

Antimicrobial Resistance, Virulence and Plasmid Prediction using Whole Genome Sequences (WGS)

Introduction to Bioinformatics – Module 1



**Public Health Alliance for
Genomic Epidemiology**

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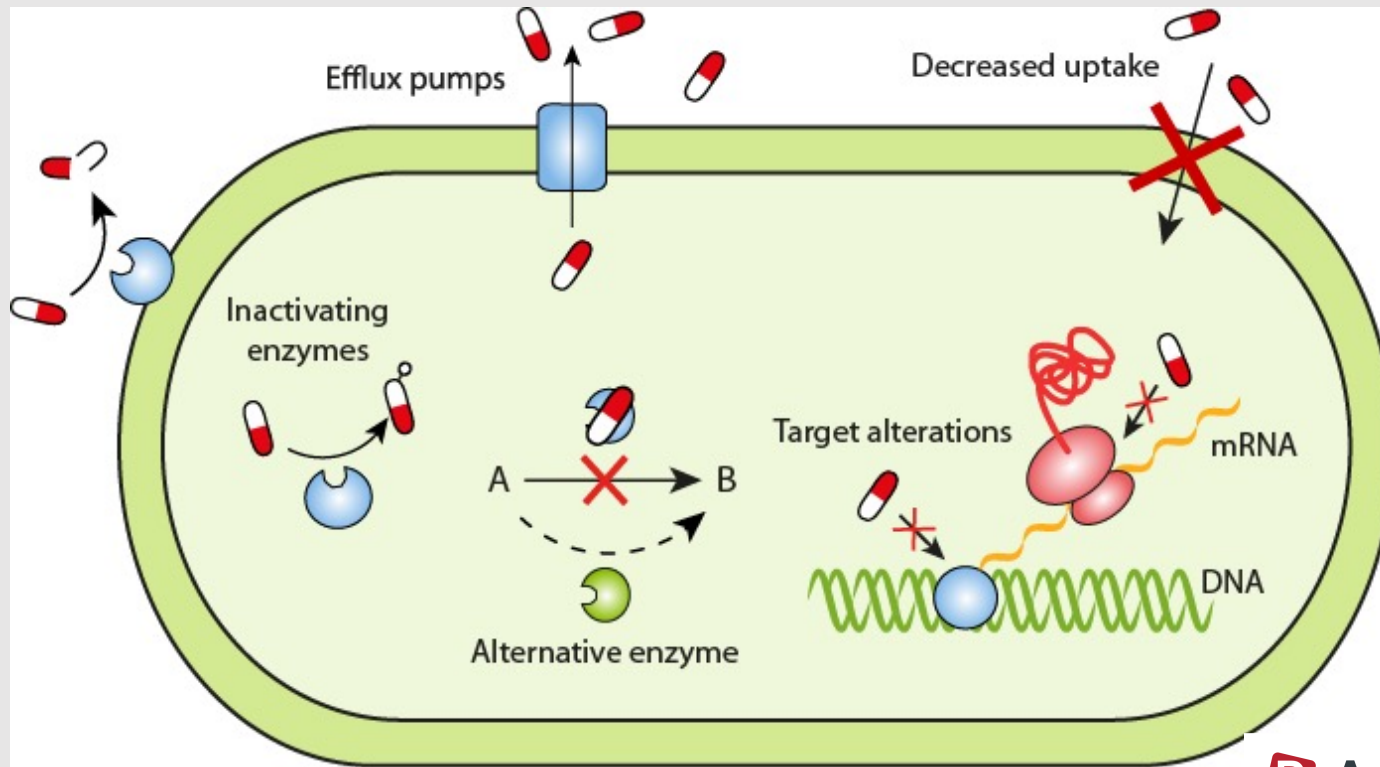


Introduction

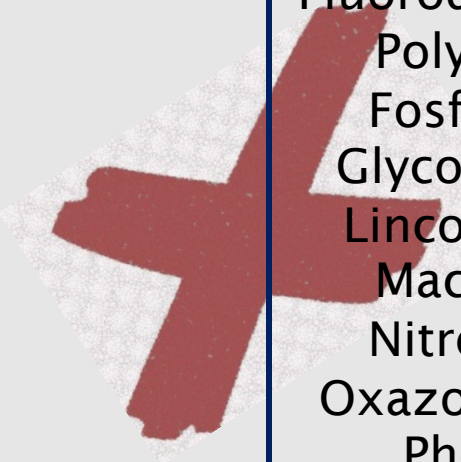
- Antimicrobial resistance (AMR) is a "silent pandemic"
 - Resistance to last resort antibiotics
 - Depleting antibiotics reserves
- Virulence factors promote pathogenicity in bacteria
- Likely mobility of AMR/Virulence genes by plasmids



Mechanism of antibiotic resistance



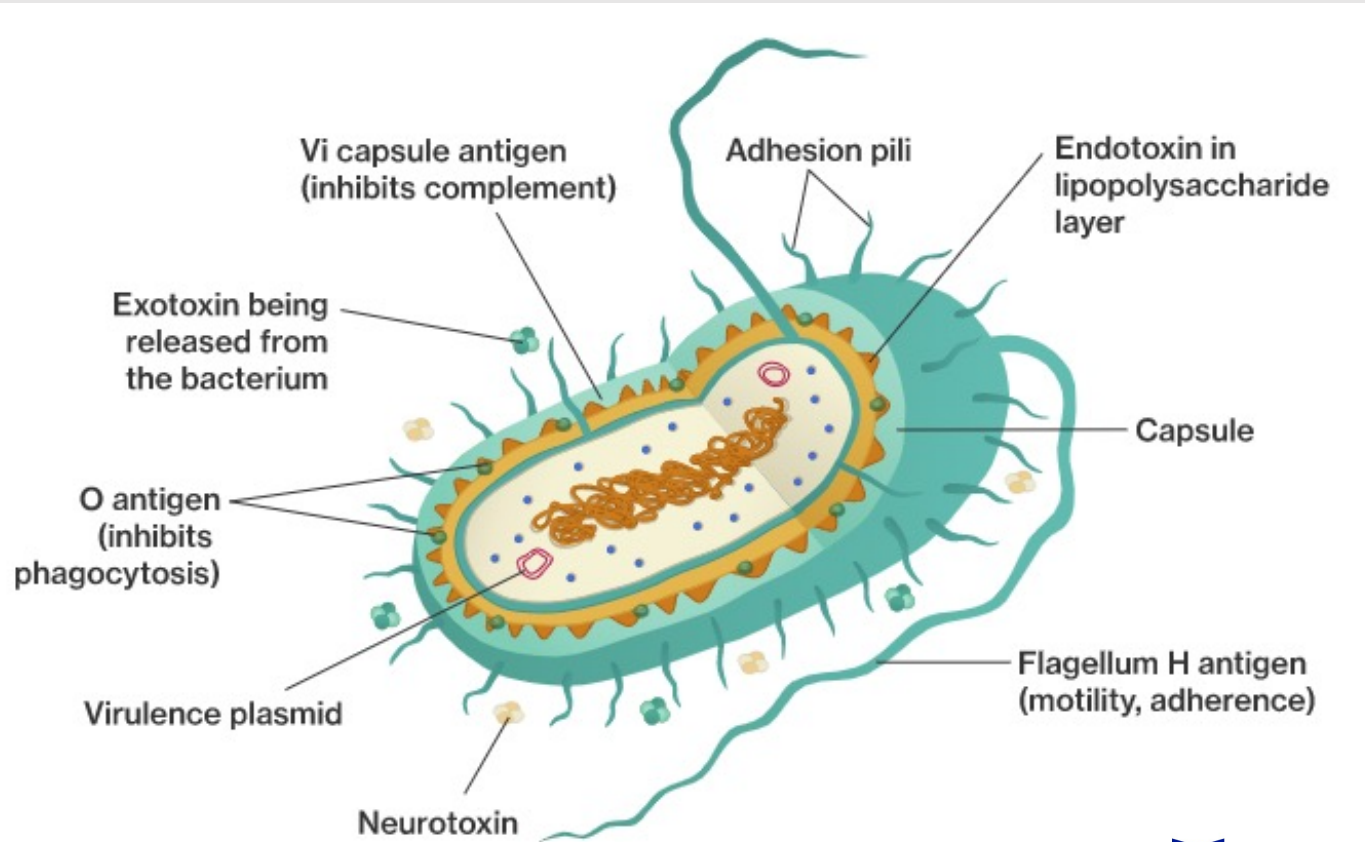
ReAct



Aminoglycosides
Ansamycin
Beta-lactams
Fluoroquinolones
Polymixins
Fosfomycin
Glycopeptides
Lincosamides
Macrolides
Nitrofurans
Oxazolidinones
Phenicol
Quinolones
Sulphonamides
Trimethoprim
Tetracyclines
etc...



Virulence factors in bacteria



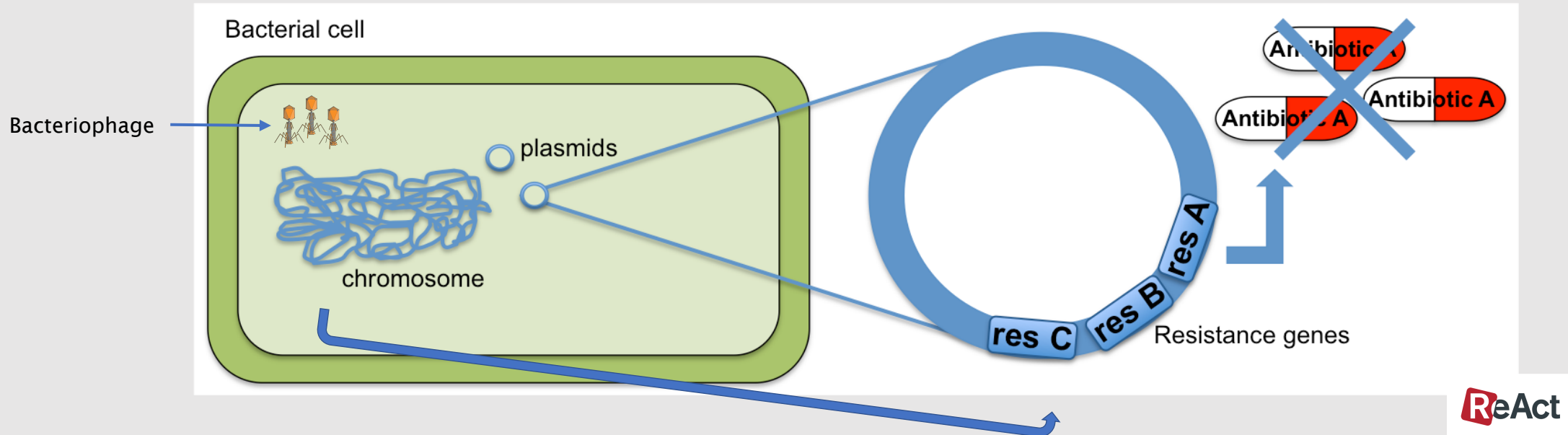
e.g., Capsules
Adhesins
Toxins



1. Invade host
2. Cause disease
3. Evade host defences



DNA structures in a bacterial cell



ReAct

A Bacterial cell may harbour genetic materials in chromosomal or plasmid DNA that promote antibiotic resistance and/or pathogenicity.

Bioinformatic analysis for detection of antimicrobial resistance genes, virulence determinants and plasmid DNA from Whole Genome Sequence Data.



How is AMR predicted using WGS?

Alignment and mapping of WGS against a reference database

- Determine antimicrobial resistance genes
- Chromosomal/point mutations in the sequence data

Databases

- Contain a total of all existing AMR genes
- Contain all existing chromosomal mutations specifying AMR.
- Mutations that lead to porin modifications and efflux pumps that can mediate AMR.



Sequence alignment

- Sequence alignment are core of bioinformatics
- Matching between query and reference sequence

Query seq	ACTGACTT	GC ^A CTGC	TGACGAAT	CTA [↓] CACGAG
Target seq	ACTGACTT	GCT ^T CTGC	TGA_GAAT	CTATCACGAG

Biological Event.	Conservation	Substitution	Insertion	Deletion
Alignment interpretation	Match	Mismatch	Gap	Gap

CACTTACTAGACTGTTAGACTAGCAGATACAG^GACTAGACGAGCATCAT
CTGTCACTTACTAGACTGTTAGACTAGC_GATACAG^CACTAGACGAGCATCATGGACTA

Coverage (width): 83%

Identity: 96%



Some tools used in the prediction of AMR

- DTU CGE (ResFinder)
- ARIBA
- ARG-ANNOT
- MEGARes
- NCBI AMRFinder
- ABRICATE
- CARD
- ARDB
- Galileo AMR etc.

These tools may differ in many ways e.g.,

- Accessibility
- Formats of input and output
- Level of complexity
- User friendliness
- Computational power
- Time to complete analysis

*PHA4GE hAMRonization of output data



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ResFinder 4.1 AMR Prediction

<https://cge.cbs.dtu.dk/services/ResFinder/>

Easy-to-use
Freely available
Good graphical-user interface

Updates

ResFinder and PointFinder software: [\(2021-05-27\)](#)
ResFinder database: [\(2021-04-20\)](#)
PointFinder database: [\(2021-02-01\)](#)

A database is only as good as its
content, hence, need for regular
updates.

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Overview of genes

Article abstract

ResFinder 4.1

ResFinder identifies acquired genes and/or finds chromosomal mutations mediating antimicrobial resistance in total or partial DNA sequence of bacteria.

The database is curated by:
Frank Møller Aarestrup
(click to contact)

Updates

ResFinder and PointFinder software: [\(2021-05-27\)](#)
ResFinder database: [\(2021-04-20\)](#)
PointFinder database: [\(2021-02-01\)](#)

Chromosomal point mutations

Acquired antimicrobial resistance genes

Select species

Campylobacter spp.*

*Chromosomal point mutation database exists

Select type of your reads

Assembled Genome/Contigs

If you get an "Access forbidden. Error 403": Make sure the start of the web address is https and not just http. Fix it by clicking [here](#).

Isolate File

Name	Size	Progress	Status

Upload

Remove



ResFinder 4.1 AMR Prediction

complete			
Antimicrobial	Class	WGS-predicted phenotype	Genetic background
vancomycin	glycopeptide	No resistance	blaSHV-70 (blaSHV-70_DQ013287), blaSHV-13 (blaSHV-13_AF164577), blaSHV-11 (blaSHV-11_X98101)
mupirocin	pseudomonic acid	No resistance	
tobramycin	aminoglycoside	No resistance	
virginiamycin m	streptogramin a	No resistance	
isepamicin	aminoglycoside	No resistance	
virginiamycin s	streptogramin b	No resistance	
hydrogen peroxide	peroxides	No resistance	
butirosin	aminoglycoside	No resistance	
ampicillin	beta-lactam	Resistant	sul1 (sul1_X15024), sul2 (sul2_AY034138)
astromicin	aminoglycoside	No resistance	
lividomycin	aminoglycoside	No resistance	OqxA (OqxA_EU370913), OqxB (OqxB_EU370913), dfrA15 (dfrA15_AF221900)
sulfamethoxazole	folate pathway antagonist	Resistant	
temocillin	beta-lactam	No resistance	acrR (p.R173G), acrR (p.F172S), acrR (p.K201M), acrR (p.F197I), acrR (p.P161R), acrR (p.G164_None491del), acrR (p.L195V)
trimethoprim	folate pathway antagonist	Resistant	
oleandomycin	macrolide	No resistance	
unknown macrolide	macrolide	No resistance	
florfenicol	phenicol	No resistance	fosA (fosA_ACZD01000244)
fluoroquinolone	under_development	Resistant	
quinupristin	streptogramin b	No resistance	blaSHV-11 (blaSHV-11_X98101)
fosfomycin	fosfomycin	Resistant	
cephalothin	beta-lactam	Resistant	



Colour Schemes: ResFinder 4.1 AMR Prediction

Polymyxin									
Resistance gene	Identity	Alignment Length/Gene Length	Position in reference	Contig or Depth	Position in contig	Phenotype	PMID	Accession no.	Notes
mcr-10	100.0	1620/1620	1..1620	NODE_17_lengt h_49013_cov_8. 249131	29724..31343	colistin	32116151	MN179494	

Dark green

- Indicates perfect match for the gene.
- Identity is 100% and the sequence length matches perfectly with reference gene in database



Colour Schemes: ResFinder 4.1 AMR Prediction

Beta-lactam									
Resistance gene	Identity	Alignment Length/Gene Length	Position in reference	Contig or Depth	Position in contig	Phenotype	PMID	Accession no.	Notes
blaSHV-70	99.8838559814	861/861	1..861	NODE_56_length_2052_cov_11.077829	173..1033	amoxicillin,ampicillin,aztreonam,cefepime,cefotaxime,ceftazidime,ceftiaxone,piperacillin,ticarcillin	16563703	DQ013287	Class A

Light green

Indicate imperfect identity due to variations in the bases of the query sequence compared to reference in database.



Colour Schemes: ResFinder 4.1 AMR Prediction

Aminoglycoside									
Resistance gene	Identity	Alignment Length/Gene Length	Position in reference	Contig or Depth	Position in contig	Phenotype	PMID	Accession no.	Notes
aph(3'')-Ib	100.0	803/804	2..804	NODE_17_lengt h_49013_cov_8. 249131	38899..39701	streptomycin	9687410	AF024602	Alternative name strA, orfH

Grey

Indicate imperfect alignment of query and reference gene
in database

Alignment is shorter than reference gene in database



Genotypic prediction of antimicrobial resistance phenotype in *E. coli*

Research

Open Access

Use of whole genome sequencing of commensal *Escherichia coli* in pigs for antimicrobial resistance surveillance, United Kingdom, 2018 | Check for updates

Like 0

Download

Emma Stubberfield¹, Manal AbuOun¹, Ellie Sayers^{1,2}, Heather M O'Connor³, Roderick M Card¹, Muna F Anjum¹

Stubberfield et al., 2019

- Source: caecal content of pigs
- Sample size: 515 *E. coli* isolates
- Exposed to 9 antimicrobials across 7 drug classes



Genotypic prediction of AMR phenotype in *E. coli*

Antibiotic	Ciprofloxacin	Cefotaxime	Ceftazidime	Gentamycin	Florfenicol	Ampicillin	Apramycin	SXT	Tetracycline	Overall
Specificity	96.1%	99.8%	99.8%	100.0%	100.0%	99.1%	100.0%	93.3%	95.0%	98.9%
Sensitivity	85.2%	90.0%	61.6%	95.7%	75.0%	95.9%	100.0%	88.7%	98.0%	90.7%
Kappa Correlation	0.814	0.9431	0.726	0.976	0.848	0.945	1.000	0.812	0.930	0.914

Results

- WGS/MIC specificity of 99%,
- Predictive value of a positive test was 98%
- kappa value of 0.914
- The majority of discrepancies were from false negative correlations.

isolates with reduced susceptibility lacking a relevant genetic resistance determinant, suggesting unknown resistance genes may be present in these bacteria.



Prediction of virulence determinants and plasmid replicons types

WGS sequence data are examined for;

- Presence of absence of virulence determinants
- plasmid replicon types

Uses similar alignment and mapping of genes in a
database



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VirulenceFinder 2.0

Virulence Genes Prediction

<https://cge.cbs.dtu.dk/services/VirulenceFinder/>

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VirulenceFinder 2.0

[Service](#)[Instructions](#)[Output](#)[Article abstract](#)[Version history](#)

Software version: 2.0.3 ([2020-05-21](#))

Database version: ([2020-05-29](#))

The database is curated by:
Flemming Scheutz, SSI
([click to contact](#))

Select species

Listeria
S. aureus
Escherichia coli
Enterococcus

Select threshold for %ID

90 % ▼

Select minimum length

60 % ▼

Select type of your reads

Only data from one single isolate should be uploaded. If raw sequencing reads are uploaded KMA will be used for mapping. KMA supports the following sequencing platforms: Illumina, Ion Torrent, Roche 454, SOLiD, Oxford Nanopore, and PacBio.

Assembled or Draft Genome/Contigs ▼

Choose File(s)

Name

Size

Progress

Status

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PlasmidFinder 2.1

Plasmid Replicon Prediction

<https://cge.cbs.dtu.dk/services/PlasmidFinder/>

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Article abstract

PlasmidFinder 2.1

Software version: [2.0.1 \(2020-07-01\)](#)

Database version: [\(2021-01-13\)](#)

[Test sequence](#)

The database is curated by:
Henrik Hasman and Alessandra Carattoli
(click to contact)

Select database

Select multiple items, with Ctrl-Click (or Cmd-Click on Mac)

Gram Positive
Enterobacteriaceae

Select threshold for minimum % identity

95 %

Select minimum % coverage

60 %

Select type of your reads

Only data from one single isolate should be uploaded. If raw sequencing reads are uploaded KMA will be used for mapping. KMA supports the following sequencing platforms: Illumina, Ion Torrent, Roche454, SOLiD, Oxford Nanopore, and PacBio.

Assembled or Draft Genome/Contigs* (fasta)

Please note that "Assembled Genomes/Contigs" should be selected if you have already assembled your short sequencing reads into one continuous genome or into several contigs. "Assembled Genomes/Contigs" is defined as one or several contigs in one FASTA file (one entry per contig). It is indifferent which type of short sequence reads were used to produce the genome/contigs.

 Isolate File

Name

Size

Progress

Status

 Upload

 Remove



Benefits of WGS AMR, Virulence and Plasmid prediction

- They are vigorous and phenotypically confirmed methods that have been developed and improved over decades
- Provides more information (e.g., description) about detected genes in query sequence
- It can be used to detect new and emerging clones of antimicrobial resistance
- Phenotypic AMR aren't flawless
- Important tool in AMR surveillance.



Limitations to WGS prediction

Detect presence or absence

- no quantitative measurement of resistance/virulence
- genes might not be expressed in host

Detection of plasmid replicons may require further analysis to verify its carriage.



Limitations to WGS prediction

Your results are only as good as the content of database

No gene = no AMR /virulence
True or False?

Kieffer *et al.*, 2018

126 pig sample

8 positive for colistin resistant E. coli

1 possessed *mcr-3*

MIC = 4 μ g/ml

ECOFF= 2 μ g/ml



4 μ g/ml

8 μ g/ml

Limitations associated with
laboratory determination of MIC

A novel resistance gene not yet
in the database



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Thank you



The
**Fleming
Fund**

