laboratory studies reveal that ET-1 might cause arrhythmic effects in CHF settings [33], besides being involved in cardiac remodelling via fibroblast activation, inflammation of the heart and activation by renin–angiotensin–aldosterone system stimulation [34, 347]. In fact, ECE inhibition promoted the inactivation of the renin–angiotensin–aldosterone system in the CHF milieu [348]. Human studies prompted to clarify the functional link between CHF and ET system activation and progression to be a compensatory neuro-humoral adaptation [342, 344, 349], being patients with the highest ET-1 levels the ones with worse prognosis [344]. Blocking the ET system has turned into a main target for therapeutical treatments.

Preclinical studies with rats demonstrated the use of ERA, namely, BQ-123 to enhance myocardial function and viability [350, 351]. In humans, intravenous administration of BQ-123 [352] or bosentan [353] diminished systemic and pulmonary vascular resistance and BP. Although CHF treatment with ERA was promising, none of the four multicenter, controlled randomised clinical trials was prosperous [354]. Dhaun et al. thoroughly review the discontinued trials [355]. Moreover, acute heart failure portrayed similar results. Initial studies with ERA did in fact propose a haemodynamic advantage, ultimately showing no major effect [356–358]. Whether lower drug doses would have displayed beneficial

effects, currently remains unknown [359]. Jankowich et al. have thoroughly reviewed the potential use of ET-1 as CVD prognosis on account of more personalised treatment schemes [30]. In stable angina patients, big ET-1 high levels were indicative of cardiovascular events such as non-fatal myocardial infarction and stroke, showing a diminished event-free survival [360]. ET-1 may also predict post-myocardial infarction phenomena, namely, the absence of tissue perfusion after percutaneous coronary intervention therapy or even mortality [361]. However, C-terminal proET-1 levels anticipated heart failure in high-risk cardiovascular subjects but not in low-risk patients [362]. Other studies of coronary revascularisation or cardiac catheterisation have also failed to determine cardiovascular-related events [363, 364]. Thus, even if ET-1 peptide levels may support the prediction of cardiovascular events in patients with stable coronary artery disease, this requires further study in multiple cohorts and populations in order for ET-1 to allow more personalised therapy regimens.

ET-1 and Takotsubo syndrome

A group of Japanese cardiologists identified a disease characterised by akinesia/hypokinasia of the distal LV areas with basal normokinesia, which they named "Takotsubo syndrome" [365]. Years later, the event of Takotsubo syndrome (TS) was confirmed [366] as a syndrome caused by microvascular dysfunction resulting in temporary wall motion irregularities with a characteristic ballooning in the LV [365]. It is also referred to as stress cardiomyopathy [367] or "broken heart syndrome" [368]. It is rather a rare disease, estimated in 2% of patients with an early diagnosis of ACS [369], 90% of which are postmenopausal women with normal coronary arteries [370–372]. Major clinical indications of TS are contractile dysfunction commonly affecting the apical heart section, in the absence of coronary thrombosis [365]. Moreover, cardiac contractility unusual disruption influences 28–40% of TS patients, affecting other heart areas [373–376]. Aetiology of TS is normally physical or emotional stress, reporting 39-55% physical stress-related cases compared to 17–33% emotional stressors [377, 378]. Indeed, norepinephrine serum levels were increased in TS patients suffering from emotional stress [379]. Taken that stress prompts TS, cortisol and catecholamine levels should be investigated. In fact, TS patients' plasma epinephrine levels were increased in the subacute phase [380]. Furthermore, an increase in norepinephrine levels was more evident in TS patients during a mental stress exam than in control subjects [381]. Also, epineprhine blood levels in TS patients were increased compared to ACS patients [382]. Further groups have reaffirmed increased levels of both norepinephrine and epinephrine in TS patients' plasma [383], in addition



to elevated cortisol levels in TS patients [384]. Besides the catecholamine apparent trigger of TS, other humour factors such as ET-1 may also be implicated in TS pathogenesis. It has been described that ET-1 causes coronary artery spasm [385–387], being the small diameter arteries the most sensitive [385]. It also contributes to microvascular myocardial dysfunction [388, 389]. Moreover, ET exerts a positive inotropic effect [390, 391] and has been seen to reduce the contractile function of isolated mouse cardiomyocytes [392]. In a study, ET-1 levels in blood plasma were twofold higher than in healthy volunteers, a signature of circulating micro-RNAs differentiates takotsubo cardiomyopathy from AMI; however, other investigators claim no differences in ET-1 levels in TS patients compared to their matching age, gender and risk factor group between comparable groups, as seen in a clinical study [393]. Furthermore, administration of ERA has reported an increased survival rate in heart failure rats [351, 394], suggesting ET-1 as a trigger for TS, although further research must be conducted.

ET-1 and pregnancy complications

ET receptor expression is changing during normal pregnancy as revealed by the increased expression of ETA and ETB receptors in the uterus of pregnant versus non-pregnant women [395–397]. However, most studies addressing ET-1 functional role in BP control originated from animal studies. ET-1 supports utero-placental vasculature contractile tone, which decreases near pregnancy term [398]. Activation of ETB receptor in rats is required for optimal pregnancy outcomes [399]. During normal pregnancies, endogenous progesterone and oestrogen levels are increased, while circulating ET-1 levels are reduced [152]. ET-1 plays an important role in embryonic development, as disruption of the edn1 gene or endothelin receptor A may result in a hypomorphic pharyngeal skeleton or skeletal element fusions [12]. Endothelin ligands and receptors are exclusive to vertebrates and manage to control neural crest cell development [12].

Pregnancy-related hypertension includes many disorders such as eclampsia and preeclampsia (PE) (attenuated progesterone levels), gestational hypertension (GH) and haemolysis, elevated liver enzymes and low platelets (HELLP) syndrome [9, 11, 400]. PE is a multisystem disorder associated with increased renal vascular resistance and elevated BP, and endothelial dysfunction is one main cause of both maternal and foetal morbidity and mortality worldwide, whose underlying mechanisms are hardly understood [9, 401]. The hypoxic placenta releases anti-angiogenic factors (sFlt-1 and soluble endoglin), cell-free nucleic acids, free radicals and proinflammatory mediators and major effector ET-1 [402], which disturb the balance between endothelium-derived constricting (TXA2, angiotensin and ET-1) and relaxing factors

(PG12 and NO) [9]. These circulating cytokines stimulate further ET-1 production by EC [10]. Many groups reported increased ET-1 plasma levels of preeclamptic women in comparison to normotensive pregnancy controls [403, 404]. ET-1 umbilical vein concentrations in PE were higher than in normal pregnancies [405]. In fact, progesterone complementation to umbilical vascular EC exposed to serum from preeclamptic women weakened secretion of ET-1 in humans [406]. Other studies have shown increased plasma ET-1 levels in umbilical cord cells and renal tissues throughout late PE, suggesting that ET-1 is involved in the progression of PE rather than in the initial phases [10, 407]. Many studies have shown similar ET-1 levels in normal pregnancies and PE, presenting high levels just in severe PE and HELLP syndrome [408]. PE represents a risk for long-term diseases for mothers and their babies, such as renal, cardiovascular, CVS or neurological diseases [11].

277

Experimental models of PE have also revealed increased ET-1 tissue levels in both the kidney and placenta [404]. Aggarwal et al. reported a link between the elevation of sFlt-1, sEng and ET-1 in PE maternal circulation which suggests the secondary mediation of ET-1 in PE pathogenesis to these anti-angiogenic factors released by the placenta [409]. ET-1 precursor, pepro-ET-1 mRA elevated tissue levels, has also been associated with PE in experimental models. Placental ischaemic rats expressed increased pepro-ET-1 in the renal cortex and medulla as opposed to normal pregnancy animals [410]. Moreover, infusion with pro-inflammatory cytokine, TNF- α , mediated hypertension by inducing pepro-ET-1 gene expression in the placenta, kidney and maternal vasculature of pregnant rats [411]. An additional proposed mechanism for elevated ET-1 levels in PE is through activation of matrix metalloproteinase (MMP). MMP has been reported to be increased in women affected by PE and increased MMP-2 expression increased an increase in big ET-1 conversion, hence increasing vasoconstriction [412]. Another study revealed increased vascular and serum MMP-1 in women with PE, which might promote hypertension development in the mother [413]. Considerably, ETA receptor blockade enables protection against PE [414-417]. ETA receptor antagonist was infused in a rat model of PE, resulting in a rise in BP, demonstrating the key role of ET-1 [410]. Nevertheless, ETA receptors are vital for foetal development during the first trimester; thus, their use should be restricted. ETA receptor antagonists that do not transgress the placental barrier would be an alternative.

Furthermore, endothelial dysfunction, characterised by imbalances of vasoconstrictor/vasodilator factors, involves increases in blood coagulation potential (hypercoagulation) leading to uterine vessel vasoconstriction, platelet aggregation activation and resulting in final miscarriage [418]. Dubyk et al. compared ET-1 and NO serum levels in pregnancies with a risk of miscarriage, spontaneous abortion and



non-developing pregnancies versus physiological pregnancies [418]. They reported a significant increase in ET-1 and decreased NO levels in all groups compared to the control group. This indicated that endothelial dysfunction was likely the cause of miscarriage in these women, serving ET-1 and NO as potential markers for endothelial dysfunction [418].

Prenatal death increases with the presence of pregnancy complications such as polyhydramnios or oligohydramnios [419, 420]. Oligohydramnios refers to a diminished volume of amniotic fluid than anticipated for gestational age which may have a foetal, placental or maternal cause or even be idiopathic [421]. Pregnancies with oligohydramnios complications might be at risk of pulmonary hypoplasia, foetal deformation or umbilical cord compression [421]. Elevated ET-1,2 concentrations were observed in pregnancies affected by oligohydramnios [422]. Moreover, studies measuring ET-1,2 levels in umbilical venous blood collected at delivery revealed increased levels in oligogydramnios infants [423]. Thus, increased levels of ET-1 in the feotus can result in oligohydramnios. A study observed lower ET-1 levels in oligohydramnios twins in respect of twins with polyhydramnios, suggesting a critical role of this hormone in amniotic fluid volume regulation [424]. No correlation between gestational age and ET-1 amniotic fluid [424] nor foetal ET-1,2 [423, 425] concentration was observed. Other studies revealed higher ET-1 levels at pregnancy term compared to at midtrimester pregnancy [426, 427]. ET-1 might affect amniotic fluid volume regulation by causing the release of vasopressin, natriuretic peptide, aldosterone and diminishing renal perfusion [422]. Further investigations would elucidate the exact ET-1 implications in pregnancy complications.

Multi-faceted involvement of ET-1 in migraine attacks

Migraine is a neurovascular disorder acknowledged since olden times which is greatly prevalent in society, affecting 1 in 10 people worldwide [428-430]. Migraine pathophysiology theories are very vast. Thus, whether the vasculature plays a primary role in the inception and maintenance of migraines remains uncertain [431–433]. Most studies emphasise vasodilatory mediators when researching migraines; however, vasoconstrictors and their effects must be considered. In particular, diminished levels of NO vasodilatant metabolites and an enhancement of ET-1 have been detected in migraine patients [434]. Studies report elevated baseline levels of ET-1 compared to control subjects [434–437]. ET-1 regulated cerebral blood flow and its receptors have been identified in the endothelium and VSMCs across the CNS and in the arterial system [70, 71]. Interestingly, plasma levels of ET-1 were found to be elevated in the early stages of migraine attacks and promptly

diminished at the onset of the headache [435]. Gallai et al. also observed increased ET-1 plasma levels in the ictal phase of migraine [438]. However, contradicting data were found, revealing no significant ET-1 concentration changes during migraines [439, 440]. All studies demonstrated increased levels in venous blood during migraine attacks. Hypoxia has also been seen to increase ET-1 expression, eliciting migraine attacks with and without aura [441, 442]. One of the primary factors for migraine aura might be cortical spreading depression [443, 444], along with the cause for migraine headache [445, 446]. ET-1-induced CSD may be mediated by microinfarction on account of vasoconstriction [447]. CSD can be triggered by ischaemia or mechanical, electrical and chemical cortical stimulation [448], and has been detected in association with vascular responses throughout migraine attacks with aura [443, 449]. Moreover, migraine aura attacks are improbably linked to neuronal damage [450], as well as neuroimaging data to be clear from sublicinal infarcts or white matter hyperintensities [451]. Furthermore, ET-1 is related to nociception in the nervous system [452], and it is seen to induce pain and cause sensitisation to distinct nociceptive stimuli in the human peripheral nervous system [115]. It triggers the release of endogenous, migraine-inducing molecules such as NO [453] and calcitonin gene-related peptides [454], which have been shown to initiate migraine attacks in clinical studies [455]. ET-1 might influence and generate migraines with aura by originating a cascade comprising migraine-triggering substances. Broadly recognised migraine prophylaxis drugs are beta-blockers and the ACE inhibitor lisinopril which reduce both ET-1 synthesis and release in human EC [456, 457]. An AngII type 1 receptor blocker decreases ET-1 concentration in essential hypertension patients [458]. A randomised clinical trial with combined ETA/ETB receptor antagonist bosentan was ineffective in the acute treatment of migraine [459]. This study failed during migraine attacks, but ET-1 antagonists might be effective in migraine prophylaxis or when administered at the outset of attacks.

ET-1 in ischaemic stroke

In humans, ischaemic stroke is the second leading cause of death and disability globally [460]. Based on the area and size of the brain injury, patients normally suffer from lifetime impairments, affecting from cognitive, sensory and motor to behavioural and communicative functions [461]. The majority of stroke cases results from transient or permanent obstruction of the cerebral blood vessel, depriving the brain of energy and oxygen [462]. The ischaemic cascade initiates the formation of ROS, accumulates calcium intracellularly, releases glutamate and induces inflammatory processes, resulting in tissue injury (infarction) [462]).



ET-1 also induces neuronal damage [463], augments blood-brain barrier permeability and enhances vasospasm related to subarachnoid haemorrhage (SAH) [464]. ET-1 levels were seen to be elevated in both plasma and brain tissue in ischaemic stroke patients [236, 463, 465, 466]. In acute ischaemic stroke patients, plasma ET-1 levels were increased, being more marked within the initial 24 h after stroke onset, correlating to neurological damage severity [466]. Another study observed a correlation between Big ET concentrations and their specific clinical outcome (high levels: poor prognosis/low levels: more favourable outcome) [467]. ET-1, as the potent and long-lasting vasoconstrictor that it is, has been used to induce focal ischaemia in animal models, resulting in afflicted pure-motor and sensorimotor conducts which are reliant on the area of ischaemic insult in these models [468-471]. ET-1 can be applied to cortical surfaces [25], resulting in the dose-dependent ischaemic lesion with marginal ischaemic edema [468, 472], or directly onto exposed middle cerebral artery [473] as an intracranial injection [472]. Increased potential of ET-1 when delivered to conscious rats in relation to anesthesised ones has been proven [474]. Models of anterior cerebral artery occlusion and white matter ischaemia in the internal capsule have also been conducted with ET-1 [475, 476]. Furthermore, some studies utilising ET-1 to induce cerebral ischaemia models have been developed in non-human primates, in specific in marmoset monkeys [477, 478]. Its administration generated dose-dependent decreases of vessel calibre in middle cerebral arteries, succeeded by progressive reperfusion [477]. ET-1-treated marmosets displayed pronounced contralateral motor deficits in grip force [477]. Dai et al. revealed ET-1 potential to induce transient ischaemic stroke in rhesus monkeys and generate focal ischaemia in non-human primates, making it a compelling stroke and post-stroke brain repair model [479]. In fact, ET receptor antagonists have been reported to exert protective effects in animal models of stroke [93, 480, 481].

ET-1 model advantages are the low mortality rates, less invasive technique and conceivable induction of direct focal ischaemia in both superficial and deep brain areas. In contrast, the ET-1 model presents limitations related to astrocytes and neuron production of ET-1 receptor and ECE [482], which may induce astrocytosis [483]. Thus, it is suggested that ET-1 may be involved in CVS disease pathogenesis, pointing future directions towards the employment of endothelin as an early predictive factor for patients undergoing an ischaemic stroke.

ET-1 and neurodegenerative disorders

279

Neurodegenerative diseases such as AD are characterised by a loss of neurons in the brain, which may result in loss of memory and cognitive function deficits [484]. There are a number of factors involved in AD neuronal degenerative alterations such as beta-amyloid deposition, pro-inflammatory cytokine/chemokine secretion and microtubule destabilisation [485]. Vascular dysfunction plays a main role in AD progression [486], and ET-1 has been described to be increased in the cerebral cortex and cerebral blood vessels in AD [39, 486, 487]. Increases in amyloid beta-protein (Aβ) indirectly stimulate ET-1 production [486, 488, 489]. Cerebral vasculature of mice infused with Aß increased ET-1 production [490], and human neuroblastoma and brain microvascular EC exposed to Aβ increased ECE-1 and -2 secretion, resulting in elevated ET-1 secretion [39, 486, 491]. Astrocytes also regulate ET-1 expression in AD and other brain disorders [39, 40, 492]. Furthermore, increased ET-1 has been reported in a number of neurodegenerative diseases, such as multiple sclerosis, Parkinson's disease or amyotrophic lateral sclerosis [41, 492, 493]. For instance, ALS is characterised by progressive loss of motor neurons and astrogliosis. In ALS, a neuro-inflammatory reaction takes place in glial cells, namely astrocytes and microglia [41]. Ranno et al. examined the expression of ET-1 in both spinal cords of SOD1-G93A mouse model of familiar ALS and ALS patients and reported increased ET-1 expression in both cases. In their in vitro mixed spinal cord culture model with reactive astrocytes, ET-1 exposure exerted harmful effects on MNs which were concentration and time dependent [41]. The following study investigated the underlying mechanisms of ET-1 toxic effects on MNs cultures. Their results suggest that ET-1 toxicity does not directly result from oxidative stress or COX-2 activation but requires NO and is mediated by phosphoinositide 3-kinase (PI3K) diminished pathway activation [493]. They also observed that microglia cells are not involved in ET-1 detrimental effects on MNs [493]. Thus, ET-1 signalling may be a fitting therapeutic approach to hinder MN degeneration in ALS disease.

ET-1 in cancer

ET-1 participates in tumour growth, cell proliferation and other aspects of cancer progression in a variety of tumours [494]. In cancer types such as breast, colon, pancreatic and prostate cancer and human oral squamous carcinoma cell lines, ET-1 protein or mRNA secretion was increased [495–497]. Two distinct classifications of endothelin-derived tumours have been described. Both tumour groups hypersecrete ET-1; however, one upregulates ETA receptors and



downregulates ETB receptors slightly, such as ovarian, colon, prostate, pancreatic and renal cell carcinoma, whereas the other upregulates ETB receptors and downregulates ETA receptors, like breast and lung cancers [494]. The endothelin axis results in activation of atypical proliferation, alteration of nociceptive stimuli, apoptosis evasion, angiogenesis, cell proliferation, immune modulation and metastasis invasion, by triggering multiple signalling pathways [44]. ET-1 mitogenic activity may be increased by the interplay with growth factors such as EGF, insulin, insulin-like growth factor, TGF, PDGF, basic fibroblast growth factor and IL-6 [44].

Ovarian carcinoma ET-1 and ETA are overexpressed in a great number of primary and metastatic ovarian cancers, relating also with progressive stages of cancer. In fact, increased ET-1 levels were found in ascites of patients with epithelial OC, in which ETA exerts pleiotropic effects such as survival, migration and invasion [45]. Gene expression analysis studies revealed ETA as a metastasis-related gene [498] greatly expressed in post-chemotherapy specimens in relation to untreated primary ovarian carcinomas [499]. Besides ETA, EGFR is also overexpressed in OC, being usually linked to poor prognosis and related to tumour resistance to chemotherapy, thus making it a prominent therapeutic target [500, 501]. This knowledge promoted the investigation of EGFR inhibitor gefitinib (ZD1839, Iressa) along with ETA antagonist ZD4054, which revealed an enhanced efficacy, resulting in partial or complete tumour regression of ovarian carcinoma xenografts followed by decreased vascularisation, VEGF, MAPK, EGFR, matrix metalloproteinase-2 (MMP-2) and Ki-67 [502]. Kajiyama et al. observed how ovarian carcinoma cells overexpressing neutral endopeptidase (cell surface aminopeptidase that degrades ET-1 amongst other peptides) presented diminished ET-1 levels, cell proliferation, viability and invasiveness [503]. Overexpression of NEP in vivo showed reduced tumorigenesis, suggesting the use of NEP as a suppressor of ovarian carcinoma progression by targeting ET-1. In another study, patient-derived xenografts treated by dual ET-1R antagonist macitentan in combination with cisplatinum showed shutting down of the β-arr1-mediated YAP/mutp53 transcriptional programme (its activation correlates with the worst cancer prognosis) accompanied with anticancer effects in high-grade serous OC [504]. Chellini et al. (2019) evaluated signalling network for adhesion components, cytoskeletal remodelling and ECM degradation in OC. It has been concluded that the ET-1 receptor regulates extracellular matrix degradation and consequent metastatic spread in OC via β-arr1/IQGAP1 signalling pathway [46].

Breast carcinoma Increased expression of ET-1 and its receptors correlates with the progression of the disease and its malign potential [505]. Also, ECE-1 overexpression in

breast cancers has been linked to adverse outcomes [506], whereas increased expression of ETA relates to low disease-free survival and overall survival. Increased ETA expression in breast cancer relates to chemotherapy resistance, thus serving ETA as a predictive marker for chemotherapy therapy outcomes [505]. Overexpression of ET-1 has been detected in numerous diseases [20]; indeed, in breast carcinoma, its increased expression has been associated with poor disease prognosis [507]. High levels of blood ET-1 result in overall hypoxic effects, which consecutively lead to metastatic disorders [47, 48, 508]. Additionally, ET-1 blood patterns are not well understood in regard to the distinct breast cancer subtypes.

A recent work reported increased circulating levels of ET-1 in breast cancer also correlated to LV remodelling in this patient cohort compared to healthy controls [509]. In vitro studies have reported a correlation of ET-1 axis expression levels and breast cancer cell line invading potential [510]. Grimshaw et al. established an in vitro invasive breast tumour cell phenotype by exposing tumour cells to ET-1, through the action of ETA and ETB receptors and elevated MMP activity [511]. In a model of breast carcinoma, the use of dual endothelin receptor antagonist (ERA), bosentan, inhibited bone metastasis, as well as tumour growth and vascularisation [512]. Another study reported high relevance of the ETB receptor in breast carcinogenesis. Silencing the EDNRB gene modulates invasiveness in breast cancer cells towards ET-3. ETB receptor isoform specific controls breast cancer cell invasiveness and isoform- and subtype-specific dissimilarities in patient survival [513]. Consequently, knockdown of the ETB receptor by a specific shRNA significantly suppresses the proliferation and metastatic parameters in MDA-MB-231 and BT549 cells and upregulates apoptosis [49]. In consensus, suppression of ETB receptor expression inhibited cancer growth in the mice xenograft model.

Prostate carcinoma The ET axis has been described to contribute to prostate cancer pathophysiology. ET-1 in healthy prostate subjects is produced by epithelial cells and maximum levels are detected in seminal fluid. In prostate carcinoma, ET-1 local levels are increased due to a decrease in ET-1 clearance pathway constituents [514]. ETA increased expression is seen in primary and metastatic prostate carcinomas, correlating with tumour development and grade, where ET-1 exerts cancer progression actions. In a study where ETA antagonist atrasentan was administered to prostate cancer patients, they revealed a reduction in pain and prostate cancer progression biomarkers [515]. Another ETA orally active antagonist has been researched in phase II clinical trial, revealing general improvement when administered to metastatic hormone-refractory prostate cancer patients [516]. Silencing ET-1 by RNAi significantly suppresses



the progression and invasion of PC-3 prostate cancer cells. These effects affect several signalling pathways including Erk1/2/Bcl-2/Caspase-3, PI3K/Akt/Caspase-3 and MMPs (MMP-2 and MMP-9) [517]. To this end, the role of the proteasome inhibitor bortezomib in ET-1 was evaluated, observing an induction of cell signalling in prostate cancer. ET-1 supplement to PC-3 cells decreased Bad, p53, p21 and p27 expression, and increased IL-8, VEGF, proteasomal activity and NFkappaB levels. Furthermore, these pro-neoplastic effects of ET-1 in PC-3 cells were reversed by the treatment with bortezomib which suggests its distinct role in the regulation of cancer cell proliferation and apoptosis [518].

Colon carcinoma ET-1, ETA and ECE-1 are actively expressed in colon adenocarcinoma cells in comparison to non-cancerous cells [519]. ET-1 expression is elevated in the majority of primary human colon cancers [520]. Moreover, ET-1 by inhibiting β -catenin signalling is capable of recovering colon cancer cells from growth arrest and apoptosis, revealing ET-1 oncogenic function in colon carcinoma [520]. In vitro studies revealed upregulation of ETA expression in all cell sorts compared to healthy colon cells and described ET-1 as a mitogen for colorectal cancer cells. Cancer-associated blood vessels and fibroblasts revealed increased ETA binding while having downregulated ETB, which prevailed in the non-cancerous colon [519]. This shift might induce ETA-related ET-1 colorectal cancer growth and neovascularisation, suggesting the benefit of ETA receptor antagonist as colon carcinoma adjunct therapy [521]. Wang et al. [522] reported that the ETA receptor stimulates colon tumorigenesis via an increase in cell proliferation and migration. Mechanistic evaluation of this effect revealed ET-1-induced YAP/TAZ dephosphorylation and transcriptional activation in multiple colon cancer cells. In detail, ETA receptor activation stimulates G protein Gαq/11 and Rho GTPase pathway and suppresses the Hippo pathway that collectively leads to ETA receptor-induced carcinogenesis via YAP/TAZ activation. The role of the ET axis and its downstream signalling in metastatic colon carcinogenesis was investigated in a clinical study [50]. The authors evaluated the mRNA expression of 36 genes linked with the ET axis using 18 non-metastatic and 20 metastatic colon carcinomas in comparison with normal colon mucosa. Data showed that the majority of genes in the ET axis are overexpressed (17 from 36) in malignant colon tissue. It has been concluded that suppression of PTEN expression may boost a malignant phenotype in colon carcinogenesis.

Other cancers ET-1 has also been detected in the lungs, cervix, colon, melanoma, pancreatic, glioblastoma, neuroblastoma and bladder cancers. In this regard, the ETB receptor represents a potential prognostic marker for lung

adenocarcinoma patients. ETB receptor may act via regulation of the ERK signalling pathway in lung adenocarcinoma [523]. In aggressive cervical squamous cell carcinoma, overexpressed ET-1 contributes to the angiogenesis signalling pathways [43]. Clinical data indicate that a MAPK pathway inhibitors combined with ETB receptor antagonists could have a synergistic anticancer effect in melanoma patients with upregulated MAPK signalling [524]. It has been documented that ET-1 stimulates MAP kinase and AP-1 signalling pathways, increasing the expression of MMP-9 and MMP-13 and consequently activating cell migration in human glioblastoma [525]. Pancreatic ductal adenocarcinoma is related to increased levels of ET-1 [526, 527], revealing likewise overexpression of ET axis components such as ETB, in PDAC tissues [528]. A recent study observed the expression of ET axis constituents in pancreatic acinar and islet cells compared to the minor levels in pancreatic ducts of control mice [298].

Many investigations were carried out with ET-1 receptor antagonists, range from preclinical to clinical trials reaching phases II and III with either selective or specific ETA or dual ETA/ETB antagonists or even selective ETB agonists IRL-1620 [297, 529, 530]. ERAs' action is exerted firstly on ET-1, which in turn blocks autocrine and paracrine pathways, hampering tumour growth and angiogenic effects, making them promising tools for cancer therapeutics [531]. IRL-1620 may be employed to enhance drug delivery (in chemotherapy and radiation therapy) due to the increase in tumour perfusion which leads to enhanced therapeutical effectiveness [297, 529]. Given that ETA-specific antagonists in cancer have portrayed unfavourable results, a potential combination of ETA/ETB blockage with macitentan may be advisable for cancer, as observed in preclinical studies [297]. In fact, the concomitant enhancement of antimumoral immune reactions and suppression of tumour cell invasiveness would make macitentan a strong candidate for cancer therapy [297]. It is critical to comprehend the entire range of molecules and pathways activated by ET-1 in order to design targeted therapies. Additionally, investigating novel combination strategies by merging macitentan with other molecular target therapies, chemotherapy or radiation therapy is required.

ET-1 and individual COVID-19 outcomes

The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) poses a hazard to individuals with chronic health afflictions which are more prone to progress to a lifemenacing stage characterised by inflammatory lung processes and cytokine outbursts [61].



Recent findings hypothesise receptor-interacting protein kinase 3 [532] oligomerisation of open reading frame (ORF)-3a to drive necrotic cell death, leading to tissue injury and detrimental inflammation [60]. More precisely, ET-1 production in the dendritic cell has been seen to be stimulated by necroptosis (regulated necrosis) [533]. Abdul et al. described that blockage of ET receptors with bosentan impaired the necroptosis pathway along with increased brain microvascular endothelial cell migration, proposing that ET-1 activates programmed cell death pathways under inflammatory stimuli [533].

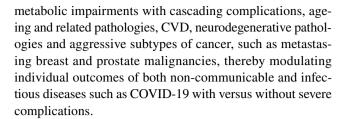
As previously noted, ET-1 detrimental action is exerted via ETA present on pulmonary smooth muscle cells and ETB in vascular wall muscle cells which are upregulated in the context of systemic and pulmonary hypertension [534]. Thus, in this context, ERAs have been validated to be used for treating cases of pulmonary arterial hypertension (PAH) [62]. This is the case of bosentan, known for decreasing profibrotic and pro-inflammatory cytokines (interleukin (IL)-2, -6, -8 and interferon-γ) in patients with scleroderma [535]. Furthermore, bosentan has proven efficacy against some viruses together with exerting anti-IL-6 effects [536, 537]. In a case report of influenza A-related acute respiratory distress syndrome, bosentan administration led to symptom amelioration along with mechanical ventilation weaning [538]. Furthermore, other studies have also used ERAs as cancer therapeutic drugs, diminishing the effect of ET-1 on tumour progression, by constraining epidermal growth factor receptor and blocking angiogenic effects [531]. Also, ETA receptor antagonists like ERAs have been used for the treatment of chronic kidney diseases along with hypertension by opposing ET-1 vasoconstricting effects [72, 531]. These results suggested a potential use of ERAs as a drug against SARS-CoV-2 inflicted hypertension and hypoxic vasoconstriction [63]. However, further tools for improved identification, intervention and prognosis of COVID-19 patients are needed, along with further research on potentially involved mechanisms.

ET-1 as the target in the framework of 3P medicine: status quo and outlook

Depending on the pathway, the level of release and corresponding targets, ET-1 is involved in the regulation of physical and mental well-being; female and male health; modulation of senses, pain and stress reactions; drug sensitivity; healing processes; amongst others.

Endothelin in disease prediction

Shifted ET-1 homeostasis may influence and predict development and progression of suboptimal health conditions,



Endothelin in disease prevention early in life

The most prominent examples are vascular dysregulation, systemic vasoconstriction and arterial stiffness linked to elevated ET-1 levels, which individually may lead to cascading pathologies discussed in this article. Physical training has proven to decrease BP and enhance endothelial function in hypertensive and healthy subjects [32, 539] and it has improved arterial stiffness by elevating vasodilation/ NO availability and reducing vasoconstriction/ET-1 level in adults [540]. Aerobic exercise reduces ROS and increases NO bioavailability improving the balance between NO and ET-1 [2]. Thereby, DNA methylation of EDN1, NOS2, ALU and TNF genes is crucial in reducing systemic BP, meanwhile implementing continuous aerobic physical training [2]. A study which examined the effect of combined resistance and aerobic exercise (CRAE) training in obese pre-hypertensive girls on BP, body composition, blood nitrite/nitrate (circulating NO marker) and ET-1 levels [1] has demonstrated that CRAE training managed to reduce SBP, arterial stiffness, increased total nitrite/nitrate and decreased ET-1 levels and decreased percentage of body fat and increased lean body mass in obese adolescent girls with pre-hypertension. These results convey that CRAE-mediated decrease in BP is possibly due to an improvement in vascular endothelial cell function, as evidenced by the elevated blood nitrite/nitrate and diminished ET-1 levels, and would be a potential therapeutic treatment for high BP, preventing prospective CVD [1]. In consensus, other studies demonstrated how physical training decreases ET-1 and increases NO circulating levels collectively leading to reduced BP [541, 542]. To this end, increased vasodilatation by the NO bioavailability reduces arterial stiffness through a structural alteration of the endothelium that in turn reduces BP in prehypertensive patients [1].

Endothelin in the prevention of accelerated ageing

Approaching disease prevention must be the initial phase to achieve a healthy/physiologic ageing [177]. Endothelin activation along with inflammatory processes is related to a greater incidence of chronic diseases in the elderly compared to the younger population [543]. Thus, endothelial therapy and physical exercise are preventive procedures enabling a favourable ageing process [91]. Physical activity has been



shown to have comparable effects as drug treatments [544], since it enhances the release and bioactivity of endothelial NO, which in turn reduces the production and biological activity of ET-1 [177, 545, 546]. Some other related effects are anti-hypertensive, anti-inflammatory and anti-diabetic functions [547, 548]. The term endothelial therapy comprises preventive efforts, physical training and potential use of endothelial factor treatment in an effort to improve standard endothelial homeostasis [177, 549, 550]. In fact, elderly women exposed to endurance training presented reduced ET-1 circulating levels [205]. Training also dampens ET-1 vascular tone rise in the elderly [179, 551], along with a suppressed age-related BP increase [545, 546]. Indeed, there is a range of studies assessing ET-1-induced disease prevention, and ERAs have specifically been studied to prevent severe disease progression, delay disease onset or as potential therapeutics. In fact, ETA-specific ambrisentan has been proven to be advantageous in preventing vasoconstriction and cellular proliferation mediated by VSMC ETA receptors, concomitantly maintaining ETB receptor vasodilator function [552, 553]. Macitentan as dual ERA inhibits ET-1 binding to both ETA and ETB, differing from bosentan and ambrisentan in its slow receptor dissociation characteristics at a cellular level [554, 555]. It appears that macitentan is able to block ET-1-promoted signalling in a more effective manner than other ERA [556]. ERA may be used for the prevention of fibrotic disease development. In fact, bosentan is known to inhibit ECM deposition and ET-1-induced fibroblast proliferation, besides being able to reduce pulmonary, cardiac, hepatic and renal fibrosis in many disease models by endothelin axis activation [557]. The hepatic fibrogenic response was also diminished in liver disease experimental models by inhibiting endothelin signalling with ERA [284, 285]. Furthermore, ET-1 is believed to be an essential intermediary in cerebral vasospasm subsequent to a SAH, given that animal and human studies report increased blood and cerebrospinal fluid ET-1 levels [558]. ET-1 and NO interplay is crucial in preserving cerebral vascular dilation and cerebral blood flow upon SAH. Vasospasm can be prevented by either ET-1 antagonist or ECE inhibitor administration, being ETA receptor antagonist clazosentan favourable in preventing or reversing cerebral vasospasm [559]. In SAH experimental animal models, clazosentan has been seen to decrease or reverse cerebral vasospam [560]. Further reports in experimental SAH rabbits, rats, dogs and monkey models have proven the prevention or reduction of vasospam following endothelin antagonist administration [561]. Despite ERAs are a promising therapeutical treatment for many diseases, additional studies are required to assess the benefits and safety of ERA treatment along with the validity of combination therapy schemes.

Endothelin, targeted therapy and personalised medicine

Since ET-1 discovery, the therapeutic strategy has evolved around the application of either ETA receptor (ambrisentan) or ETA/ETB dual blockage (bosentan and macitentan) by using orally administered small molecules antagonists [562]. There has been recent advancement in this field with the use of selective peptide agonists and antagonists along with monoclonal antibody antagonists, potentially broadening therapeutical extent to further pathophysiological settings. The use of ETA antagonist ambrisentan together with the phosphodiesterase (PDE) 5 inhibitor tadalafil was proven to enhance PAH treatment [563]. Combining two vasoconstrictive sites such as angiotensin AT1 and ETA antagonist, sparsentan has become another innovative approach [564]. In addition, single-nucleotide polymorphism (SNP) has been studied to validate its genome implication, as it is related to numerous vascular diseases where elevated big ET precursor plasma levels have been identified [565]. This SNP would potentially enable patient stratification for assignation to ET treatment and allow for a more personalised therapy [565].

Monoclonal antibodies (MABs) MABs have been widely researched as they are extremely selective for their targeted protein and present an extended plasma half-life [566]. Although manufacturing costs of MABs are quite substantial, they provide a broader range of treatment approaches compared to small molecules [567]. ETB receptor MAB, rendomab-B1 (RB1), was more effective than BQ788 competing for ET-1 [568]. RB1 was tested in melanoma cells, exerting low affinity for ETB receptors displayed in this cancer, which suggests diversity in ETB tumour subtypes [56]. A novel MAB, rendomab-B4 (RB4), was reported to attach to ETB receptor from three distinct melanoma cell lines but not to ETB on human embryonic kidney or native receptors in human umbilical vein endothelial cells [56]. Thus, RB1 and -B4 binding qualities present disparities that may be related to post-translational modifications [569]. ETA receptor MAB, rendomab-A63 (RA63), presents a sub-nanomolar binding affinity, while not being modified by an excess of ET-1 [570]. RA63 also binds ETA present on glioma stem cell surface [571]. As a result of MAB increased half-life, they may be a suitable therapeutic scheme considering ET-1 sustained vasoconstrictor activity, which has been suggested to bind irreversibly to ETA receptors. However, a study revealed that ET-1 binding cleavage from cloned ETA receptors although slow compared to further vasoactive elements was not irreversible [572]. ET-1 prolonged effect could be reversed both in vivo and in vitro by small-molecule antagonists [573, 574].



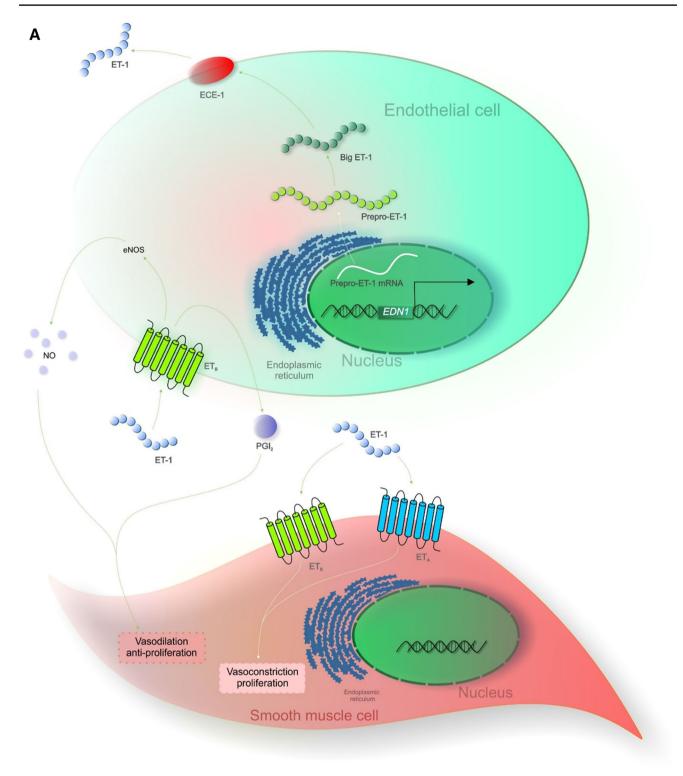


Fig. 1 A Schematic presentation of the endothelin-1 production and functionality. Abbreviations: ETA, endothelin A receptor; ETB, endothelin B receptor; ET-1, endothelin-1; PGI2, prostacyclin; eNOS, endothelial nitric oxide synthase, ECE-1, endothelin-converting

enzyme 1; EDN1, endothelin 1 gene; NO, nitric oxide. ${\bf B}$ Endothelin-1 involvement into a spectrum of pathophysiological processes and disorders



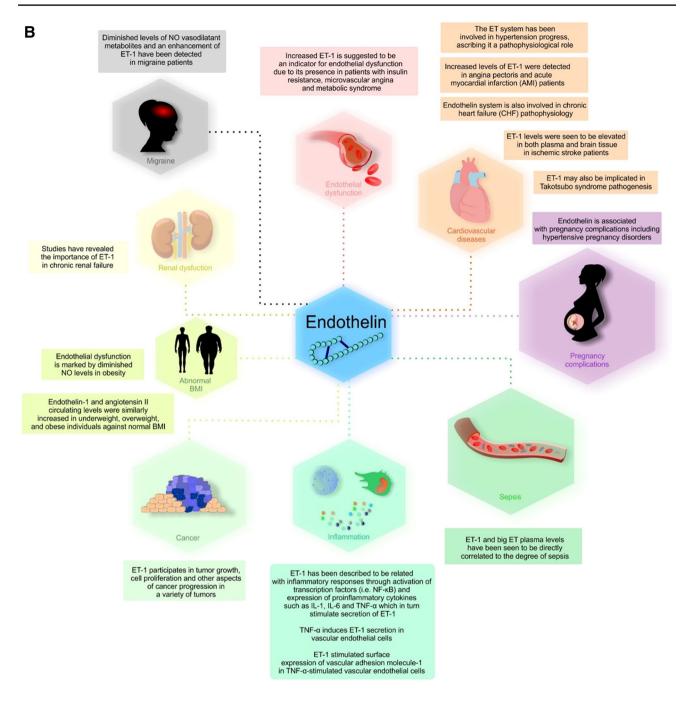


Fig. 1 (continued)

Dual AT1/ETA receptor antagonist Preclinical studies demonstrated how the combination of angiotensin AT1 and ETA receptor antagonist blockage resulted in more effective in reducing BP in hypertension animal models than each antagonist alone [575]. Thus, sparsentan (BMS-346567) is an orally active dual AT1/ETA antagonist, which presents a similar increased affinity for both receptors. Preclinical data seem to indicate an effective blockage of direct vascular actions of both vasoconstricting molecules; however, their effects on downstream signalling and side effects

comparison are not yet apparent [576]. Phase II trials with sparsentan are being conducted to evaluate focal segmental glomerulosclerosis treatment [577].

ETA antagonist and PDE5 inhibitor combination Usually, monotherapy utilising ET receptor antagonists or PDE5 inhibitors is standard when treating PAH patients [578]; however, preclinical studies have revealed that a dual combination may be advantagenous [579]. A murine study stated a mitigation of ET-1 constricted isolated rat pulmonary



arteries after treatment with ETA antagonist ambrisentan and PDE5 inhibitor tadalafil [579]. Translational studies reported a reduced risk of clinical-failure events when compared to either monotherapy treatment [563, 580]. Nevertheless, increased frequency of undesirable adverse effects appeared with the inhibitor combination therapy, possibly being related to ETB receptor activation [581].

The aim of personalised medicine is to adjust drug therapies and individualise patient treatment. SNP detection may result in an association between the patients' genetics and a target against the presumed disease trigger. Gupta et al. determined an ordinary A/G SNP (rs9349379) in the Phosphatase And Actin Regulator 1 (PHACTR1) gene, which regulated edn1 gene expression of ET-1 along with being involved in endothelial cell survival and tubule formation [565]. They speculated that big ET-1 amounts would be more elevated in patients with the minor and most common allele (G/G) [582], intermediate in those with A/G and minimal with A/A at rs9349379, and revealed a considerable relation between G genotype and an increase of 20% in plasma levels. Thus, individuals with G/G allele would present increased ET-1 levels, leading to enhanced vasoconstriction and contributing to angina and vasospasm in coronary artery disease patients [565]. Furthermore, G/G SNP allele patients are more susceptible to ETA antagonists than A/G or A/A. Thus, as demonstrated, SNP testing would be valuable in allowing patient stratification to allocate them with the optimal therapy and personalising endothelin treatment within this field.

The involvement of endothelin-1 in physiologic processes and diseases relevant for predictive diagnostics targeted prevention and personalisation of treatment algorithms are summarised in Fig. 1A and B.

Author contributions O.G. has concepted and coordinated the study. The manuscript was drafted by A.T.C. and B.L. M.S. has concepted and created illustrations. P.K. has contributed with his expertise in cancer research and prevention. FAG has contributed to the final version. All authors have read and agreed to the published version of the manuscript.

Funding Open Access funding enabled and organized by Projekt DEAL. Open Access funding enabled and organised by Projekt DEAL. The present study was supported by the LISPER project (grant Nr. 313011V446) in a bilateral agreement with the European Association for Predictive, Preventive and Personalised Medicine.

Data availability Not applicable.

Code availability Not applicable.



Compliance with ethical standards

Ethics approval Not applicable.

Consent to participate Not applicable.

Consent for publication Not applicable.

Conflict of interest The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- Son WM, Sung KD, Bharath LP, Choi KJ, Park SY. Combined exercise training reduces blood pressure, arterial stiffness, and insulin resistance in obese prehypertensive adolescent girls. Clin Exp Hypertens. 2017;39:546–52. https://doi.org/10.1080/10641 963.2017.1288742.
- Ferrari L, Vicenzi M, Tarantini L, Barretta F, Sironi S, Baccarelli AA, Guazzi M, Bollati V. Effects of physical exercise on endothelial function and DNA methylation. Int J Environ Res Public Health. 2019; 16. https://doi.org/10.3390/ijerph16142530
- Yammine L, Kang DH, Baun MM, Meininger JC. Endothelin-1 and psychosocial risk factors for cardiovascular disease: a systematic review. Psychosom Med. 2014;76:109–21. https://doi. org/10.1097/psy.0000000000000026.
- Zhang H, Cao Y, Pei H, Wang H, Ma L, Wang Z, Diao X, Yang Y, Liu N, Wei Y, et al. Shenmayizhi formula combined with Ginkgo extract tablets for the treatment of vascular dementia: a randomized, double-blind, controlled trial. Evid Based Complement Alternat Med. 2020;2020:8312347. https://doi.org/10.1155/2020/8312347.
- Lekontseva O, Chakrabarti S, Davidge ST. Endothelin in the female vasculature: a role in aging? Am J Physiol Regul Integr Comp Physiol. 2010;298:R509–16. https://doi.org/10.1152/ajpre gu.00656.2009.
- de Oliveira AA, Nunes KP. Hypertension and erectile dysfunction: breaking down the challenges. Am J Hypertens. 2020;34:134–42. https://doi.org/10.1093/ajh/hpaa143.
- Kucera R, Pecen L, Topolcan O, Dahal AR, Costigliola V, Giordano FA, Golubnitschaja O. Prostate cancer management: long-term beliefs, epidemic developments in the early twenty-first century and 3PM dimensional solutions. EPMA J. 2020;11(3):399-418. https://doi.org/10.1007/ s13167-020-00214-1.
- 8. Haynes WG, Webb DJ. Endothelin as a regulator of cardiovascular function in health and disease. J Hypertens. 1998;16(8):1081–98. https://doi.org/10.1097/00004872-199816080-00001.

- Sava RI, March KL, Pepine CJ. Hypertension in pregnancy: taking cues from pathophysiology for clinical practice. Clin Cardiol. 2018;41:220–7. https://doi.org/10.1002/clc.22892.
- Qu H, Khalil RA. Vascular mechanisms and molecular targets in hypertensive pregnancy and preeclampsia. Am J Physiol Heart Circ Physiol. 2020;319:H661–81. https://doi.org/10.1152/ajphe art.00202.2020.
- Bakrania BA, Spradley FT, Drummond HA, LaMarca B, Ryan MJ, Granger JP. Preeclampsia: linking placental ischemia with maternal endothelial and vascular dysfunction. Compr Physiol. 2020;11:1315–49. https://doi.org/10.1002/cphy.c200008.
- Square TA, Jandzik D, Massey JL, Romášek M, Stein HP, Hansen AW, Purkayastha A, Cattell MV, Medeiros DM. Evolution of the endothelin pathway drove neural crest cell diversification. Nature. 2020;585:563–8. https://doi.org/10.1038/s41586-020-2720-z.
- Avishai E, Yeghiazaryan K, Golubnitschaja O. Impaired wound healing: facts and hypotheses for multi-professional considerations in predictive, preventive and personalised medicine. EPMA J. 2017;8:23–33. https://doi.org/10.1007/s13167-017-0081-y.
- Khimji AK, Rockey DC. Endothelin and hepatic wound healing. Pharmacol Res. 2011;63:512–8. https://doi.org/10.1016/j.phrs. 2011.03.005.
- Woo KY. Chronic wound-associated pain, psychological stress, and wound healing. Surg Technol Int. 2012;22:57–65.
- Stolzenburg-Veeser L, Golubnitschaja O. Mini-encyclopaedia of the wound healing - Opportunities for integrating multi-omic approaches into medical practice. J Proteom. 2018;188:71–84. https://doi.org/10.1016/j.jprot.2017.07.017.
- Bryche B, Le Bourhis M, Congar P, Martin C, Rampin O, Meunier N. Endothelin impacts on olfactory processing in rats. Behav Brain Res. 2019;362:1–6. https://doi.org/10.1016/j.bbr.2018.12.048.
- Konieczka K, Ritch R, Traverso CE, Kim DM, Kook MS, Gallino A, Golubnitschaja O, Erb C, Reitsamer HA, Kida T, et al. Flammer syndrome. EPMA J. 2014;5:11. https://doi.org/10.1186/ 1878-5085-5-11.
- Golubnitschaja O, Flammer J. Individualised patient profile: clinical utility of Flammer syndrome phenotype and general lessons for predictive, preventive and personalised medicine. EPMA J. 2018;9:15–20. https://doi.org/10.1007/s13167-018-0127-9.
- Golubnitschaja O. Feeling cold and other underestimated symptoms in breast cancer: anecdotes or individual profiles for advanced patient stratification? EPMA J. 2017;8:17–22. https:// doi.org/10.1007/s13167-017-0086-6.
- Goncharenko V, Bubnov R, Polivka J Jr, Zubor P, Biringer K, Bielik T, Kuhn W, Golubnitschaja O. Vaginal dryness: individualised patient profiles, risks and mitigating measures. EPMA J. 2019;10(1):73–9. https://doi.org/10.1007/s13167-019-00164-3.
- Kunin A, Sargheini N, Birkenbihl C, Moiseeva N, Fröhlich H, Golubnitschaja O. Voice perturbations under the stress overload in young individuals: phenotyping and suboptimal health as predictors for cascading pathologies. EPMA J. 2020;11(4):1–11. https://doi.org/10.1007/s13167-020-00229-8.
- Kunin A, Polivka J Jr, Moiseeva N, Golubnitschaja O. "Dry mouth" and "Flammer" syndromes-neglected risks in adolescents and new concepts by predictive, preventive and personalised approach. EPMA J. 2018;9(3):307–17. https://doi.org/10. 1007/s13167-018-0145-7.
- Golubnitschaja O. Flammer Syndrome From Phenotype to Associated Pathologies, Prediction, Prevention and Personalisation V.11, 2019 (eBook). https://doi.org/10.1007/ 978-3-030-13550-8.
- Harman D. Aging and disease: extending functional life span. Ann N Y Acad Sci. 1996;786:321–36. https://doi.org/10.1111/j. 1749-6632.1996.tb39074.x.

- Csiszar A, Wang M, Lakatta EG, Ungvari Z. Inflammation and endothelial dysfunction during aging: role of NF-kappaB. J Appl Physiol. 1985;2008(105):1333–41. https://doi.org/10.1152/jappl physiol.90470.2008.
- Michaud M, Balardy L, Moulis G, Gaudin C, Peyrot C, Vellas B, Cesari M, Nourhashemi F. Proinflammatory cytokines, aging, and age-related diseases. J Am Med Dir Assoc. 2013;14:877–82. https://doi.org/10.1016/j.jamda.2013.05.009.
- 28. Donato AJ, Gano LB, Eskurza I, Silver AE, Gates PE, Jablonski K, Seals DR. Vascular endothelial dysfunction with aging: endothelin-1 and endothelial nitric oxide synthase. Am J Physiol Heart Circ Physiol. 2009;297:H425–32. https://doi.org/10.1152/ajpheart.00689.2008.
- 29. Eroglu E, Kocyigit I, Lindholm B. The endothelin system as target for therapeutic interventions in cardiovascular and renal disease. Clin Chim Acta. 2020;506:92–106. https://doi.org/10.1016/j.cca.2020.03.008.
- Jankowich M, Choudhary G. Endothelin-1 levels and cardiovascular events. Trends Cardiovasc Med. 2020;30:1–8. https://doi. org/10.1016/j.tcm.2019.01.007.
- Pacher R, Stanek B, Hülsmann M, Koller-Strametz J, Berger R, Schuller M, Hartter E, Ogris E, Frey B, Heinz G, et al. Prognostic impact of big endothelin-1 plasma concentrations compared with invasive hemodynamic evaluation in severe heart failure. J Am Coll Cardiol. 1996;27:633–41. https://doi.org/10.1016/0735-1097(95)00520-x.
- Beck DT, Casey DP, Martin JS, Emerson BD, Braith RW. Exercise training improves endothelial function in young prehypertensives. Exp Biol Med (Maywood). 2013;238:433–41. https://doi.org/10.1177/1535370213477600.
- Aronson D, Burger AJ. Neurohumoral activation and ventricular arrhythmias in patients with decompensated congestive heart failure: role of endothelin. Pacing Clin Electrophysiol. 2003;26:703–10. https://doi.org/10.1046/j.1460-9592.2003.00120 x
- Yang LL, Gros R, Kabir MG, Sadi A, Gotlieb AI, Husain M, Stewart DJ. Conditional cardiac overexpression of endothelin-1 induces inflammation and dilated cardiomyopathy in mice. Circulation. 2004;109:255–61. https://doi.org/10.1161/01.Cir.00001 05701.98663.D4.
- Polivka J Jr, Polivka J, Pesta M, Rohan V, Celedova L, Mahajani S, Topolcan O, Golubnitschaja O. Risks associated with the stroke predisposition at young age: facts and hypotheses in light of individualized predictive and preventive approach. EPMA J. 2019;10(1):81–99. https://doi.org/10.1007/s13167-019-00162-5.
- 36. Yudkin JS, Eringa E, Stehouwer CD. "Vasocrine" signalling from perivascular fat: a mechanism linking insulin resistance to vascular disease. Lancet. 2005;365:1817–20. https://doi.org/10.1016/s0140-6736(05)66585-3.
- Sarafidis PA, Bakris GL. Insulin and endothelin: an interplay contributing to hypertension development? J Clin Endocrinol Metab. 2007;92:379–85. https://doi.org/10.1210/jc.2006-1819.
- Higashi Y, Sasaki S, Nakagawa K, Kimura M, Noma K, Sasaki S, Hara K, Matsuura H, Goto C, Oshima T, et al. Low body mass index is a risk factor for impaired endothelium-dependent vasodilation in humans: role of nitric oxide and oxidative stress. J Am Coll Cardiol. 2003;42:256–63. https://doi.org/10.1016/s0735-1097(03)00630-2.
- Palmer JC, Barker R, Kehoe PG, Love S. Endothelin-1 is elevated in Alzheimer's disease and upregulated by amyloid-β.
 J Alzheimers Dis. 2012;29:853–61. https://doi.org/10.3233/jad-2012-111760.
- D'Haeseleer M, Beelen R, Fierens Y, Cambron M, Vanbinst A-M, Verborgh C, Demey J, De Keyser J. Cerebral hypoperfusion in multiple sclerosis is reversible and mediated by endothelin-1.



Proc Natl Acad Sci USA. 2013;110:5654–8. https://doi.org/10.1073/pnas.1222560110.

- Ranno E, D'Antoni S, Spatuzza M, Berretta A, Laureanti F, Bonaccorso CM, Pellitteri R, Longone P, Spalloni A, Iyer AM, et al. Endothelin-1 is over-expressed in amyotrophic lateral sclerosis and induces motor neuron cell death. Neurobiol Dis. 2014;65:160–71. https://doi.org/10.1016/j.nbd.2014.01.002.
- 42. Pek SLT, Lim SC, Ang K, Kwan PY, Tang WE, Sum CF, Tavintharan S. Endothelin-1 predicts incident diabetic peripheral neuropathy in Type 2 Diabetes: a cohort study. Eur J Endocrinol. 2020;182(4):429–38. https://doi.org/10.1530/EJE-19-0523.
- 43. Fathy A, Abdelrahman ÅE. EZH2, Endothelin-1, and CD34 as biomarkers of aggressive cervical squamous cell carcinoma: an immunohistochemical study. Turk Patoloji Derg. 2018;34:150–7. https://doi.org/10.5146/tjpath.2018.01425.
- Nelson J, Bagnato A, Battistini B, Nisen P. The endothelin axis: emerging role in cancer. Nat Rev Cancer. 2003;3:110–6. https://doi.org/10.1038/nrc990.
- 45. Bagnato A, Spinella F, Rosanò L. Emerging role of the endothelin axis in ovarian tumor progression. Endocr Relat Cancer. 2005;12:761–72. https://doi.org/10.1677/erc.1.01077.
- Chellini L, Caprara V, Spadaro F, Sestito R, Bagnato A, Rosanò L. Regulation of extracellular matrix degradation and metastatic spread by IQGAP1 through endothelin-1 receptor signalling in ovarian cancer. Matrix Biol. 2019;81:17–33. https://doi.org/10.1016/j.matbio.2018.10.005.
- Cox TR, Rumney RMH, Schoof EM, Perryman L, Høye AM, Agrawal A, Bird D, Latif NA, Forrest H, Evans HR, et al. The hypoxic cancer secretome induces pre-metastatic bone lesions through lysyl oxidase. Nature. 2015;522:106–10. https://doi.org/ 10.1038/nature14492.
- Vanharanta S. A hypoxic ticket to the bone metastatic niche. Breast Cancer Res. 2015;17:122. https://doi.org/10.1186/ s13058-015-0635-7.
- Gu X, Han S, Cui M, Xue J, Ai L, Sun L, Zhu X, Wang Y, Liu C. Knockdown of endothelin receptor B inhibits the progression of triple-negative breast cancer. Ann N Y Acad Sci. 2019;1448:5– 18. https://doi.org/10.1111/nyas.14039.
- Sørby LA, Kleiveland CR, Andersen SN, Bukholm IR, Jacobsen MB. The endothelin axis in the metastatic process of colon carcinoma. Anticancer Res. 2011;31:861–9.
- Hasue F, Kuwaki T, Yamada H, Fukuda Y, Shimoyama M. Inhibitory actions of endothelin-1 on pain processing. J Cardiovasc Pharmacol. 2004;44(Suppl 1):S318–20. https://doi.org/10.1097/01.fjc.0000166271.40044.0c.
- Tocci P, Cianfrocca R, Sestito R, Rosanò L, Di Castro V, Blandino G, Bagnato A. Endothelin-1 axis fosters YAPinduced chemotherapy escape in ovarian cancer. Cancer Lett. 2020;492:84–95. https://doi.org/10.1016/j.canlet.2020.08.026.
- Rehberg B, Mathivon S, Combescure C, Mercier Y, Savoldelli GL. Prediction of acute postoperative pain following breast cancer surgery using the pain sensitivity questionnaire: a cohort study. Clin J Pain. 2017;33:57–66. https://doi.org/10.1097/ajp. 000000000000000380.
- Hans G, Deseure K, Robert D, De Hert S. Neurosensory changes in a human model of endothelin-1 induced pain: a behavioral study. Neurosci Lett. 2007;418:117–21. https://doi.org/10.1016/j. neulet.2007.03.008.
- Lutz BM, Wu S, Gu X, Atianjoh FE, Li Z, Fox BM, Pollock DM, Tao YX. Endothelin type A receptors mediate pain in a mouse model of sickle cell disease. Haematologica. 2018;103:1124–35. https://doi.org/10.3324/haematol.2017.187013.
- 56. Borrull A, Allard B, Wijkhuisen A, Herbet A, Lamourette P, Birouk W, Leiber D, Tanfin Z, Ducancel F, Boquet D, et al. Rendomab B4, a monoclonal antibody that discriminates the human endothelin B receptor of melanoma cells and inhibits

- their migration. MAbs. 2016;8:1371–85. https://doi.org/10.1080/19420862.2016.1208865.
- Chen Y, Su X, Qin Q, Yu Y, Jia M, Kong L, Zhang H, Li H. Metformin inhibited homocysteine-induced upregulation of endothelin receptors through the Sirt1/NF-kappaB signaling pathway in vascular smooth muscle cells. Vascul Pharmacol. 2020;124:106613.
- Mahdi MR, Georges RB, Ali DM, Bedeer RF, Eltahry HM, Gabr AHZ, Berger MR. Modulation of the endothelin system in colorectal cancer liver metastasis: influence of epigenetic mechanisms? Front Pharmacol. 2020;11:180. https://doi.org/10.3389/fphar.2020.00180.
- Naya M, Aikawa T, Manabe O, Obara M, Koyanagawa K, Katoh C, Tamaki N. Elevated serum endothelin-1 is an independent predictor of coronary microvascular dysfunction in non-obstructive territories in patients with coronary artery disease. Heart Vessels. 2021. https://doi.org/10.1007/s00380-020-01767-x.
- Badagliacca R, Sciomer S, Petrosillo N. Endothelin receptor antagonists for pulmonary arterial hypertension and COVID-19: Friend or foe? J Heart Lung Transplant. 2020;39:729–30. https:// doi.org/10.1016/j.healun.2020.04.007.
- Guo YR, Cao QD, Hong ZS, Tan YY, Chen SD, Jin HJ, Tan KS, Wang DY, Yan Y. The origin, transmission and clinical therapies on coronavirus disease 2019 (COVID-19) outbreak - an update on the status. Mil Med Res. 2020;7:11. https://doi.org/10.1186/ s40779-020-00240-0.
- Karmouty-Quintana H, Thandavarayan RA, Keller SP, Sahay S, Pandit LM, Akkanti B. Emerging mechanisms of pulmonary vasoconstriction in SARS-CoV-2-induced acute respiratory distress syndrome (ARDS) and potential therapeutic targets. Int J Mol Sci. 2020; 21. https://doi.org/10.3390/ijms21218081
- Mihalopoulos M, Dogra N, Mohamed N, Badani K, Kyprianou N. COVID-19 and kidney disease: molecular determinants and clinical implications in renal cancer. Eur Urol Focus. 2020;6:1086–96. https://doi.org/10.1016/j.euf.2020.06.002.
- Hickey KA, Rubanyi G, Paul RJ, Highsmith RF. Characterization of a coronary vasoconstrictor produced by cultured endothelial cells. Am J Physiol. 1985;248:C550–6. https://doi.org/10.1152/ ajpcell.1985.248.5.C550.
- O'Brien RF, Robbins RJ, McMurtry IF. Endothelial cells in culture produce a vasoconstrictor substance. J Cell Physiol. 1987;132:263–70. https://doi.org/10.1002/jcp.1041320210.
- Yanagisawa M, Kurihara H, Kimura S, Tomobe Y, Kobayashi M, Mitsui Y, Yazaki Y, Goto K, Masaki T. A novel potent vasoconstrictor peptide produced by vascular endothelial cells. Nature. 1988;332:411–5. https://doi.org/10.1038/332411a0.
- Kloog Y, Ambar I, Sokolovsky M, Kochva E, Wollberg Z, Bdolah A. Sarafotoxin, a novel vasoconstrictor peptide: phosphoinositide hydrolysis in rat heart and brain. Science. 1988;242:268–70. https://doi.org/10.1126/science.2845579.
- Takasaki C, Tamiya N, Bdolah A, Wollberg Z, Kochva E. Sarafotoxins S6: several isotoxins from Atractaspis engaddensis (burrowing asp) venom that affect the heart. Toxicon. 1988;26:543–8. https://doi.org/10.1016/0041-0101(88)90234-6.
- Inoue A, Yanagisawa M, Kimura S, Kasuya Y, Miyauchi T, Goto K, Masaki T. The human endothelin family: three structurally and pharmacologically distinct isopeptides predicted by three separate genes. Proc Natl Acad Sci U S A. 1989;86:2863–7. https://doi.org/10.1073/pnas.86.8.2863.
- Arai H, Hori S, Aramori I, Ohkubo H, Nakanishi S. Cloning and expression of a cDNA encoding an endothelin receptor. Nature. 1990;348:730–2. https://doi.org/10.1038/348730a0.
- 71. Sakurai T, Yanagisawa M, Takuwa Y, Miyazaki H, Kimura S, Goto K, Masaki T. Cloning of a cDNA encoding a non-isopeptide-selective subtype of the endothelin receptor. Nature. 1990;348:732–5. https://doi.org/10.1038/348732a0.



- Houde M, Desbiens L, D'Orléans-Juste P. Endothelin-1: biosynthesis, signaling and vasoreactivity. Adv Pharmacol. 2016;77:143–75. https://doi.org/10.1016/bs.apha.2016.05.002.
- 73. <(1992). IL-4 and IL-2 promote human T-cell proliferation through symmetrical but independent pathways. Clinical Immunology and Immunopathology, 64(3), 210–217. doi1.pdf>.
- Barton M, Yanagisawa M. Endothelin: 20 years from discovery to therapy. Can J Physiol Pharmacol. 2008;86:485–98. https:// doi.org/10.1139/y08-059.
- Barton M, Tharaux P-L. Endothelin and the podocyte. Clin Kidney J. 2012;5:17–27. https://doi.org/10.1093/ckj/sfs001.
- Barton M, Yanagisawa M. Endothelin: 30 years from discovery to therapy. Hypertension. 2019;74:1232–65. https://doi.org/10. 1161/hypertensionaha.119.12105.
- Warner TD, Allcock GH, Corder R, Vane JR. Use of the endothelin antagonists BQ-123 and PD 142893 to reveal three endothelin receptors mediating smooth muscle contraction and the release of EDRF. Br J Pharmacol. 1993;110:777–82. https://doi.org/10.1111/j.1476-5381.1993.tb13879.x.
- Malek AM, Zhang J, Jiang J, Alper SL, Izumo S. Endothelin-1 gene suppression by shear stress: pharmacological evaluation of the role of tyrosine kinase, intracellular calcium, cytoskeleton, and mechanosensitive channels. J Mol Cell Cardiol. 1999;31:387–99. https://doi.org/10.1006/jmcc.1998.0873.
- Masaki T, Sawamura T. Endothelin and endothelial dysfunction. Proceedings of the Japan Academy. Series B, Physical and biological sciences. 2006; 82:17–24. https://doi.org/10.2183/pjab. 82.17
- Blais V, Fugère M, Denault JB, Klarskov K, Day R, Leduc R. Processing of proendothelin-1 by members of the subtilisin-like pro-protein convertase family. FEBS Lett. 2002;524:43–8. https://doi.org/10.1016/s0014-5793(02)02998-8.
- Shimada K, Matsushita Y, Wakabayashi K, Takahashi M, Matsubara A, Iijima Y, Tanzawa K. Cloning and functional expression of human endothelin-converting enzyme cDNA. Biochem Biophys Res Commun. 1995;207:807–12. https://doi.org/10.1006/bbrc.1995.1258.
- 82. Emoto N, Yanagisawa M. Endothelin-converting enzyme-2 is a membrane-bound, phosphoramidon-sensitive metalloprotease with acidic pH optimum. J Biol Chem. 1995;270:15262–8. https://doi.org/10.1074/jbc.270.25.15262.
- 83. Corder R. Identity of Endothelin-converting enzyme and other targets for the therapeutic regulation of endothelin biosynthesis. In; 2001, 35–67.
- Battistini B, D'Orléans-Juste P, Sirois P. Endothelins: circulating plasma levels and presence in other biologic fluids. Lab Invest. 1993;68:600–28.
- Masaki T. Historical review: endothelin. Trends Pharmacol Sci. 2004;25:219–24. https://doi.org/10.1016/j.tips.2004.02.008.
- Stow LR, Gumz ML, Lynch IJ, Greenlee MM, Rudin A, Cain BD, Wingo CS. Aldosterone modulates steroid receptor binding to the endothelin-1 gene (edn1). J Biol Chem. 2009;284:30087– 96. https://doi.org/10.1074/jbc.M109.030718.
- 87. Todd-Turla KM, Zhu XL, Shu X, Chen M, Yu T, Smart A, Killen PD, Fejes-Toth G, Briggs JP, Schnermann JB. Synthesis and secretion of endothelin in a cortical collecting duct cell line. Am J Physiol. 1996;271:F330–9. https://doi.org/10.1152/ajprenal. 1996.271.2.F330.
- 88. Saito S, Hirata Y, Imai T, Marumo F. Autocrine regulation of the endothelin-1 gene in rat endothelial cells. J Cardiovasc Pharmacol. 1995;26(Suppl 3):S84–7.
- Pégorier S, Arouche N, Dombret MC, Aubier M, Pretolani M. Augmented epithelial endothelin-1 expression in refractory asthma. J Allergy Clin Immunol. 2007;120:1301–7. https://doi. org/10.1016/j.jaci.2007.09.023.

- Ergul A, Grubbs A, Zhang Y, Spinale F. Selective upregulation of endothelin converting enzyme-1a in the human failing heart. J Cardiac Fail. 2000;6:314–20. https://doi.org/10.1054/jcaf.2000. 19227.
- 91. Aharinejad S, Krenn K, Paulus P, Schäfer R, Zuckermann A, Grimm M, Abraham D. Differential role of TGF-beta1/bFGF and ET-1 in graft fibrosis in heart failure patients. Am J Transplant. 2005;5:2185–92. https://doi.org/10.1111/j.1600-6143.2005.
- Strzalka-Mrozik B, Nowak A, Gola J, Kowalczyk M, Kapral M, Mazurek U. Factors associated with changes in endothelin-1 gene expression in patients with diabetic retinopathy in type 2 diabetes mellitus. Mol Vis. 2010;16:1272–9.
- 93. Donckier JE, Michel L, Delos M, Havaux X, Van Beneden R. Interrelated overexpression of endothelial and inducible nitric oxide synthases, endothelin-1 and angiogenic factors in human papillary thyroid carcinoma. Clin Endocrinol (Oxf). 2006;64:703–10. https://doi.org/10.1111/j.1365-2265.2006. 02535.x.
- 94. Emori T, Hirata Y, Imai T, Ohta K, Kanno K, Eguchi S, Marumo F. Cellular mechanism of thrombin on endothelin-1 biosynthesis and release in bovine endothelial cell. Biochem Pharmacol. 1992;44:2409–11. https://doi.org/10.1016/0006-2952(92)90687-e.
- Kurihara H, Yoshizumi M, Sugiyama T, Takaku F, Yanagisawa M, Masaki T, Hamaoki M, Kato H, Yazaki Y. Transforming growth factor-beta stimulates the expression of endothelin mRNA by vascular endothelial cells. Biochem Biophys Res Commun. 1989;159:1435–40. https://doi.org/10.1016/0006-291x(89)92270-5.
- Yoshimoto S, Ishizaki Y, Sasaki T, Murota S. Effect of carbon dioxide and oxygen on endothelin production by cultured porcine cerebral endothelial cells. Stroke. 1991;22:378–83. https://doi.org/10.1161/01.str.22.3.378.
- 97. Verhaar MC, Strachan FE, Newby DE, Cruden NL, Koomans HA, Rabelink TJ, Webb DJ. Endothelin-A receptor antagonist-mediated vasodilatation is attenuated by inhibition of nitric oxide synthesis and by endothelin-B receptor blockade. Circulation. 1998;97:752–6. https://doi.org/10.1161/01.cir.97.8.752.
- Haynes WG, Ferro CJ, O'Kane KP, Somerville D, Lomax CC, Webb DJ. Systemic endothelin receptor blockade decreases peripheral vascular resistance and blood pressure in humans. Circulation. 1996;93:1860–70. https://doi.org/10.1161/01.cir. 93.10.1860.
- Kurihara Y, Kurihara H, Suzuki H, Kodama T, Maemura K, Nagai R, Oda H, Kuwaki T, Cao WH, Kamada N, et al. Elevated blood pressure and craniofacial abnormalities in mice deficient in endothelin-1. Nature. 1994;368:703–10. https://doi.org/10.1038/368703a0.
- 100. Kuwaki T, Kurihara H, Cao WH, Kurihara Y, Unekawa M, Yazaki Y, Kumada M. Physiological role of brain endothelin in the central autonomic control: from neuron to knockout mouse. Prog Neurobiol. 1997;51:545–79. https://doi.org/10.1016/s0301-0082(96)00063-9.
- Kurihara H, Kurihara Y, Yazaki Y. Lessons from gene deletion of endothelin systems. In; 2001, 141–54.
- 102. Baynash AG, Hosoda K, Giaid A, Richardson JA, Emoto N, Hammer RE, Yanagisawa M. Interaction of endothelin-3 with endothelin-B receptor is essential for development of epidermal melanocytes and enteric neurons. Cell. 1994;79:1277–85. https://doi.org/10.1016/0092-8674(94)90018-3.
- 103. Gardiner SM, Kemp PA, Bennett T. Inhibition by phosphor amidon of the regional haemodynamic effects of proendothelin-2 and -3 in conscious rats. Br J Pharmacol. 1992;107:584–90.



- 104. Uchide T, Masuda H, Lee YS, Makiyama Y, Mitsui Y, Saida K. Fluctuating gene expression and localized cellular distribution of vasoactive intestinal contractor (VIC) in mouse uterus. J Histochem Cytochem. 2000;48:699–707. https://doi.org/10.1177/002215540004800514.
- 105. de la Monte SM, Quertermous T, Hong CC, Bloch KD. Regional and maturation-associated expression of endothelin 2 in rat gastrointestinal tract. J Histochem Cytochem. 1995;43:203–9. https://doi.org/10.1177/43.2.7822776.
- Takizawa S, Uchide T, Adur J, Kozakai T, Kotake-Nara E, Quan J, Saida K. Differential expression of endothelin-2 along the mouse intestinal tract. J Mol Endocrinol. 2005;35:201–9. https://doi.org/10.1677/jme.1.01787.
- 107. Masuo Y, Ishikawa Y, Kozakai T, Uchide T, Komatsu Y, Saida K. Vasoactive intestinal contractor/endothelin-2 gene expression in the murine central nervous system. Biochem Biophys Res Commun. 2003;300:661–8. https://doi.org/10.1016/S0006-291X(02)02872-3.
- 108. Palanisamy GS, Cheon YP, Kim J, Kannan A, Li Q, Sato M, Mantena SR, Sitruk-Ware RL, Bagchi MK, Bagchi IC. A novel pathway involving progesterone receptor, endothelin-2, and endothelin receptor B controls ovulation in mice. Mol Endocrinol. 2006;20:2784–95. https://doi.org/10.1210/me. 2006-0093.
- Karet FE, Davenport AP. Localization of endothelin peptides in human kidney. Kidney Int. 1996;49:382–7. https://doi.org/10. 1038/ki.1996.56.
- Plumpton C, Champeney R, Ashby MJ, Kuc RE, Davenport AP. Characterization of endothelin isoforms in human heart: endothelin-2 demonstrated. J Cardiovasc Pharmacol. 1993;22(Suppl 8):S26–8. https://doi.org/10.1097/00005344-199322008-00009.
- Howard PG, Plumpton C, Davenport AP. Anatomical localization and pharmacological activity of mature endothelins and their precursors in human vascular tissue. J Hypertens. 1992;10:1379–86. https://doi.org/10.1097/00004872-199211000-00010.
- 112. Marciniak SJ, Plumpton C, Barker PJ, Huskisson NS, Davenport AP. Localization of immunoreactive endothelin and proendothelin in the human lung. Pulm Pharmacol. 1992;5:175–82. https://doi.org/10.1016/0952-0600(92)90038-i.
- Matsumoto H, Suzuki N, Kitada C, Fujino M. Endothelin family peptides in human plasma and urine: their molecular forms and concentrations. Peptides. 1994;15:505–10. https://doi.org/10.1016/0196-9781(94)90213-5.
- 114. Suzuki N, Matsumoto H, Miyauchi T, Kitada C, Tsuda M, Goto K, Masaki T, Fujino M. Sandwich-enzyme immunoassays for endothelin family peptides. J Cardiovasc Pharmacol. 1991;17(Suppl 7):S420–2. https://doi.org/10.1097/00005344-199100177-00119.
- Davenport AP, Maguire JJ. Endothelin. Handb Exp Pharmacol. 2006:295–329. https://doi.org/10.1007/3-540-32967-6_9
- Ishida N, Tsujioka K, Tomoi M, Saida K, Mitsui Y. Differential activities of two distinct endothelin family peptides on ileum and coronary artery. FEBS Lett. 1989;247:337–40. https://doi.org/10. 1016/0014-5793(89)81365-1.
- 117. Chang I, Bramall AN, Baynash AG, Rattner A, Rakheja D, Post M, Joza S, McKerlie C, Stewart DJ, McInnes RR, et al. Endothelin-2 deficiency causes growth retardation, hypothermia, and emphysema in mice. J Clin Invest. 2013;123:2643–53. https://doi.org/10.1172/jci66735.
- 118. Haq A, Kayali M, Hammami MM, Jaroudi K, al-Sedairy ST. Immunoreactive endothelin-1, endothelin-2 and big endothelin-1 in follicular fluids of women undergoing ovulation induction for in-vitro fertilization. Hum Reprod. 1996; 11:269–73. https://doi. org/10.1093/humrep/11.2.269
- Antonio J. Gene Expression of Endothelin-1 in the Porcine Ovary: Follicular Development. Biol Reprod. 2000;63:1377–82.

- https://doi.org/10.1043/0006-3363(2000)063%3c1377: GEOEIT%3e2.0.CO;2.
- 120. Shirasuna K, Watanabe S, Oki N, Wijayagunawardane MP, Matsui M, Ohtani M, Miyamoto A. A cooperative action of endothelin-1 with prostaglandin F(2alpha) on luteal function in the cow. Domest Anim Endocrinol. 2006;31:186–96. https://doi.org/10.1016/j.domaniend.2005.10.004.
- 121. Uchide T, Masuda H, Mitsui Y, Saida K. Gene expression of vasoactive intestinal contractor/endothelin-2 in ovary, uterus and embryo: comprehensive gene expression profiles of the endothelin ligand-receptor system revealed by semi-quantitative reverse transcription-polymerase chain reaction analysis in adult mouse tissues and during late embryonic development. J Mol Endocrinol. 1999;22:161–71. https://doi.org/10.1677/jme.0.0220161.
- Ko C, Gieske MC, Al-Alem L, Hahn Y, Su W, Gong MC, Iglarz M, Koo Y. Endothelin-2 in ovarian follicle rupture. Endocrinology. 2006;147:1770–9. https://doi.org/10.1210/en.2005-1228.
- 123. Elferink JG, de Koster BM. The effect of endothelin-2 (ET-2) on migration and changes in cytosolic free calcium of neutrophils. Naunyn Schmiedebergs Arch Pharmacol. 1996;353:130–5. https://doi.org/10.1007/bf00168749.
- Grimshaw MJ, Wilson JL, Balkwill FR. Endothelin-2 is a macrophage chemoattractant: implications for macrophage distribution in tumors. Eur J Immunol. 2002;32:2393–400. https://doi.org/10.1002/1521-4141(200209)32:9%3c2393::Aid-immu2393%3e3.0.Co;2-4.
- Davenport AP. International Union of Pharmacology. XXIX.
 Update on endothelin receptor nomenclature. Pharmacol Rev. 2002; 54:219–26. https://doi.org/10.1124/pr.54.2.219
- 126. Gardiner SM, Kemp PA, Compton AM, Bennett T. Coeliac haemodynamic effects of endothelin-1, endothelin-3, proendothelin-1 [1-38] and proendothelin-3 [1-41] in conscious rats. Br J Pharmacol. 1992;106:483–8. https://doi.org/10.1111/j.1476-5381.1992.tb14360.x.
- Lee S, Lin M, Mele A, Cao Y, Farmar J, Russo D, Redman C. Proteolytic processing of big endothelin-3 by the kell blood group protein. Blood. 1999;94:1440–50.
- 128. Clapéron A, Rose C, Gane P, Collec E, Bertrand O, Ouimet T. The Kell protein of the common K2 phenotype is a catalytically active metalloprotease, whereas the rare Kell K1 antigen is inactive. Identification of novel substrates for the Kell protein. J Biol Chem. 2005; 280:21272–83. https://doi.org/10.1074/jbc.M500100200
- 129. Onda H, Ohkubo S, Ogi K, Kosaka T, Kimura C, Matsumoto H, Suzuki N, Fujino M. One of the endothelin gene family, endothelin 3 gene, is expressed in the placenta. FEBS Lett. 1990;261:327–30. https://doi.org/10.1016/0014-5793(90) 80583-5.
- Takahashi K, Ghatei MA, Jones PM, Murphy JK, Lam HC, O'Halloran DJ, Bloom SR. Endothelin in human brain and pituitary gland: comparison with rat. J Cardiovasc Pharmacol. 1991;17(Suppl 7):S101–3. https://doi.org/10.1097/00005344-199100177-00026.
- Davenport AP, Hoskins SL, Kuc RE, Plumpton C. Differential distribution of endothelin peptides and receptors in human adrenal gland. Histochem J. 1996;28:779–89. https://doi.org/10.1007/ bf02272151.
- Matsumoto H, Suzuki N, Onda H, Fujino M. Abundance of endothelin-3 in rat intestine, pituitary gland and brain. Biochem Biophys Res Commun. 1989;164:74–80. https://doi.org/10.1016/ 0006-291x(89)91684-7.
- 133. Shiba R, Sakurai T, Yamada G, Morimoto H, Saito A, Masaki T, Goto K. Cloning and expression of rat preproendothelin-3 cDNA. Biochem Biophys Res Commun. 1992;186:588–94. https://doi.org/10.1016/S0006-291X(05)80849-6.



 Liu Y, Yamada H, Ochi J. Immunocytochemical studies on endothelin in mast cells and macrophages in the rat gastrointestinal tract. Histochem Cell Biol. 1998;109:301–7. https://doi.org/ 10.1007/s004180050230.

- 135. Miyauchi Y, Sakai S, Maeda S, Shimojo N, Watanabe S, Honma S, Kuga K, Aonuma K, Miyauchi T. Increased plasma levels of big-endothelin-2 and big-endothelin-3 in patients with end-stage renal disease. Life Sci. 2012;91:729–32. https://doi.org/10.1016/j.lfs.2012.08.008.
- Gupta RM, Libby P, Barton M. Linking regulation of nitric oxide to endothelin-1: The Yin and Yang of vascular tone in the atherosclerotic plaque. Atherosclerosis. 2020;292:201–3. https://doi. org/10.1016/j.atherosclerosis.2019.11.001.
- Vanhoutte PM, Zhao Y, Xu A, Leung SW. Thirty years of saying no: sources, fate, actions, and misfortunes of the endothelium-derived vasodilator mediator. Circ Res. 2016;119:375–96. https://doi.org/10.1161/circresaha.116.306531.
- Mitsutomi N, Akashi C, Odagiri J, Matsumura Y. Effects of endogenous and exogenous nitric oxide on endothelin-1 production in cultured vascular endothelial cells. Eur J Pharmacol. 1999;364:65–73. https://doi.org/10.1016/s0014-2999(98) 00806-1.
- 139. Oelze M, Knorr M, Kröller-Schön S, Kossmann S, Gottschlich A, Rümmler R, Schuff A, Daub S, Doppler C, Kleinert H, et al. Chronic therapy with isosorbide-5-mononitrate causes endothelial dysfunction, oxidative stress, and a marked increase in vascular endothelin-1 expression. Eur Heart J. 2013;34:3206–16. https://doi.org/10.1093/eurheartj/ehs100.
- 140. Barton M, Haudenschild CC, d'Uscio LV, Shaw S, Münter K, Lüscher TF. Endothelin ETA receptor blockade restores NOmediated endothelial function and inhibits atherosclerosis in apolipoprotein E-deficient mice. Proc Natl Acad Sci U S A. 1998;95:14367–72. https://doi.org/10.1073/pnas.95.24.14367.
- Endemann DH, Schiffrin EL. Endothelial dysfunction. J Am Soc Nephrol. 2004;15:1983–92. https://doi.org/10.1097/01. Asn.0000132474.50966.Da.
- 142. Aflyatumova GN, Nigmatullina RR, Sadykova DI, Chibireva MD, Fugetto F, Serra R. Endothelin-1, nitric oxide, serotonin and high blood pressure in male adolescents. Vasc Health Risk Manag. 2018;14:213–23. https://doi.org/10.2147/VHRM. S170317.
- 143. Głowińska B, Urban M, Hryniewicz A, Peczyńska J, Florys B, Al-Hwish M. Endothelin-1 plasma concentration in children and adolescents with atherogenic risk factors. Kardiol Pol. 2004:61:329–38.
- 144. Tostes RC, Fortes ZB, Callera GE, Montezano AC, Touyz RM, Webb RC, Carvalho MH. Endothelin, sex and hypertension. Clin Sci (Lond). 2008;114:85–97. https://doi.org/10.1042/cs20070169.
- 145. Miller VM, Mulvagh SL. Sex steroids and endothelial function: translating basic science to clinical practice. Trends Pharmacol Sci. 2007;28:263–70. https://doi.org/10.1016/j.tips.2007.04. 004.
- 146. Polderman KH, Stehouwer CD, van Kamp GJ, Dekker GA, Verheugt FW, Gooren LJ. Influence of sex hormones on plasma endothelin levels. Ann Intern Med. 1993;118:429–32. https:// doi.org/10.7326/0003-4819-118-6-199303150-00006.
- 147. David FL, Carvalho MH, Cobra AL, Nigro D, Fortes ZB, Rebouças NA, Tostes RC. Ovarian hormones modulate endothelin-1 vascular reactivity and mRNA expression in DOCA-salt hypertensive rats. Hypertension. 2001;38:692–6. https://doi.org/10.1161/01.hyp.38.3.692.
- 148. Miyauchi T, Yanagisawa M, Iida K, Ajisaka R, Suzuki N, Fujino M, Goto K, Masaki T, Sugishita Y. Age- and sex-related variation of plasma endothelin-1 concentration in normal and

- hypertensive subjects. Am Heart J. 1992;123:1092–3. https://doi.org/10.1016/0002-8703(92)90734-d.
- 149. Polderman KH, Stehouwer CD, van Kamp GJ, Schalkwijk CG, Gooren LJ. Modulation of plasma endothelin levels by the menstrual cycle. Metabolism: clinical and experimental. 2000; 49:648–50. https://doi.org/10.1016/s0026-0495(00)80042-6
- Gohar EY, Yusuf C, Pollock DM. Ovarian hormones modulate endothelin A and B receptor expression. Life Sci. 2016;159:148– 52. https://doi.org/10.1016/j.lfs.2016.01.010.
- 151. Carbonne B, Mignot TM, Tsatsaris V, Ferré F. Changes in plasma and amniotic fluid endothelin levels during pregnancy: facts or artefacts? European Journal of Obstetrics & Gynecology and Reproductive Biology. 1998;76:15–9. https://doi.org/10.1016/ S0301-2115(97)00141-3.
- 152. Lygnos MC, Pappa KI, Papadaki HA, Relakis C, Koumantakis E, Anagnou NP, Eliopoulos GD. Changes in maternal plasma levels of VEGF, bFGF, TGF-beta1, ET-1 and sKL during uncomplicated pregnancy, hypertensive pregnancy and gestational diabetes. Vivo. 2006;20:157–63.
- 153. Morey AK, Razandi M, Pedram A, Hu RM, Prins BA, Levin ER. Oestrogen and progesterone inhibit the stimulated production of endothelin-1. Biochem J. 1998;330(Pt 3):1097–105. https://doi. org/10.1042/bj3301097.
- 154. Wilbert-Lampen U, Seliger C, Trapp A, Straube F, Plasse A. Female sex hormones decrease constitutive endothelin-1 release via endothelial sigma-1/cocaine receptors: an action independent of the steroid hormone receptors. Endothelium. 2005;12:185–91. https://doi.org/10.1080/10623320500227275.
- 155. van Kesteren PJ, Kooistra T, Lansink M, van Kamp GJ, Asscheman H, Gooren LJ, Emeis JJ, Vischer UM, Stehouwer CD. The effects of sex steroids on plasma levels of marker proteins of endothelial cell functioning. Thromb Haemost. 1998;79:1029–33.
- 156. Ajayi AA, Ogungbade GO, Okorodudu AO. Sex hormone regulation of systemic endothelial and renal microvascular reactivity in type-2 diabetes: studies in gonadectomized and sham-operated Zucker diabetic rats. Eur J Clin Invest. 2004;34:349–57. https://doi.org/10.1111/j.1365-2362.2004.01339.x.
- 157. Rossignoli Pde S, De Labio RW, Payão SL, Pereira OC, Chies AB. Orchidectomy enhances the expression of endothelin-1 and ETB receptors in rat portal vein. J Smooth Muscle Res. 2014;50:85–92. https://doi.org/10.1540/jsmr.50.85.
- Wedgwood S, Dettman RW, Black SM. ET-1 stimulates pulmonary arterial smooth muscle cell proliferation via induction of reactive oxygen species. Am J Physiol Lung Cell Mol Physiol. 2001;281:L1058–67. https://doi.org/10.1152/ajplung.2001.281.5. L1058.
- 159. Kähler J, Mendel S, Weckmüller J, Orzechowski HD, Mittmann C, Köster R, Paul M, Meinertz T, Münzel T. Oxidative stress increases synthesis of big endothelin-1 by activation of the endothelin-1 promoter. J Mol Cell Cardiol. 2000;32:1429–37. https://doi.org/10.1006/jmcc.2000.1178.
- 160. Carneiro FS, Nunes KP, Giachini FR, Lima VV, Carneiro ZN, Nogueira EF, Leite R, Ergul A, Rainey WE, Clinton Webb R, et al. Activation of the ET-1/ETA pathway contributes to erectile dysfunction associated with mineralocorticoid hypertension. J Sex Med. 2008;5:2793–807. https://doi.org/10.1111/j.1743-6109.2008.01009.x.
- Carneiro FS, Giachini FR, Lima VV, Carneiro ZN, Nunes KP, Ergul A, Leite R, Tostes RC, Webb RC. DOCA-salt treatment enhances responses to endothelin-1 in murine corpus cavernosum. Can J Physiol Pharmacol. 2008;86:320–8. https://doi.org/ 10.1139/y08-031.
- Holmquist F, Kirkeby HJ, Larsson B, Forman A, Alm P, Andersson KE. Functional effects, binding sites and immunolocalization



of endothelin-1 in isolated penile tissues from man and rabbit. J Pharmacol Exp Ther. 1992;261:795–802.

- Zhao W, Christ GJ. Endothelin-1 as a putative modulator of erectile dysfunction. II. Calcium mobilization in cultured human corporal smooth muscle cells. J Urol. 1995; 154:1571–9.
- 164. Ergul A, Shoemaker K, Puett D, Tackett RL. Gender differences in the expression of endothelin receptors in human saphenous veins in vitro. J Pharmacol Exp Ther. 1998;285:511–7.
- Kittikulsuth W, Pollock JS, Pollock DM. Sex differences in renal medullary endothelin receptor function in angiotensin II hypertensive rats. Hypertension. 2011;58:212–8. https://doi.org/10. 1161/hypertensionaha.111.172734.
- Ahnstedt H, Cao L, Krause DN, Warfvinge K, Säveland H, Nilsson OG, Edvinsson L. Male-female differences in upregulation of vasoconstrictor responses in human cerebral arteries. PLoS ONE. 2013;8: e62698. https://doi.org/10.1371/journal.pone.0062698.
- 167. Fortes ZB, Nigro D, Scivoletto R, Carvalho MH. Influence of sex on the reactivity to endothelin-1 and noradrenaline in spontaneously hypertensive rats. Clin Exp Hypertens A. 1991;13:807–16. https://doi.org/10.3109/10641969109042084.
- Tostes RC, David FL, Carvalho MH, Nigro D, Scivoletto R, Fortes ZB. Gender differences in vascular reactivity to endothelin-1 in deoxycorticosterone-salt hypertensive rats. J Cardiovasc Pharmacol. 2000;36:S99-101. https://doi.org/10.1097/00005344-200036051-00032.
- 169. Stauffer BL, Westby CM, Greiner JJ, Van Guilder GP, Desouza CA. Sex differences in endothelin-1-mediated vasoconstrictor tone in middle-aged and older adults. Am J Physiol Regul Integr Comp Physiol. 2010;298:R261–5. https://doi.org/10.1152/ajpregu.00626.2009.
- Kellogg DL, Jr., Liu Y, Pérgola PE. Selected contribution: gender differences in the endothelin-B receptor contribution to basal cutaneous vascular tone in humans. J Appl Physiol (1985). 2001; 91:2407–11; discussion 389–90. https://doi.org/10.1152/jappl. 2001.91.5.2407
- Zimmerman MA, Sullivan JC. Hypertension: what's sex got to do with it? Physiology (Bethesda). 2013;28:234–44. https://doi. org/10.1152/physiol.00013.2013.
- 172. Sullivan JC, Pollock JS, Pollock DM. Superoxide-dependent hypertension in male and female endothelin B receptor-deficient rats. Exp Biol Med (Maywood). 2006;231:818–23.
- 173. Taylor TA, Gariepy CE, Pollock DM, Pollock JS. Gender differences in ET and NOS systems in ETB receptor-deficient rats: effect of a high salt diet. Hypertension. 2003;41:657–62. https://doi.org/10.1161/01.Hyp.0000048193.85814.78.
- Nakano D, Pollock DM. Contribution of endothelin A receptors in endothelin 1-dependent natriuresis in female rats. Hypertension. 2009;53:324–30. https://doi.org/10.1161/hypertensionaha. 108.123687.
- 175. Jin C, Speed JS, Hyndman KA, O'Connor PM, Pollock DM. Sex differences in ET-1 receptor expression and Ca2+ signaling in the IMCD. Am J Physiol Renal Physiol. 2013;305:F1099–104. https://doi.org/10.1152/ajprenal.00400.2013.
- Mokdad AH, Marks JS, Stroup DF, Gerberding JL. Actual causes of death in the United States, 2000. JAMA. 2004;291:1238–45. https://doi.org/10.1001/jama.291.10.1238.
- Barton M. Prevention and endothelial therapy of coronary artery disease. Curr Opin Pharmacol. 2013;13:226–41. https://doi.org/ 10.1016/j.coph.2013.05.005.
- 178. Williams CS, Mann M, DuBois RN. The role of cyclooxygenases in inflammation, cancer, and development. Oncogene. 1999;18:7908–16. https://doi.org/10.1038/sj.onc.1203286.
- 179. Van Guilder GP, Westby CM, Greiner JJ, Stauffer BL, DeSouza CA. Endothelin-1 vasoconstrictor tone increases with age in healthy men but can be reduced by regular aerobic exercise.

- Hypertension. 2007;50:403–9. https://doi.org/10.1161/hypertensionaha.107.088294.
- 180. Goettsch W, Lattmann T, Amann K, Szibor M, Morawietz H, Münter K, Müller SP, Shaw S, Barton M. Increased expression of endothelin-1 and inducible nitric oxide synthase isoform II in aging arteries in vivo: implications for atherosclerosis. Biochem Biophys Res Commun. 2001;280:908–13. https://doi.org/10.1006/bbrc.2000.4180.
- 181. Ishihata A, Katano Y. Investigation of differentially expressed genes in the ventricular myocardium of senescent rats. Ann N Y Acad Sci. 2006;1067:142–51. https://doi.org/10.1196/annals. 1354.017.
- 182. Meyer MR, Fredette NC, Barton M, Prossnitz ER. Regulation of vascular smooth muscle tone by adipose-derived contracting factor. PloS one. 2013; 8:e79245-e. https://doi.org/10.1371/journ al.pone.0079245
- 183. Barton M, Shaw S, d'Uscio LV, Moreau P, Lüscher TF. Angiotensin II increases vascular and renal endothelin-1 and functional endothelin converting enzyme activity in vivo: role of ETA receptors for endothelin regulation. Biochem Biophys Res Commun. 1997;238:861–5. https://doi.org/10.1006/bbrc.1997.7394
- 184. Ortmann J, Amann K, Brandes RP, Kretzler M, Münter K, Parekh N, Traupe T, Lange M, Lattmann T, Barton M. Role of podocytes for reversal of glomerulosclerosis and proteinuria in the aging kidney after endothelin inhibition. Hypertension. 2004;44:974–81. https://doi.org/10.1161/01.HYP.0000149249.09147.b4.
- 185. Wang X, Guo Z, Ding Z, Khaidakov M, Lin J, Xu Z, Sharma SG, Jiwani S, Mehta JL. Endothelin-1 upregulation mediates aging-related cardiac fibrosis. J Mol Cell Cardiol. 2015;80:101–9. https://doi.org/10.1016/j.yjmcc.2015.01.001.
- 186. Totan Y, Koca C, Erdurmuş M, Keskin U, Yiğitoğlu R. Endothelin-1 and nitric oxide levels in exudative age-related macular degeneration. J Ophthalmic Vis Res. 2015;10:151–4. https://doi.org/10.4103/2008-322x.163765.
- Sakai T, Matsushima M, Tsuneoka H. Endothelin-1 in neovascular AMD. Ophthalmology. 2011;118:1217.e1. https://doi.org/ 10.1016/j.ophtha.2011.01.033.
- 188. Cabral T, Lima LH, Mello LGM, Polido J, Correa ÉP, Oshima A, Duong J, Serracarbassa P, Regatieri CV, Mahajan VB, et al. Bevacizumab injection in patients with neovascular age-related macular degeneration increases angiogenic biomarkers. Ophthalmol Retina. 2018;2:31–7. https://doi.org/10.1016/j.oret.2017.04.004
- 189. Taniguchi T, Shimazawa M, Sasaoka M, Shimazaki A, Hara H. Endothelin-1 impairs retrograde axonal transport and leads to axonal injury in rat optic nerve. Curr Neurovasc Res. 2006;3:81– 8. https://doi.org/10.2174/156720206776875867.
- Stokely ME, Yorio T, King MA. Endothelin-1 modulates anterograde fast axonal transport in the central nervous system. J Neurosci Res. 2005;79:598–607. https://doi.org/10.1002/jnr.20383.
- 191. Li S, Zhang A, Cao W, Sun X. Elevated plasma endothelin-1 levels in normal tension glaucoma and primary open-angle glaucoma: a meta-analysis. J Ophthalmol. 2016;2016:2678017. https://doi.org/10.1155/2016/2678017.
- 192. Emre M, Orgül S, Haufschild T, Shaw SG, Flammer J. Increased plasma endothelin-1 levels in patients with progressive open angle glaucoma. Br J Ophthalmol. 2005;89:60–3. https://doi. org/10.1136/bjo.2004.046755.
- López-Riquelme N, Villalba C, Tormo C, Belmonte A, Fernandez C, Torralba G, Hernández F. Endothelin-1 levels and biomarkers of oxidative stress in glaucoma patients. Int Ophthalmol. 2015;35:527–32. https://doi.org/10.1007/s10792-014-9979-8.
- 194. Liu Y, Han B, Li F, Liu H. Endothelin-1 concentration in aqueous humor predicts postoperative late low intraocular pressure in



- primary open-angle glaucoma after trabeculectomy. J Glaucoma. 2019;28:633–6. https://doi.org/10.1097/ijg.000000000000001254.
- Barton M. Cholesterol and atherosclerosis: modulation by oestrogen. Curr Opin Lipidol. 2013;24:214–20. https://doi.org/10.1097/MOL.0b013e3283613a94.
- Barton M. Mechanisms and therapy of atherosclerosis and its clinical complications. Curr Opin Pharmacol. 2013;13:149–53. https://doi.org/10.1016/j.coph.2013.05.001.
- Barton M, Meyer MR. Postmenopausal hypertension: mechanisms and therapy. Hypertension. 2009;54:11–8. https://doi.org/10.1161/hypertensionaha.108.120022.
- Störk S, van der Schouw YT, Grobbee DE, Bots ML. Estrogen, inflammation and cardiovascular risk in women: a critical appraisal. Trends Endocrinol Metab. 2004;15:66–72. https://doi.org/10.1016/j.tem.2004.01.005.
- Coylewright M, Reckelhoff JF, Ouyang P. Menopause and hypertension: an age-old debate. Hypertension. 2008;51:952–9. https://doi.org/10.1161/hypertensionaha.107.105742.
- Lewis SJ. Cardiovascular disease in postmenopausal women: myths and reality. Am J Cardiol. 2002; 89:5E-10E; discussion E-1E. https://doi.org/10.1016/s0002-9149(02)02403-7
- Orshal JM, Khalil RA. Gender, sex hormones, and vascular tone.
 Am J Physiol Regul Integr Comp Physiol. 2004;286:R233–49. https://doi.org/10.1152/ajpregu.00338.2003.
- Stanhewicz AE, Wenner MM, Stachenfeld NS. Sex differences in endothelial function important to vascular health and overall cardiovascular disease risk across the lifespan. Am J Physiol Heart Circ Physiol. 2018;315:H1569–88. https://doi.org/10.1152/ajphe art.00396.2018.
- 203. Gavin KM, Seals DR, Silver AE, Moreau KL. Vascular endothelial estrogen receptor alpha is modulated by estrogen status and related to endothelial function and endothelial nitric oxide synthase in healthy women. J Clin Endocrinol Metab. 2009;94:3513–20. https://doi.org/10.1210/jc.2009-0278.
- Wenner MM, Sebzda KN, Kuczmarski AV, Pohlig RT, Edwards DG. ET(B) receptor contribution to vascular dysfunction in postmenopausal women. Am J Physiol Regul Integr Comp Physiol. 2017;313:R51–7. https://doi.org/10.1152/ajpregu.00410.2016.
- Maeda S, Tanabe T, Miyauchi T, Otsuki T, Sugawara J, Iemitsu M, Kuno S, Ajisaka R, Yamaguchi I, Matsuda M. Aerobic exercise training reduces plasma endothelin-1 concentration in older women. J Appl Physiol. 1985;2003(95):336–41. https://doi.org/10.1152/japplphysiol.01016.2002.
- Lekontseva O, Jiang Y, Davidge ST. Estrogen replacement increases matrix metalloproteinase contribution to vasoconstriction in a rat model of menopause. J Hypertens. 2009;27:1602–8. https://doi.org/10.1097/HJH.0b013e32832c41b5.
- 207. Tan Z, Wang T-H, Yang D, Fu X-D, Pan J-Y. Mechanisms of 17β-estradiol on the production of ET-1 in ovariectomized rats. Life Sci. 2003;73:2665–74. https://doi.org/10.1016/S0024-3205(03)00605-2.
- Maturana MA, Breda V, Lhullier F, Spritzer PM. Relationship between endogenous testosterone and cardiovascular risk in early postmenopausal women. Metabolism. 2008;57:961–5. https:// doi.org/10.1016/j.metabol.2008.02.012.
- Tilly N, Schneider JG, Leidig-Bruckner G, Sommer U, Kasperk C. Endothelin-1 levels in patients with disorders of the thyroid gland. Exp Clin Endocrinol Diabetes. 2003;111:80–4. https://doi.org/10.1055/s-2003-39234.
- Eguchi K, Kawakami A, Nakashima M, Ida H, Sakito S, Sakai M, Terada K, Kawabe Y, Fukuda T, Ishikawa N, et al. Stimulation of mitogenesis in human thyroid epithelial cells by endothelin. Acta Endocrinol (Copenh). 1993;128:215–20. https://doi.org/10. 1530/acta.0.1280215.
- Letizia C, Centanni M, Cesareo R, De Ciocchis A, Cerci S, Scuro L, Fumarola A, D'Ambrosio C, Scavo D. Increased plasma

- levels of endothelin-1 in patients with hyperthyroidism. Metabolism. 1995;44:1239–42. https://doi.org/10.1016/0026-0495(95) 90022-5.
- 212. Molet S, Furukawa K, Maghazechi A, Hamid Q, Giaid A. Chemokine- and cytokine-induced expression of endothelin 1 and endothelin-converting enzyme 1 in endothelial cells. J Allergy Clin Immunol. 2000;105:333–8. https://doi.org/10.1016/s0091-6749(00)90084-8.
- Chu CH, Lee JK, Keng HM, Chuang MJ, Lu CC, Wang MC, Sun CC, Wei MC, Lam HC. Hyperthyroidism is associated with higher plasma endothelin-1 concentrations. Exp Biol Med (Maywood). 2006;231:1040–3.
- Rossi GP, Belloni AS, Nussdorfer GG, Pessina AC. Endothelin-1 and the adrenal gland. J Cardiovasc Pharmacol. 2000;35:S17-20. https://doi.org/10.1097/00005344-200000002-00005.
- 215. Rossi G, Albertin G, Belloni A, Zanin L, Biasolo MA, Prayer-Galetti T, Bader M, Nussdorfer GG, Palù G, Pessina AC. Gene expression, localization, and characterization of endothelin A and B receptors in the human adrenal cortex. J Clin Invest. 1994;94:1226–34. https://doi.org/10.1172/jci117440.
- 216. Rossi GP, Albertin G, Neri G, Andreis PG, Hofmann S, Pessina AC, Nussdorfer GG. Endothelin-1 stimulates steroid secretion of human adrenocortical cells ex vivo via both ETA and ETB receptor subtypes. J Clin Endocrinol Metab. 1997;82:3445–9. https://doi.org/10.1210/jcem.82.10.4279.
- Miller WL, Redfield MM, Burnett JC Jr. Integrated cardiac, renal, and endocrine actions of endothelin. J Clin Invest. 1989;83:317–20. https://doi.org/10.1172/jci113876.
- 218. Nussdorfer GG, Rossi GP, Belloni AS. The role of endothelins in the paracrine control of the secretion and growth of the adrenal cortex. In: Jeon KW, ed. *International Review of Cytology*. Vol. 171: Academic Press; 1997, 267–308.
- Lüscher TF. Imbalance of endothelium-derived relaxing and contracting factors. A new concept in hypertension? Am J Hypertens. 1990; 3:317–30. https://doi.org/10.1093/ajh/3.4.
 317
- 220. Chan DSM, Vieira AR, Aune D, Bandera EV, Greenwood DC, McTiernan A, Navarro Rosenblatt D, Thune I, Vieira R, Norat T. Body mass index and survival in women with breast cancer-systematic literature review and meta-analysis of 82 follow-up studies. Ann Oncol. 2014;25:1901–14. https://doi.org/10.1093/annonc/mdu042.
- 221. Ferri C, Bellini C, Desideri G, Baldoncini R, De Siati L, Santucci A. Elevated plasma endothelin-1 levels as an additional risk factor in non-obese essential hypertensive patients with metabolic abnormalities. Diabetologia. 1997;40:100–2. https://doi.org/10.1007/s001250050649.
- 222. Schinzari F, Tesauro M, Rovella V, Di Daniele N, Mores N, Veneziani A, Cardillo C. Leptin stimulates both endothelin-1 and nitric oxide activity in lean subjects but not in patients with obesity-related metabolic syndrome. J Clin Endocrinol Metab. 2013;98:1235–41. https://doi.org/10.1210/jc.2012-3424.
- 223. Considine RV, Sinha MK, Heiman ML, Kriauciunas A, Stephens TW, Nyce MR, Ohannesian JP, Marco CC, McKee LJ, Bauer TL, et al. Serum immunoreactive-leptin concentrations in normal-weight and obese humans. N Engl J Med. 1996;334:292–5. https://doi.org/10.1056/nejm199602013340503.
- 224. Mantzoros CS, Moschos S, Avramopoulos I, Kaklamani V, Liolios A, Doulgerakis DE, Griveas I, Katsilambros N, Flier JS. Leptin concentrations in relation to body mass index and the tumor necrosis factor-alpha system in humans. J Clin Endocrinol Metab. 1997;82:3408–13. https://doi.org/10.1210/jcem.82.10.4323.
- Avogaro A, de Kreutzenberg SV. Mechanisms of endothelial dysfunction in obesity. Clin Chim Acta. 2005;360:9–26. https:// doi.org/10.1016/j.cccn.2005.04.020.



 Poirier P, Eckel RH. Obesity and cardiovascular disease. Curr Atheroscler Rep. 2002;4:448–53. https://doi.org/10.1007/ s11883-002-0049-8.

- Van Guilder GP, Stauffer BL, Greiner JJ, Desouza CA. Impaired endothelium-dependent vasodilation in overweight and obese adult humans is not limited to muscarinic receptor agonists. Am J Physiol Heart Circ Physiol. 2008;294:H1685–92. https://doi. org/10.1152/ajpheart.01281.2007.
- Yoon N, Lteif AA, Han K, Mather KJ. Endothelin contributes differently to peripheral vascular tone and blood pressure in human obesity and diabetes. J Am Soc Hypertens. 2008;2:182–91. https://doi.org/10.1016/j.jash.2007.10.006.
- 229. Campia U, Tesauro M, Cardillo C. Human obesity and endothelium-dependent responsiveness. Br J Pharmacol. 2012;165:561– 73. https://doi.org/10.1111/j.1476-5381.2011.01661.x.
- Steinberg HO, Chaker H, Leaming R, Johnson A, Brechtel G, Baron AD. Obesity/insulin resistance is associated with endothelial dysfunction. Implications for the syndrome of insulin resistance. The Journal of clinical investigation. 1996; 97:2601–10. https://doi.org/10.1172/JCI118709
- Perticone F, Ceravolo R, Candigliota M, Ventura G, Iacopino S, Sinopoli F, Mattioli PL. Obesity and body fat distribution induce endothelial dysfunction by oxidative stress: protective effect of vitamin C. Diabetes. 2001;50:159–65. https://doi.org/10.2337/ diabetes.50.1.159.
- Van Guilder GP, Hoetzer GL, Dengel DR, Stauffer BL, DeSouza CA. Impaired endothelium-dependent vasodilation in normotensive and normoglycemic obese adult humans. J Cardiovasc Pharmacol. 2006;47:310–3. https://doi.org/10.1097/01.fic.0000205097.29946.d3.
- 233. Bigornia SJ, Mott MM, Hess DT, Apovian CM, McDonnell ME, Duess MA, Kluge MA, Fiscale AJ, Vita JA, Gokce N. Long-term successful weight loss improves vascular endothelial function in severely obese individuals. Obesity (Silver Spring). 2010;18:754–9. https://doi.org/10.1038/oby.2009.482.
- 234. Woo KS, Chook P, Yu CW, Sung RY, Qiao M, Leung SS, Lam CW, Metreweli C, Celermajer DS. Effects of diet and exercise on obesity-related vascular dysfunction in children. Circulation. 2004;109:1981–6. https://doi.org/10.1161/01.Cir.00001 26599.47470.Be.
- 235. Woo KS, Chook P, Yu CW, Sung RY, Qiao M, Leung SS, Lam CW, Metreweli C, Celermajer DS. Overweight in children is associated with arterial endothelial dysfunction and intima-media thickening. Int J Obes Relat Metab Disord. 2004;28:852–7. https://doi.org/10.1038/sj.ijo.0802539.
- Francischetti EA, Genelhu VA. Obesity-hypertension: an ongoing pandemic. Int J Clin Pract. 2007;61:269–80. https://doi.org/10.1111/j.1742-1241.2006.01262.x.
- Cardillo C, Kilcoyne CM, Cannon RO 3rd, Panza JA. Interactions between nitric oxide and endothelin in the regulation of vascular tone of human resistance vessels in vivo. Hypertension. 2000;35:1237–41. https://doi.org/10.1161/01.hyp.35.6. 1237.
- 238. Levin ER. Endothelins. N Engl J Med. 1995;333:356–63. https://doi.org/10.1056/nejm199508103330607.
- 239. Mather KJ, Lteif A, Steinberg HO, Baron AD. Interactions between endothelin and nitric oxide in the regulation of vascular tone in obesity and diabetes. Diabetes. 2004;53:2060–6. https://doi.org/10.2337/diabetes.53.8.2060.
- 240. Cardillo C, Kilcoyne CM, Waclawiw M, Cannon RO 3rd, Panza JA. Role of endothelin in the increased vascular tone of patients with essential hypertension. Hypertension. 1999;33:753–8. https://doi.org/10.1161/01.hyp.33.2.753.
- Cardillo C, Campia U, Iantorno M, Panza JA. Enhanced vascular activity of endogenous endothelin-1 in obese hypertensive

- patients. Hypertension. 2004;43:36–40. https://doi.org/10.1161/01.Hyp.0000103868.45064.81.
- 242. Weil BR, Westby CM, Van Guilder GP, Greiner JJ, Stauffer BL, DeSouza CA. Enhanced endothelin-1 system activity with overweight and obesity. Am J Physiol Heart Circ Physiol. 2011;301:H689–95. https://doi.org/10.1152/ajpheart.00206.
- 243. Hamdy O, Ledbury S, Mullooly C, Jarema C, Porter S, Ovalle K, Moussa A, Caselli A, Caballero AE, Economides PA, et al. Lifestyle modification improves endothelial function in obese subjects with the insulin resistance syndrome. Diabetes Care. 2003;26:2119–25. https://doi.org/10.2337/diacare.26.7.2119.
- 244. Maeda S, Jesmin S, Iemitsu M, Otsuki T, Matsuo T, Ohkawara K, Nakata Y, Tanaka K, Goto K, Miyauchi T. Weight loss reduces plasma endothelin-1 concentration in obese men. Exp Biol Med (Maywood). 2006;231:1044–7.
- Liang L, Yu J, Zhou W, Liu N, E LL, Wang DS, Liu H. Endothelin-1 stimulates proinflammatory cytokine expression in human periodontal ligament cells via mitogen-activated protein kinase pathway. J Periodontol. 2014; 85:618–26. https://doi.org/10. 1902/jop.2013.130195
- 246. Virdis A, Schiffrin EL. Vascular inflammation: a role in vascular disease in hypertension? 2003.
- Parameswaran N, Patial S. Tumor necrosis factor-α signaling in macrophages. Crit Rev Eukaryot Gene Expr. 2010;20:87–103. https://doi.org/10.1615/critreveukargeneexpr.v20.i2.10.
- 248. Corder R, Carrier M, Khan N, Klemm P, Vane JR. Cytokine regulation of endothelin-1 release from bovine aortic endothelial cells. J Cardiovasc Pharmacol. 1995;26(Suppl 3):S56–8.
- Kahaleh MB, Fan PS. Effect of cytokines on the production of endothelin by endothelial cells. Clin Exp Rheumatol. 1997:15:163-7.
- 250. Shinagawa S, Okazaki T, Ikeda M, Yudoh K, Kisanuki YY, Yanagisawa M, Kawahata K, Ozaki S. T cells upon activation promote endothelin 1 production in monocytes via IFN-γ and TNF-α. Sci Rep. 2017;7:14500. https://doi.org/10.1038/s41598-017-14202-5.
- Mosser DM, Edwards JP. Exploring the full spectrum of macrophage activation. Nat Rev Immunol. 2008;8:958–69. https://doi.org/10.1038/nri2448.
- Wynn TA, Chawla A, Pollard JW. Macrophage biology in development, homeostasis and disease. Nature. 2013;496:445–55. https://doi.org/10.1038/nature12034.
- Ozdemir R, Parlakpinar H, Polat A, Colak C, Ermis N, Acet A. Selective endothelin a (ETA) receptor antagonist (BQ-123) reduces both myocardial infarct size and oxidant injury. Toxicology. 2006;219:142–9. https://doi.org/10.1016/j.tox.2005.11.022.
- 254. Ford RL, Mains IM, Hilton EJ, Reeves ST, Stroud RE, Crawford FA Jr, Ikonomidis JS, Spinale FG. Endothelin-A receptor inhibition after cardiopulmonary bypass: cytokines and receptor activation. Ann Thorac Surg. 2008;86:1576–83. https://doi.org/10.1016/j.athoracsur.2008.06.076.
- 255. Tonari M, Kurimoto T, Horie T, Sugiyama T, Ikeda T, Oku H. Blocking endothelin-B receptors rescues retinal ganglion cells from optic nerve injury through suppression of neuroinflammation. Invest Ophthalmol Vis Sci. 2012;53:3490–500. https://doi.org/10.1167/jovs.11-9415.
- 256. Piechota-Polanczyk A, Kleniewska P, Gorąca A. The influence of ETA and ETB receptor blockers on LPS-induced oxidative stress and NF-κB signaling pathway in heart. Gen Physiol Biophys. 2012;31:271–8. https://doi.org/10.4149/gpb_2012_031.
- 257. Lin C-C, Hsieh H-L, Shih R-H, Chi P-L, Cheng S-E, Yang C-M. Up-regulation of COX-2/PGE2 by endothelin-1 via MAPK-dependent NF-κB pathway in mouse brain microvascular endothelial cells. Cell communication and signaling: CCS. 2013; 11:8-. https://doi.org/10.1186/1478-811X-11-8



- 258. Donate PB, Cunha TM, Verri WA Jr, Junta CM, Lima FO, Vieira SM, Peres RS, Bombonato-Prado KF, Louzada P Jr, Ferreira SH, et al. Bosentan, an endothelin receptor antagonist, ameliorates collagen-induced arthritis: the role of TNF-α in the induction of endothelin system genes. Inflamm Res. 2012;61:337–48. https://doi.org/10.1007/s00011-011-0415-5.
- Piechota A, Goraca A. Influence of nuclear factor-κB inhibition on endothelin-1 induced lung edema and oxidative stress in rats. J Physiol Pharmacol. 2011;62:183–8.
- Kleniewska P, Piechota-Polanczyk A, Michalski L, Michalska M, Balcerczak E, Zebrowska M, Goraca A. Influence of block of NF-kappa B signaling pathway on oxidative stress in the liver homogenates. Oxid Med Cell Longev. 2013;2013: 308358. https://doi.org/10.1155/2013/308358.
- Ishizuka T, Takamizawa-Matsumoto M, Suzuki K, Kurita A. Endothelin-1 enhances vascular cell adhesion molecule-1 expression in tumor necrosis factor alpha-stimulated vascular endothelial cells. Eur J Pharmacol. 1999;369:237–45. https://doi.org/10.1016/s0014-2999(99)00042-4.
- 262. Li L, Chu Y, Fink GD, Engelhardt JF, Heistad DD, Chen AF. Endothelin-1 stimulates arterial VCAM-1 expression via NADPH oxidase-derived superoxide in mineralocorticoid hypertension. Hypertension. 2003;42:997–1003. https://doi.org/10. 1161/01.Hyp.0000095980.43859.59.
- Blankenberg S, Barbaux S, Tiret L. Adhesion molecules and atherosclerosis. Atherosclerosis. 2003;170:191–203. https://doi.org/10.1016/s0021-9150(03)00097-2.
- 264. Anggrahini DW, Emoto N, Nakayama K, Widyantoro B, Adiarto S, Iwasa N, Nonaka H, Rikitake Y, Kisanuki YY, Yanagisawa M, et al. Vascular endothelial cell-derived endothelin-1 mediates vascular inflammation and neointima formation following blood flow cessation. Cardiovasc Res. 2009;82:143–51. https://doi.org/10.1093/cvr/cvp026.
- Freund-Michel V, Guibert C, Dubois M, Courtois A, Marthan R, Savineau JP, Muller B. Reactive oxygen species as therapeutic targets in pulmonary hypertension. Ther Adv Respir Dis. 2013;7:175–200. https://doi.org/10.1177/1753465812472940.
- 266. Li MW, Mian MO, Barhoumi T, Rehman A, Mann K, Paradis P, Schiffrin EL. Endothelin-1 overexpression exacerbates atherosclerosis and induces aortic aneurysms in apolipoprotein E knockout mice. Arterioscler Thromb Vasc Biol. 2013;33:2306–15. https://doi.org/10.1161/atvbaha.113.302028.
- Mian MO, Idris-Khodja N, Li MW, Leibowitz A, Paradis P, Rautureau Y, Schiffrin EL. Preservation of endothelium-dependent relaxation in atherosclerotic mice with endothelium-restricted endothelin-1 overexpression. J Pharmacol Exp Ther. 2013;347:30–7. https://doi.org/10.1124/jpet.113.206532.
- Kitada K, Ohkita M Fau Matsumura Y, Matsumura Y. Pathological Importance of the Endothelin-1/ET(B) Receptor system on vascular diseases.
- 269. Kitada K, Yui N Fau Matsumoto C, Matsumoto C Fau Mori T, Mori T Fau - Ohkita M, Ohkita M Fau - Matsumura Y, Matsumura Y. Inhibition of endothelin ETB receptor system aggravates neointimal hyperplasia after balloon injury of rat carotid artery.
- Eming SA, Krieg T, Davidson JM. Inflammation in wound repair: molecular and cellular mechanisms. J Invest Dermatol. 2007;127:514–25. https://doi.org/10.1038/sj.jid.5700701.
- Martin P, Nunan R. Cellular and molecular mechanisms of repair in acute and chronic wound healing. Br J Dermatol. 2015;173:370–8. https://doi.org/10.1111/bjd.13954.
- Flammer J, Konieczka K. Retinal venous pressure: the role of endothelin. EPMA J. 2015;6:21. https://doi.org/10.1186/ s13167-015-0043-1.
- 273. Asbert M, Ginès A, Ginès P, Jiménez W, Clària J, Saló J, Arroyo V, Rivera F, Rodés J. Circulating levels of endothelin in cirrhosis.

- Gastroenterology. 1993;104:1485–91. https://doi.org/10.1016/0016-5085(93)90360-o.
- 274. Moore K, Wendon J, Frazer M, Karani J, Williams R, Badr K. Plasma endothelin immunoreactivity in liver disease and the hepatorenal syndrome. N Engl J Med. 1992;327:1774–8. https://doi.org/10.1056/neim199212173272502.
- 275. Alam I, Bass NM, Bacchetti P, Gee L, Rockey DC. Hepatic tissue endothelin-1 levels in chronic liver disease correlate with disease severity and ascites. Am J Gastroenterol. 2000;95:199–203. https://doi.org/10.1111/j.1572-0241.2000.01684.x.
- 276. Rockey DC, Fouassier L, Chung JJ, Carayon A, Vallee P, Rey C, Housset C. Cellular localization of endothelin-1 and increased production in liver injury in the rat: potential for autocrine and paracrine effects on stellate cells. Hepatology. 1998;27:472–80. https://doi.org/10.1002/hep.510270222.
- 277. Zhan S, Chan C-C, Serdar B, Rockey DC. Fibronectin stimulates endothelin-1 synthesis in rat hepatic myofibroblasts via a Src/ERK-regulated signaling pathway. Gastroenterology. 2009;136:2345-55.e554. https://doi.org/10.1053/j.gastro.2009.01.062.
- Shao R, Shi Z, Gotwals PJ, Koteliansky VE, George J, Rockey DC. Cell and molecular regulation of endothelin-1 production during hepatic wound healing. Mol Biol Cell. 2003;14:2327–41. https://doi.org/10.1091/mbc.02-06-0093.
- Shao R, Rockey DC. Effects of endothelins on hepatic stellate cell synthesis of endothelin-1 during hepatic wound healing. J Cell Physiol. 2002;191:342–50. https://doi.org/10.1002/jcp. 10110.
- Housset C, Rockey DC, Bissell DM. Endothelin receptors in rat liver: lipocytes as a contractile target for endothelin 1. Proc Natl Acad Sci U S A. 1993;90:9266–70. https://doi.org/10.1073/pnas. 90.20.9266.
- 281. Pinzani M, Marra F. Cytokine receptors and signaling in hepatic stellate cells. Semin Liver Dis. 2001;21:397–416. https://doi.org/10.1055/s-2001-17554.
- Friedman SL. Liver fibrosis from bench to bedside. J Hepatol. 2003;38(Suppl 1):S38-53. https://doi.org/10.1016/s0168-8278(02)00429-4.
- Pinzani M, Marra F, Carloni V. Signal transduction in hepatic stellate cells. Liver. 1998;18:2–13. https://doi.org/10.1111/j. 1600-0676.1998.tb00120.x.
- Rockey DC, Chung JJ. Endothelin antagonism in experimental hepatic fibrosis. Implications for endothelin in the pathogenesis of wound healing. The Journal of clinical investigation. 1996; 98:1381–8. https://doi.org/10.1172/JCI118925
- 285. Feng HQ, Weymouth ND, Rockey DC. Endothelin antagonism in portal hypertensive mice: implications for endothelin receptor-specific signaling in liver disease. Am J Physiol Gastrointest Liver Physiol. 2009;297:G27-33. https://doi.org/10.1152/ajpgi.90405.2008.
- Thirunavukkarasu C, Yang Y, Subbotin VM, Harvey SAK, Fung J, Gandhi CR. Endothelin receptor antagonist TAK-044 arrests and reverses the development of carbon tetrachloride induced cirrhosis in rats. Gut. 2004;53:1010–9. https://doi.org/ 10.1136/gut.2003.026534.
- Cavasin MA, Semus H, Pitts K, Peng Y, Sandoval J, Chapo J, Plato CF. Acute effects of endothelin receptor antagonists on hepatic hemodynamics of cirrhotic and noncirrhotic rats. Can J Physiol Pharmacol. 2010;88:636–43. https://doi.org/10.1139/ y10-038.
- 288. Gouadon E, Meunier N, Grebert D, Durieux D, Baly C, Salesse R, Caillol M, Congar P. Endothelin evokes distinct calcium transients in neuronal and non-neuronal cells of rat olfactory mucosa primary cultures. Neuroscience. 2010;165:584–600. https://doi.org/10.1016/j.neuroscience.2009.10.046.



289. Le Bourhis M, Rimbaud S, Grebert D, Congar P, Meunier N. Endothelin uncouples gap junctions in sustentacular cells and olfactory ensheathing cells of the olfactory mucosa. Eur J Neurosci. 2014;40:2878–87. https://doi.org/10.1111/ejn.12665.

- Blomstrand F, Venance L, Sirén AL, Ezan P, Hanse E, Glowinski J, Ehrenreich H, Giaume C. Endothelins regulate astrocyte gap junctions in rat hippocampal slices. Eur J Neurosci. 2004;19:1005–15. https://doi.org/10.1111/j.0953-816x.2004. 03197.x.
- Laziz I, Larbi A, Grebert D, Sautel M, Congar P, Lacroix MC, Salesse R, Meunier N. Endothelin as a neuroprotective factor in the olfactory epithelium. Neuroscience. 2011;172:20–9. https://doi.org/10.1016/j.neuroscience.2010.10.063.
- Todorova MG, Josifova T, Konieczka K. Endothelin-1 plasma levels in patients with both retinitis pigmentosa and flammer syndrome. Klin Monbl Augenheilkd. 2015;232:514–8. https:// doi.org/10.1055/s-0035-1545674.
- 293. Yamamoto T, Shimoyama N, Asano H, Mizuguchi T. Analysis of the role of endothelin-A and endothelin-B receptors on nociceptive information transmission in the spinal cord with FR139317, an endothelin-A receptor antagonist, and sarafotoxin S6c, an endothelin-B receptor agonist. J Pharmacol Exp Ther. 1994;271:156–63.
- Nikolov R, Semkova I, Maslarova J, Moyanova S. Antinociceptive effect of centrally administered endothelin-1 and endothelin-3 in the mouse. Methods Find Exp Clin Pharmacol. 1993;15:447–53.
- 295. Jarvis MF, Wessale JL, Zhu CZ, Lynch JJ, Dayton BD, Calzadilla SV, Padley RJ, Opgenorth TJ, Kowaluk EA. ABT-627, an endothelin ET(A) receptor-selective antagonist, attenuates tactile allodynia in a diabetic rat model of neuropathic pain. Eur J Pharmacol. 2000;388:29–35. https://doi.org/10.1016/s0014-2999(99) 00865-1.
- 296. Tocci P, Blandino G, Bagnato A. YAP and endothelin-1 signaling: an emerging alliance in cancer. J Exp Clin Cancer Res. 2021;40:27. https://doi.org/10.1186/s13046-021-01827-8.
- Rosanò L, Spinella F, Bagnato A. Endothelin 1 in cancer: biological implications and therapeutic opportunities. Nat Rev Cancer. 2013;13:637–51. https://doi.org/10.1038/nrc3546.
- 298. Gupta S, Prajapati A, Gulati M, Gautam S, Kumar S, Dalal V, Talmon G, Rachagani S, Jain M. Irreversible and sustained upregulation of endothelin axis during oncogene-associated pancreatic inflammation and cancer. Neoplasia (New York, N.Y.). 2020; 22:98 110.
- Ahn HM, Kim DG, Kim YJ. Blockade of endothelin receptor A enhances the therapeutic efficacy of gemcitabine in pancreatic cancer cells. Biochem Biophys Res Commun. 2020;527:568–73. https://doi.org/10.1016/j.bbrc.2020.04.118.
- 300. Rosanò L, Cianfrocca R, Tocci P, Spinella F, Di Castro V, Caprara V, Semprucci E, Ferrandina G, Natali PG, Bagnato A. Endothelin A receptor/β-arrestin signaling to the Wnt pathway renders ovarian cancer cells resistant to chemotherapy. Cancer Res. 2014;74:7453–64. https://doi.org/10.1158/0008-5472. Can-13-3133.
- Lheureux S, Gourley C, Vergote I, Oza AM. Epithelial ovarian cancer. Lancet. 2019;393:1240–53. https://doi.org/10.1016/s0140-6736(18)32552-2.
- Armbruster S, Coleman RL, Rauh-Hain JA. Management and treatment of recurrent epithelial ovarian cancer. Hematol Oncol Clin North Am. 2018;32:965–82. https://doi.org/10.1016/j.hoc. 2018.07.005.
- von K\u00e4nel R. Psychosocial stress and cardiovascular risk: current opinion. Swiss Med Wkly. 2012;142: w13502. https://doi.org/10. 4414/smw.2012.13502.
- 304. Yusuf S, Hawken S, Ounpuu S, Dans T, Avezum A, Lanas F, McQueen M, Budaj A, Pais P, Varigos J, et al. Effect of

- potentially modifiable risk factors associated with myocardial infarction in 52 countries (the INTERHEART study): case-control study. Lancet. 2004;364:937–52. https://doi.org/10.1016/s0140-6736(04)17018-9.
- Rozanski A, Blumenthal JA, Davidson KW, Saab PG, Kubzansky L. The epidemiology, pathophysiology, and management of psychosocial risk factors in cardiac practice: the emerging field of behavioral cardiology. J Am Coll Cardiol. 2005;45:637–51. https://doi.org/10.1016/j.jacc.2004.12.005.
- Hartley B, Treiber F, Ludwig D, Kapuku G. Correlates of femoral artery flow mediated dilation in a multi-ethnic sample of 12- to 26-year-olds. Ethn Dis. 2004;14:227–32.
- 307. Treiber FA, Kapuku GK, Davis H, Pollock JS, Pollock DM. Plasma endothelin-1 release during acute stress: role of ethnicity and sex. Psychosom Med. 2002;64:707–13. https://doi.org/10.1097/01.psy.0000021952.59258.1c.
- Webb HE, Garten RS, McMinn DR, Beckman JL, Kamimori GH, Acevedo EO. Stress hormones and vascular function in firefighters during concurrent challenges. Biol Psychol. 2011;87:152–60. https://doi.org/10.1016/j.biopsycho.2011.02.024.
- Burg MM, Martens EJ, Collins D, Soufer R. Depression predicts elevated endothelin-1 in patients with coronary artery disease. Psychosom Med. 2011;73:2–6. https://doi.org/10.1097/PSY. 0b013e3181fdfb25.
- Wagner JA, Tennen H, Finan PH, White WB, Burg MM, Ghuman N. Lifetime history of depression, type 2 diabetes, and endothelial reactivity to acute stress in postmenopausal women.
 Int J Behav Med. 2012;19:503–11. https://doi.org/10.1007/s12529-011-9190-5.
- Burg MM, Soufer A, Lampert R, Collins D, Soufer R. Autonomic contribution to endothelin-1 increase during laboratory angerrecall stress in patients with coronary artery disease. Mol Med. 2011;17:495–501. https://doi.org/10.2119/molmed.2010.00083.
- 312. Lobo A, Launer LJ, Fratiglioni L, Andersen K, Di Carlo A, Breteler MM, Copeland JR, Dartigues JF, Jagger C, Martinez-Lage J, et al. Prevalence of dementia and major subtypes in Europe: a collaborative study of population-based cohorts. Neurologic Diseases in the Elderly Research Group. Neurology. 2000; 54:S4–9.
- Wolters FJ, Ikram MA. Epidemiology of vascular dementia. Arterioscler Thromb Vasc Biol. 2019;39:1542–9. https://doi. org/10.1161/atvbaha.119.311908.
- Rizzi L, Rosset I, Roriz-Cruz M. Global epidemiology of dementia: Alzheimer's and vascular types. Biomed Res Int. 2014;2014: 908915. https://doi.org/10.1155/2014/908915.
- 315. Briyal S, Philip T, Gulati A. Endothelin-A receptor antagonists prevent amyloid-β-induced increase in ETA receptor expression, oxidative stress, and cognitive impairment. J Alzheimers Dis. 2011;23:491–503. https://doi.org/10.3233/jad-2010-101245.
- 316. O'Brien JT, Erkinjuntti T, Reisberg B, Roman G, Sawada T, Pantoni L, Bowler JV, Ballard C, DeCarli C, Gorelick PB, et al. Vascular cognitive impairment. Lancet Neurol. 2003;2:89–98. https://doi.org/10.1016/s1474-4422(03)00305-3.
- Matsuzawa Y, Lerman A. Endothelial dysfunction and coronary artery disease: assessment, prognosis, and treatment.
 Coron Artery Dis. 2014;25:713–24. https://doi.org/10.1097/mca.0000000000000178.
- 318. Miyauchi T, Yanagisawa M, Tomizawa T, Sugishita Y, Suzuki N, Fujino M, Ajisaka R, Goto K, Masaki T. Increased plasma concentrations of endothelin-1 and big endothelin-1 in acute myocardial infarction. Lancet. 1989;2:53–4. https://doi.org/10.1016/s0140-6736(89)90303-6.
- 319. Suzuki N, Miyauchi T, Tomobe Y, Matsumoto H, Goto K, Masaki T, Fujino M. Plasma concentrations of endothelin-1 in spontaneously hypertensive rats and DOCA-salt hypertensive



- rats. Biochem Biophys Res Commun. 1990;167:941–7. https://doi.org/10.1016/0006-291x(90)90614-s.
- 320. Kohno M, Murakawa K, Horio T, Yokokawa K, Yasunari K, Fukui T, Takeda T. Plasma immunoreactive endothelin-1 in experimental malignant hypertension. Hypertension. 1991;18:93–100. https://doi.org/10.1161/01.hyp.18.1.93.
- Schiffrin EL, Larivière R, Li JS, Sventek P. Enhanced expression of the endothelin-1 gene in blood vessels of DOCA-salt hypertensive rats: correlation with vascular structure. J Vasc Res. 1996;33:235–48. https://doi.org/10.1159/000159151.
- 322. Vierhapper H, Wagner O, Nowotny P, Waldhäusl W. Effect of endothelin-1 in man. Circulation. 1990;81:1415–8. https://doi.org/10.1161/01.cir.81.4.1415.
- 323. Krum H, Viskoper RJ, Lacourciere Y, Budde M, Charlon V. The effect of an endothelin-receptor antagonist, bosentan, on blood pressure in patients with essential hypertension. Bosentan Hypertension Investigators. N Engl J Med. 1998; 338:784–90. https://doi.org/10.1056/nejm199803193381202
- 324. Yuan W, Cheng G, Li B, Li Y, Lu S, Liu D, Xiao J, Zhao Z. Endothelin-receptor antagonist can reduce blood pressure in patients with hypertension: a meta-analysis. Blood Press. 2017;26:139–49. https://doi.org/10.1080/08037051.2016. 1208730.
- 325. Cardillo C, Kilcoyne CM, Cannon RO 3rd, Panza JA. Increased activity of endogenous endothelin in patients with hypercholesterolemia. J Am Coll Cardiol. 2000;36:1483–8. https://doi.org/10.1016/s0735-1097(00)00910-4.
- 326. Ihling C, Szombathy T, Bohrmann B, Brockhaus M, Schaefer HE, Loeffler BM. Coexpression of endothelin-converting enzyme-1 and endothelin-1 in different stages of human atherosclerosis. Circulation. 2001;104:864–9. https://doi.org/10.1161/hc3301.094742.
- 327. Rossi GP, Colonna S, Pavan E, Albertin G, Della Rocca F, Gerosa G, Casarotto D, Sartore S, Pauletto P, Pessina AC. Endothelin-1 and its mRNA in the wall layers of human arteries ex vivo. Circulation. 1999;99:1147–55. https://doi.org/10.1161/01.cir.99.9.1147.
- 328. Kowala MC, Rose PM, Stein PD, Goller N, Recce R, Beyer S, Valentine M, Barton D, Durham SK. Selective blockade of the endothelin subtype A receptor decreases early atherosclerosis in hamsters fed cholesterol. Am J Pathol. 1995;146:819–26.
- Boulanger CM, Tanner FC, Béa ML, Hahn AW, Werner A, Lüscher TF. Oxidized low density lipoproteins induce mRNA expression and release of endothelin from human and porcine endothelium. Circ Res. 1992;70:1191–7. https://doi.org/10.1161/ 01.res.70.6.1191.
- Roberts WC. Atherosclerotic risk factors—are there ten or is there only one? Am J Cardiol. 1989;64:552—4. https://doi.org/10.1016/0002-9149(89)90444-x.
- Lerman A, Edwards BS, Hallett JW, Heublein DM, Sandberg SM, Burnett JC Jr. Circulating and tissue endothelin immunoreactivity in advanced atherosclerosis. N Engl J Med. 1991;325:997–1001. https://doi.org/10.1056/nejm19911003325 1404.
- Shah R. Endothelins in health and disease. Eur J Intern Med. 2007;18:272–82. https://doi.org/10.1016/j.ejim.2007.04.002.
- 333. Homma S, Miyauchi T, Sugishita Y, Goto K, Sato M, Ohshima N. Vasoconstrictor effects of endothelin-1 on myocardium microcirculation studied by the Langendorff perfusion method: differential sensitivities among microvessels. Microvasc Res. 1992;43:205–17. https://doi.org/10.1016/0026-2862(92)90017-j.
- Thengchaisri N, Hein TW, Ren Y, Kuo L. Endothelin-1 impairs coronary arteriolar dilation: role of p38 kinase-mediated superoxide production from NADPH oxidase. J Mol Cell Cardiol. 2015;86:75–84. https://doi.org/10.1016/j.yjmcc.2015.07.014.

- Clozel M, Qiu C, Qiu CS, Hess P, Clozel JP. Short-term endothelin receptor blockade with tezosentan has both immediate and long-term beneficial effects in rats with myocardial infarction. J Am Coll Cardiol. 2002;39:142–7. https://doi.org/10.1016/s0735-1097(01)01692-8.
- Stewart DJ, Kubac G, Costello KB, Cernacek P. Increased plasma endothelin-1 in the early hours of acute myocardial infarction. J Am Coll Cardiol. 1991;18:38–43. https://doi.org/10.1016/s0735-1097(10)80214-1.
- Omland T, Lie RT, Aakvaag A, Aarsland T, Dickstein K. Plasma endothelin determination as a prognostic indicator of 1-year mortality after acute myocardial infarction. Circulation. 1994;89:1573–9. https://doi.org/10.1161/01.cir.89.4.1573.
- 338. Wenzel RR, Fleisch M, Shaw S, Noll G, Kaufmann U, Schmitt R, Jones CR, Clozel M, Meier B, Lüscher TF. Hemodynamic and coronary effects of the endothelin antagonist bosentan in patients with coronary artery disease. Circulation. 1998;98:2235–40. https://doi.org/10.1161/01.cir.98.21.2235.
- 339. MacCarthy PA, Pegge NC, Prendergast BD, Shah AM, Groves PH. The physiological role of endogenous endothelin in the regulation of human coronary vasomotor tone. J Am Coll Cardiol. 2001;37:137–43. https://doi.org/10.1016/s0735-1097(00) 01042-1.
- 340. Adlbrecht C, Andreas M, Redwan B, Distelmaier K, Mascherbauer J, Kaider A, Wolzt M, Tilea IA, Neunteufl T, Delle-Karth G, et al. Systemic endothelin receptor blockade in ST-segment elevation acute coronary syndrome protects the microvasculature: a randomised pilot study. EuroIntervention. 2012;7:1386–95. https://doi.org/10.4244/eijv7i12a218.
- McMurray JJ, Ray SG, Abdullah I, Dargie HJ, Morton JJ. Plasma endothelin in chronic heart failure. Circulation. 1992;85:1374–9. https://doi.org/10.1161/01.cir.85.4.1374.
- 342. Parker JD, Thiessen JJ. Increased endothelin-1 production in patients with chronic heart failure. American Journal of Physiology-Heart and Circulatory Physiology. 2004;286:H1141–5. https://doi.org/10.1152/ajpheart.00239.2001.
- 343. Tsutamoto T, Wada A, Maeda Y, Adachi T, Kinoshita M. Relation between endothelin-1 spillover in the lungs and pulmonary vascular resistance in patients with chronic heart failure. J Am Coll Cardiol. 1994;23:1427–33. https://doi.org/10.1016/0735-1097(94)90387-5.
- 344. Pousset F, Isnard R, Lechat P, Kalotka H, Carayon A, Maistre G, Escolano S, Thomas D, Komajda M. Prognostic value of plasma endothelin-1 in patients with chronic heart failure. Eur Heart J. 1997;18:254–8. https://doi.org/10.1093/oxfordjour nals.eurheartj.a015228.
- 345. Hülsmann M, Stanek B, Frey B, Sturm B, Putz D, Kos T, Berger R, Woloszczuk W, Putz D, Kos T, et al. Value of cardiopulmonary exercise testing and big endothelin plasma levels to predict short-term prognosis of patients with chronic heart failure. J Am Coll Cardiol. 1998;32:1695–700. https://doi.org/10.1016/s0735-1097(98)00437-9.
- 346. Gray GA, Webb DJ. The endothelin system and its potential as a therapeutic target in cardiovascular disease. Pharmacol Ther. 1996;72:109–48. https://doi.org/10.1016/s0163-7258(96) 00101-5.
- 347. Choukroun G, Hajjar R, Kyriakis JM, Bonventre JV, Rosenzweig A, Force T. Role of the stress-activated protein kinases in endothelin-induced cardiomyocyte hypertrophy. J Clin Invest. 1998;102:1311–20. https://doi.org/10.1172/jci3512.
- 348. Wada A, Tsutamoto T, Ohnishi M, Sawaki M, Fukai D, Maeda Y, Kinoshita M. Effects of a specific endothelin-converting enzyme inhibitor on cardiac, renal, and neurohumoral functions in congestive heart failure: comparison of effects with those of endothelin A receptor antagonism. Circulation. 1999;99:570–7. https://doi.org/10.1161/01.cir.99.4.570.



349. Rich S, McLaughlin VV. Endothelin receptor blockers in cardiovascular disease. Circulation. 2003;108:2184–90. https:// doi.org/10.1161/01.Cir.0000094397.19932.78.

- 350. Sakai S, Miyauchi T, Sakurai T, Kasuya Y, Ihara M, Yamaguchi I, Goto K, Sugishita Y. Endogenous endothelin-1 participates in the maintenance of cardiac function in rats with congestive heart failure. Marked increase in endothelin-1 production in the failing heart. Circulation. 1996; 93:1214–22. https://doi.org/10.1161/01.cir.93.6.1214
- 351. Sakai S, Miyauchi T, Kobayashi M, Yamaguchi I, Goto K, Sugishita Y. Inhibition of myocardial endothelin pathway improves long-term survival in heart failure. Nature. 1996;384:353–5. https://doi.org/10.1038/384353a0.
- Cowburn PJ, Cleland JG, McArthur JD, MacLean MR, McMurray JJ, Dargie HJ. Short-term haemodynamic effects of BQ-123, a selective endothelin ET(A)-receptor antagonist, in chronic heart failure. Lancet. 1998;352:201–2. https://doi.org/10.1016/s0140-6736(05)77807-7.
- 353. Kiowski W, Sütsch G, Hunziker P, Müller P, Kim J, Oechslin E, Schmitt R, Jones R, Bertel O. Evidence for endothelin-1-mediated vasoconstriction in severe chronic heart failure. Lancet. 1995;346:732–6. https://doi.org/10.1016/s0140-6736(95) 91504-4.
- Kelland NF, Webb DJ. Clinical trials of endothelin antagonists in heart failure: publication is good for the public health. Heart (British Cardiac Society). 2007;93:2–4. https://doi.org/10.1136/ hrt.2006.089250.
- Dhaun N, Webb DJ. Endothelins in cardiovascular biology and therapeutics. Nat Rev Cardiol. 2019;16:491–502. https://doi.org/ 10.1038/s41569-019-0176-3.
- 356. Kaluski E, Kobrin I, Zimlichman R, Marmor A, Krakov O, Milo O, Frey A, Kaplan S, Krakover R, Caspi A, et al. RITZ-5: randomized intravenous TeZosentan (an endothelin-A/B antagonist) for the treatment of pulmonary edema: a prospective, multicenter, double-blind, placebo-controlled study. J Am Coll Cardiol. 2003;41:204–10. https://doi.org/10.1016/s0735-1097(02) 02708-0.
- 357. Davison BA, Takagi K, Senger S, Koch G, Metra M, Kimmoun A, Mebazaa A, Voors AA, Nielsen OW, Chioncel O, et al. Megatrials in heart failure: effects of dilution in examination of new therapies. Eur J Heart Fail. 2020;22:1698–707. https://doi.org/10.1002/ejhf.1780.
- 358. O'Connor CM, Gattis WA, Adams KF Jr, Hasselblad V, Chandler B, Frey A, Kobrin I, Rainisio M, Shah MR, Teerlink J, et al. Tezosentan in patients with acute heart failure and acute coronary syndromes: results of the Randomized Intravenous TeZosentan Study (RITZ-4). J Am Coll Cardiol. 2003;41:1452–7. https://doi.org/10.1016/s0735-1097(03)00194-3.
- 359. Kelland NF, Webb DJ. Clinical trials of endothelin antagonists in heart failure: a question of dose? Exp Biol Med (Maywood). 2006:231:696–9.
- 360. Zhou BY, Guo YL, Wu NQ, Zhu CG, Gao Y, Qing P, Li XL, Wang Y, Dong Q, Liu G, et al. Plasma big endothelin-1 levels at admission and future cardiovascular outcomes: a cohort study in patients with stable coronary artery disease. Int J Cardiol. 2017;230:76–9. https://doi.org/10.1016/j.ijcard.2016.12.082.
- 361. Gao R, Wang J, Zhang S, Yang G, Gao Z, Chen X. The value of combining plasma D-dimer and endothelin-1 levels to predict noreflow after percutaneous coronary intervention of ST-segment elevation in acute myocardial infarction patients with a type 2 diabetes mellitus history. Med Sci Monit. 2018; 24:3549–56. https://doi.org/10.12659/msm.908980
- 362. Brouwers FP, van Gilst WH, Damman K, van den Berg MP, Gansevoort RT, Bakker SJ, Hillege HL, van Veldhuisen DJ, van der Harst P, de Boer RA. Clinical risk stratification optimizes value of biomarkers to predict new-onset heart failure

- in a community-based cohort. Circ Heart Fail. 2014;7:723–31. https://doi.org/10.1161/circheartfailure.114.001185.
- 363. Cuculi F, Lim CC, van Gaal W, Testa L, Westaby S, Arnold JR, Neubauer S, Banning AP. Systemic levels of endothelin correlate with systemic inflammation and not with myocardial injury or left ventricular ejection fraction in patients undergoing percutaneous coronary intervention and on-pump coronary artery bypass grafting. Interact Cardiovasc Thorac Surg. 2011;13:585–9. https://doi.org/10.1510/icvts.2011.278192.
- 364. Mayyas F, Al-Jarrah M, Ibrahim K, Mfady D, Van Wagoner DR. The significance of circulating endothelin-1 as a predictor of coronary artery disease status and clinical outcomes following coronary artery catheterization. Cardiovasc Pathol. 2015;24:19– 25. https://doi.org/10.1016/j.carpath.2014.08.004.
- 365. Kurisu S, Sato H, Kawagoe T, Ishihara M, Shimatani Y, Nishioka K, Kono Y, Umemura T, Nakamura S. Tako-tsubo-like left ventricular dysfunction with ST-segment elevation: a novel cardiac syndrome mimicking acute myocardial infarction. Am Heart J. 2002;143:448–55. https://doi.org/10.1067/mhj.2002.120403.
- 366. Tsuchihashi K, Ueshima K, Uchida T, Oh-mura N, Kimura K, Owa M, Yoshiyama M, Miyazaki S, Haze K, Ogawa H, et al. Transient left ventricular apical ballooning without coronary artery stenosis: a novel heart syndrome mimicking acute myocardial infarction. Angina Pectoris-Myocardial Infarction Investigations in Japan. J Am Coll Cardiol. 2001; 38:11–8. https://doi.org/10.1016/s0735-1097(01)01316-x
- Pavin D, Le Breton H, Daubert C. Human stress cardiomyopathy mimicking acute myocardial syndrome. Heart. 1997;78:509–11. https://doi.org/10.1136/hrt.78.5.509.
- 368. Lateef F. The, "broken heart sydrome": you're likely to have it only once! Singapore Med J. 2010;51:e76–8.
- Schlossbauer SA, Ghadri JR, Scherff F, Templin C. The challenge of Takotsubo syndrome: heterogeneity of clinical features.
 Swiss Med Wkly. 2017;147: w14490. https://doi.org/10.4414/smw.2017.14490.
- Thygesen K, Alpert JS, Jaffe AS, Chaitman BR, Bax JJ, Morrow DA, White HD. Fourth Universal Definition of Myocardial Infarction (2018). Circulation. 2018;138:e618–51. https://doi.org/10.1161/cir.0000000000000017.
- Pelliccia F, Sinagra G, Elliott P, Parodi G, Basso C, Camici PG. Takotsubo is not a cardiomyopathy. Int J Cardiol. 2018;254:250–3. https://doi.org/10.1016/j.ijcard.2017.12.009.
- 372. Templin C, Ghadri JR, Diekmann J, Napp LC, Bataiosu DR, Jaguszewski M, Cammann VL, Sarcon A, Geyer V, Neumann CA, et al. Clinical features and outcomes of takotsubo (stress) cardiomyopathy. N Engl J Med. 2015;373:929–38. https://doi.org/10.1056/NEJMoa1406761.
- 373. Kurowski V, Kaiser A, von Hof K, Killermann DP, Mayer B, Hartmann F, Schunkert H, Radke PW. Apical and midventricular transient left ventricular dysfunction syndrome (tako-tsubo cardiomyopathy): frequency, mechanisms, and prognosis. Chest. 2007;132:809–16. https://doi.org/10.1378/chest.07-0608.
- Stiermaier T, Möller C, Graf T, Eitel C, Desch S, Thiele H, Eitel I. Prognostic usefulness of the ballooning pattern in patients with takotsubo cardiomyopathy. Am J Cardiol. 2016;118:1737–41. https://doi.org/10.1016/j.amjcard.2016.08.055.
- 375. Medina de Chazal H, Del Buono MG, Keyser-Marcus L, Ma L, Moeller FG, Berrocal D, Abbate A. Stress cardiomyopathy diagnosis and treatment: JACC State-of-the-Art Review. J Am Coll Cardiol. 2018; 72:1955–71. https://doi.org/10.1016/j.jacc. 2018.07.072
- 376. Del Buono MG, Potere N, Chiabrando JG, Bressi E, Abbate A. Takotsubo syndrome: diagnostic work-up and clues into differential diagnosis. Curr Opin Cardiol. 2019;34:673–86. https://doi.org/10.1097/hco.00000000000000672.



- Konstantinos G, El-Battrawy I, Schramm K, Uzair A, Hoffmann U, Martin B, Ibrahim A. Comparison and outcome analysis of patients with takotsubo cardiomyopathy triggered by emotional stress or physical stress. Front Psychol. 2017;8:527. https://doi.org/10.3389/fpsyg.2017.00527.
- 378. Kagiyama N, Okura H, Matsue Y, Tamada T, Imai K, Yamada R, Kume T, Hayashida A, Neishi Y, Yoshida K. Multiple unfavorable echocardiographic findings in takotsubo cardiomyopathy are associated with increased in-hospital events and mortality. J Am Soc Echocardiogr. 2016;29:1179–87. https://doi.org/10.1016/j.echo.2016.08.021.
- Sobue Y, Watanabe E, Ichikawa T, Koshikawa M, Yamamoto M, Harada M, Ozaki Y. Physically triggered takotsubo cardiomyopathy has a higher in-hospital mortality rate. Int J Cardiol. 2017;235:87–93. https://doi.org/10.1016/j.ijcard.2017.02.090.
- 380. Christensen TE, Bang LE, Holmvang L, Skovgaard DC, Oturai DB, Søholm H, Thomsen JH, Andersson HB, Ghotbi AA, Ihlemann N, et al. (123)I-MIBG Scintigraphy in the subacute state of takotsubo cardiomyopathy. JACC Cardiovasc Imaging. 2016;9:982–90. https://doi.org/10.1016/j.jcmg.2016.01.028.
- Smeijers L, Szabó BM, van Dammen L, Wonnink W, Jakobs BS, Bosch JA, Kop WJ. Emotional, neurohormonal, and hemodynamic responses to mental stress in Tako-Tsubo cardiomyopathy. Am J Cardiol. 2015;115:1580–6. https://doi.org/10.1016/j.amjca rd.2015.02.064.
- 382. Núñez-Gil IJ, Bernardo E, Feltes G, Escaned J, Mejía-Rentería HD, De Agustín JA, Vivas D, Nombela-Franco L, Jiménez-Quevedo P, Macaya C, et al. Platelet function in takotsubo cardiomyopathy. J Thromb Thrombolysis. 2015;39:452–8. https:// doi.org/10.1007/s11239-014-1109-y.
- 383. Uchida Y, Egami H, Uchida Y, Sakurai T, Kanai M, Shirai S, Nakagawa O, Oshima T. Possible participation of endothelial cell apoptosis of coronary microvessels in the genesis of takotsubo cardiomyopathy. Clin Cardiol. 2010;33:371–7. https://doi.org/10.1002/clc.20777.
- 384. Lee SJ, Kang JG, Ryu OH, Kim CS, Ihm SH, Choi MG, Yoo HJ, Hong KS. The relationship of thyroid hormone status with myocardial function in stress cardiomyopathy. Eur J Endocrinol. 2009;160:799–806. https://doi.org/10.1530/eje-08-0808.
- 385. Tippins JR, Antoniw JW, Maseri A. Endothelin-1 is a potent constrictor in conductive and resistive coronary arteries. J Cardiovasc Pharmacol. 1989;13(Suppl 5):S177–9. https://doi.org/10.1097/00005344-198900135-00048.
- 386. Kurihara H, Yoshizumi M, Sugiyama T, Yamaoki K, Nagai R, Takaku F, Satoh H, Inui J, Yanagisawa M, Masaki T, et al. The possible role of endothelin-1 in the pathogenesis of coronary vasospasm. J Cardiovasc Pharmacol. 1989; 13 Suppl 5:S132–7; discussion S42. https://doi.org/10.1097/00005344-19890 0135-00033
- 387. Matsuyama K, Yasue H, Okumura K, Saito Y, Nakao K, Shirakami G, Imura H. Increased plasma level of endothelin-1-like immunoreactivity during coronary spasm in patients with coronary spastic angina. Am J Cardiol. 1991;68:991–5. https://doi.org/10.1016/0002-9149(91)90484-3.
- Madias JE. Takotsubo syndrome and coronary microcirculation dysfunction: Vasospasm or damage due to adjacent cardiomyocyte injury and/or myocardial edema? Int J Cardiol. 2016;215:90–1. https://doi.org/10.1016/j.ijcard.2016.04.117.
- 389. Kaski JC, Russo G. Cardiac syndrome X: an overview. Hosp Pract (1995). 2000; 35:75–6, 9–82, 5–8 passim. https://doi.org/ 10.3810/hp.2000.02.183
- McClellan G, Weisberg A, Winegrad S. Endothelin regulation of cardiac contractility in absence of added endothelin. Am J Physiol. 1995;268:H1621–7. https://doi.org/10.1152/ajpheart. 1995.268.4.H1621.

- 391. Miyauchi T, Goto K. Heart failure and endothelin receptor antagonists. Trends Pharmacol Sci. 1999;20:210–7. https://doi.org/10.1016/S0165-6147(99)01297-3.
- Nishimaru K, Miura Y, Endoh M. Mechanisms of endothelin-1-induced decrease in contractility in adult mouse ventricular myocytes. Br J Pharmacol. 2007;152:456–63. https://doi.org/ 10.1038/sj.bjp.0707392.
- 393. Naegele M, Flammer AJ, Enseleit F, Roas S, Frank M, Hirt A, Kaiser P, Cantatore S, Templin C, Fröhlich G, et al. Endothelial function and sympathetic nervous system activity in patients with Takotsubo syndrome. Int J Cardiol. 2016;224:226–30. https://doi.org/10.1016/j.ijcard.2016.09.008.
- 394. Mulder P, Richard V, Derumeaux G, Hogie M, Henry JP, Lallemand F, Compagnon P, Macé B, Comoy E, Letac B, et al. Role of endogenous endothelin in chronic heart failure: effect of long-term treatment with an endothelin antagonist on survival, hemodynamics, and cardiac remodeling. Circulation. 1997;96:1976–82. https://doi.org/10.1161/01.cir.96.6.1976.
- 395. Wolff K, Faxén M, Lunell NO, Nisell H, Lindblom B. Endothelin receptor type A and B gene expression in human nonpregnant, term pregnant, and preeclamptic uterus. Am J Obstet Gynecol. 1996;175:1295–300. https://doi.org/10.1016/s0002-9378(96)70044-3.
- 396. Wolff K, Nisell H, Modin A, Lundberg JM, Lunell NO, Lindblom B. Contractile effects of endothelin 1 and endothelin 3 on myometrium and small intramyometrial arteries of pregnant women at term. Gynecol Obstet Invest. 1993;36:166–71. https://doi.org/10.1159/000292619.
- 397. Osada K, Tsunoda H, Miyauchi T, Sugishita Y, Kubo T, Goto K. Pregnancy increases ET-1-induced contraction and changes receptor subtypes in uterine smooth muscle in humans. Am J Physiol. 1997;272:R541–8. https://doi.org/10.1152/ajpregu. 1997.272.2.R541.
- 398. Ajne G, Nisell H, Wolff K, Jansson T. The role of endogenous endothelin in the regulation of uteroplacental and renal blood flow during pregnancy in conscious rats. Placenta. 2003;24:813-8. https://doi.org/10.1016/s0143-4004(03) 00113-9.
- Madsen KM, Neerhof MG, Wessale JL, Thaete LG. Influence of ET(B) receptor antagonism on pregnancy outcome in rats. J Soc Gynecol Investig. 2001;8:239

 44. https://doi.org/10.1016/ s1071-5576(01)00120-4.
- Breathett K, Muhlestein D, Foraker R, Gulati M. Differences in preeclampsia rates between African American and Caucasian women: trends from the National Hospital Discharge Survey. J Womens Health (Larchmt). 2014;23:886–93. https://doi.org/10. 1089/jwh.2014.4749.
- 401. Palei AC, Spradley FT, Warrington JP, George EM, Granger JP. Pathophysiology of hypertension in pre-eclampsia: a lesson in integrative physiology. Acta Physiol (Oxf). 2013;208:224–33. https://doi.org/10.1111/apha.12106.
- Goulopoulou S, Davidge ST. Molecular mechanisms of maternal vascular dysfunction in preeclampsia. Trends Mol Med. 2015;21:88–97. https://doi.org/10.1016/j.molmed.2014.11.009.
- 403. George EM, Palei AC, Granger JP. Endothelin as a final common pathway in the pathophysiology of preeclampsia: therapeutic implications. Curr Opin Nephrol Hypertens. 2012;21:157–62. https://doi.org/10.1097/MNH.0b013e328350094b.
- 404. LaMarca B, Parrish M, Ray LF, Murphy SR, Roberts L, Glover P, Wallukat G, Wenzel K, Cockrell K, Martin JN, Jr., et al. Hypertension in response to autoantibodies to the angiotensin II type I receptor (AT1-AA) in pregnant rats: role of endothelin-1. Hypertension (Dallas, Tex.: 1979). 2009; 54:905–9. https://doi.org/10.1161/HYPERTENSIONAHA.109.137935



405. Mastrogiannis DS, O'Brien WF, Krammer J, Benoit R. Potential role of endothelin-1 in normal and hypertensive pregnancies. Am J Obstet Gynecol. 1991;165:1711–6. https://doi.org/10.1016/ 0002-9378(91)90020-r.

- 406. Kiprono LV, Wallace K, Moseley J, Martin J Jr, Lamarca B. Progesterone blunts vascular endothelial cell secretion of endothelin-1 in response to placental ischemia. Am J Obstet Gynecol. 2013;209(44):e1-6. https://doi.org/10.1016/j.ajog.2013.03.032.
- Taylor RN, Varma M, Teng NN, Roberts JM. Women with preeclampsia have higher plasma endothelin levels than women with normal pregnancies. J Clin Endocrinol Metab. 1990;71:1675–7. https://doi.org/10.1210/jcem-71-6-1675.
- 408. Karakus S, Bozoklu Akkar O, Yildiz C, Sancakdar E, Cetin M, Cetin A. Serum levels of ET-1, M30, and angiopoietins-1 and -2 in HELLP syndrome and preeclampsia compared to controls. Arch Gynecol Obstet. 2016;293:351–9. https://doi.org/10.1007/s00404-015-3803-1.
- 409. Aggarwal PK, Chandel N, Jain V, Jha V. The relationship between circulating endothelin-1, soluble fms-like tyrosine kinase-1 and soluble endoglin in preeclampsia. J Hum Hypertens. 2012;26:236–41. https://doi.org/10.1038/jhh.2011.29.
- 410. Alexander BT, Rinewalt AN, Cockrell KL, Massey MB, Bennett WA, Granger JP. Endothelin type a receptor blockade attenuates the hypertension in response to chronic reductions in uterine perfusion pressure. Hypertension. 2001;37:485–9. https://doi.org/10.1161/01.hyp.37.2.485.
- 411. LaMarca BD, Ryan MJ, Gilbert JS, Murphy SR, Granger JP. Inflammatory cytokines in the pathophysiology of hypertension during preeclampsia. Curr Hypertens Rep. 2007;9:480–5. https://doi.org/10.1007/s11906-007-0088-1.
- 412. Abdalvand A, Morton JS, Bourque SL, Quon AL, Davidge ST. Matrix metalloproteinase enhances big-endothelin-1 constriction in mesenteric vessels of pregnant rats with reduced uterine blood flow. Hypertension. 2013;61:488–93. https://doi.org/10.1161/hypertensionaha.111.00055.
- 413. Nugent WH, Mishra N, Strauss JF 3rd, Walsh SW. Matrix metal-loproteinase 1 causes vasoconstriction and enhances vessel reactivity to angiotensin II via protease-activated receptor 1. Reprod Sci. 2016;23:542–8. https://doi.org/10.1177/1933719115607998.
- 414. Kernochan LE, Tran BN, Tangkijvanich P, Melton AC, Tam SP, Yee HF Jr. Endothelin-1 stimulates human colonic myofibroblast contraction and migration. Gut. 2002;50:65–70. https://doi.org/ 10.1136/gut.50.1.65.
- 415. Murphy SR, LaMarca BBD, Cockrell K, Granger JP. Role of endothelin in mediating soluble fms-like tyrosine kinase 1-induced hypertension in pregnant rats. Hypertension (Dallas, Tex.: 1979). 2010; 55:394–8. https://doi.org/10.1161/HYPER TENSIONAHA.109.141473
- 416. George EM, Granger JP. Endothelin: key mediator of hypertension in preeclampsia. Am J Hypertens. 2011;24:964–9. https://doi.org/10.1038/ajh.2011.99.
- 417. Saleh L, Danser JA, van den Meiracker AH. Role of endothelin in preeclampsia and hypertension following antiangiogenesis treatment. Curr Opin Nephrol Hypertens. 2016;25:94–9. https://doi.org/10.1097/mnh.000000000000197.
- 418. Dubyk LV, Yuzko OM, Chernetska NV, Kolotylo TR. Early pregnancy and endothelial dysfunction. Wiad Lek. 2020;73:743-5.
- Golan A, Lin G, Evron S, Arieli S, Niv D, David MP. Oligohydramnios: maternal complications and fetal outcome in 145 cases. Gynecol Obstet Invest. 1994;37:91–5. https://doi.org/10.1159/000292532.
- 420. Hamza A, Herr D, Solomayer EF, Meyberg-Solomayer G. Polyhydramnios: causes, diagnosis and therapy. Geburtshilfe Frauenheilkd. 2013;73:1241–6. https://doi.org/10.1055/s-0033-1360163.

- Keilman C, Shanks AL. Oligohydramnios. [Updated 2020 Sep 15]. . StatPearls [Internet] Treasure Island (FL): StatPearls 2020
- 422. Harvey-Wilkes KB, Nielsen HC, D'Alton ME. Elevated endothelin levels are associated with increased placental resistance. Am J Obstet Gynecol. 1996;174:1599–604. https://doi. org/10.1016/s0002-9378(96)70613-0.
- McQeen J, Kingdom JC, Connell JM, Whittle MJ. Fetal endothelin levels and placental vascular endothelin receptors in intrauterine growth retardation. Obstet Gynecol. 1993;82:992–8.
- 424. Bajoria R, Ward S, Chatterjee R. Brain natriuretic peptide and endothelin-1 in the pathogenesis of polyhydramnios-oligohydramnios in monochorionic twins. Am J Obstet Gynecol. 2003;189:189–94. https://doi.org/10.1067/mob.2003.340.
- 425. Isozaki-Fukuda Y, Kojima T, Hirata Y, Ono A, Sawaragi S, Sawaragi I, Kobayashi Y. Plasma immunoreactive endothelin-1 concentration in human fetal blood: its relation to asphyxia. Pediatr Res. 1991;30:244–7. https://doi.org/10.1203/00006450-19910 9000-00008.
- Itoh H, Sagawa N, Hasegawa M, Okagaki A, Inamori K, Ihara Y, Mori T, Ogawa Y, Suga S, Mukoyama M, et al. Brain natriuretic peptide is present in the human amniotic fluid and is secreted from amnion cells. J Clin Endocrinol Metab. 1993;76:907–11. https://doi.org/10.1210/jcem.76.4.8473404.
- 427. Sagawa N, Hasegawa M, Itoh H, Nanno H, Mori T, Yano J, Yoshimasa T, Nakao K. Current topic: the role of amniotic endothelin in human pregnancy. Placenta. 1994;15:565–75. https://doi.org/10.1016/s0143-4004(05)80404-7.
- 428. Smitherman TA, Burch R, Sheikh H, Loder E. The prevalence, impact, and treatment of migraine and severe headaches in the United States: a review of statistics from national surveillance studies. Headache. 2013;53:427–36. https://doi.org/10.1111/head.12074.
- 429. Burch RC, Loder S, Loder E, Smitherman TA. The prevalence and burden of migraine and severe headache in the United States: updated statistics from government health surveillance studies. Headache. 2015;55:21–34. https://doi.org/10.1111/head.12482.
- 430. Woldeamanuel YW, Cowan RP. Migraine affects 1 in 10 people worldwide featuring recent rise: a systematic review and meta-analysis of community-based studies involving 6 million participants. J Neurol Sci. 2017;372:307–15. https://doi.org/10.1016/j.jns.2016.11.071.
- 431. Goadsby PJ. The vascular theory of migraine—a great story wrecked by the facts. Brain. 2009;132:6–7. https://doi.org/10.1093/brain/awn321.
- Messlinger K. Migraine: where and how does the pain originate? Exp Brain Res. 2009;196:179–93. https://doi.org/10.1007/s00221-009-1756-y.
- Shevel E. The extracranial vascular theory of migraine–a great story confirmed by the facts. Headache. 2011;51:409–17. https:// doi.org/10.1111/j.1526-4610.2011.01844.x.
- 434. Bianchi A, Pitari G, Amenta V, Giuliano F, Gallina M, Costa R, Ferlito S. Endothelial, haemostatic and haemorheological modifications in migraineurs. Artery. 1996;22:93–100.
- Kallela M, Färkkilä M, Saijonmaa O, Fyhrquist F. Endothelin in migraine patients. Cephalalgia. 1998;18:329–32. https://doi.org/ 10.1046/j.1468-2982.1998.1806329.x.
- 436. Färkkilä M, Palo J, Saijonmaa O, Fyhrquist F. Raised plasma endothelin during acute migraine attack. Cephalalgia. 1992; 12:383–4; discussion 40. https://doi.org/10.1111/j.1468-2982. 1992.00383.x
- 437. Hamed SA, Hamed EA, Ezz Eldin AM, Mahmoud NM. Vascular risk factors, endothelial function, and carotid thickness in patients with migraine: relationship to atherosclerosis. J Stroke Cerebrovasc Dis. 2010;19:92–103. https://doi.org/10.1016/j.jstro kecerebrovasdis.2009.04.007.



- 438. Gallai V, Sarchielli P, Firenze C, Trequattrini A, Paciaroni M, Usai F, Palumbo R. Endothelin 1 in migraine and tension-type headache. Acta Neurol Scand. 1994;89:47–55. https://doi.org/10.1111/j.1600-0404.1994.tb01632.x.
- Hasselblatt M, Köhler J, Volles E, Ehrenreich H. Simultaneous monitoring of endothelin-1 and vasopressin plasma levels in migraine. NeuroReport. 1999;10:423–5. https://doi.org/10.1097/ 00001756-199902050-00039.
- 440. Nattero G, Mengozzi G, Inconis T, Paradisi L. Nitric oxide, endothelin-1, and transcranial Doppler in migraine. Findings in interictal conditions and during migraine attack. Headache. 1996; 36:307–11. https://doi.org/10.1046/j.1526-4610.1996.3605307.x
- 441. Arngrim N, Schytz HW, Britze J, Amin FM, Vestergaard MB, Hougaard A, Wolfram F, de Koning PJ, Olsen KS, Secher NH, et al. Migraine induced by hypoxia: an MRI spectroscopy and angiography study. Brain. 2016;139:723–37. https://doi.org/10.1093/brain/awv359.
- Kourembanas S, Marsden PA, McQuillan LP, Faller DV. Hypoxia induces endothelin gene expression and secretion in cultured human endothelium. J Clin Invest. 1991;88:1054–7. https://doi. org/10.1172/jci115367.
- 443. Hadjikhani N, Sanchez Del Rio M, Wu O, Schwartz D, Bakker D, Fischl B, Kwong KK, Cutrer FM, Rosen BR, Tootell RB, et al. Mechanisms of migraine aura revealed by functional MRI in human visual cortex. Proc Natl Acad Sci USA. 2001;98:4687–92. https://doi.org/10.1073/pnas.071582498.
- Ayata C. Cortical spreading depression triggers migraine attack: pro. Headache. 2010;50:725–30. https://doi.org/10.1111/j.1526-4610.2010.01647.x.
- 445. Bolay H, Reuter U, Dunn AK, Huang Z, Boas DA, Moskowitz MA. Intrinsic brain activity triggers trigeminal meningeal afferents in a migraine model. Nat Med. 2002;8:136–42. https://doi.org/10.1038/nm0202-136.
- 446. Zhang X, Levy D, Kainz V, Noseda R, Jakubowski M, Burstein R. Activation of central trigeminovascular neurons by cortical spreading depression. Ann Neurol. 2011;69:855–65. https://doi.org/10.1002/ana.22329.
- 447. Dreier JP, Kleeberg J, Alam M, Major S, Kohl-Bareis M, Petzold GC, Victorov I, Dirnagl U, Obrenovitch TP, Priller J. Endothelin-1-induced spreading depression in rats is associated with a microarea of selective neuronal necrosis. Exp Biol Med (Maywood). 2007;232:204–13.
- 448. Ayata C, Lauritzen M. Spreading depression, spreading depolarizations, and the cerebral vasculature. Physiol Rev. 2015;95:953–93. https://doi.org/10.1152/physrev.00027.2014.
- 449. Olesen J, Friberg L, Olsen TS, Iversen HK, Lassen NA, Andersen AR, Karle A. Timing and topography of cerebral blood flow, aura, and headache during migraine attacks. Ann Neurol. 1990;28:791–8. https://doi.org/10.1002/ana.410280610
- Olesen J. The International Classification of Headache Disorders, 3rd edition (beta version). Cephalalgia. 2013; 33:629–808. https://doi.org/10.1177/0333102413485658
- 451. Gaist D, Garde E, Blaabjerg M, Nielsen HH, Krøigård T, Østergaard K, Møller HS, Hjelmborg J, Madsen CG, Iversen P, et al. Migraine with aura and risk of silent brain infarcts and white matter hyperintensities: an MRI study. Brain. 2016;139:2015–23. https://doi.org/10.1093/brain/aww099.
- 452. Smith TP, Haymond T, Smith SN, Sweitzer SM. Evidence for the endothelin system as an emerging therapeutic target for the treatment of chronic pain. J Pain Res. 2014;7:531–45. https:// doi.org/10.2147/JPR.S65923.
- 453. Bourque SL, Davidge ST, Adams MA. The interaction between endothelin-1 and nitric oxide in the vasculature: new perspectives. Am J Physiol Regul Integr Comp Physiol.

- 2011;300:R1288-95. https://doi.org/10.1152/ajpregu.00397.2010.
- 454. Wang Y, Wang DH. Prevention of endothelin-1-induced increases in blood pressure: role of endogenous CGRP. Am J Physiol Heart Circ Physiol. 2004;287:H1868–74. https://doi.org/10.1152/ajpheart.00241.2004.
- 455. Schytz HW, Schoonman GG, Ashina M. What have we learnt from triggering migraine? Curr Opin Neurol. 2010;23:259–65. https://doi.org/10.1097/WCO.0b013e328337b884.
- 456. Garlichs CD, Zhang H, Mügge A, Daniel WG. Beta-blockers reduce the release and synthesis of endothelin-1 in human endothelial cells. Eur J Clin Invest. 1999;29:12–6. https://doi.org/10.1046/j.1365-2362.1999.00407.x.
- 457. Desideri G, Grassi D, Croce G, Bocale R, Tiberti S, Evangelista S, Necozione S, Di Orio F, Ferri C. Different effects of angiotensin converting enzyme inhibitors on endothelin-1 and nitric oxide balance in human vascular endothelial cells: evidence of an oxidant-sensitive pathway. Mediators Inflamm. 2008;2008; 305087. https://doi.org/10.1155/2008/305087.
- Ghiadoni L, Virdis A, Magagna A, Taddei S. Effect of the angiotensin II type 1 receptor blocker essential hypertension. Online. 2000:501–6.
- 459. May A, Gijsman HJ, Wallnöfer A, Jones R, Diener HC, Ferrari MD. Endothelin antagonist bosentan blocks neurogenic inflammation, but is not effective in aborting migraine attacks. Pain. 1996;67:375–8. https://doi.org/10.1016/0304-3959(96)03137-5.
- Di Carlo A. Human and economic burden of stroke. Age Ageing. 2009;38:4–5. https://doi.org/10.1093/ageing/afn282.
- Benowitz LI, Carmichael ST. Promoting axonal rewiring to improve outcome after stroke. Neurobiol Dis. 2010;37:259–66. https://doi.org/10.1016/j.nbd.2009.11.009.
- Lipton P. Ischemic cell death in brain neurons. Physiol Rev. 1999;79:1431–568. https://doi.org/10.1152/physrev.1999.79.4. 1431.
- 463. Macrae IM, Robinson MJ, Graham DI, Reid JL, McCulloch J. Endothelin-1-induced reductions in cerebral blood flow: dose dependency, time course, and neuropathological consequences. J Cereb Blood Flow Metab. 1993;13:276–84. https://doi.org/10. 1038/jcbfm.1993.34.
- 464. Kessler IM, Pacheco YG, Lozzi SP, de Araújo AS, Jr., Onishi FJ, de Mello PA. Endothelin-1 levels in plasma and cerebrospinal fluid of patients with cerebral vasospasm after aneurysmal subarachnoid hemorrhage. Surg Neurol. 2005; 64 Suppl 1:S1:2–5; discussion S1:5. https://doi.org/10.1016/j.surneu.2005.04.014
- Benigni A, Remuzzi G. Endothelin antagonists. Lancet. 1999;353:133–8. https://doi.org/10.1016/s0140-6736(98) 09423-9.
- Ziv I, Fleminger G, Djaldetti R, Achiron A, Melamed E, Sokolovsky M. Increased plasma endothelin-1 in acute ischemic stroke. Stroke. 1992;23:1014–6. https://doi.org/10.1161/01.str. 23.7.1014.
- 467. Hamann GF, Isenberg E, Strittmatter M, Moili R, Schimrigk K. Big-endothelin in acute ischemic stroke. J Stroke Cerebrovasc Dis. 1993;3:256–60. https://doi.org/10.1016/s1052-3057(10) 80070-9.
- 468. Fuxe K, Bjelke B, Andbjer B, Grahn H, Rimondini R, Agnati LF. Endothelin-1 induced lesions of the frontoparietal cortex of the rat. A possible model of focal cortical ischemia. Neuroreport. 1997; 8:2623–9. https://doi.org/10.1097/00001756-199707280-00040
- 469. Abeysinghe HCS, Bokhari L, Dusting GJ, Roulston CL. Brain remodelling following endothelin-1 induced stroke in conscious rats. PloS one. 2014; 9:e97007-e. https://doi.org/10.1371/journ al.pone.0097007
- Blasi F, Whalen MJ, Ayata C. Lasting pure-motor deficits after focal posterior internal capsule white-matter infarcts in rats. J



Cereb Blood Flow Metab. 2015;35:977–84. https://doi.org/10.1038/jcbfm.2015.7.

- Tennant KA, Jones TA. Sensorimotor behavioral effects of endothelin-1 induced small cortical infarcts in C57BL/6 mice. J Neurosci Methods. 2009;181:18–26. https://doi.org/10.1016/j. ineumeth.2009.04.009.
- 472. Hughes PM, Anthony DC, Ruddin M, Botham MS, Rankine EL, Sablone M, Baumann D, Mir AK, Perry VH. Focal lesions in the rat central nervous system induced by endothelin-1. J Neuropathol Exp Neurol. 2003;62:1276–86. https://doi.org/10.1093/inen/62.12.1276.
- 473. Robinson MJ, Macrae IM, Todd M, Reid JL, McCulloch J. Reduction of local cerebral blood flow to pathological levels by endothelin-1 applied to the middle cerebral artery in the rat. Neurosci Lett. 1990;118:269–72. https://doi.org/10.1016/0304-3940(90)90644-o.
- 474. Bogaert L, Scheller D, Moonen J, Sarre S, Smolders I, Ebinger G, Michotte Y. Neurochemical changes and laser Doppler flowmetry in the endothelin-1 rat model for focal cerebral ischemia. Brain Res. 2000;887:266–75. https://doi.org/10.1016/s0006-8993(00) 02959-0.
- 475. Frost SB, Barbay S, Mumert ML, Stowe AM, Nudo RJ. An animal model of capsular infarct: endothelin-1 injections in the rat. Behav Brain Res. 2006;169:206–11. https://doi.org/10.1016/j.bbr.2006.01.014.
- 476. Ward NM, Sharkey J, Marston HM, Brown VJ. Simple and choice reaction-time performance following occlusion of the anterior cerebral arteries in the rat. Exp Brain Res. 1998;123:269–81. https://doi.org/10.1007/s002210050569.
- 477. Virley D, Hadingham SJ, Roberts JC, Farnfield B, Elliott H, Whelan G, Golder J, David C, Parsons AA, Hunter AJ. A new primate model of focal stroke: endothelin-1-induced middle cerebral artery occlusion and reperfusion in the common marmoset. J Cereb Blood Flow Metab. 2004;24:24–41. https://doi.org/10.1097/01.Wcb.0000095801.98378.4a.
- 478. Teo L, Bourne JA. A reproducible and translatable model of focal ischemia in the visual cortex of infant and adult marmoset monkeys. Brain Pathol. 2014;24:459–74. https://doi.org/10.1111/ bpa.12129.
- 479. Dai P, Huang H, Zhang L, He J, Zhao X, Yang F, Zhao N, Yang J, Ge L, Lin Y, et al. A pilot study on transient ischemic stroke induced with endothelin-1 in the rhesus monkeys. Sci Rep. 2017;7:45097. https://doi.org/10.1038/srep45097.
- 480. Vatter H, Zimmermann M, Tesanovic V, Raabe A, Seifert V, Schilling L. Cerebrovascular characterization of clazosentan, the first nonpeptide endothelin receptor antagonist shown to be clinically effective for the treatment of cerebral vasospasm. Part II: effect on endothelin(B) receptor-mediated relaxation. J Neurosurg. 2005; 102:1108–14. https://doi.org/10.3171/jns.2005.102.6. 1108
- 481. Matsuo Y, Mihara S, Ninomiya M, Fujimoto M. Protective effect of endothelin type A receptor antagonist on brain edema and injury after transient middle cerebral artery occlusion in rats. Stroke. 2001;32:2143–8. https://doi.org/10.1161/hs0901.94259.
- 482. Nakagomi S, Kiryu-Seo S, Kiyama H. Endothelin-converting enzymes and endothelin receptor B messenger RNAs are expressed in different neural cell species and these messenger RNAs are coordinately induced in neurons and astrocytes respectively following nerve injury. Neuroscience. 2000;101:441–9. https://doi.org/10.1016/s0306-4522(00)00345-6.
- 483. Uesugi M, Kasuya Y, Hayashi K, Goto K. SB209670, a potent endothelin receptor antagonist, prevents or delays axonal degeneration after spinal cord injury. Brain Res. 1998;786:235–9. https://doi.org/10.1016/s0006-8993(97)01431-5.

- 484. Ubhi K, Masliah E. Alzheimer's disease: recent advances and future perspectives. J Alzheimers Dis. 2013;33(Suppl 1):S185– 94. https://doi.org/10.3233/jad-2012-129028.
- 485. Wang DB, Dayton RD, Zweig RM, Klein RL. Transcriptome analysis of a tau overexpression model in rats implicates an early pro-inflammatory response. Exp Neurol. 2010;224:197–206. https://doi.org/10.1016/j.expneurol.2010.03.011.
- 486. Palmer JC, Tayler HM, Love S. Endothelin-converting enzyme-1 activity, endothelin-1 production, and free radical-dependent vasoconstriction in Alzheimer's disease. J Alzheimers Dis. 2013;36:577–87. https://doi.org/10.3233/jad-130383.
- Luo J, Grammas P. Endothelin-1 is elevated in Alzheimer's disease brain microvessels and is neuroprotective. J Alzheimers Dis. 2010;21:887–96. https://doi.org/10.3233/jad-2010-091486.
- 488. Pacheco-Quinto J, Eckman EA. Endothelin-converting enzymes degrade intracellular β-amyloid produced within the endosomal/lysosomal pathway and autophagosomes. J Biol Chem. 2013;288:5606–15. https://doi.org/10.1074/jbc.M112.422964.
- 489. Pacheco-Quinto J, Herdt A, Eckman CB, Eckman EA. Endothelin-converting enzymes and related metalloproteases in Alzheimer's disease. Journal of Alzheimer's disease: JAD. 2013;33(Suppl 1):S101-10. https://doi.org/10.3233/ JAD-2012-129043.
- 490. Paris D, Humphrey J, Quadros A, Patel N, Crescentini R, Crawford F, Mullan M. Vasoactive effects of A beta in isolated human cerebrovessels and in a transgenic mouse model of Alzheimer's disease: role of inflammation. Neurol Res. 2003;25:642–51. https://doi.org/10.1179/016164103101201940.
- 491. Palmer JC, Baig S, Kehoe PG, Love S. Endothelin-converting enzyme-2 is increased in Alzheimer's disease and up-regulated by Abeta. Am J Pathol. 2009;175:262–70. https://doi.org/10. 2353/ajpath.2009.081054.
- 492. Hammond TR, Gadea A, Dupree J, Kerninon C, Nait-Oumesmar B, Aguirre A, Gallo V. Astrocyte-derived endothelin-1 inhibits remyelination through notch activation. Neuron. 2014;81:588–602. https://doi.org/10.1016/j.neuron.2013.11.015.
- 493. D'Antoni S, Ranno E, Spatuzza M, Cavallaro S, Catania MV. Endothelin-1 induces degeneration of cultured motor neurons through a mechanism mediated by nitric oxide and PI3K/Akt pathway. Neurotox Res. 2017;32:58–70. https://doi.org/10.1007/ s12640-017-9711-3.
- 494. Nelson JB, Hedican SP, George DJ, Reddi AH, Piantadosi S, Eisenberger MA, Simons JW. Identification of endothelin-1 in the pathophysiology of metastatic adenocarcinoma of the prostate. Nat Med. 1995;1:944–9. https://doi.org/10.1038/ nm0995-944.
- 495. Lalich M, McNeel DG, Wilding G, Liu G. Endothelin receptor antagonists in cancer therapy. Cancer Invest. 2007;25:785–94. https://doi.org/10.1080/07357900701522588.
- 496. Pirtskhalaishvili G, Nelson JB. Endothelium-derived factors as paracrine mediators of prostate cancer progression. Prostate. 2000;44:77–87. https://doi.org/10.1002/1097-0045(20000615) 44:1%3c77::aid-pros10%3e3.0.co;2-g.
- Quang PN, Schmidt BL. Peripheral endothelin B receptor agonist-induced antinociception involves endogenous opioids in mice. Pain. 2010;149:254–62. https://doi.org/10.1016/j.pain. 2010.02.009.
- 498. Donninger H, Bonome T, Radonovich M, Pise-Masison CA, Brady J, Shih JH, Barrett JC, Birrer MJ. Whole genome expression profiling of advance stage papillary serous ovarian cancer reveals activated pathways. Oncogene. 2004;23:8065–77. https:// doi.org/10.1038/sj.onc.1207959.
- 499. Jazaeri AA, Awtrey CS, Chandramouli GV, Chuang YE, Khan J, Sotiriou C, Aprelikova O, Yee CJ, Zorn KK, Birrer MJ, et al. Gene expression profiles associated with response to chemotherapy in epithelial ovarian cancers. Clin Cancer



- Res. 2005;11:6300–10. https://doi.org/10.1158/1078-0432. Ccr-04-2682.
- 500. Scambia G, Benedetti-Panici P, Ferrandina G, Distefano M, Salerno G, Romanini ME, Fagotti A, Mancuso S. Epidermal growth factor, oestrogen and progesterone receptor expression in primary ovarian cancer: correlation with clinical outcome and response to chemotherapy. Br J Cancer. 1995;72:361–6. https://doi.org/10.1038/bjc.1995.339.
- Wakeling AE, Guy SP, Woodburn JR, Ashton SE, Curry BJ, Barker AJ, Gibson KH. ZD1839 (Iressa): an orally active inhibitor of epidermal growth factor signaling with potential for cancer therapy. Cancer Res. 2002;62:5749

 –54.
- 502. Rosanò L, Di Castro V, Spinella F, Tortora G, Nicotra MR, Natali PG, Bagnato A. Combined targeting of endothelin A receptor and epidermal growth factor receptor in ovarian cancer shows enhanced antitumor activity. Cancer Res. 2007;67:6351–9. https://doi.org/10.1158/0008-5472.Can-07-0883.
- Kajiyama H, Shibata K, Terauchi M, Morita T, Ino K, Mizutani S, Kikkawa F. Neutral endopeptidase 24.11/CD10 suppresses progressive potential in ovarian carcinoma in vitro and in vivo. Clin Cancer Res. 2005; 11:1798–808. https://doi.org/10.1158/1078-0432.Ccr-04-2395
- 504. Tocci P, Cianfrocca R, Di Castro V, Rosanò L, Sacconi A, Donzelli S, Bonfiglio S, Bucci G, Vizza E, Ferrandina G, et al. β-arrestin1/YAP/mutant p53 complexes orchestrate the endothelin A receptor signaling in high-grade serous ovarian cancer. Nat Commun. 2019;10:3196. https://doi.org/10.1038/s41467-019-11045-8.
- Smollich M, Wülfing P. The endothelin axis: a novel target for pharmacotherapy of female malignancies. Curr Vasc Pharmacol. 2007;5:239–48. https://doi.org/10.2174/157016107781024082.
- 506. Smollich M, Götte M, Yip GW, Yong ES, Kersting C, Fischgräbe J, Radke I, Kiesel L, Wülfing P. On the role of endothelin-converting enzyme-1 (ECE-1) and neprilysin in human breast cancer. Breast Cancer Res Treat. 2007;106:361–9. https://doi.org/10.1007/s10549-007-9516-9.
- 507. Wülfing P, Diallo R, Kersting C, Wülfing C, Poremba C, Rody A, Greb RR, Böcker W, Kiesel L. Expression of endothelin-1, endothelin-A, and endothelin-B receptor in human breast cancer and correlation with long-term follow-up. Clin Cancer Res. 2003;9:4125–31.
- Flammer J, Konieczka K, Flammer AJ. The primary vascular dysregulation syndrome: implications for eye diseases. EPMA J. 2013;4:14. https://doi.org/10.1186/1878-5085-4-14.
- 509. Maayah ZH, Takahara S, Alam AS, Ferdaoussi M, Sutendra G, El-Kadi AOS, Mackey JR, Pituskin E, Paterson DI, Dyck JRB. Breast cancer diagnosis is associated with relative left ventricular hypertrophy and elevated endothelin-1 signaling. BMC Cancer. 2020;20:751. https://doi.org/10.1186/s12885-020-07217-1.
- 510. Hagemann T, Binder C, Binder L, Pukrop T, Trümper L, Grimshaw MJ. Expression of endothelins and their receptors promotes an invasive phenotype of breast tumor cells but is insufficient to induce invasion in benign cells. DNA Cell Biol. 2005;24:766–76. https://doi.org/10.1089/dna.2005.24.766.
- Grimshaw MJ, Hagemann T, Ayhan A, Gillett CE, Binder C, Balkwill FR. A role for endothelin-2 and its receptors in breast tumor cell invasion. Cancer Res. 2004;64:2461–8. https://doi. org/10.1158/0008-5472.can-03-1069.
- 512. Dréau D, Karaa A, Culberson C, Wyan H, McKillop IH, Clemens MG. Bosentan inhibits tumor vascularization and bone metastasis in an immunocompetent skin-fold chamber model of breast carcinoma cell metastasis. Clin Exp Metastasis. 2006;23:41–53. https://doi.org/10.1007/s10585-006-9016-z.
- 513. Halaka M, Hired ZA, Rutledge GE, Hedgepath CM, Anderson MP, St John H, Do JM, Majmudar PR, Walker C, Alawawdeh A, et al. Differences in endothelin B receptor isoforms expression

- and function in breast cancer cells. J Cancer. 2020;11:2688–701. https://doi.org/10.7150/jca.41004.
- 514. Bagnato A, Spinella F, Rosanò L. The endothelin axis in cancer: the promise and the challenges of molecularly targeted therapy. Can J Physiol Pharmacol. 2008;86:473–84. https://doi.org/10. 1139/y08-058.
- Carducci MA, Jimeno A. Targeting bone metastasis in prostate cancer with endothelin receptor antagonists. Clin Cancer Res. 2006;12:6296s-s6300. https://doi.org/10.1158/1078-0432. Ccr-06-0929.
- 516. James ND, Borre M, Zonnenberg B, Beuzeboc P, Morris T, Phung D. ZD4054, a potent, specific endothelin A receptor antagonist, improves overall survival in pain-free or mildly symptomatic patients with hormone-resistant prostate cancer (HRPC) and bone metastases. Eur. J. Cancer. 2007; 5:3-. https://doi.org/10.1016/S1359-6349(07)70096-4
- Zhang HJ, Qian WQ, Chen R, Sun ZQ, Song JD, Sheng L. New therapeutic schedule for prostatic cancer-3 cells with ET-1 RNAi and Endostar. Asian Pac J Cancer Prev. 2014;15:10079–83. https://doi.org/10.7314/apjcp.2014.15.23.10079.
- Tsapakidis K, Vlachostergios PJ, Voutsadakis IA, Befani CD, Patrikidou A, Hatzidaki E, Daliani DD, Moutzouris G, Liakos P, Papandreou CN. Bortezomib reverses the proliferative and antiapoptotic effect of neuropeptides on prostate cancer cells. Int J Urol. 2012;19:565–74. https://doi.org/10.1111/j.1442-2042. 2012.02967.x.
- 519. Egidy G, Juillerat-Jeanneret L, Jeannin JF, Korth P, Bosman FT, Pinet F. Modulation of human colon tumor-stromal interactions by the endothelin system. Am J Pathol. 2000;157:1863–74. https://doi.org/10.1016/s0002-9440(10)64825-0.
- Kim TH, Xiong H, Zhang Z, Ren B. beta-Catenin activates the growth factor endothelin-1 in colon cancer cells. Oncogene. 2005;24:597–604. https://doi.org/10.1038/sj.onc.1208237.
- Hoosein MM, Dashwood MR, Dawas K, Ali HM, Grant K, Savage F, Taylor I, Loizidou M. Altered endothelin receptor subtypes in colorectal cancer. Eur J Gastroenterol Hepatol. 2007;19:775–82. https://doi.org/10.1097/MEG.0b013e3282c563de.
- 522. Wang Z, Liu P, Zhou X, Wang T, Feng X, Sun YP, Xiong Y, Yuan HX, Guan KL. Endothelin promotes colorectal tumorigenesis by activating YAP/TAZ. Cancer Res. 2017;77:2413–23. https://doi.org/10.1158/0008-5472.Can-16-3229.
- 523. Wei F, Ge Y, Li W, Wang X, Chen B. Role of endothelin receptor type B (EDNRB) in lung adenocarcinoma. Thoracic cancer. 2020;11:1885–90. https://doi.org/10.1111/1759-7714.13474.
- 524. Schäfer A, Haenig B, Erupathil J, Strickner P, Sabato D, Welford RWD, Klaeylé L, Simon E, Krepler C, Brafford P, et al. Inhibition of endothelin-B receptor signaling synergizes with MAPK pathway inhibitors in BRAF mutated melanoma. Oncogene. 2021;40:1659–73. https://doi.org/10.1038/s41388-020-01628-x.
- 525. Hsieh WT, Yeh WL, Cheng RY, Lin C, Tsai CF, Huang BR, Wu CY, Lin HY, Huang SS, Lu DY. Exogenous endothelin-1 induces cell migration and matrix metalloproteinase expression in U251 human glioblastoma multiforme. J Neurooncol. 2014;118:257–69. https://doi.org/10.1007/s11060-014-1442-1.
- 526. Kusuhara M, Yamaguchi K, Nagasaki K, Hayashi C, Suzaki A, Hori S, Handa S, Nakamura Y, Abe K. Production of endothelin in human cancer cell lines. Cancer Res. 1990;50:3257–61.
- 527. Oikawa T, Kushuhara M, Ishikawa S, Hitomi J, Kono A, Iwanaga T, Yamaguchi K. Production of endothelin-1 and thrombomodulin by human pancreatic cancer cells. Br J Cancer. 1994;69:1059–64. https://doi.org/10.1038/bjc.1994.208.
- Cook N, Brais R, Qian W, Hak CC, Corrie PG. Endothelin-1 and endothelin B receptor expression in pancreatic adenocarcinoma. J Clin Pathol. 2015;68:309–13. https://doi.org/10.1136/ jclinpath-2014-202521.



 Rajeshkumar NV, Matwyshyn G, Gulati A. IRL-1620, a tumor selective vasodilator, augments the uptake and efficacy of chemotherapeutic agents in prostate tumor rats. Prostate. 2007;67:701– 13. https://doi.org/10.1002/pros.20556.

- Bagnato A, Rosanò L. The endothelin axis in cancer. Int J Biochem Cell Biol. 2008;40:1443–51. https://doi.org/10.1016/j.biocel.2008.01.022.
- Enevoldsen FC, Sahana J, Wehland M, Grimm D, Infanger M, Krüger M. Endothelin receptor antagonists: status quo and future perspectives for targeted therapy. J Clin Med. 2020;9:824. https:// doi.org/10.3390/jcm9030824.
- Moriwaki K, Chan FK-M. RIP3: a molecular switch for necrosis and inflammation. Genes & development. 2013; 27:1640–9. https://doi.org/10.1101/gad.223321.113
- 533. Abdul Y, Ward R, Dong G, Ergul A. Lipopolysaccharide-induced necroptosis of brain microvascular endothelial cells can be prevented by inhibition of endothelin receptors. Physiol Res. 2018; 67:S227-s36. https://doi.org/10.33549/physiolres.933842
- 534. Iglarz M, Steiner P, Wanner D, Rey M, Hess P, Clozel M. Vascular effects of endothelin receptor antagonists depends on their selectivity for ETA versus ETB receptors and on the functionality of endothelial ETB receptors. J Cardiovasc Pharmacol. 2015;66:332–7. https://doi.org/10.1097/fjc.00000000000000283.
- 535. Bellisai F, Morozzi G, Scaccia F, Chellini F, Simpatico A, Pecetti G, Galeazzi M. Evaluation of the effect of Bosentan treatment on proinflammatory cytokine serum levels in patients affected by Systemic Sclerosis. Int J Immunopathol Pharmacol. 2011;24:261–4. https://doi.org/10.1177/039463201102400134.
- Tcherakian C, Rivaud E, Catherinot E, Zucman D, Metivier AC, Couderc LJ. Pulmonary arterial hypertension related to HIV: is inflammation related to IL-6 the cornerstone? Rev Pneumol Clin. 2011;67:250–7. https://doi.org/10.1016/j.pneumo.2011.06.006.
- 537. Funke C, Farr M, Werner B, Dittmann S, Überla K, Piper C, Niehaus K, Horstkotte D. Antiviral effect of bosentan and valsartan during coxsackievirus B3 infection of human endothelial cells. J Gen Virol. 2010;91:1959–70. https://doi.org/10.1099/vir.0.020065-0.
- 538. Guo Q, Huang JA, Fraidenburg DR. Bosentan as rescue treatment in refractory hypoxemia and pulmonary hypertension in a patient with ARDS and H7N9 influenza virus infection. Lung. 2014;192:635–6. https://doi.org/10.1007/s00408-014-9602-9.
- Braith RW, Stewart KJ. Resistance exercise training: its role in the prevention of cardiovascular disease. Circulation. 2006;113:2642–50. https://doi.org/10.1161/circulationaha.105. 584060.
- Kang SJ, Jung SL. Effect of 12 weeks aerobic exercise on obesity index, insulin resistance, cardiovascular disease risk factors and exercise capacity in obese adolescent. Exerc Sci. 2010; 19:277– 88. https://doi.org/10.15857/ksep.2010.19.3.277
- 541. Dhaun N, Goddard J, Kohan DE, Pollock DM, Schiffrin EL, Webb DJ. Role of endothelin-1 in clinical hypertension: 20 years on. Hypertension. 2008;52:452–9. https://doi.org/10.1161/hypertensionaha.108.117366.
- 542. Parrinello G, Scaglione R, Pinto A, Corrao S, Cecala M, Di Silvestre G, Amato P, Licata A, Licata G. Central obesity and hypertension: the role of plasma endothelin. Am J Hypertens. 1996;9:1186–91. https://doi.org/10.1016/s0895-7061(96) 00259-2.
- 543. Barton M, Nett P, Amann K, Teixeira M. Anti-inflammatory effects of endothelin receptor antagonists and their importance for treating human disease. In; 2010.
- 544. Wienbergen H, Hambrecht R. Physical exercise and its effects on coronary artery disease. Curr Opin Pharmacol. 2013;13:218–25. https://doi.org/10.1016/j.coph.2012.12.003.
- 545. Franklin SS, Pierce GL. Cardiorespiratory fitness and the attenuation of age-related rise in blood pressure: an important

- role for effective primordial prevention. J Am Coll Cardiol. 2014;64:1254–6. https://doi.org/10.1016/j.jacc.2014.06.1183.
- 546. Liu J, Sui X, Lavie CJ, Zhou H, Park YM, Cai B, Liu J, Blair SN. Effects of cardiorespiratory fitness on blood pressure trajectory with aging in a cohort of healthy men. J Am Coll Cardiol. 2014;64:1245–53. https://doi.org/10.1016/j.jacc.2014.06.1184.
- 547. Caulin-Glaser T, Falko J, Hindman L, La Londe M, Snow R. Cardiac rehabilitation is associated with an improvement in C-reactive protein levels in both men and women with cardio-vascular disease. J Cardiopulm Rehabil. 2005; 25:332–6; quiz 7–8. https://doi.org/10.1097/00008483-200511000-00003
- Gielen S, Walther C, Schuler G, Hambrecht R. Anti-inflammatory effects of physical exercise. A new mechanism to explain the benefits of cardiac rehabilitation? J Cardiopulm Rehabil. 2005; 25:339–42. https://doi.org/10.1097/00008483-200511000-00005
- Barton M, Haudenschild CC. Endothelium and atherogenesis: endothelial therapy revisited. J Cardiovasc Pharmacol. 2001;38(Suppl 2):S23–5. https://doi.org/10.1097/00005344-200111002-00007.
- Traupe T, Ortmann J, Münter K, Barton M. Endothelial therapy of atherosclerosis and its risk factors. Curr Vasc Pharmacol. 2003;1:111–21. https://doi.org/10.2174/1570161033476763.
- 551. Thijssen DH, Rongen GA, van Dijk A, Smits P, Hopman MT. Enhanced endothelin-1-mediated leg vascular tone in healthy older subjects. J Appl Physiol. 1985;2007(103):852–7. https:// doi.org/10.1152/japplphysiol.00357.2007.
- Peacock AJ, Zamboni W, Vizza CD. Ambrisentan for the treatment of adults with pulmonary arterial hypertension: a review. Curr Med Res Opin. 2015;31:1793–807. https://doi.org/10.1185/03007995.2015.1074890.
- 553. Chaumais MC, Guignabert C, Savale L, Jaïs X, Boucly A, Montani D, Simonneau G, Humbert M, Sitbon O. Clinical pharmacology of endothelin receptor antagonists used in the treatment of pulmonary arterial hypertension. Am J Cardiovasc Drugs. 2015;15:13–26. https://doi.org/10.1007/s40256-014-0095-y.
- 554. Gatfield J, Mueller Grandjean C, Sasse T, Clozel M, Nayler O. Slow receptor dissociation kinetics differentiate macitentan from other endothelin receptor antagonists in pulmonary arterial smooth muscle cells. PLoS ONE. 2012;7: e47662. https://doi.org/10.1371/journal.pone.0047662.
- 555. Sidharta PN, Krähenbühl S, Dingemanse J. Pharmacokinetic and pharmacodynamic evaluation of macitentan, a novel endothelin receptor antagonist for the treatment of pulmonary arterial hypertension. Expert Opin Drug Metab Toxicol. 2015;11:437–49. https://doi.org/10.1517/17425255.2015.1000859.
- 556. Bedan M, Grimm D, Wehland M, Simonsen U, Infanger M, Krüger M. A focus on macitentan in the treatment of pulmonary arterial hypertension. Basic Clin Pharmacol Toxicol. 2018;123:103–13. https://doi.org/10.1111/bcpt.13033.
- Clozel M, Salloukh H. Role of endothelin in fibrosis and antifibrotic potential of bosentan. Ann Med. 2005;37:2–12. https:// doi.org/10.1080/07853890410018925.
- 558. Zimmermann M, Seifert V. Endothelin and subarachnoid hemorrhage: an overview. Neurosurgery. 1998; 43:863–75; discussion 75–6. https://doi.org/10.1097/00006123-199810000-00083
- Chow M, Dumont AS, Kassell NF. Endothelin receptor antagonists and cerebral vasospasm: an update. Neurosurgery. 2002;
 51:1333–41; discussion 42.
- Macdonald RL. Clazosentan: an endothelin receptor antagonist for treatment of vasospasm after subarachnoid hemorrhage. Expert Opin Investig Drugs. 2008;17:1761–7. https://doi.org/ 10.1517/13543784.17.11.1761.
- Armstead WM. Endothelins and the role of endothelin antagonists in the management of posttraumatic vasospasm. Curr Pharm Des. 2004;10:2185–92. https://doi.org/10.2174/1381612043 384178.



 Maguire JJ, Davenport AP. Endothelin@25 - new agonists, antagonists, inhibitors and emerging research frontiers: IUPHAR Review 12. Br J Pharmacol. 2014;171:5555–72. https://doi.org/ 10.1111/bph.12874.

- 563. Galiè N, Barberà JA, Frost AE, Ghofrani H-A, Hoeper MM, McLaughlin VV, Peacock AJ, Simonneau G, Vachiery J-L, Grünig E, et al. Initial use of ambrisentan plus tadalafil in pulmonary arterial hypertension. N Engl J Med. 2015;373:834–44. https://doi.org/10.1056/NEJMoa1413687.
- 564. Komers R, Gipson DS, Nelson P, Adler S, Srivastava T, Derebail VK, Meyers KE, Pergola P, MacNally ME, Hunt JL, et al. Efficacy and safety of sparsentan compared with irbesartan in patients with primary focal segmental glomerulosclerosis: randomized, controlled trial design (DUET). Kidney Int Rep. 2017;2:654–64. https://doi.org/10.1016/j.ekir.2017.02.019.
- 565. Gupta RM, Hadaya J, Trehan A, Zekavat SM, Roselli C, Klarin D, Emdin CA, Hilvering CRE, Bianchi V, Mueller C, et al. A genetic variant associated with five vascular diseases is a distal regulator of endothelin-1 gene expression. Cell. 2017;170:522-33.e15. https://doi.org/10.1016/j.cell.2017.06.049.
- Hutchings CJ, Koglin M, Olson WC, Marshall FH. Opportunities for therapeutic antibodies directed at G-protein-coupled receptors. Nat Rev Drug Discov. 2017;16:787–810. https://doi.org/10. 1038/nrd.2017.91.
- Davenport AP, Kuc RE, Southan C, Maguire JJ. New drugs and emerging therapeutic targets in the endothelin signaling pathway and prospects for personalized precision medicine. Physiol Res. 2018; 67:S37-s54. https://doi.org/10.33549/physiolres.933872
- 568. Allard B, Wijkhuisen A, Borrull A, Deshayes F, Priam F, Lamourette P, Ducancel F, Boquet D, Couraud JY. Generation and characterization of rendomab-B1, a monoclonal antibody displaying potent and specific antagonism of the human endothelin B receptor. MAbs. 2013;5:56–69. https://doi.org/10.4161/mabs. 22696.
- Davenport AP, Hyndman KA, Dhaun N, Southan C, Kohan DE, Pollock JS, Pollock DM, Webb DJ, Maguire JJ. Endothelin. Pharmacol Rev. 2016;68:357–418. https://doi.org/10.1124/pr. 115.011833.
- 570. Herbet A, Costa N, Leventoux N, Mabondzo A, Couraud JY, Borrull A, Hugnot JP, Boquet D. Antibodies targeting human endothelin-1 receptors reveal different conformational states in cancer cells. Physiol Res. 2018; 67:S257-s64. https://doi.org/10.33549/physiolres.933848
- 571. Harland SP, Kuc RE, Pickard JD, Davenport AP. Expression of endothelin(A) receptors in human gliomas and meningiomas, with high affinity for the selective antagonist PD156707. Neurosurgery. 1998; 43:890–8; discussion 8–9. https://doi.org/10.1097/ 00006123-199810000-00097
- 572. Blandin V, Vigne P, Breittmayer JP, Frelin C. Allosteric inhibition of endothelin ETA receptors by 3, 5-dibromosalicylic acid. Mol Pharmacol. 2000;58:1461–9. https://doi.org/10.1124/mol. 58.6.1461.
- 573. Warner TD, Allcock GH, Vane JR. Reversal of established responses to endothelin-1 in vivo and in vitro by the endothelin

- receptor antagonists, BQ-123 and PD 145065. Br J Pharmacol. 1994;112:207–13. https://doi.org/10.1111/j.1476-5381.1994. tb13053.x.
- 574. Pierre LN, Davenport AP. Blockade and reversal of endothelininduced constriction in pial arteries from human brain. Stroke. 1999;30:638–43. https://doi.org/10.1161/01.str.30.3.638.
- 575. Kowala MC, Murugesan N, Tellew J, Carlson K, Monshizadegan H, Ryan C, Gu Z, Kane B, Fadnis L, Baska RA, et al. Novel dual action AT1 and ETA receptor antagonists reduce blood pressure in experimental hypertension. J Pharmacol Exp Ther. 2004;309:275–84. https://doi.org/10.1124/jpet.103.055855.
- 576. Murugesan N, Gu Z, Fadnis L, Tellew JE, Baska RA, Yang Y, Beyer SM, Monshizadegan H, Dickinson KE, Valentine MT, et al. Dual angiotensin II and endothelin A receptor antagonists: synthesis of 2'-substituted N-3-isoxazolyl biphenylsulfonamides with improved potency and pharmacokinetics. J Med Chem. 2005;48:171–9. https://doi.org/10.1021/jm049548x.
- 577. Trachtman H, Nelson P, Adler S, Campbell KN, Chaudhuri A, Derebail VK, Gambaro G, Gesualdo L, Gipson DS, Hogan J, et al. DUET: a phase 2 study evaluating the efficacy and safety of sparsentan in patients with FSGS. J Am Soc Nephrol. 2018;29:2745–54. https://doi.org/10.1681/asn.2018010091.
- 578. Kuntz M, Leiva-Juarez MM, Luthra S. Systematic review of randomized controlled trials of endothelin receptor antagonists for pulmonary arterial hypertension. Lung. 2016;194:723–32. https://doi.org/10.1007/s00408-016-9928-6.
- 579. Liang F, Yang S, Yao L, Belardinelli L, Shryock J. Ambrisentan and tadalafil synergistically relax endothelin-induced contraction of rat pulmonary arteries. Hypertension. 2012;59:705–11. https://doi.org/10.1161/hypertensionaha.111.182261.
- 580. Hoeper MM, McLaughlin VV, Barberá JA, Frost AE, Ghofrani HA, Peacock AJ, Simonneau G, Rosenkranz S, Oudiz RJ, White RJ, et al. Initial combination therapy with ambrisentan and tadalafil and mortality in patients with pulmonary arterial hypertension: a secondary analysis of the results from the randomised, controlled AMBITION study. Lancet Respir Med. 2016;4:894–901. https://doi.org/10.1016/s2213-2600(16)30307-1.
- 581. Mercurio V, Mukherjee M, Tedford RJ, Zamanian RT, Khair RM, Sato T, Minai OA, Torres F, Girgis RE, Chin K, et al. Improvement in right ventricular strain with ambrisentan and tadalafil upfront therapy in scleroderma-associated pulmonary arterial hypertension. Am J Respir Crit Care Med. 2018;197:388–91. https://doi.org/10.1164/rccm.201704-0789LE.
- 582. Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, Korbel JO, Marchini JL, McCarthy S, McVean GA, Abecasis GR. A global reference for human genetic variation. Nature. 2015;526:68–74. https://doi.org/10.1038/nature15393.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

