

## OPEN ACCESS

Citation: Wala M, Kołodziejek J, Patykowski J (2021) Nitrogen signals and their ecological significance for seed germination of ten psammophilous plant species from European dry acidic grasslands. PLoS ONE 16(1): e0244737. https://doi.org/10.1371/journal.pone.0244737

**Editor:** Yajuan Zhu, Chinese Academy of Forestry, CHINA

Received: July 30, 2020
Accepted: December 16, 2020
Published: January 4, 2021

Copyright: © 2021 Wala et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All relevant data are within the manuscript and its Supporting Information files.

**Funding:** The authors received no specific funding for this work.

**Competing interests:** The authors have declared that no competing interests exist.

RESEARCH ARTICLE

# Nitrogen signals and their ecological significance for seed germination of ten psammophilous plant species from European dry acidic grasslands

Mateusz Wala 61\*, Jeremi Kołodziejek1, Jacek Patykowski2

- 1 Department of Geobotany and Plant Ecology, Faculty of Biology and Environmental Protection, University of Lodz, Łódź, Poland, 2 Komandorska, Łódź, Poland
- \* mateusz.wala@biol.uni.lodz.pl

## Abstract

The presented study evaluated effects of potassium nitrate (KNO<sub>3</sub>), ammonium nitrate (NH<sub>4</sub> NO<sub>3</sub>) and ammonium chloride (NH<sub>4</sub>CI) on the germination-related characteristics of 10 species from European dry acidic grasslands. Germination was studied under controlled laboratory conditions. The seeds were subjected to KNO<sub>3</sub>, NH<sub>4</sub> NO<sub>3</sub> and NH<sub>4</sub>Cl in four doses (1, 10, 50 and 100 mM) and to distilled water. Final germination percentage, index of germination velocity and index of germination synchrony were determined. Content of nitrogen in the soil probed from the site of seeds collection was also analyzed. Significant effects of type of the nitrogen compounds and their concentrations were observed. High concentrations of nitrogen-containing salts inhibited completion of germination in almost all species. Helichrysum arenarium and Hypericum perforatum showed preference for NH<sub>4</sub><sup>+</sup> over NO<sub>3</sub><sup>-</sup>, whereas Arnoseris minima, Alyssum montanum, Jasione montana and Spergula morisonii showed preference for NO<sub>3</sub> over NH<sub>4</sub> . Centaurea scabiosa, C. stoebe and Hypochaeris radicata had no preference and wide tolerance to the type of nitrogen-containing compound. Echium vulgare showed differential response hard for interpretation. A. montanum and J. montana showed stenotopic behavior in terms of nitrogen-related conditions. It is proposed that nitrogen-rich soil gaps favor establishment of more nitro-tolerant plant species (e.g. C. scabiosa, C. stoebe and H. radicata) as compared to nitrogen-poor ones.

#### Introduction

Dry acid grasslands typically occur on free-draining soils overlying acid rocks or superficial deposits such as sands and gravels, which were deposited mostly during and after the last ice age [1]. High surface temperatures, low water storage, low nutrient contents, low organic matter content and litter cover on sandy substrate are the main characteristics of this habitat [2, 3]. The psammophilous grassland from *Spergulo morisonii-Corynephoretum canescentis* (Tx. 1928) Libb. 1933 (syn. *Corniculario aculeatae-Corynephoretum canescentis* Steffen 1931) [4] is a most common plant association formed on those poor and acidic sands. They are composed

of pioneer plant species with wide and low/specialized environmental requirements. According to previous studies [4], the Spergulo-Corynephoretum is the only association within the Corynephorion canescentis Klika 1931 (order Corynephoretalia canescentis, Klika 1934, class Koelerio-Corynephoretea Klika in Klika & Novak 1941). The most characteristic herb and grass species for this association are Achillea millefolium L., Corynephorus canescens (L.) P. Beauv., Helichrysum arenarium (L.) Moench, Pilosella officinarum Vaill. (syn. Hieracium pilosella L.), Hypochaeris radicata L., Jasione montana L., Rumex acetosella L., Scleranthus perennis L., Spergula morisonii Boreau, Teesdalia nudicaulis (L.) W. T. Aiton and Veronica dillenii Crantz [4, 5].

Re-colonizing dynamics in dry acidic grasslands are strongly determined by regeneration characteristics of each species, such as life cycle, dispersal abilities and germination pattern [2]. Thus, it can be expected that at least some vegetation pattern changes observed in field can be explained by differences in germination requirements. Low water holding capacity of the soil and possibly soil nitrogen (N) status are the main factors limiting completion of germination and seedling survival in this kind of environments [6]. Total N concentrations recorded in the soils of Spergulo-Corynephoretum vary between 0.01 and 0.08% of soil dry weight (SDW) (equal to 100-800 mg kg<sup>-1</sup> SDW [2]) which suggests its poor nutritional quality. Regarding N type, there are two major mineral forms available to plants, i.e. ammonium (NH<sub>4</sub><sup>+</sup>) and nitrate (NO<sub>3</sub><sup>-</sup>). On inland sand dunes, NH<sub>4</sub><sup>+</sup> soil concentration is typically higher than that of NO<sub>3</sub><sup>-</sup> [7–9]. Low net N mineralization and greater concentration of NH<sub>4</sub><sup>+</sup> than NO<sub>3</sub><sup>-</sup> are connected with N circulation process and result from halted nitrification process in pioneer sand-settled communities [9]. Thus, acidic soils provide NH<sub>4</sub><sup>+</sup> as the predominant chemical form of N and, interestingly, this form of N is preferred by acidophilous plant species, while more or less calciphilous species prefer  $NO_3^-$  [10]. However, this is totally opposite to molecular basis of N uptake, where easier scavenging of NO<sub>3</sub> by plants can be observed in acidic soils due to greater availability of H<sup>+</sup> for NO<sub>3</sub><sup>-</sup>/H<sup>+</sup> symport mechanism [11].

The composition and diversity of species as well as primary productivity of terrestrial ecosystems are strongly affected by the sources of limiting nutrients, e.g. N [12]. It is an essential macronutrient for plants and is required for many primary and secondary metabolic processes. Regarding germination process, N may modulate germination in many plant species, both alone and in combination with other abiotic factors, e.g. changing temperature or lightdark cycle [13-15]. While the stimulating role of NO<sub>3</sub> on completion of seed germination was well recognized [15, 16], mode of action of NH<sub>4</sub><sup>+</sup> is more enigmatic. NH<sub>4</sub><sup>+</sup>-driven alternations of seed metabolism during germination are associated with decelerated mobilization of seed resources [17] which may reduce germination-related parameters [18] however NH<sub>4</sub><sup>+</sup>-tolerant plants are less susceptible than intolerant ones. These harmful effects are related to toxic action of NH<sub>4</sub><sup>+</sup> [19], especially in tissues with not overdriven metabolism, where NH<sub>4</sub><sup>+</sup> cannot be directly incorporated into bio-molecules. It must be mentioned that besides N availability, pH is another major factor limiting plant growth on acid soils. As an example, pH of the soils on which dry inland grasslands (Spergulo-Corynephoretum) are settled ranges 3.6–6.8 [2]. As plants are able to absorb N in cationic (e.g. NH<sub>4</sub><sup>+</sup>) and anionic (e.g. NO<sub>3</sub><sup>-</sup>) forms, uptake of nitrogen-containing compounds influences the pH in rooting substratum [20, 21]. It is known that pH values of 1-100 mM solutions of KNO<sub>3</sub>, NH<sub>4</sub>NO<sub>3</sub> and NH<sub>4</sub>Cl are slightly different [14]. Therefore, uptake and utilization of N by plant seeds and then seedlings can lead to gradual depletion of N sources on a local scale which may shift the pH status and create microhabitats, especially in sandy soils lacking developed sorption complex and when speciescharacteristic N uptake pattern can be observed.

Only a few studies focused on the germination requirements of plant species in dry sandy grasslands [22, 23]. Differential effect of NO<sub>3</sub><sup>-</sup> and NH<sub>4</sub><sup>+</sup> on plant growth and yield [18] as

well as the importance of N for development and persistence of acidic dry grasslands were analyzed [2] however the comparative studies on influence of N type and dose on completion of germination are not very common in literature. It can be hypothesized that exposure of seeds to exogenous N-containing salts can significantly affect ability of seeds to complete germination. Furthermore, it can be expected that  $\mathrm{NH_4}^+$  and  $\mathrm{NO_3}^-$  exert different effects on completion of seed germination. It is also not known if the studied plant species are similar in germination requirements. As an output of this study we intended to answer the following questions: 1) is nitrogen a factor controlling seed-based revegetation and 2) is the ability to complete germination under wide range of conditions species dependent. The hypotheses tested are that 1) nitrogen type and dose affect germination-related characteristics of the studied species and 2) responses to nitrogen type and dose vary among plant species.

#### Material and methods

## Selection of species and collection of seeds

Seeds of 10 species (nomenclature follows The Plant List [24]) native to central Europe were used in this study: Alyssum montanum L., Arnoseris minima (L.) Schweigg. & Körte, Centaurea stoebe L., C. scabiosa L., Echium vulgare L., Helichrysum arenarium (L.) Moench, L., Hypochaeris radicata L., Hypericum perforatum L., Jasione montana L. and Spergula morisonii Boreau (syn. S. vernalis Willd.) (Table 1). These species are from 6 families (Brassicaceae, Asteraceae, Boraginaceae, Hypericaceae, Campanulaceae and Caryophyllaceae) with the majority from Asteraceae. The species we studied were selected based on their Ellenberg Indicator Values [25] for soil reaction (R, Table 1). All species we studied are light-demanding,

Table 1. List of the studied species, their seed characteristics and habitat preferences of mature plants.

Species	Abbreviation	Family	Growth form	Seed size [mm] <sup>a</sup>	Seed weight [mg] <sup>b</sup>	ight [mg] <sup>b</sup> Ellenberg Index <sup>c</sup>					
						L	Т	K	F	R	N
Alyssum montanum L.	Amo	Brassicaceae	Perennial herb	1.5-1.9 x 1.1-1.3	$0.368 \pm 0.028$	9	6	4	2	7	1
Arnoseris minima (L.) Schweigg. & Körte	Ami	Asteraceae	Annual herb	1.6-2.2 x 0.6-0.7	$0.215 \pm 0.019$	7	6	2	4	3	3
Centaurea scabiosa L.	Csc	Asteraceae	Perennial herb	4.5-5.0 x 2.0-2.2	$3.095 \pm 0.133$	7	0	3	3	8	4
C. stoebe Tausch	Cst	Asteraceae	Biennial herb	2.5-3.0 x 1.2-1.4	$1.023 \pm 0.034$	8	7	6	2	8	3
Echium vulgare L.	Evu	Boraginaceae	Biennial herb	2.4-2.8 x 1.5-1.8	$1.940 \pm 0.104$	9	6	3	4	8	4
Helichrysum arenarium (L.) Moench	Har	Asteraceae	Perennial herb	0.9-1.1 x 0.4-0.5	$0.043 \pm 0.005$	8	6	7	2	5	1
Hypericum perforatum L.	Нре	Hypericaceae	Perennial herb	1.0-1.2 x 0.5-0.6	$0.060 \pm 0.008$	7	6	5	4	6	3
Hypochaeris radicata L.	Hra	Asteraceae	Perennial herb	5.0-10.0 x 0.5-0.6	$0.578 \pm 0.038$	8	5	3	5	4	3
Jasione montana L.	Jmo	Campanulaceae	Annual/biennial herb	0.6-0.8 x 0.2-0.3	$0.018 \pm 0.005$	7	6	3	3	3	2
Spergula morisonii Boreau	Smo	Caryophyllaceae	Annual herb	1.3-1.7 x 1.3-1.6	$0.293 \pm 0.024$	9	5	4	3	0	2

Nomenclature of studied taxa follows the Plant List [24]

chabitat preference of mature plants follows 1–9 point scale proposed by Ellenberg [25]. L–light requirements ranging 7–9, where 7 denotes semi-light conditions (c.a. 30% of relative illumination) and 9 denotes full light conditions (> 50% of relative illumination); T–temperature requirements ranging 5–7, where 5 denotes species preferring moderately cool to warm conditions (characteristic of montane and submontane conditions, mostly southern Fennoscandia) and 7 denotes species preferring warm conditions (characteristic of North European Plain); K–continentiality requirements ranging 2–7, where 2 denotes atlantic conditions and 7 denotes subcontinental conditions; F–soil moisture requirements ranging 2–5, where 2 denotes dry and extremely dry soils and 5 denotes moist soils; R–soil pH requirements ranging 3–8, where 3 denotes slightly acidic to slightly basic soils and 8 denotes average basic soils originating from limestones. N–nitrogen availability requirements ranging 1–4, where 1 denotes extremely infertile soils and 4 denotes slightly and intermediately fertile soils; 0 –indifferent behavior, wide amplitude or unequal behavior in different areas.

https://doi.org/10.1371/journal.pone.0244737.t001

<sup>&</sup>lt;sup>a</sup>seed size follows literature [28]

bseed weight measured in this study (determined by weighing 100 or 500 air-dried seeds, expressed as mean (SD); n = 4)

require relatively high temperature, medium soil moisture and have low N indicators of ecological behavior, typical of N low habitats (Table 1) [25]. The species we selected were among the most representative and abundant in the patches of *Spergulo-Corynephoretum* located in the eastern village periphery of Niewiesz in central Poland (19°83'N, 10°83'E at 149 m a.s.l.). The studied plant community is settled on Entic Podzol soil. The local climate is temperate (annual precipitation of 587.2 mm and a temperature range of –2.5 to 22.4°C [26]), and the seasons are clearly differentiated. Meteorological data based on 10-year observations (2000–2010) indicated that the mean annual temperature was 8.8°C.

Mature seeds of all species were hand-harvested at the end of the summer of 2016, coinciding with their dispersal period. The seeds were taken from at least 10 individuals of one single, large and representative population. The seeds were collected on privately-owned property, where no specific permissions are required. The seeds were stored dry in paper bags at 20–22°C (relative humidity of 30%) for two weeks and then used for experiments within one month. Stratification of the seeds was not conducted due to the fact that fresh seeds of the species we studied are able to efficiently complete germination [27]. At this stage, the seeds were inspected under binocular magnifying glass; all malformed, wrinkled, damaged or discolored ones were removed from the seed pool. After inspection, the seeds were weighed. Dimensions of the seeds were taken from the literature [28].

## General germination procedures

Ability of the seeds to complete germination was tested in glass Petri dishes ( $\emptyset = 5$  cm). The seeds of each species were mixed before use to fulfil the randomization requirement. The seeds (25 per one Petri dish) were placed on double-layer filter paper and moistened with 3 ml of the tested solutions. To maintain stable and homogenous conditions all dishes were sealed with parafilm. The moisture level was monitored daily and the seeds were re-watered when any loss was observed. Germination test was conducted under 16/8 h photoperiod (light was supplied with white fluorescent tubes with a photon flux density of 52 μmol m<sup>-2</sup>·s<sup>-1</sup>) at 23°C. Close to natural photoperiod was used, due to the fact that the seeds of the species we studied complete germination only under sufficient light conditions [27, 29-31]. The temperature of 23°C was listed as optimal for completion of germination for many herbaceous plant species from the temperate region (including wild-living rangeland species [29, 32] and species from dry acidic grasslands) and represents the mean temperatures at the Lodz Weather Station during June and July, when most seeds germinate in natural habitats. Furthermore, it allows direct comparisons of many species subjected to various chemical treatments [33, 34]. Experiments were repeated four times (n = 4). Due to the fact that completion of germination of the studied species reached plateau after 12 days in the preliminary test, it was recorded daily for 14 days. The seeds that completed germination were removed from the Petri dishes. An emerged radicle was a sign of completed germination. Germination-related traits were determined on the basis of the number of viable seeds. Dead seeds (with soft and brown embryo) were not included in the calculations.

## Treatments and experimental setup

The seeds were treated with 1, 10, 50 or 100 mM solutions of KNO<sub>3</sub>, NH<sub>4</sub>NO<sub>3</sub> or NH<sub>4</sub>Cl. For control, distilled water was used (conductivity < 0.06  $\mu$ S cm<sup>-1</sup>). Thus, the experimental design consisted of 13 experimental groups for each species. By such a range of nitrogen types and doses we intended to simulate conditions which can be observed in nature as well as to create extreme situations which cannot be encountered there. The seeds subjected to distilled water (0 mM of N) germinated under absolute non-N conditions which are very rare in the field.

Table 2. Acidity (pH) of water solutions of nitrogen-containing salts used in this study.

Nitrogen type			pН	
	1 mM	10 mM	50 mM	100 mM
KNO <sub>3</sub>	5.13	5.76	5.86	6.01
NH <sub>4</sub> NO <sub>3</sub>	5.50	6.36	6.80	6.80
NH <sub>4</sub> Cl	5.10	5.65	5.83	5.80

The data follows literature [14].

https://doi.org/10.1371/journal.pone.0244737.t002

Treatment with 100 mM of N was used to simulate the conditions with exceeding concentration of N which is not likely in sand-settled plant communities. Solutions of 1, 10 and 50 mM simulated very poor (regarding N concentration) conditions, perturbed soil with exposed deeper layers (where N content is c.a. 0.02% kg<sup>-1</sup> soil) and typical soil surface of sand soil, e.g. Podzol (where N content is c.a. 0.09% kg<sup>-1</sup> soil), respectively. Furthermore, previous studies indicated that 0–100 mM range of solutions of N-containing salts is suitable for studies on germination requirements [35]. According to literature [14], pH values of the tested solutions are slightly different (Table 2).

## Determination of germination related parameters

To estimate reactions of the studied species to nitrogen type and dose, final germination percentage (FGP), index of germination velocity (IGV, known as modified Timson's index [36]) and index of germination synchrony (IGS, known as Z value [37]) were calculated. Higher values of FGP (where minimum is 0% and maximum is 100%) indicate greater ability to complete germination. Higher values of IGV (where minimum is 0 and maximum is 100) denote more rapid completion of germination. Higher values of IGS (where minimum is 0 and maximum is 1) indicate greater synchronization of germination. The germination-related parameters were calculated using the germinationmetrics package v. 0.1.3 [38] run in R software (v. 3.5.2, 64 bit version [39]) with the functions implemented therein.

#### Analysis of the soil from the habitat of the studied species

The soil analyzed in this study was probed from the same site from which the seeds were collected. Four representative plots  $(1 \times 1 \text{ m})$  were designated. Subsequently, one soil subsample (0-15 cm) soil depth core;  $\emptyset = 2 \text{ cm}$ ) from each plot was collected. All subsamples were pooled into a single bulk sample.

The soil analysis was conducted by Regional Chemical and Agricultural Station in Łódź according to the certified norms and procedures used in Poland. Soil pH $_{\rm KCl}$  was measured using the potentiometric method according to the PN-ISO 10390:1997 norm. Total N was determined by the titrimetric Kjeldahl method (procedure PB 49 ed. 2) and N-NO $_3$  and N-NH $_4$  were determined using the continuous flow analysis (CFA) coupled with spectrophotometric detection (procedure PB 50 ed. 1).

#### Statistical analysis

One-way ANOVA analyses were conducted do detect differences between groups. For further analysis, the conservative Bonferroni post-hoc test was used to detect differences between mean values. This test was selected because it allows to compare large numbers of groups without erroneous results of hypothesis testing. Differences were considered significant at P < 0.05. To estimate influence of the studied factors, two-way ANOVA and three-way

ANOVA were used for analysis among each species and for analysis of all species together, respectively. All analyses were conducted using StatisticaTM v. 13.3 (Tibco Software Inc.).

#### Results

## Germination-related parameters

The species we studied showed generally differential reactions to the examined nitrogen compounds and their concentrations. In the absence of any form of N, we observed a wide range of responses regarding final germination percentage (FGP). The seeds subjected to distilled water showed low FGP, ranging 9–26% (*A. minima, E. vulgare, H. arenarium, H. perforatum* and *S. morisonii*), medium FGP, ranging 49–56% (*C. scabiosa* and *J. montana*) or very high FGP, ranging 80–82% (*C. stoebe* and *H. radicata*) (Fig 1). Interestingly, we were unable to record any completion of germination of *A. montanum* seeds subjected to this treatment (Fig 1). Similar trend was observed for index of germination velocity (IGV) describing germination speed (Fig 2) and index of germination synchrony (IGS) indicating synchronization of germination (Fig 3).

Regarding preferences of N type as a germination stimulator, we were able to distinguish three major groups of plant species. The first group (*H. arenarium* and *H. perforatum*) preferred NH<sub>4</sub><sup>+</sup> over NO<sub>3</sub><sup>-</sup> as a signal for completion of germination (Fig 1). The second group (*A. montanum*, *J. montana* and *S. morisonii*) is composed of species preferring NO<sub>3</sub><sup>-</sup> over NH<sub>4</sub><sup>+</sup> as a stimulant for completion of germination (Fig 1). The third group (*C. scabiosa*, *C. stoebe*, *H. radicata*) is composed of species with no preference and wide tolerance to N type (Fig 1). *E. vulgare* and *A. minima* showed differential response, hard for interpretation.

Two of the studied species showed stenotopic behavior (Fig 1). In the case of A. montanum, a relatively narrow window for stimulation of germination process ranging 1–50 mM of NH<sub>4</sub>NO<sub>3</sub> and 10–50 mM KNO<sub>3</sub> was observed while J. montana was able to complete germination in solutions ranging 0–10 mM N.

Two-way ANOVA showed that N type and dose had significant influence on FGP and IGV (with very few exceptions of *C. stoebe* and *E. vulgare*) (Table 3). All interactions of N type and dose had significant influence on FGP and IGV of all species (Table 3). According to two-way ANOVA, N type significantly influenced IGS only in *C. stoebe*, *H. arenarium*, *H. radicata* and *J. montana*, whereas influence of N dose on IGS was insignificant only in *C. scabiosa* (Table 3). Interactions of N type and dose had no influence on IGS of *C. scabiosa* and *E. vulgare* (Table 3).

According to three-way ANOVA, all analyzed factors (species, N type and N dose) significantly affected the measured germination-related traits (Table 4).

#### Nitrogen status of the soil from the collection site of seeds

Soil analysis (Table 5) showed that acidity of the tested Entic Podzol was typical of this type of soil (pH<sub>KCl</sub> = 5.7) and met the requirements of the studied plant community. Analysis also showed that the soil contained relatively low total N. Mineral forms of N (N-NO<sub>3</sub><sup>-</sup> and N-NH<sub>4</sub><sup>+</sup>) did not contribute substantially to the total N pool (0.99% of total N). Molar N-NH<sub>4</sub><sup>+</sup>: N-NO<sub>3</sub><sup>-</sup> ratio of 1.57 showed that N-NH<sub>4</sub><sup>+</sup> was a predominant N form in the studied soil.

#### **Discussion**

According to our results, it is expected that increasing N deposition in soil causes completion of germination-dependent drift in seedling recruitments. As soil N content rises up to maximal

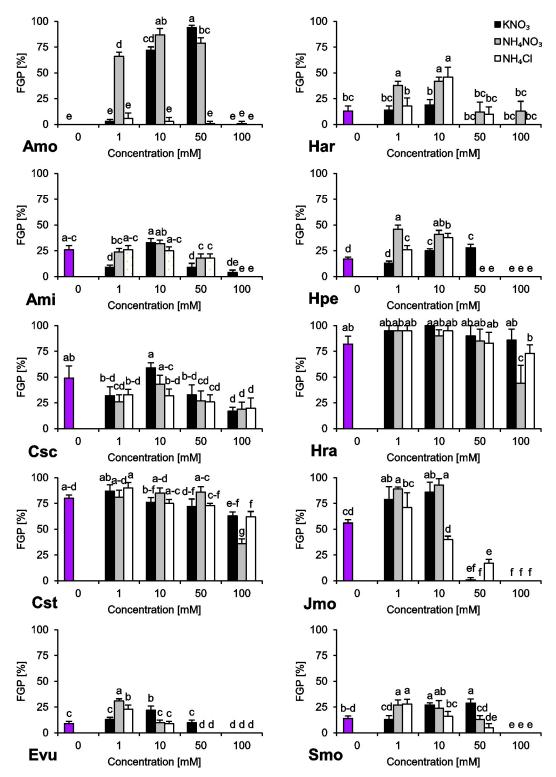


Fig 1. Final Germination Percentage (FGP) of ten studied species subjected to different nitrogen types and doses. The seeds were incubated at 23 °C under a 16/8 h photoperiod for 14 days. Values presented are the mean (SD) (n = 4). Different letters indicate significant differences between groups (ANOVA, Bonferroni's post-hoc test at P < 0.05).

https://doi.org/10.1371/journal.pone.0244737.g001

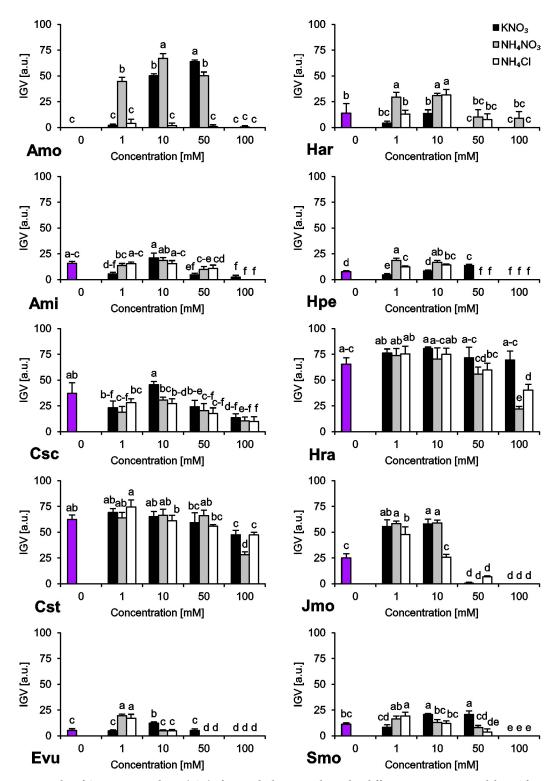


Fig 2. Index of Germination Velocity (IGV) of ten studied species subjected to different nitrogen types and doses. The seeds were incubated at 23°C under a 14 h photoperiod for 16/8 days. Values presented are the mean (SD) (n = 4). Different letters indicate significant differences between groups (ANOVA, Bonferroni's post-hoc test at P < 0.05).

https://doi.org/10.1371/journal.pone.0244737.g002

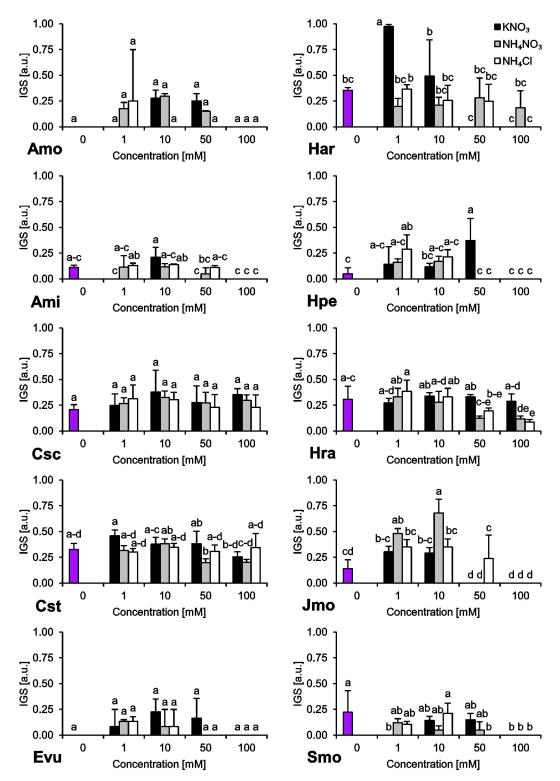


Fig 3. Index of Germination Synchrony (IGS) of ten studied species subjected to different nitrogen types and doses. The seeds were incubated at 23 °C under a 16/8 h photoperiod for 14 days. Values presented are the mean (SD) (n = 4). Different letters indicate significant differences between groups (ANOVA, Bonferroni's post-hoc test at P < 0.05).

https://doi.org/10.1371/journal.pone.0244737.g003

Table 3. Results of two-way ANOVA (F value) showing influence of nitrogen type and dose on Final Germination Percentage (FGP), Index of Germination Velocity (IGV) and Index of Germination Synchrony (IGS) of studied species.

Species	Nitrogen	Nitrogen	NT x ND	
	Type (NT) <sup>a</sup>	Dose (ND) <sup>b</sup>		
	(df=2)	(df=3)	(df=6)	
		FGP		
Amo	1078.7***	716.7***	269.2***	
Ami	9.1***	159.2***	14.2***	
Csc	4.7*	24.3***	3.5**	
Cst	1.6n.s.	89.0***	17.9***	
Evu	10.0***	278.9***	46.8***	
Har	34.3***	67.4***	5.2***	
Нре	24.0***	462.5***	112.0***	
Hra	8.8***	21.5***	4.3**	
Jmo	17.8***	500.1***	24.9***	
Smo	7.1**	88.1***	19.6***	
		IGV		
Amo	911.8***	610.7***	233.1***	
Ami	3.9***	113.0***	9.9***	
Csc	8.8***	46.2***	4.4**	
Cst	3.0n.s.	68.6***	8.4***	
Evu	0.9n.s.	239.6***	59.9***	
Har	55.5***	70.4***	6.5***	
Нре	24.7***	468.0***	164.3***	
Hra	30.4***	54.4***	8.4***	
Imo	36.2***	850.2***	33.7***	
Smo	11.5***	107.8***	28.2***	
		IGS		
Amo	1.7n.s.	3.7*	3.1*	
Ami	3.2n.s.	22.5***	5.2***	
Csc	0.6n.s.	1.0n.s.	0.5n.s.	
Cst	7.8**	6.2**	3.6**	
Evu	1.9n.s.	3.7*	1.3n.s.	
Har	5.6**	21.6***	12.4***	
Нре	2.6n.s.	10.5***	7.5***	
Hra	8.9***	18.8***	5.9***	
Jmo	10.9***	75.8***	9.1***	
Smo	1.1n.s.	18.1***	10.6***	

<sup>&</sup>lt;sup>a</sup>KNO<sub>3</sub>, NH<sub>4</sub>NO<sub>3</sub> or NH<sub>4</sub>Cl

https://doi.org/10.1371/journal.pone.0244737.t003

values observed in inland sand dunes, disappearance of *E. vulgare*, *H. perforatum* and *J. montana* can be predicted. Increase in N can cause alternations in micro-habitats, negatively affecting ability to complete germination of *A. minima*, *A. montanum*, *H. arenarium* and *S. morisonii* seeds. This leads to the conclusion, that germination requirements of these species

<sup>&</sup>lt;sup>b</sup>1, 10, 50 or 100 mM solutions.

 $<sup>^*</sup>P < 0.05$ 

 $<sup>^{**}</sup>P < 0.01$ 

<sup>\*\*\*</sup>P < 0.001, ns-not significant.

Table 4. Results of three-way ANOVA (F value) showing effects of studied factors on germination-related parameters.

Factor	df		F values and significance	
		FGP	IGV	IGS
(S) Species <sup>a</sup>	9	1060.4***	1420.0***	49.7***
(NT) Nitrogen Type <sup>b</sup>	2	88.9***	80.2***	6.0**.
(ND) Nitrogen Dose <sup>c</sup>	3	879.5***	932.6***	89.5***
S x NT	18	51.0***	63.1***	3.6***
S x ND	27	69.6***	69.3***	6.6***
NT x ND	6	59.8***	63.5***	2.2*
S x NT x ND	54	18.9***	18.1***	6.1***

<sup>a</sup>ten studied species (see Table 1 for details)

https://doi.org/10.1371/journal.pone.0244737.t004

are fitted to low N status of poor soils (e.g. Podzols). Similar conclusion was drawn regarding some woodland species from Spain [14] and some species from temperate thermophilous oak forest [40]. Only a few of the species we studied (C. scabiosa, C. stoebe and H. radicata) can efficiently complete germination under high N concentrations, even in 100 mM NH<sub>4</sub>NO<sub>3</sub> solution. As C. stoebe [41] and H. radicata [42, 43] were listed as invasive species, these results partially explain their invasiveness but also show their capability of fast and successful establishment in a wide range of environmental conditions, including inland sand dunes (also those in advanced succession stage). In the case of C. scabiosa, ability to complete germination in a wide range of conditions observed in our study supports the conception of 'stress-tolerance' strategy of this species [44], but probably not inclinations for invasive behavior (due to stable FGP < 50%). It must be however highlighted that data pertaining to multi-species comparisons should be interpreted with caution due to possible dissonance between results of experiments conducted under a given conditions and individual optima of the species. Low germination indices observed in the presented study in several species subjected to control conditions suggest that in vitro conditions (physical and/or chemical factors) do not fully match environmental optimum for completion of germination [35].

The most abundant species (frequently only a few) in a given plant community (e.g. psammophilous grassland on acidic soils) can be defined as 'core species', while the less abundant

Table 5. Nitrogen (N) status of Entic Podzol probed from site of seeds collection.

N-related	Entic Podzol (pH <sub>KCl</sub> = 5.7)  Unit of measurement		
soil characteristic			
	[mg kg <sup>-1</sup> DSW]	[mmol kg <sup>-1</sup> DSW]	
Total N	640 ± 90	$4.571 \pm 0.006$	
N-NO <sub>3</sub>	$4.34 \pm 0.81$	$0.070 \pm 0.013$	
N-NH <sub>4</sub> <sup>+</sup>	$2.01 \pm 0.47$	$0.112 \pm 0.026$	
NH <sub>4</sub> <sup>+</sup> :NO <sub>3</sub> <sup>-</sup> ratio	0.46	1.57	

Values presented are the mean (SD). Molar equivalent of each N-related characteristic was calculated basing on the respective weight value. DSW-dry soil weight.

https://doi.org/10.1371/journal.pone.0244737.t005

bKNO3, NH4NO3 or NH4Cl

c1, 10, 50 or 100 mM solutions.

<sup>\*</sup>P < 0.05

<sup>\*\*</sup>P < 0.01

<sup>\*\*\*</sup>P < 0.001, n.s.–not significant.

ones are 'satellite species' [45]. One might expect that species with wide ecological optimum, tolerant to insufficient level of resources or their excess are 'core species' and the other ones are 'specialists', However, meta-analyses in this field do not support the abovementioned assumption [2]. *J. montana*, *H. arenarium* and particularly *S. morisonii*, the species occurring in substantial numbers in inland sand grasslands [4], showed relatively low completion of germination in the narrow range of conditions, whereas *C. scabiosa*, *C. stoebe* and *H. radicata*, the species persisting on acidic psammophilous grasslands in lesser number [4], showed cosmopolitan reaction in terms of N requirements. This shows that, in extremely poor environments of inland sand grasslands, wide tolerance to environmental N-dependent stimuli at germination stage does not guarantee survival and persistence. As an evolutionary solution to ensure successful establishment in optimal habitats, the seeds of plant species from Asteraceae (e.g. *C. scabiosa*, *C. stoebe*, *H. arenarium*, *H. radicata*) are equipped with pappus allowing distribution by wind (anemochory) [46] and by animals (exozoochory) [47] which is beneficial in the open ecosystem. Such an adaptation allows at least some of the produced seeds to colonize more favorable areas.

Our results suggest that A. montanum does not tolerate NH<sub>4</sub><sup>+</sup> as a sole N source, but this effect can be greatly minimized by an equivalent amount of NO<sub>3</sub>, or even under specific circumstances (10 mM), additional NH<sub>4</sub><sup>+</sup> can be beneficial. The presented study strongly suggests meso-stenotopic behavior of this species regarding both N type and dose. Surprisingly, we did not record completion of germination of this species in the control treatment ( $H_2O$ ). It is in contrast to the study [48] showing high completion of germination of seeds in distilled water. Such diametrical difference may be due to provenance of seeds, altered environmental conditions affecting development of seeds or to genetic differences between populations from various geographic locations, as it was reported for Rumex crispus L. [49]. However, the results similar to ours were presented before [22] regarding the study conducted in 32/20°C thermoperiod in 12/12 h photoperiod. In the same study lower temperature (20/10 and 8/4°C) greatly promoted completion of germination (100 and 56% respectively), which indicates thermoinhibition of germination process in this species. This shows that completion of germination in A. montanum is under thermal control (this species needs at least cooler night temperature or temperatures characteristic of early spring) but adequate N type and dose substitute for those requirements. Very similar abolition of thermoinhibition by NO<sub>3</sub><sup>-</sup> treatment was presented in the studies on Lactuca sativa L. [50]. The authors pointed that the stimulant role of NO<sub>3</sub><sup>-</sup> consisted in induction of NO-dependent signaling transduction pathways and suppression of ABA-dependent regulation of dormancy. This mutual antagonism of NO<sub>3</sub><sup>-</sup>-dependent induction of germination and ABA-dependent suppression was widely described before [51]. We can also see strong ecological advantage of this adaptation, as it allows to avoid failure of establishment, e.g. when seeds are deeply buried in sandy soil (low N status [52]) or sown on bare sand (temperature exceeding thermal requirements for completion of germination; even up to 70°C [2]). Our observation can also partially explain why in central Europe (e.g. Poland) A. montanum can be found on fluvial sands [53], which contain greater amount of nutrients (from alluvial deposits) than aeolic ones [54]. However, A. montanum is often assigned as a species from alkaline xerothermic grasslands, thus in our opinion habitat preference of this species needs further elucidation.

It was repeatedly showed that increasing N status in soil reduced species richness in semiarid grasslands [55]. Most species we studied have low or very low N requirements and completed germination mostly at low levels of N, but some completion of germination was also observed in the solutions with N concentrations exceeding environmental values. As it was showed by numerous evaluations, completion of germination under supraoptimal load of N might lead to exhaustion of seed pool in seed bank due to high mortality at seedling stage under mismatched environmental conditions [56]. Among the species we studied only *H. perforatum* and *J. montana* showed substantial differences pertaining to synchronization of completion of germination (1 mM NH<sub>4</sub>Cl and 1–10 mM of NH<sub>4</sub>NO<sub>3</sub>, respectively). It can be assumed that synchronic completion of germination under such conditions is associated with some kind of adaptation allowing avoidance of the above mentioned seed bank depletion or that it is an efficient mechanism permitting rapid and massive establishment in N-poor vegetation gaps [57]. Completion of germination under very poor conditions suggests that *J. montana* is an oligo-stenotopic plant species. Considering this information, germination pattern of *J. montana* seems to indicate advanced adaptation allowing successful establishment on inland sand dunes. It was shown in field evaluations, where *J. montana* was showed to occur more frequently in low N-deposition sites [58].

Shift from N-poor to relatively N-rich conditions is a natural repercussion of succession in psammophilous grasslands due to deposition of plant biomass [2, 59] which causes gradual development of mineral horizon [9]. Organic forms of N are released from reservoir of plant biomass via microbial metabolism [52] and mineralized throughout succession of acidic grassland [9, 52]. It is worth mentioning that N cycling process in Podzol soils (where mineral forms of N account only 1% of total N [60]) is influenced by many physical and chemical soil traits [2, 52, 61]. It makes hard to predict quantitative equilibrium between  $NO_3^-$  and  $NH_4^+$  forms in soil. However,  $NH_4^+$  is adsorbed mostly on silt and clay fractions, whilst  $NO_3^-$  remains in a soil solution [62] thus is better available for plants (e.g. during seed imbibition) but is also more likely to percolate to ground water with precipitation [2]. In context of presented results it can be stated that the species persisting in acidic dry grasslands need systematic perturbations uncovering poorer soil layers, free of N depositions.

High light level and light heterogeneity are characteristic of loose-grassland-type vegetation [2, 63]. Light conditions in this type of environments are not strongly differentiated, thus soil seems to be the major factor affecting plant functioning in this community [2]. Perceiving soil nutritional feature through the prism of the N status, we concluded that Entic Podzol, on which the studied community was settled, represented upper values of conditions characteristic of dry acidic grasslands [2]. However, they are still relatively low and indicates poor nutritional quality. It can be even further diminished due to aeolian processes [2]. Furthermore, in this type of environments, microbial nitrification is halted at early succession stages due to low pH of soil and symbiotic N fixation becomes significant only at advanced stages of community development due to establishment of *Trifolium* and *Medicago* species [2]. Due to that, complex transformation of inland sand dunes causes gradual increase in N content in soil up to the stage of succession dominated by grasses, which mechanically stabilize soil, allow development of mineral horizon and deposition of nutrients. This can, however, eliminate species adapted to poor soils.

Summarizing, our study clearly showed that both N type and dose affected the ability of the studied species to complete germination (question 1). Furthermore, each species we studied showed different reaction to N type and dose, which suggests individual requirements for N type and dose to complete germination (question 2). Considering biology of the studied species, it can be seen that plants from dry psammophilous grasslands show two different types of adaptation, i.e. some species are adapted to complete germination on N-poor soils (true pioneer species), whilst only a few of them are able to withstand gradual eutrophication of the occupied habitat. It implies that advancing N deposition on dry psammophilous grasslands (both from natural and anthropogenic sources) may lead to loss of potential niches suitable for propagation of pioneer plants and may reduce genetic and species diversity. Last but not least, influence of progressive homogenization of edaphic conditions on the ability of seeds to complete germination (in terms of increasing N availability) is yet another mechanism that is likely

to contribute to secondary succession in dry psammophilous grasslands. Thus, natural as well as anthropogenic processes associated with enhanced N deposition (e.g. due to increased atmospheric N load) shift the environmental balance, favoring establishment of mesophilic species and causing gradual loss of diversity of pioneer plants.

# **Supporting information**

**S1** File. Results of experiment. (XLSX)

# **Acknowledgments**

The authors want to thank M. Fronczak for her linguistic correction of the manuscript.

#### **Author Contributions**

Conceptualization: Mateusz Wala, Jeremi Kołodziejek.

**Data curation:** Mateusz Wala, Jeremi Kołodziejek, Jacek Patykowski. **Formal analysis:** Mateusz Wala, Jeremi Kołodziejek, Jacek Patykowski.

Investigation: Mateusz Wala, Jeremi Kołodziejek, Jacek Patykowski. Methodology: Mateusz Wala, Jeremi Kołodziejek, Jacek Patykowski.

Project administration: Jeremi Kołodziejek.

Resources: Jeremi Kołodziejek, Jacek Patykowski.

Software: Mateusz Wala.

Supervision: Jeremi Kołodziejek, Jacek Patykowski.

Validation: Mateusz Wala, Jeremi Kołodziejek, Jacek Patykowski.

Visualization: Mateusz Wala.

Writing – original draft: Mateusz Wala, Jeremi Kołodziejek, Jacek Patykowski.
Writing – review & editing: Mateusz Wala, Jeremi Kołodziejek, Jacek Patykowski.

#### References

- Bateman MD, Godby SP. Late-Holocene inland dune activity in the UK: a case study from Breckland, East Anglia. Holocene 2004; 14: 579–588. https://doi.org/10.1191/0959683604hl735rp
- Jentsch A, Beyschlag W. Vegetation ecology of dry acidic grasslands in the lowland area of central Europe. Flora 2003; 198: 3–25. https://doi.org/10.1078/0367-2530-00071
- Ödman AM, Schnoor TK, Ripa J, Olsson PA. Soil disturbance as a restoration measure in dry sandy grasslands. Biodivers Conserv. 2012; 21: 1921–1935. https://doi.org/10.1007/s10531-012-0292-4
- 4. Czyżewska M. Syntaksonomia śródlądowych, pionierskich muraw napiaskowych. (Syntaxonomy of inland and pioneer psammophilous grasslands) Monogr. Bot. 1992; 74: 1–174.
- Jentsch A. Disturbance driven vegetation dynamics. Concepts from biogeography to community ecology, and experimental evidence from dry acidic grasslands in central Europe. Diss. Bot. 2004; 384: 1–218.
- Leuschner C, Ellenberg H. Sand dunes and their vegetation series. In: Leuschner C, Ellenberg H, editors. Ecology of Central European Non-Forest Vegetation: Coastal to Alpine, Natural to Man-Made Habitats. Vegetation Ecology of Central Europe, Volume II. Cham: Springer International Publishing Switzerland; 2017. pp. 63–115. https://doi.org/10.1007/978-3-319-43048-5
- Gigon A, Rorison I. The response of some ecologically distinct plant species to nitrate-and to ammonium-nitrogen. J. Ecol. 1972; 60: 93–102. https://doi.org/10.2307/2258043

- Storm C, Herget I, Kappes J, Vormwald B. N\u00e4hrstoff\u00f6kologische Untersuchungen im Darmstadt\u00e4Dieburger Sandgebiet in (teilweise ruderalisierten) Sandpionierfluren und-rasen. Botanik und Naturschutz in Hessen 1998; 10: 41\u00e46.
- Sparrius LB, Kooijman AM. Nitrogen deposition and soil carbon content affect nitrogen mineralization during primary succession in acid inland drift sand vegetation. Plant Soil 2013; 364: 219–228. https:// doi.org/10.1007/s11104-012-1351-z
- Bartelheimer M, Poschlod P. The response of grassland species to nitrate versus ammonium coincides with their pH optima. J. Veget. Sci. 2014; 25: 760–770. https://doi.org/10.1111/jvs.12124
- Fan X, Naz M, Fan X, Xuan W, Miller AJ, Xu G. Plant nitrate transporters: from gene function to application. J. Exp. Bot. 2017; 68: 2463–2475. https://doi.org/10.1093/jxb/erx011 PMID: 28158856
- Chapin FS. The mineral nutrition of wild plants. Annu. Rev. Ecol. Syst. 1980; 11: 233–260. https://doi. org/10.1146/annurev.es.11.110180.001313
- **13.** Baskin CC, Baskin JM. Seeds: Ecology, biogeography and evolution of dormancy and germination. Lexington: Academic Press; 1998.
- Pérez-Fernández MA, Rodríguez-Echeverría S. Effect of smoke, charred wood, and nitrogenous compounds on seed germination of ten species from woodland in centralwestern Spain. J. Chem. Ecol. 2003; 29: 237–251. https://doi.org/10.1023/a:1021997118146 PMID: 12647865
- Duermeyer L, Khodapanahi E, Yan D. Regulation of seed dormancy and germination by nitrate. Seed Sci. Res. 2018; 28: 50–157. https://doi.org/10.1017/s096025851800020x
- Penfield S. Seed dormancy and germination. Curr. Biol. 2017; 27: 874–878. <a href="https://doi.org/10.1016/j.cub.2017.05.050">https://doi.org/10.1016/j.cub.2017.05.050</a> PMID: 28898656
- Liu Y, Sun J, Tian Z, Hakeem A, Wang F, Jiang D, et al. Physiological responses of wheat (*Triticum aestivum* L.) germination to elevated ammonium concentrations: reserve mobilization, sugar utilization, and antioxidant metabolism. Plant Growth Regul. 2017; 81: 209–220. <a href="https://doi.org/10.1007/s10725-016-0198-3">https://doi.org/10.1007/s10725-016-0198-3</a>
- **18.** Britto DT, Kronzucker HJ. NH4+ toxicity in higher plants: a critical review. J. Plant Physiol. 2002; 159: 567–584. https://doi.org/10.1078/0176-1617-0774
- Barker AV, Bryson GM. Nitrogen. In: Barker AV, Pilbeam DJ, editors. Handbook of plant nutrition. Boca Raton: CRC Press; 2007. pp. 21–50.
- Imas P, Bar-Yosef B, Kafkafi U, Ganmore-Neumann R. Release of carboxylic anions and protons by tomato roots in response to ammonium nitrate ration and pH in nutrient solution. Plant Soil. 1997; 191: 24–34. https://doi.org/10.1023/A:1004214814504
- Savvas D. Passam HC, Olympios C. Effect of ammonium nitrogen on lettuce grown on pumice in a closed hydroponic system. Hort. Sci. 2006; 41: 1667–1673. <a href="https://doi.org/10.21273/HORTSCI.41.7.1667">https://doi.org/10.21273/HORTSCI.41.7.1667</a>
- 22. Partzsch M, Piesch C, Hensen IA. Comparative study of germination biology and plant performance in two dry grassland species. Folia Geobot. 2011; 46: 35–48. https://doi.org/10.1007/s12224-010-9084-3
- 23. Abedi M. Seed ecology in dry sandy grasslands—an approach to patterns and mechanisms. Ph.D. Thesis, University of Regensburg. 2013.
- 24. The Plant List Version 1.1. [cited 26th August 2019]. Database: Plant taxonomy base [Internet]. Available from: http://www.theplantlist.org/
- **25.** Ellenberg H. Indicator values of vascular plants in central Europe 1. Indicator values of vascular plants not including Rubus. Scripta Geobot. 1991; 18: 9–166.
- Kołodziejek J, Glińska S, Michlewska S. Seasonal leaf dimorphism in Potentilla argentea L. var. tenuiloba (Jord.) Sw. (Rosaceae). Acta Bot. Croat. 2015; 74: 53–70. https://doi.org/10.1515/botcro-2015-0012
- Abedi M, Bartelheimer M, Poschlod P. Aluminium toxic effects on seedling root survival affect plant composition along soil reaction gradients—a case study in dry sandy grasslands. J. Veget. Sci. 2013; 24: 1074–1085. https://doi.org/10.1111/jvs.12016
- Bojňnanský V, Fargašová A. Atlas of seeds and fruits of central and east-European flora: the Carpathian Mountains region. Dordrecht: Springer; 2007. https://doi.org/10.1007/978-1-4020-5362-7
- 29. Deno NC. Seed germination: theory and practice. Second Edition. State College: Pennsylvania State University; 1993.
- Çirak C, Ayan AK, Kevseroğlu K. The effects of light and some presoaking treatments on germination rate of St. John's worth (*Hypericum perforatum* L.) seeds. Pak. J. Biol. Sci. 2004; 7: 182–186. https:// doi.org/10.3923/pjbs.2004.182.186
- 31. Koutsovoulou K, Daws MI, Thanos CA. Campanulaceae: A family with small seeds that require light for germination. Ann. Bot. 2014; 113: 135–143. https://doi.org/10.1093/aob/mct250 PMID: 24232382

- Dürr C, Dickie JB, Yang X. Ranges of critical temperature and water potential values for the germination of species worldwide: Contribution to a seed trait database. Agric. For. Meteorol. 2015; 200: 222–232. https://doi.org/10.1016/j.agrformet.2014.09.024
- 33. Wierzbicka M, Obodzińska J. The effect of lead on seed imbibition and germination in different plant species. Plant Sci. 1998; 137: 155–171. https://doi.org/10.1016/S0168-9452(98)00138-1
- Mojzes A, Kalapos T. Plant-derived smoke stimulates germination of four herbaceous species common in temperate regions of Europe. Plant Ecol. 2014; 215: 411–415. https://doi.org/10.1007/s11258-014-0311-5
- Pérez-Fernández MA, Rodríguez-Echeverría S, Calvo-Magro E, David-Antonio C. Germination of selected perennial plant species from western Spain under nitrogen, light and wet and dry heat treatments. J. Mediterranean Ecol. 2003; 4: 23–33.
- Khan MA, Ungar IA. Effects of thermoperiod on recovery of seed germination of halophytes from saline conditions. Am. J. Bot. 1997; 84: 279–283. https://doi.org/10.2307/2446089 PMID: 21712207
- Ranal MA, Santana DGDE. How and why to measure the germination process? Revista Brasil Bot. 2006; 29: 1–11. https://doi.org/10.1590/S0100-84042006000100002
- **38.** Aravind J, Vimala Devi S, Radhamani J, Jacob SR, Kalyani Srinivasan. Germinationmetrics: seed germination indices and curve fitting. R package version 0.1.3. [cited 11 March 2019]. Available from: https://github.com/aravind-j/germinationmetricshttps://cran.r-project.org/package=germinationmetrics.
- 39. R Core Team R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. [cited 11 March 2019]. Available from: <a href="https://www.R-project.org/">https://www.R-project.org/</a>.
- Kołodziejek J, Patykowski J, Wala M. Effect of light, gibberellic acid and nitrogen source on germination of eight taxa from dissapearing European temperate forest, *Potentillo albae-Quercetum*. Sci. Rep. 2017; 7: 13924. https://doi.org/10.1038/s41598-017-13101-z PMID: 29066749
- Henery M, Bowman G, Mráz P, Treier UA, Gex-Fabry E, Schaffner U, et al. Evidence for a combination of pre-adapted traits and rapid adaptive change in the invasive plant *Centaurea stoebe*. J. Ecol. 2010; 98: 800–813. https://doi.org/10.1111/j.1365-2745.2010.01672.x
- Ortiz MÁ, Tremetsberger K, Terrab A, Stuessy TF, García-Castaňo L, Urtubey E, et al. Phylogeography
  of the invasive weed *Hypochaeris radicata* (Asteraceae): From Moroccan origin to worldwide introduced
  populations. Mol. Ecol. 2008; 17: 3654–3667. https://doi.org/10.1111/j.1365-294X.2008.03835.x
   PMID: 18662226
- Schoenfelder AC, Bishop JG, Martinson HM, Fagan WF. Resource use efficiency and community effects of invasive *Hypochaeris radicata* (Asteraceae) during primary succession. Am. J. Bot. 2010; 97: 1772–1779. https://doi.org/10.3732/ajb.0900383 PMID: 21616816
- 44. Moora M, Öpik M, Zobel M. Performance of two *Centaurea* species in response to different root-associated microbial communities and to alterations in nutrient availability. Ann. Bot. Fenn. 2004; 41: 263–271. https://doi.org/10.3732/ajb.0900383 PMID: 21616816
- Ozinga WA. Assembly of plant communities in fragmented landscapes: the role of dispersal. Ph.D. Thesis, Wageningen University, 2008.
- 46. Sheldon JC, Burrows FM. The dispersal effectiveness of the achene–pappus units of selected Compositae in steady winds with convection. New Phytol. 1973; 72: 665–675. https://doi.org/10.1111/j.1469-8137.1973.tb04415.x
- Miguel MF, Lortie CJ, Callaway RM, Hierro JL. Competition does not come at the expense of colonization in seed morphs with increased size and dispersal. Am. J. Bot. 2017; 104: 1323–1333. https://doi.org/10.3732/ajb.1700266 PMID: 29885233
- **48.** Ouzounidou G. Effect of copper on germination and seedling growth of *Minuartia, Silene, Alyssum* and *Thlaspi.* Biol. Plant. 1995; 37: 411–416. https://doi.org/10.1007/BF02913990
- Pérez-Fernández M, Elliott CP, Valentine A, Oyola JA. Seed provenance determines germination responses of *Rumex crispus* (L.) under water stress and nutrient availability. J. Plant. Ecol. 2019; 12: 949–969. https://doi.org/10.1093/jpe/rtz034
- Dong T, Tong J, Xiao L. Nitrate, abscisic acid and gibberellin interactions on the thermoinhibition of lettuce seed germination. Plant Growth Regul. 2012; 66: 191–202. https://doi.org/10.1007/s10725-011-9643-5
- Nonogaki H. Seed biology updates
   –highlights and new discoveries in seed dormancy and germination.
   Front. Plant Sci. 2017; 8: 524. https://doi.org/10.3389/fpls.2017.00524 PMID: 28443117
- Brankatschk R, Fischer T, Veste M. Succession of N cycling processes in biological soil crusts on a Central European inland dune. FEMS Microbiol. Ecol. 2013; 83: 149–160. https://doi.org/10.1111/j. 1574-6941.2012.01459.x PMID: 22816620
- Kucharczyk M. Phytogeographical Roles of Lowland Rivers on the Example of the Middle Vistula. Lublin, Poland. Lublin: Maria Sklodowska-Curie University Press, 2003.

- 54. Belnap J, Munson SM, Field JP. Aeolian and fluvial processes in dryland regions: The need for integrated studies. Ecohydrology 2011; 4: 615–622. https://doi.org/10.1002/eco.258
- Zhong M, Miao Y, Han S, Wang D. Nitrogen addition decreases seed germination in a temperate steppe. Ecol. Evol. 2019; 9: 8441–8449. https://doi.org/10.1002/ece3.5151 PMID: 31410252
- 56. Bird EJ, Choi YD. Response of native plants to elevated soil nitrogen in the sand dunes of Lake Michigan, USA. Biol. Conserv. 2017; 212: 398–405. https://doi.org/10.1016/j.biocon.2016.12.001
- Jiménez-Alfaro B, Silveira FAO, Fidelis A, Poschlod P, Commander LE. Seed germination traits can contribute better to plant community ecology. J. Veg. Sci. 2016; 27: 637–645. https://doi.org/10.1111/jvs.12375
- Sparrius LB, Sevink J, Kooijman AM. Effects of nitrogen deposition on soil and vegetation in primary succession stages in inland drift sands. Plant Soil 2012; 353: 261–272. https://doi.org/10.1007/s11104-011-1029-y
- 59. Jones DL, Shannon D, Murphy DV, Farrar J. Role of dissolved organic nitrogen (DON) in soil N cycling in grassland soils. Soil Biol. Biochem. 2004; 36: 749–756. https://doi.org/10.1016/j.soilbio.2004.01.003
- 60. Mocek A. Gleboznawstwo (Soil Science). Warszawa: Wydawnictwo Naukowe PWN, 2015.
- Rennenberg H, Dannenmann M, Gessler A, Kreuzwieser J, Simon J, Papen H. Nitrogen balance in forest soils: Nutritional limitation of plants under climate change stresses. Plant Biol. 2009; 11: 4–23. https://doi.org/10.1111/j.1438-8677.2009.00241.x PMID: 19778364
- Rossi N, Ciavatta C, Vittori Antisari L. Seasonal pattern of nitrate losses from cultivated soil with subsurface drainage. Water Air Soil Pollut. 1991; 60: 1–10. https://doi.org/10.1007/BF00293961
- 63. Bakker C, Blair JM, Knapp AK. Does resource availability, resource heterogeneity or species turnover mediate changes in plant species richness in grazed grasslands? Oecologia 2003; 137: 385–391. https://doi.org/10.1007/s00442-003-1360-y PMID: 12955488