

Class 10: Structural Bioinformatics

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What is in the PDB Database?

The main repository of biomolecular structure info is the PDB < www.rcsb.org >.

Lets see what this database contains:

```
stats <- read.csv("pdb_stats.csv", row.names=1)
stats
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	161,663	12,592	12,337	200	74	32
Protein/Oligosaccharide	9,348	2,167	34	8	2	0
Protein/NA	8,404	3,924	286	7	0	0
Nucleic acid (only)	2,758	125	1,477	14	3	1
Other	164	9	33	0	0	0
Oligosaccharide (only)	11	0	6	1	0	4
Total						
Protein (only)	186,898					
Protein/Oligosaccharide	11,559					
Protein/NA	12,621					
Nucleic acid (only)	4,378					
Other	206					
Oligosaccharide (only)	22					

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

We have to get rid of the commas. Which function can do so?

```
x <- stats$X.ray
sum(as.numeric(gsub(",", "", x)))
```

```
[1] 182348
```

I can turn this into a function and then use `apply()` to work on the entire table of data.

```
sumcomma <- function(x) {  
  sum( as.numeric(gsub(",", "", x)) )  
}  
  
sumcomma(stats$X.ray)
```

```
[1] 182348
```

```
n.total <- sumcomma(stats$Total)  
n.total
```

```
[1] 215684
```

```
sumcomma(stats$EM)
```

```
[1] 18817
```

```
apply(stats, 2, sumcomma)
```

X.ray	EM	NMR	Multiple.methods
182348	18817	14173	230
Neutron	Other	Total	
79	37	215684	

```
apply(stats, 2, sumcomma) / sumcomma(stats$Total)
```

X.ray	EM	NMR	Multiple.methods
0.8454405519	0.0872433746	0.0657118748	0.0010663749
Neutron	Other	Total	
0.0003662766	0.0001715473	1.0000000000	

Q2: What proportion of structures in the PDB are protein?

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

```
186898/248805733 * 100
```

```
[1] 0.07511804
```

Visualizing the HIV-1 Protease Structure

Mol* (“Mol-Star”) viewer is now everywhere.

I want to insert my image from Mol* here.

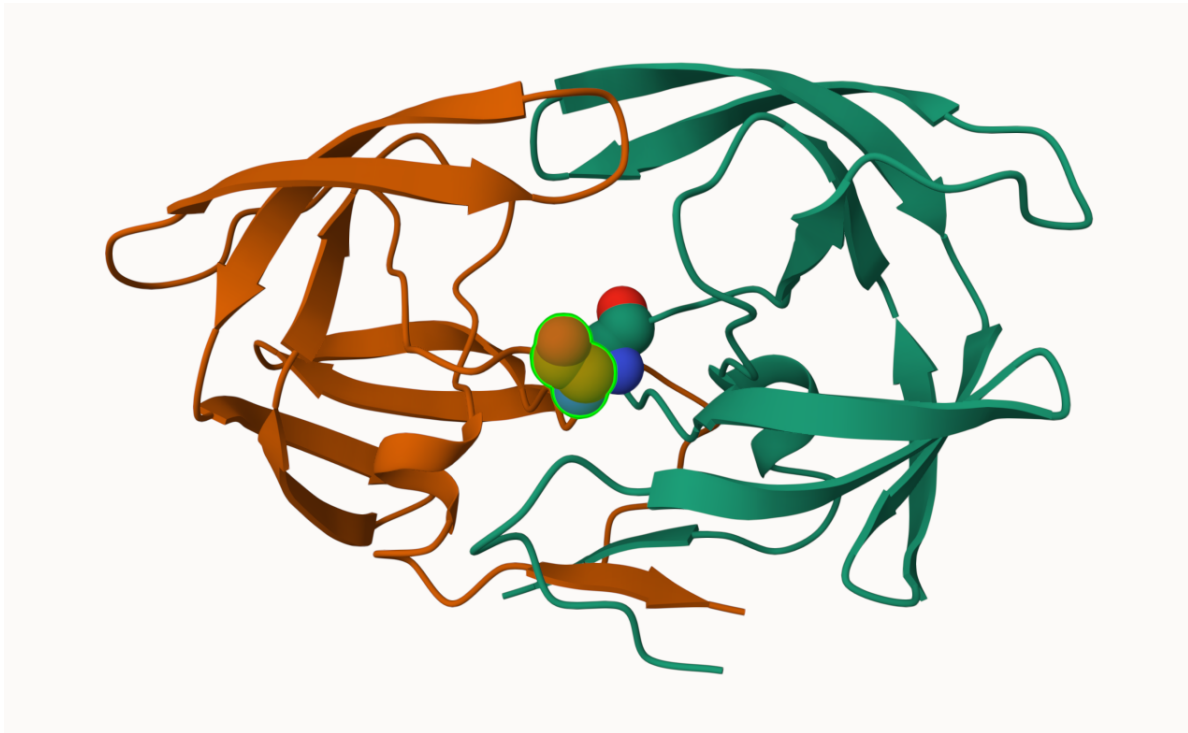


Figure 1: My first molecular image

Working with the bio3d package

```
library(bio3d)
```

```
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

pdb

```
Call: read.pdb(file = "1hsg")
```

Total Models#: 1

Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)

Non-protein/nucleic resid values: [HOH (127), MK1 (1)]

Protein sequence:

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

head(pdb\$atom)

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40
	segid elesy charge												
1	<NA>		N	<NA>									
2	<NA>		C	<NA>									
3	<NA>		C	<NA>									
4	<NA>		O	<NA>									
5	<NA>		C	<NA>									
6	<NA>		C	<NA>									

```
pdbseq(pdb)
```

```
 1  2  3  4  5  6  7  8  9 10 11 12 13 14 15 16 17 18 19 20
"P" "Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G" "G" "Q" "L" "K"
21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40
"E" "A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M" "S" "L" "P" "G"
41 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60
"R" "W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V" "R" "Q" "Y" "D"
61 62 63 64 65 66 67 68 69 70 71 72 73 74 75 76 77 78 79 80
"Q" "I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L" "V" "G" "P" "T"
81 82 83 84 85 86 87 88 89 90 91 92 93 94 95 96 97 98 99  1
"P" "V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T" "L" "N" "F" "P"
 2  3  4  5  6  7  8  9 10 11 12 13 14 15 16 17 18 19 20 21
"Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G" "G" "Q" "L" "K" "E"
22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41
"A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M" "S" "L" "P" "G" "R"
42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60 61
"W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V" "R" "Q" "Y" "D" "Q"
62 63 64 65 66 67 68 69 70 71 72 73 74 75 76 77 78 79 80 81
"I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L" "V" "G" "P" "T" "P"
82 83 84 85 86 87 88 89 90 91 92 93 94 95 96 97 98 99
"V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T" "L" "N" "F"
```

Predicting functional motions of a single structure

We can do bioinformatics predictions of functional motions (ie. flexibility/dynamics).

```
pdb <- read.pdb("6s36")
```

Note: Accessing on-line PDB file

PDB has ALT records, taking A only, rm.alt=TRUE

```
pdb
```

```
Call: read.pdb(file = "6s36")
```

```
Total Models#: 1
```

Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)

Non-protein/nucleic resid values: [CL (3), HOH (238), MG (2), NA (1)]

Protein sequence:

MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIMDAGKLV
DELVIALVKERIAQEDCRNGFLDGFRTIPQADAMKEAGINVDYVLEFDVPDELIVDKI
VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQM
TAPLIG
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG

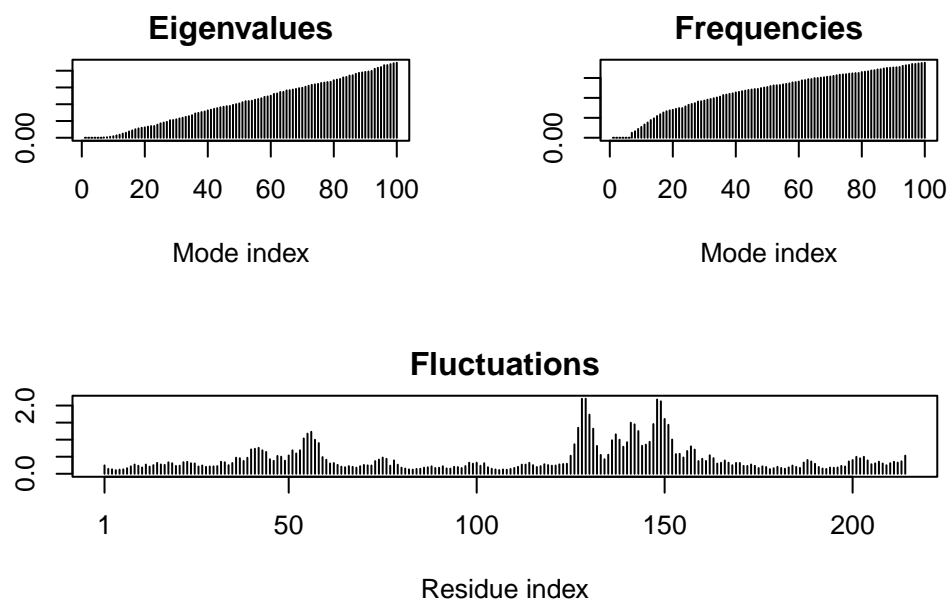
+ attr: atom, xyz, seqres, helix, sheet,
calpha, remark, call

```
m <- nma(pdb)
```

Building Hessian... Done in 0.03 seconds.

Diagonalizing Hessian... Done in 0.35 seconds.

```
plot(m)
```



```
mktrj(m, file="adk_m7.pdb")
```