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| Title: | Defining the cytosolic pathway of glutathione degradation in <i>Arabidopsis thaliana</i> : Role of the ChaC/GCG family of γ -glutamyl cyclotransferases as glutathione-degrading enzymes and AtLAP1 as the Cys-Gly peptidase |
| Authors: | Kumar, Shailesh (/jspui/browse?type=author&value=Kumar%2C+Shailesh) Kaur, Amandeep (/jspui/browse?type=author&value=Kaur%2C+Amandeep) Chattopadhyay, B. (/jspui/browse?type=author&value=Chattopadhyay%2C+B.) Bachhawat, A.K. (/jspui/browse?type=author&value=Bachhawat%2C+A.K.) |
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| Abstract: | <p>Glutathione homeostasis is critical to plant life and its adaptation to stress. The γ-glutamyl cycle of glutathione biosynthesis and degradation plays a pre-eminent role in glutathione homeostasis. The genes encoding two enzymatic steps of glutathione degradation, the γ-glutamyl cyclotransferase (GGCT; acting on γ-glutamyl amino acids) and the Cys-Gly dipeptidase, have, however, lacked identification. We have investigated the family of GGCTs in <i>Arabidopsis thaliana</i>. We show through in vivo functional assays in yeast that all three members of the ChaC/GCG subfamily show significant activity towards glutathione but no detectable activity towards γ-glutamyl methionine. Biochemical characterization of the purified recombinant enzymes GGCT2;2 and GGCT2;3 further confirmed that they act specifically to degrade glutathione to yield 5-oxoproline and Cys-Gly peptide and show no significant activity towards γ-glutamyl cysteine. The K_m for glutathione was 1.7 and 4.96 mM for GGCT2;2 and GGCT2;3 respectively and was physiologically relevant. Evaluation of representative members of other subfamilies indicates the absence of GGCTs from plants showing significant activity towards γ-glutamyl-amino acids as envisaged in the classical γ-glutamyl cycle. To identify the Cys-Gly peptidase, we evaluated leucine aminopeptidases (LAPs) as candidate enzymes. The cytosolic AtLAP1 (<i>A. thaliana</i> leucine aminopeptidase 1) and the putative chloroplastic AtLAP3 displayed activity towards Cys-Gly peptide through in vivo functional assays in yeast. Biochemical characterization of the in vitro purified hexameric AtLAP1 enzyme revealed a K_m for Cys-Gly of 1.3 mM that was physiologically relevant and indicated that AtLAP1 represents a cytosolic Cys-Gly peptidase activity of <i>A. thaliana</i>. The studies provide new insights into the functioning of the γ-glutamyl cycle in plants. © The Authors Journal compilation</p> |
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