

SAnDReS User Guide

Statistical Analysis of Docking Results and Scoring functions

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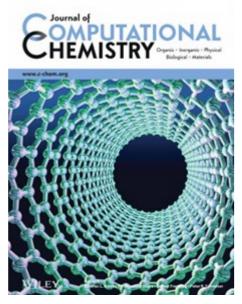
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1. International Research Team

SAnDReS 2.0 has the contribution of researchers from six countries and ten different affiliations (photos in alphabetical order). Click on the photo to access Scopus author profile.



2. How to Cite SAnDReS 2.0



de Azevedo, W. F., Jr; Quiroga, R.; Villarreal, M. A.; da Silveira, N. J. F.; Bitencourt-Ferreira, G.; da Silva, A. D.; Veit-Acosta, M.; Oliveira, P. R.; Tutone, M.; Biziukova, N.; Poroikov, V.; Tarasova, O.; Baud, S. SAnDReS 2.0: Development of Machine-Learning Models to Explore the Scoring Function Space. *J. Comput. Chem.* **2024**, *45* (27), 2333–2346. https://doi.org/10.1002/jcc.27449.

3. Conventions and Availability

This User Guide shows how to install and use the attributes of the SAnDReS program (Version 2.0.0). This guide includes the capabilities of the program, how to apply these capabilities, and how to install SAnDReS on Linux.

Here, we have the following typographical conventions:

Arial font with italic

Indicates filenames and folders (directories) in the main text.

Courier New font with Italic

Used for Linux commands, PDB (Protein Data Bank) listings, command lines, and commands to be typed by the user.

SAnDReS is open-source software and freely distributed under GNU General PublicLicense v3.0 (GPL-3.0 License). Its code is available to download on GitHub (https://github.com/azevedolab/sandres).

The following sections describe the installation guidelines and tutorials.

4. Introduction

SAnDReS (Statistical Analysis of Docking Results and Scoring Functions) (de Azevedo et al., 2024) draws inspiration from several protein systems we studied in the last decades. These projects began in the 1990s with pioneering studies focused on intermolecular interactions between cyclin-dependent kinase (CDK) (EC 2.7.11.22) and inhibitors (de Azevedo et al., 1996; de Azevedo et al., 1997). The continuing studies of protein-ligand interactions made it clear the necessity of computational tools to address these complex systems. SAnDReS is our approach to build models which expand our understanding of intermolecular interactions.

SAnDReS is a free and open-source (GPL-3.0 License) computational environment for the development of machine-learning models (Bitencourt-Ferreira & de Azevedo, 2019; Bitencourt-Ferreira et al., 2021; Bitencourt-Ferreira, Rizzotto et al., 2021) for the prediction of ligand-binding affinity (Xavier et al., 2016; Bitencourt-Ferreira & de Azevedo, 2019; Veit-Acosta & de Azevedo, 2021). We developed SAnDReS using Python 3 programming language and Pandas, SciPy, NumPy, Scikit-Learn (Pedregosa et al., 2011), and Matplotlib libraries as a computational tool to explore the Scoring Function Space concept (Ross et al., 2013; Heck et al., 2017; Bitencourt-Ferreira & de Azevedo, 2019).

SAnDReS 2.0.0 (de Azevedo et al., 2024) brings the most advanced tools for protein-ligand docking simulation and machine-learning modeling. We have the newest version of AutoDock Vina (Trott & Olson, 2010; Eberhardt et al., 2021), available in October 2024 (version 1.2.3), as a docking engine. Also, SAnDReS 2.0.0 uses Scikit-Learn (Pedregosa et al., 2011) to generate machine learning models taking terms in the AutoDock Vina scoring function and descriptors. SAnDReS-generated models predict binding affinity for a specific protein system with superior performance compared to classical scoring functions and other machine-learning scoring functions (K_{DEEP} (Jiménez et al., 2018), CSM-lig (Pires & Ascher, 2016), and $\Delta VinaRF_{20}$ (Wang & Zhang, 2017)). In summary, SAnDReS can design a scoring function adequate to the protein system of your interest. SAnDReS focuses on developing a machine-learning model targeted to one protein system but it can also build universal scoring functions.

You need Python 3 installed on your computer to run SAnDReS 2.0.0. In addition, you need Pandas, Matplotlib, NumPy, Scikit-Learn, and SciPy. It is also necessary to have AutoDockFR-AutoDock for Flexible Receptors (ADFRsuite) (Ravindranath et al., 2015). You can make the installation faster by installing Anaconda.

4.1. Funding

The Brazilian National Council for Scientific and Technological Development (CNPq) (Process 306298/2022-8) supports this research project. This study was financed in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brazil (CAPES) – Finance Code 001. MVA acknowledges Diether Haenicke Scholarship from Western Michigan University. OT, NB, and VP thank the Program for Basic Research in the Russian Federation for a long-term period 2021–2030 (project No. 122030100170-5). R.Q and M.A.V thank Secyt-UNC for their financial support.

5. Installing SAnDReS (Linux)

You should type all commands shown here in a Linux terminal. The easiest way to open a Linux terminal is to use the Ctrl+Alt+T key combination.

Step 1. Download <u>Anaconda Installer for Linux</u> or newer. Go to the directory where you have the installer file and type the following commands:

```
chmod u+x Anaconda3-2021.11-Linux-x86_64.sh ./Anaconda3-2021.11-Linux-x86 64.sh
```

Follow the instructions of the installer. You may use a newer installer, but be sure to have the right installer in the above command lines.

Step 2. Download ADFRsuite version 1.0 (<u>ADFRsuite 1.0 Linux 64 installer app</u>). Type the following commands:

```
cd ~
cp Downloads/ADFRsuite_Linux-x86_64_1.0_install .
chmod a+x ADFRsuite_Linux-x86_64_1.0_install
./ADFRsuite Linux-x86_64_1.0_install
```

Follow the instructions of the installer. You need to add the path of ADFRsuite to your .bashrc (e.g., PATH="/home/walter/ADFRsuite-1.0/bin:\$PATH").

Step 3. To run SAnDReS properly, you need <u>Scikit-Learn</u> 1.5.2. To be sure you have version 1.5.2, open a terminal, and type the following commands:

```
python3 -m pip uninstall scikit-learn
python3 -m pip install scikit-learn==1.5.2
```

Step 4. Download SAnDReS (https://github.com/azevedolab/sandres/raw/master/sandres2.zip). Copy the sandres2 zipped directory (sandres2.zip) to wherever you want it and unzip the zipped directory.

Type the following command:

```
unzip sandres2.zip
```

Then, change to *sandres2* directory and type:

```
python3 sandres2.py
```

Now you have the GUI window for SAnDReS.

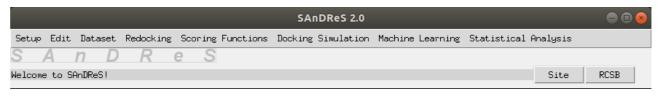


Figure 1. SAnDReS Main Menu.

Have a good SAnDReS session!

5.1. Colophon

We performed machine-learning modeling and protein–ligand docking simulations reported on this user guide using a Notebook PC with 8GB of memory, a 500 GB SSD, a GPU GeForce® GTX 1650, and an Intel® Core® i5-10300H @ 2.50 GHz processor running Linux Ubuntu 18.04.

5.2. AutoDock Vina 1.2

You may have slightly different docking results using AutoDock Vina 1.2 (Eberhardt et al., 2021) integrated to SAnDReS 2.0 due to difference in hardware and Linux flavor. Even when you use the same set of input files.

6. Tutorial 01: CDK2 Docked Structures with K_i Data

Here, we will take docked structures of CDK2 complexed with known inhibitors for which K_i data is available. We will employ CDK2-pose complexes to develop machine-learning models to predict pK_i. We will use binding affinity data and ligand structures available in the BindingDB (Gilson et al., 2016). We consider you have successfully installed this program as described. To run SAnDReS, go to the *sandres2* directory. You will find the results of this tutorial in <u>de Azevedo et al., 2024</u>. Type the following commands:

```
cd sandres2
python3 sandres2.py
```

It is necessary to use the directory you have SAnDReS on your computer.

Note: After SAnDReS installation, we have two files (*mlr.py* and *sandres2.py*) and the following folders in the *sandres2* directory: *datasets*, *misc*, *MLRegMPy*, and *tools*. We find auxiliary files necessary to run SAnDReS in the *misc* folder. We keep the project directories in the *datasets* folder, one project directory for each protein system.

6.1. Setup

The idea of this tutorial is to use the experimental information and build machine-learning models. We focus on inhibition constant (K_i) data. We will employ docked poses to build our machine learning models. We will assess the machine-learning models using DOME metrics (Walsh et al., 2021). Here, we will define the project directory and copy the files to run this tutorial (Tutorial 01). SAnDReS has predefined ligand data with experimental binding affinities (inhibition constant (K_i), dissociation constant (K_d), and IC₅₀) and generated PDBQT files for ligand structures. We used filtered affinity data in the PDBbind version 2020 (Liu et al., 2022) to generate files with ligand information (bind_IC50.csv, bind_Kd.csv, and bind_Ki.csv). We keep the dataset the ligand structures for which we defined binding affinity not ranges (e.g., 100 <K_i< 1000 nM). The project directory is where SAnDReS keeps all files generated during its execution. We expect one for each protein system. Click Setup->Check Ligand Data.

SAnDReS will show the number of ligands available.

It is possible to add ligand data to the files <code>bind_Kd.csv</code>, <code>bind_Ki.csv</code>, and <code>bind_IC50.csv</code> found in the <code>sandres2/misc/data</code> directory. Any data included to CSV files should be followed to corresponding structural data (PDBQT files for ligands) added to <code>sandres2/misc/data/pdbqt</code> directory. If you add a ligand data to <code>bind_Ki.csv</code>, you should add the ligand structure (PDBQT format) to <code>sandres2/misc/data/pdbqt/Ki</code> directory. For example, let us suppose that you added the structure <code>XXXX</code> to the <code>bind_Ki.csv</code> file. You must add a folder named <code>XXXX</code> to the <code>sandres2/misc/data/pdbqt/Ki</code> directory with the <code>lig.pdbqt</code> file.

Next, we will enter the project directory.

We will set the project directory and copy the files to run this tutorial.

Click the following sequence in the main menu: Setup->Project Directory->Enter.

SAnDReS will open the ./misc/inputs/strdir.in file with the Fast Editor, and you should insert the directory where we will have all data related to this modeling (e.g., ./datasets/CDK2_Ki/).

After writing the project directory in the Fast Editor, click the Save and Close buttons.

Then, you click Setup->Project Directory->Make. Click the Yes option.

SAnDReS will generate a new directory named *CDK2_Ki*. You should get the following message: Successfully created the directory ./datasets/CDK2_Ki/

To create a summary of this project, click *Setup->Project Directory->Summary*.

In the Fast Editor, you add an explanation, for instance, CDK2 with Ki data.

After adding this sentence, click the Save and Close buttons.

To copy all necessary files to run this tutorial to your project directory, click *Setup->Copy Files to Run Tutorials->Tutorial 01*.

SAnDReS will download all necessary files (*cdk2_ki.mol2*, *cdk2_ki.sdf*, and *cdk2_ki.tsv*) to run this tutorial from https://github.com/azevedolab/sandres. You should get the following message: SAnDReS finished the "Copy Files to Run Tutorial 01" request!

We need to clean the downloaded files from the BindingDB to eliminate binding affinity data with undefined values for the affinity (e.g., >1000 or < 3.5).

Click Setup->BindingDB Data. Click the Yes option. Enter the parameters shown below.

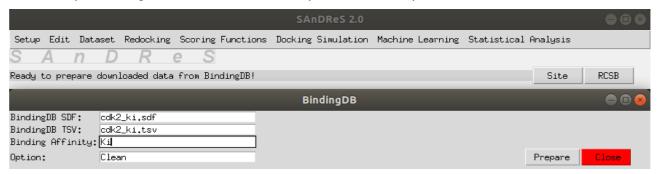


Figure 2. BindingDB Menu.

Click the *Prepare* button.

After finishing, you get the following message:

Done! SAnDReS prepared BindingDB data for machine learning modeling!

SAnDReS created the following files necessary for the modeling: affinity_BindingDB_Ki.csv and cdk2_ki_out.csv.

Click the Close button.

Now, we finished the Setup.

6.2. Dataset

Here, we will download a crystal structure from the Protein Data Bank (PDB) and generate PDBQT files for this structure (receptor.pdbqt and lig.pdbqt). We focus on carrying out docking simulations for CDK2. Our dataset has only one structure (2DS1) (Kawanishi et al., 2006). The structure 2DS1 is in the K_i data folder.

Click on Dataset->Enter PDB Access Codes.

SAnDReS will open the *pdb_codes.csv* file with the Fast Editor. Enter the PDB access code. 2DS1

Click the Save and the Close buttons.

SAnDReS saved the PDB access code in a file named *pdb_codes.csv*. If you reopen the *pdb_codes.csv* file, SAnDReS should show that you have one structure.

We will not need binding affinity data for the 2DS1 structure, but it is necessary to follow this step. SAnDReS will read the *bind_Ki.csv* file to get data (ligand number and chain) about the active ligand in the 2DS1 structure.

Click Dataset->Add->Binding Affinity Data. Click the Inhibition Constant (Ki) button. Then, click the Start button.

Once finished, you will get the following message:

SAnDReS finished the "Add Binding Affinity Data" request!

SAnDReS generated a file named *bind_Ki.csv*. Click the *Done* and the *Close* buttons.

Click Dataset->Add->Structures (PDB).

Click the with Ki data button. Then, click the Start button.

SAnDReS will start the downloading of the PDB file.

After finishing the downloading, you get the following message:

SAnDReS finished the "Add Structures (PDB)" request!

Click the *Done* button and then the *Close* button.

SAnDReS created the *pdb* folder in the project directory with a downloaded structure.

Click Dataset->Add->Structures (PDBQT). Click the Yes option.

Set the binding affinity to K_i . Choose *No* for the *Unify ligands* option.

Click the Add button, then the Start button.

After finishing this task, you will get the following message:

SAnDReS finished the "Add Structures (PDBQT)" request!

Click the *Done* and the *Close* buttons.

Click *Dataset->Check Directories for Current Dataset*. SAnDReS will check any missing directory in the *pdbqt* folder.

You get the following message:

Done! There are no missing PDBQT directories!

Click Dataset->Check Missing PDBQT Files.

SAnDReS will check any missing PDBQT files. You get the following message:

Done! No missing PDBQT files in the dataset!

We do not need to update it since there are no missing files. We have only one structure. We finished the Dataset part of this tutorial.

6.3. Redocking

We will carry out redocking of the active ligand (ligand code: CD1) for structure 2DS1.

Click Redocking->Enter Vina Parameters. SAnDReS will open the file

vina par.csv, as shown in the figure below.



Figure 3. Fast Editor with input file (vina_par.csv) for redocking.

For this tutorial, keep the parameters as shown above.

Click the Save and Close buttons.

To perform redocking, click *Redocking->Run*. Click the *Yes* option.

Then click the *Run* button. Click the *Start* button to initiate the redocking.

Once finished with the docking simulation, you have the following message:

SAnDReS finished the "Redocking->Run" request using AutoDock Vina!

Click the Done and the Close buttons.

Click Redocking->Statistical Analysis. Click the Yes option.

Once finished with the statistical analysis, you have the following message:

SAnDReS finished the "Statistical Analysis" request!

Next, we check the docking result. This part identifies unsuccessful docking simulations. We may delete these structures from the dataset or run docking simulations outside SAnDReS.

Click Redocking->Check Unsuccessful Redocking Simulations.

After checking docking simulations, you have the following message: Number of structures with unsuccessful docking simulations: 0

We finished the Redocking part of this tutorial. We do not need the Scoring Function part.

6.4. Docking Simulation

Here, we will employ AutoDock Vina 1.2 to perform docking simulations of known inhibitors against our protein target, the CDK2. AutoDock Vina is fully integrated to SAnDReS. We will use the GUI interface to prepare the input files and start docking simulation with AutoDock Vina. We have the file *cdk2_ki.mol2* with 97 molecules. We will use these docked structures to generate a machine learning model.

Table 01 shows the steps for this part of Tutorial 01.

Table 01. List of commands to run this part of Tutorial 01.

Sequence number	Commands to click
01	Docking Simulation->Enter Parameters
02	Docking Simulation->Import Ligands
03	Docking Simulation->Import Receptor
04	Docking Simulation->Run
05	Docking Simulation->Merge Results
06	Docking Simulation->Add BindingDB Data
07	Edit->Docking Simulation for Machine Learning
08	Docking Simulation->Prepare Data for Machine Learning

The details for each part follow in this section.

Click Docking Simulation->Enter Parameters.

SAnDReS opens the *sim_par.csv* file, as shown below.



Figure 4. Fast Editor with input file (sim_par.csv) for docking simulation.

Be sure that the *lig_dir* is the project directory (./datasets/CDK2_Ki/) and *lig_in* has cdk2_ki.mol2. Click the Save button and the Close button.

Click Docking Simulation->Import Ligands. Click the Yes option.

When finished the conversion, SAnDReS shows the following message: SAnDReS finished the "Docking Simulation->Import Ligands" request!

SAnDReS converted the molecules in the *cdk2_ki.mol2* file to individual PDBQT files, one for each molecule found in the *cdk2_ki.mol2* file. All PDBQT files are in the *Sim/mols* folder in the project directory.

Now, SAnDReS imports the files *config.txt* and *receptor.pdbqt*. SAnDReS will copy these files to the *Sim* folder of the project directory.

Click Docking Simulation->Import Receptor. Click the Yes option.

When finished the copying, SAnDReS shows the following message: SAnDReS finished the "Docking Simulation->Import Receptor" request!

Now, we have two additional files: *config.txt* and *receptor.pdbqt*.

You may edit the *config.txt* file by clicking *Docking Simulation->Edit config.txt*.

It is not necessary. Now, we are going to run the docking simulation.

Click *Docking Simulation->Run*. Click the *Yes* option. Click the *Run* button.

Click the *Start* button.

After finishing the simulation, we have the following message: SAnDReS finished the "Docking Simulation->Run" request!

Click the *Done* button. Click the *Close* button.

Click *Docking Simulation->Merge Results*. Click the Yes option.

It is going to take a while. After finishing the merging, we have the following message: SAnDReS finished the "Merge Results" request!

We generated a file with docking results (*docking_simulation.csv*). This file has descriptors for each pose calculated for all ligands used in the simulation.

Now, we add binding affinity data downloaded from the BindingDB.

Click Docking Simulation->Add BindingDB Data. Click the Yes option.

Enter parameters to have the following setup.

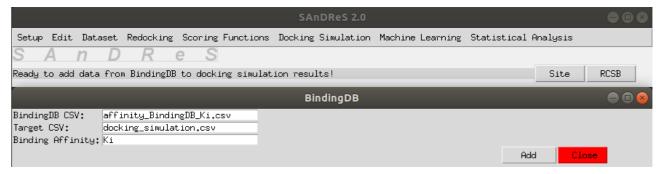


Figure 5. BindingDB menu.

Click the Add button.

We get the following message:

Done! SAnDReS added BindingDB data to docking simulation results!

Click the Close button.

Click *Edit->Docking Simulation for Machine Learning*. SAnDReS opens ./misc/inputs/ds4ml.in file. Enter *Ki* as shown below.

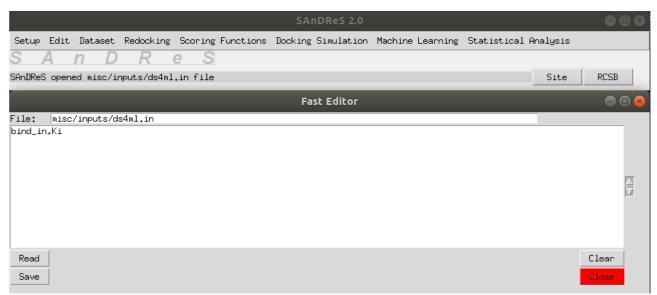


Figure 6. Fast Editor showing ds4ml.in file.

Click the Save and Close buttons.

Our last step in this part of Tutorial 01 is to prepare the docking_simulation.csv file for application of

a machine learning model.

Click Docking Simulation->Prepare Data for Machine Learning.

We get the following message:

SAnDReS finished the "Prepare Data for Machine Learning" request!

We are ready to generate our machine learning models based on docked structures.

We finished out our docking simulations.

6.5. Machine Learning

To build machine-learning models using docked structures, we will carry out the following steps. Click *Machine Learning-> Enter Parameters*.

SAnDReS opens the *mlr.in* file. In this file, we have the definition of the parameters necessary to apply the regression methods available in Scikit-Learn. We should enter parameters as shown below. The text marked in red needs updating.

```
dataset dir in,./datasets/CDK2 Ki/
sf\_file\_in, scores4xtal.csv
mlregmpy in,ml par.csv
preprocessing_in,StandardScaler
ml parameters in, ml.csv
scoring function file in, scores.csv
target_in,pKi
test_size_in,0.3
seed in, 271828
criterion in, r2
ml criterion_in,EDOME
data4criterion in, test
# Parameters for explore-sfs option
x n set in, 12
x n features in,8
```

We will focus on four lines.

```
dataset_dir_in,./datasets/CDK2_Ki/
target_in,pKi

x_n_set_in,12
x_n_features_in,8
```

The line <code>dataset_dir_in,./datasets/CDK2_Ki/</code> defines the project directory. The line <code>target_in,pKi</code> specifies the binding affinity.

The line $x_n_{set_in,12}$ shows the total number of features considered for exploring the SFS. The following line $x_n_{features_in,8}$ takes the number of features for each regression model.

Leave the parameters as indicated above.

Click the Save and the Close buttons.

SAnDReS will calculate the r^2 metric (since $criterion_in$, r2) between each feature and pK_i and select the highest correlation (r^2) features (12 features since $x_n_set_in$, 12). Next, SAnDReS will build $C_{12.8}$ x 54 models and choose the top model (highest EDOME since ml criterion in, EDOME).

Click Machine Learning->Explore->Scoring Function Space.

This process will take a while, depending on your hardware. Using an Intel Core i5-10300H processor, it took 1 hour and 50 minutes to generate 495x54 (26,730) regression models.

Note: Since exploring SFS is the most CPU-demanding of the SAnDReS tasks, we may run it outside the SAnDReS GUI. After preparing the file *mlr.in* as previously described, exit SAnDReS. To finish this SAnDReS session, click on *Setup->Exit*. Click on the Yes option. Then, open a Linux terminal and *cd* to the sandres2 directory. Then, type the following commands in a Linux terminal:

```
python3 mlr.py ./misc/data/mlr.in explore-sfs ./datasets/CDK2 Ki/mlr.log &
```

We get the following message:

Done! Finished the exploration of the Scoring Function Space.

SAnDReS generated a file named *models_test_set.csv* with the predictive performance. Below, we have the first lines of *models_test_set.csv*.

	Model features	Methods	r	p-value1	r2	rho	p-value2	MSE	RMSE	RSS	MAE	R2	DOME	EDOMEr2	EDOMErho .
1	Gauss 2 C Average Q Gauss 1 Q Torsional Torsions Repulsion	Decision Tree RegressorC\	0.792057	3.07233e-07	0.627354	0.598595	0.000602832	0.487607	0.698289	14.1406	0.519729	0.615007	0.951812	1.02673	1.03299
2	Hydrophobic S Average Q Gauss 1 Torsional Torsions N Repulsion	ExtraTreeRegressorCV	0.759859	1.7433e-06	0.577385	0.795779	2.46547e-07	0.561926	0.749617	16.2959	0.531287	0.556328	1.02031	1.1126	1.04055
3	Hydrophobic Gauss 2 C Gauss 1 Torsional Torsions N Repulsion	Decision TreeRegressorC\	0.733114	6.08882e-06	0.537456	0.739846	4.50755e-06	0.621661	0.788455	18.0282	0.531815	0.509164	1.07024	1.17742	1.1014
4	Hydrophobic Gauss 2 Hydrogen C Average Q Torsional N Repulsion	ExtraTreesRegressor	0.677421	5.42504e-05	0.458899	0.680872	4.80242e-05	0.711716	0.843632	20.6398	0.579958	0.438061	1.16784	1.296	1.21065
5	Hydrophobic Gauss 2 C Average Q Gauss 1 Torsions N Repulsion	DecisionTreeRegressor	0.700117	2.3601e-05	0.490164	0.5648	0.0014128	0.685597	0.828008	19.8823	0.622531	0.458683	1.16883	1.2881	1.24722
6	Gauss 2 Hydrogen C Gauss 1 Q Torsional N Repulsion	ExtraTreeRegressorCV	0.682258	4.57086e-05	0.465476	0.568006	0.0013082	0.685827	0.828147	19.889	0.638721	0.458502	1.17771	1.29624	1.25444
7	Gauss 2 Hydrogen C Average Q Q Torsions N Repulsion	ExtraTreeRegressor	0.716737	1.22096e-05	0.513712	0.5802	0.000969355	0.691967	0.831846	20.0671	0.636601	0.453654	1.18141	1.30162	1.25378
8	Hydrophobic Gauss 2 Hydrogen C Average Q Gauss 1 Torsions Repulsion	ExtraTreeRegressor	0.7051	1.94597e-05	0.497166	0.569597	0.00125884	0.691266	0.831424	20.0467	0.644014	0.454207	1.18487	1.30453	1.26062
9	Hydrophobic Hydrogen C Average Q Torsional Torsions N Repulsion	AdaBoostRegressor	0.656057	0.000111512	0.430411	0.643456	0.000166258	0.740791	0.860692	21.4829	0.5988	0.415104	1.20061	1.3355	1.25243
10	Hydrogen C Average Q Gauss 1 Q Torsions N Repulsion	DecisionTreeRegressor	0.674712	5.96304e-05	0.455236	0.587873	0.000797856	0.737447	0.858747	21.386	0.611424	0.417745	1.20429	1.33766	1.27285
11	Hydrophobic Gauss 2 Hydrogen Average Q Torsional Torsions N Repulsion	Extra Trees Regressor	0.650177	0.00013466	0.42273	0.663629	8.69435e-05	0.752783	0.867631	21.8307	0.592222	0.405636	1.20697	1.34538	1.25297
12	Hydrophobic Gauss 2 Hydrogen C Torsional Torsions N Repulsion	Extra Trees Regressor	0.668048	7.49442e-05	0.446289	0.677423	5.42453e-05	0.744286	0.86272	21.5843	0.606067	0.412346	1.20704	1.34249	1.2494
13	S Gauss 2 Hydrogen C Gauss 1 Torsional N Repulsion	KNeighborsRegressor	0.669339	7.17307e-05	0.448015	0.63726	0.000201034	0.717848	0.847259	20.8176	0.649126	0.43322	1.20849	1.3348	1.26176
14	Hydrophobic Gauss 2 C Average Q Torsional Torsions N Repulsion	Extra Trees Regressor	0.647083	0.000148474	0.418717	0.717576	1.17959e-05	0.769218	0.877051	22.3073	0.568849	0.39266	1.20899	1.35297	1.24154
15	Hydrophobic S Gauss 2 C Gauss 1 Torsions N Repulsion	AdaBoostRegressor	0.654445	0.000117476	0.428299	0.675013	5.90106e-05	0.751859	0.867098	21.8039	0.601255	0.406366	1.21069	1.34839	1.25355
16	Hydrophobic C Gauss 1 Q Torsional Torsions N Repulsion	AdaBoostRegressor	0.654763	0.00011628	0.428714	0.642199	0.000172848	0.73948	0.85993	21.4449	0.623369	0.41614	1.21201	1.34531	1.26372
17	Gauss 2 C Average Q Q Torsional Torsions N Repulsion	Decision Tree Regressor	0.669951	7.02509e-05	0.448834	0.651365	0.000129667	0.741728	0.861236	21.5101	0.620806	0.414365	1.21248	1.3465	1.2616
18	Hydrophobic Gauss 2 Hydrogen Average Q Q Torsions N Repulsion	ExtraTreesRegressor	0.635086	0.000214677	0.403334	0.707476	1.77249e-05	0.771848	0.878549	22.3836	0.59213	0.390584	1.22223	1.36574	1.25675
19	S Hydrogen C Gauss 1 Torsional Torsions N Repulsion	KNeighborsRegressor	0.652108	0.000126626	0.425245	0.649587	0.000137201	0.731635	0.855356	21.2174	0.657096	0.422334	1.22356	1.35307	1.27275
20	Hydrophobic S Gauss 2 Hydrogen C Torsional N Repulsion	AdaBoostRegressor	0.656593	0.00010959	0.431114	0.709371	1.64421e-05	0.748044	0.864896	21.6933	0.634272	0.409378	1.22441	1.35942	1.25843
21	Hydrophobic S Gauss 2 Hydrogen Gauss 1 Q Torsional N	KNeighborsRegressor	0.653128	0.000122559	0.426576	0.582584	0.000912918	0.730834	0.854888	21.1942	0.660866	0.422967	1.22497	1.35407	1.29413
22	Hydrophobic Gauss 2 C Gauss 1 Q Torsions N Repulsion	ExtraTreeRegressorCV	0.642244	0.000172608	0.412477	0.642505	0.000171221	0.7524	0.86741	21.8196	0.628745	0.405939	1.225	1.36145	1.2761

Figure 7. Partial view of the file models_test_set.csv.

To generate a plot to visualize the results shown in the file *models_test_set.csv*, click *Machine Learning->Plotting*.

After a few seconds, SAnDReS generates a plot named *models_test_set.pdf*. Figure 8 shows the results.

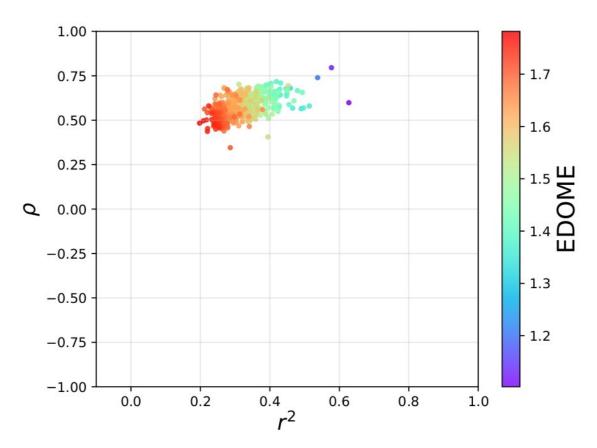


Figure 8. Predictive performance of the machine-learning models.

Taking the lowest EDOME among the top models (the first model in Figure 7), we select the DecisionTreeRegressorCV model with the following features: *Gauss 2,C,Average Q,Gauss 1,Q,Torsional,Torsions,Repulsion*

Click Machine Learning-> Enter Parameters.

Insert the chosen features in line s features in as follows.

```
s\_features\_in, \textit{Gauss 2,C,Average Q,Gauss 1,Q,Torsional,Torsions,Repulsion}
```

Insert the number of features (8) as follows:

s_n_features,8

Click the Save and the Close buttons.

Click Machine Learning->Explore->Single Set of Features.

This part is faster, and it takes only a few seconds.

We get the following message:

Done! Finished the exploration of the single set of features.

Once finished, we have a file named *scores4xtal_test_stats_analysis_models.csv*. This file has the predictive performance of 54 regression models generated using the features defined in the command line *s_features_in*. Figure 9 shows the first lines of this file.

	Feature	г	p-value(r)	r2	rho	p-value(rho)	MSE	RMSE	RSS	MAE	R2	DOME	EDOMEr2	EDOMErho	EDOME
1	Decision Tree Regressor CV	0.792057	3.07233e-07	0.627354	0.598595	0.000602832	0.487607	0.698289	14.1406	0.519729	0.615007	0.951812	1.02673	1.03299	1.1024
2	Gradient Boosting Regressor	0.50062	0.00567624	0.25062	0.68268	4.50235e-05	1.02925	1.01452	29.8482	0.627109	0.187351	1.44323	1.65629	1.4777	1.68642
3	AdaBoostRegressor	0.455675	0.0129866	0.20764	0.618197	0.000351727	1.08673	1.04246	31.5151	0.655335	0.141969	1.5008	1.72876	1.54861	1.77042
4	GradientBoostingRegressorC1	0.410927	0.0267993	0.168861	0.509422	0.00476391	1.06222	1.03064	30.8043	0.719873	0.161321	1.51123	1.72835	1.58886	1.79663
5	ExtraTreesRegressor	0.429545	0.0200474	0.184509	0.490947	0.00684595	1.08137	1.03989	31.3599	0.689716	0.146195	1.51197	1.73639	1.59537	1.80947
6	RandomForestRegressor	0.450781	0.0141217	0.203204	0.492672	0.00662363	1.09389	1.04589	31.7229	0.689193	0.136312	1.52146	1.74951	1.60381	1.82159
7	MLPRegressor	0.439823	0.0169653	0.193444	0.482818	0.0079811	1.09359	1.04575	31.7142	0.705991	0.136549	1.52891	1.75588	1.61401	1.83046
8	SGDRegressorCV	0.372018	0.0468964	0.138398	0.464836	0.0110658	1.1211	1.05882	32.512	0.717233	0.114826	1.55533	1.78958	1.64483	1.86789
9	SGDRegressor	0.430106	0.0198679	0.184992	0.484296	0.00776364	1.13058	1.06329	32.7867	0.71057	0.107347	1.55959	1.79698	1.64264	1.86952
10	VotingRegressor	0.371566	0.0471858	0.138061	0.427393	0.020748	1.14681	1.07089	33.2576	0.686912	0.0945269	1.56158	1.80511	1.66326	1.89375
11	RandomForestRegressorCV	0.341245	0.0700407	0.116448	0.434044	0.0186461	1.12598	1.06112	32.6535	0.727181	0.110974	1.56369	1.79875	1.66296	1.88569
12	Bayesian Ridge	0.417129	0.0243706	0.173996	0.485774	0.00755121	1.13706	1.06633	32.9749	0.720548	0.102225	1.56916	1.80783	1.65127	1.87954
13	AdaBoost RegressorCV	0.340979	0.0702737	0.116267	0.514494	0.00429741	1.15337	1.07395	33.4476	0.701726	0.0893539	1.57323	1.81778	1.64644	1.8815
14	TweedieRegressor	0.413776	0.0256603	0.171211	0.447592	0.014905	1.1462	1.07061	33.2397	0.727142	0.095015	1.57922	1.82014	1.67305	1.90213
15	Decision Tree Regressor	0.643114	0.000168026	0.413596	0.730035	6.9659e-06	1.11145	1.05426	32.2322	0.787715	0.122446	1.58179	1.80891	1.60466	1.82894
16	BaggingRegressor	0.391381	0.0357709	0.153179	0.387979	0.0375546	1.14161	1.06846	33.1066	0.754589	0.0986394	1.58854	1.82645	1.70236	1.92626
17	ARDRegression	0.348419	0.0639852	0.121396	0.382559	0.0405423	1.16547	1.07957	33.7988	0.739646	0.0797935	1.59979	1.84557	1.71481	1.94611
18	MLPRegressorCV	0.355002	0.0587945	0.126026	0.420495	0.0231295	1.15332	1.07393	33.4462	0.774701	0.0893938	1.60707	1.84713	1.70836	1.9359
19	TweedieRegressorCV	0.336594	0.0741978	0.113296	0.387486	0.0378187	1.18013	1.08634	34.2239	0.781406	0.0682191	1.63063	1.87807	1.74187	1.97543
20	Ridge	0.295202	0.120027	0.0871441	0.354477	0.0591955	1.2092	1.09964	35.0668	0.739516	0.0452702	1.63328	1.89185	1.75622	1.99895
21	Extra Trees Regressor CV	0.24879	0.193116	0.0618965	0.380589	0.0416747	1.21527	1.10239	35.2429	0.774828	0.0404742	1.65418	1.91233	1.76635	2.01015
22	Voting RegressorCV	0.213015	0.267233	0.0453756	0.316541	0.0943313	1.22073	1.10487	35.4012	0.770178	0.0361644	1.65617	1.91621	1.79165	2.03445
23	KNeighhorsRegressor	U 348884	0.0636072	0 12172	0.410743	0.0268744	1 228	1 10815	35 6121	0.78828	0.0304238	1 67017	1 9312	1 77107	2 0191

Figure 9. Partial view of the file scores4xtal_test_stats_analysis_models.csv.

As expected, our best regression model is on the first line of Figure 9 (DecisionTreeRegressorCV).

Note: SAnDReS may show *nan* (not a number) for statistical analysis of some regression methods. It happens due to errors generated in the Scikit-Learn (e.g., an all-zeros column taken as a feature). If you have only a few instances of this problem, you may ignore them. Otherwise, you may have to change the regression parameters. Please see Appendix.

To save your best model (DecisionTreeRegressorCV), click *Machine Learning->Models->Edit Current Model*.

SAnDReS calls Fast Editor and shows the file ./misc/data/model.in. It follows the content of the file model.in.

```
model_joblib,model_DecisionTreeRegressorCV.joblib
model_id,CDK2_Ki_DecisionTreeRegressorCV
model_stats,scores4xtal_test_DecisionTreeRegressorCV.csv
scores4xtal_test,scores4xtal_test.csv
scores4xtal_training,scores4xtal_training.csv
```

We define the joblib model in the line model_joblib,model_ DecisionTreeRegressorCV.joblib

You use the best regression model here. In this tutorial we chose DecisionTreeRegressorCV, so the *joblib* file is *model DecisionTreeRegressorCV.joblib*.

The line <code>model_id</code>, <code>CDK2_Ki_DecisionTreeRegressorCV</code> indicates the name we chosen for this regression model (also used to create a new folder into the <code>./misc/data/models/</code> directory). The

following three lines indicate the CSV files that SAnDReS will copy to the folder created into the directory named ./misc/data/models/. Here we name this folder as CDK2_Ki_DecisionTreeRegressorCV.

After entering the parameters shown above, click the Save and Close buttons of the Fast Editor.

Now we save our current model for future use. Click *Machine Learning->Models->Save Current Model*.

SAnDReS shows the following message: SAnDReS finished the "Save Current Model" request!

SAnDReS created the following folder ./misc/data/models/CDK2_Ki_DecisionTreeRegressorCV/. In this new folder, you find the following files: features.csv, model_DecisionTreeRegressorCV.joblib, score4xtal_test.csv, scores4xtal_test_DecisionTreeRegressorCV.csv, scores4xtal_training.csv, and summary.txt.

To generate a scatter plot for test set, we click *Statistical Analysis->Scatter Plot->Edit Parameters*. We update *scatter_plot_par.csv* file with the following parameters.

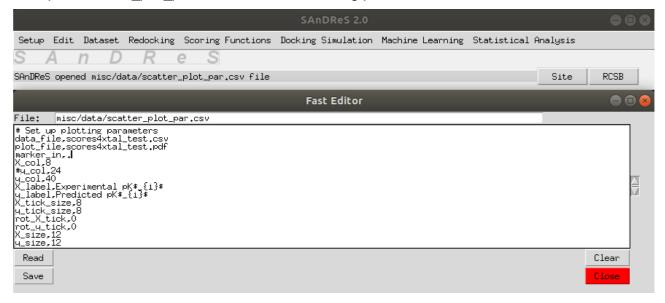


Figure 10. Fast Editor showing scatter_plot_par.csv file.

Click the Save and Close buttons.

Click Statistical Analysis->Scatter Plot->Generate. Click Plot button. Click the Close button.

We have the following plot.

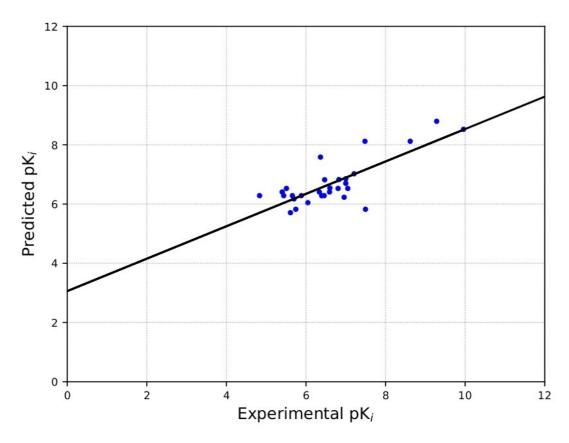


Figure 11. Scattering plot of predicted pK_i vs. experimental pK_i.

We can see the agreement between predicted and experimental values for the test set.

Now, we save our results as a zipped folder. Click Setup->Project Directory->Backup Current Project.

Click the Yes option. It may take a few minutes. After backing up the current project directory, you get the following message:

Successfully created a backup of the directory ./datasets/CDK2_Ki/

You may delete or unzip this zipped folder using additional options in the Setup menu.

To finish this session, click on Setup->Exit. Click on the Yes option.

We finished Tutorial 01.

7. Tutorial 02: AlphaFold Model of CDK19 with IC₅₀ Data

This tutorial focuses on developing a machine-learning model to predict inhibition (pIC₅₀) of cyclin-dependent kinase 19 (CDK19). We describe the results tutorial 02 in <u>de Azevedo et al., 2024</u>. So far, there is no experimental structure for this enzyme (search performed on October 1, 2024). Therefore, this tutorial will employ a model (PDB access code: (AF_AFQ9BWU1F1) generated using AlphaFold (Jumper et al., 2021). We superposed the structure of CDK19 (AF_AFQ9BWU1F1) onto the crystal structure 2DS1. Then, we transferred the inhibitor of 2DS1 (ligand CD1) to the superposed CDK19. Finally, we carried out model optimization of the CDK19-1CD structure using the minimization of sidechain positions. We employed Molegro Virtual Docker (Thomsen & Christensen, 2006) to optimize the CDK19-CD1 complex. We validated the docking (re-docking) using the protocol described in Tutorial 01 for structure 2DS1 using AutoDock Vina 1.2. The RMSD (docking) between the docked and the model of the CDK19-CD1 complex is 1.133 Å. This tutorial starts with a setup section followed by the docking simulation employing binding data available in the BindingDB. The data related to coordinate preparation and redocking (docking validation files) are available once SAnDReS copies the files for Tutorial 02.

7.1. Setup

For this part of Tutorial 02, we will define the project directory and copy the files to run this tutorial. Click the following sequence in the main menu: *Setup->Project Directory->Enter*.

After writing the project directory (./datasets/CDK19_IC50_AlphaFold/) in the Fast Editor, click the Save and Close buttons.

Then, click Setup->Project Directory->Make. Click the Yes option.

SAnDReS will generate a new directory named *CDK19_IC50_AlphaFold*. You should get the following message:

Successfully created the directory ./datasets/CDK19_IC50_AlphaFold/

To copy all necessary files to run this tutorial to your project directory, click Setup->Copy Files to Run Tutorials->Tutorial 02.

SAnDReS will copy all necessary files to run this tutorial. You should get the following message: SAnDReS finished the "Copy Files to Run Tutorial 02" request!

Click Setup->BindingDB Data. Click on the Yes button. Enter the parameters shown below.

BindingDB SDF: cdk19_ic50.sdf BindingDB TSF: cdk19_ic50.tsv

Binding Affinity: IC50

Option: Clean

Click the *Prepare* button.

After finishing, you get the following message:

Done! SAnDReS prepared BindingDB data for machine learning modeling.

SAnDReS created the following files necessary for the modeling: *affinity_BindingDB_IC50.csv* and *cdk19_IC50_out.csv*.

Click the Close button.

Now, we finished the Setup.

7.2. Docking Simulation

For this tutorial, do not run the sections Dataset and Redocking. We have all files to carry out the docking simulations using known inhibitors of CDK19. These structures are in the file $cdk19_ic50.mol2$. The commands necessary in this part of Tutorial 02 are in Table 01.

Click Docking Simulation->Enter Parameters.

SAnDReS opens the *sim_par.csv* file. Enter parameters as shown below.



Figure 12. Fast Editor with input file (sim par.csv) for the docking simulation.

Be sure that the *lig_dir* is the project directory (./datasets/CDK19_IC50_AlphaFold/) and *lig_in* has cdk19 ic50.mol2. Click the Save button and the Close button.

Click Docking Simulation->Import Ligands.

Click the Yes option.

When finished the conversion, SAnDReS shows the following message:

SAnDReS finished the "Docking Simulation->Import Ligands" request!

SAnDReS converted the molecules in the *cdk19_ic50.mol2* file to individual PDBQT files, one for each molecule found in the *cdk19_ic50.mol2* file. All PDBQT files are in the *Sim/mols* folder in the project directory.

Now, SAnDReS imports the files *config.txt* and *receptor.pdbqt*. SAnDReS will copy these files to the *Sim* folder of the project directory.

Click Docking Simulation->Import Receptor. Click the Yes option.

When finished the copying, SAnDReS shows the following message: SAnDReS finished the "Docking Simulation->Import Receptor" request!

Now, we have two additional files: *config.txt* and *receptor.pdbqt*.

You may edit the *config.txt* file by clicking *Docking Simulation->Edit config.txt*.

It is not necessary. Now, we are going to run the docking simulation.

Click Docking Simulation->Run. Click the Yes option.

Click the Run button. Click the Start button.

After finishing the simulation, we have the following message:

SAnDReS finished the "Docking Simulation->Run" request!

Click the Done button. Click the Close button.

Click *Docking Simulation->Merge Results*. Click the Yes option.

It is going to take a while. After finishing the merging, we have the following message: SAnDReS finished the "Merge Results" request!

We generated a file with results (*docking_simulation.csv*). This file has descriptors calculated for all ligands used in the simulation.

Now, we add binding affinity data downloaded from the BindingDB.

Click *Docking Simulation->Add BindingDB Data*. Click on the Yes option. We get the following popup window.

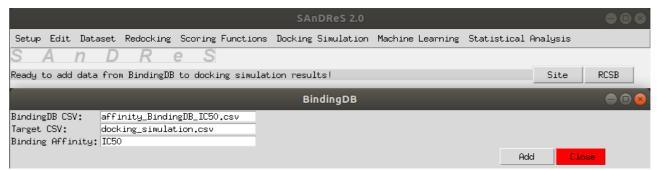


Figure 13. BindingDB menu.

Be sure to set all parameters as indicated above. Click the *Add* button.

We get the following message:

Done! SAnDReS added BindingDB data to docking simulation results!

Click the Close button.

Now, we edit the file *ds4ml.in*.

Click Edit->Docking Simulation for Machine Learning.

It should have the following line: bind in, IC50

Click the Save and Close buttons.

Our last step in this part of Tutorial 02 is to prepare the *docking_simulation.csv* file.

Click Docking Simulation->Prepare Data for Machine Learning.

We get the following message:

SAnDReS finished the "Prepare Data for Machine Learning" request!

We finished out our docking simulations.

7.3. Machine Learning

Here, we will follow the steps to build machine-learning models.

Click Machine Learning-> Enter Parameters.

SAnDReS opens the file *mlr.in*. In this file, we have the definition of the parameters necessary to apply the regression methods available in Scikit-Learn. We should enter parameters as shown below. The text marked in red needs updating.

```
dataset dir in,./datasets/CDK19 IC50 AlphaFold/
sf file in, scores4xtal.csv
mlregmpy in,ml par.csv
preprocessing\_in, StandardScaler
ml parameters in, ml.csv
scoring function file in, scores.csv
target_in,pIC50
test size in,0.3
seed in,271828
criterion in, r2
ml criterion in, EDOME
data4criterion in, test
# Parameters for explore-sfs option
x n set in, 12
x n features in,8
```

In this part of the tutorial, we focus on four lines.

```
dataset_dir_in,./datasets/CDK19_IC50_AlphaFold/
target_in,pIC50

x_n_set_in,12
x_n_features_in,8
```

The line <code>dataset_dir_in,./datasets/CDK19_IC50_AlphaFold/</code> defines the project directory. The line <code>target_in,pIC50</code> specifies the binding affinity.

The line $x_n_{set_in,12}$ shows the total number of features considered for exploring the SFS. The following line $x_n_{features_in,8}$ takes the number of features for each regression model.

Leave the parameters as indicated above.

Click the Save and the Close buttons.

Click Machine Learning->Explore->Scoring Function Space.

This process will take a while, depending on your computer. Using an Intel Core i5-10300H processor, it took 1 hour and 54 minutes to generate 495x54 (26,730) machine-learning models. After finishing all regression models, SAnDReS generates a file named *models_test_set.csv* with

the predictive performance. Below, we have the first lines of *models_test_set.csv*.

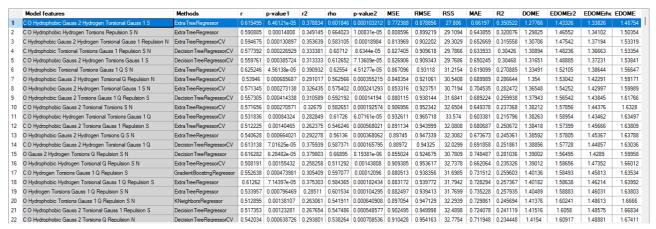


Figure 14. Partial view of the file models_test_set.csv.

To generate a plot to visualize the results in the file *models_test_set.csv*, click *Machine Learning-* >*Plotting*.

After a few seconds, SAnDReS generates a file named *models_test_set.pdf*. The following figure shows it.

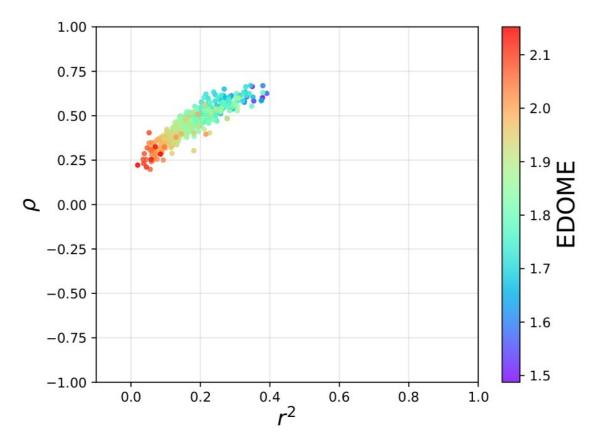


Figure 15. Predictive performance of the regression models.

Taking the lowest EDOME among the top models, we select the ExtraTreeRegressor model with the following features: *C,O,Hydrophobic,Gauss 2,Hydrogen,Torsional,Gauss 1,S*Click *Machine Learning-> Enter Parameters*.

Insert the chosen features in line $s_features_in$ as follows.

```
s features in, C,O, Hydrophobic, Gauss 2, Hydrogen, Torsional, Gauss 1,S
```

Insert the number of features (8) as follows:

```
s n features,8
```

Click the Save and the Close buttons.

Click Machine Learning->Explore->Single Set of Features.

After a few seconds, we have a file named *scores4xtal_test_stats_analysis_models.csv*. This file has the predictive performance of 54 regression models generated using the features defined in the command line *s_features_in*. The following figure shows the first lines of this file.

	Feature	r	p-value(r)	r2	rho	p-value(rho)	MSE	RMSE	RSS	MAE	R2	DOME	EDOMEr2	EDOMErho	EDOME
1	ExtraTreeRegressor	0.615495	6.46121e-05	0.378834	0.601846	0.000103212	0.772388	0.878856	27.806	0.66197	0.350522	1.27766	1.43326	1.33826	1.48754
2	RandomForestRegressor	0.361772	0.0301539	0.130879	0.426943	0.0094072	1.03436	1.01703	37.237	0.788195	0.130238	1.55309	1.78005	1.65544	1.87002
3	BaggingRegressor	0.303049	0.0723926	0.091839	0.36212	0.0299841	1.09846	1.04808	39.5447	0.794084	0.0763354	1.60692	1.85347	1.7289	1.96016
4	ExtraTreesRegressor	0.304599	0.0708735	0.0927803	0.406079	0.0139943	1.10863	1.05292	39.9109	0.814975	0.0677819	1.62538	1.87373	1.73049	1.96561
5	Voting Regressor	0.222803	0.191509	0.0496411	0.327903	0.0508978	1.13281	1.06433	40.7811	0.832548	0.047456	1.65326	1.90804	1.78466	2.02295
6	BaggingRegressorCV	0.342433	0.040918	0.117261	0.418829	0.01101	1.16812	1.0808	42.0524	0.791883	0.017761	1.66132	1.92997	1.76004	2.01558
7	ExtraTreesRegressorCV	0.337953	0.0438096	0.114212	0.367313	0.0275401	1.15957	1.07683	41.7445	0.831222	0.0249532	1.67368	1.93699	1.78928	2.0377
8	AdaBoost Regressor	0.191792	0.262466	0.0367842	0.266246	0.116514	1.17127	1.08225	42.1659	0.851775	0.01511	1.69316	1.95878	1.84532	2.0917
9	ElasticNet	nan	nan	nan	nan	nan	1.19006	1.0909	42.8423	0.875309	-0.000688334	1.71977	1.98972	nan	nan
10	Lasso	nan	nan	nan	nan	nan	1.19006	1.0909	42.8423	0.875309	-0.000688334	1.71977	1.98972	nan	nan
11	SGDRegressorCV	0.134891	0.43282	0.0181954	0.184043	0.2826	1.20462	1.09755	43.3663	0.87703	-0.0129289	1.732	2.00645	1.91458	2.16602
12	KNeighborsRegressor	0.189078	0.269406	0.0357507	0.245765	0.148509	1.24598	1.11623	44.8553	0.831231	-0.0477073	1.74201	2.03281	1.89828	2.16822
13	AdaBoost RegressorCV	0.207312	0.22505	0.0429784	0.21875	0.199923	1.23129	1.10964	44.3266	0.863338	-0.035359	1.74603	2.02992	1.91284	2.17507
14	Voting RegressorCV	0.205747	0.228648	0.042332	0.247537	0.145512	1.22945	1.10881	44.2603	0.867812	-0.0338109	1.7468	2.0298	1.90198	2.16478
15	BayesianRidge	-0.234106	0.169364	0.0548058	-0.228733	0.17965	1.22409	1.10639	44.0672	0.886943	-0.0292994	1.7522	2.03216	2.14009	2.37475
16	TheilSenRegressorCV	0.139678	0.416513	0.01951	0.296091	0.0795322	1.25903	1.12207	45.3252	0.838706	-0.0586839	1.75593	2.05039	1.89176	2.16785
17	KNeighborsRegressorCV	0.192267	0.261262	0.0369668	0.270309	0.110834	1.27116	1.12746	45.7618	0.822267	-0.0688819	1.75778	2.05726	1.90322	2.18283
18	LinearRegressionCV	0.152654	0.374098	0.0233032	0.291841	0.0841547	1.24514	1.11586	44.8249	0.872211	-0.0469975	1.76128	2.04897	1.89831	2.1679
19	RandomForestRegressorCV	0.194969	0.254492	0.0380129	0.204521	0.231496	1.25634	1.12087	45.2283	0.869125	-0.05642	1.76854	2.06004	1.93921	2.20829
20	RidgeCV	0.128785	0.454118	0.0165856	0.249211	0.14272	1.25137	1.11865	45.0493	0.877618	-0.0522398	1.76884	2.05815	1.92158	2.19082
21	Gradient Boosting Regressor	0.0632332	0.714084	0.00399843	0.140125	0.415009	1.25601	1.12072	45.2162	0.871127	-0.0561377	1.76926	2.06051	1.96715	2.23274
22	NuSVRCV	0.140122	0.41502	0.0196341	0.231824	0.17368	1.26481	1.12464	45.5331	0.88343	-0.0635389	1.78224	2.07545	1.94074	2.21305

Figure 16. Partial view of the file scores4xtal test stats analysis models.csv.

As expected, our best regression model is on the first line (ExtraTreeRegressor).

To save your best model (ExtraTreeRegressor), click *Machine Learning->Models->Edit Current Model*.

SAnDReS calls Fast Editor and shows the ./misc/data/model.in file. It follows the content of the file model.in.

```
model_joblib,model_ExtraTreeRegressor.joblib
model_id,CDK19_IC50_ExtraTreeRegressor
model_stats,scores4xtal_test_ExtraTreeRegressor.csv
scores4xtal_test,scores4xtal_test.csv
scores4xtal_training,scores4xtal_training.csv
```

After entering the parameters, click the Save and the Close buttons on the Fast Editor.

Now, we save our current model for future use. Click *Machine Learning->Models->Save Current Model*.

SAnDReS shows the following message:

SAnDReS finished the "Save Current Model" request!

SAnDReS created the folder: ./misc/data/models/CDK19_IC50_ExtraTreeRegressor/. In this folder, you find the following files: features.csv, model_ExtraTreeRegressor.joblib, score4xtal_test.csv, scores4xtal_test_ExtraTreeRegressor.csv, scores4xtal_training.csv, and summary.txt.

To generate a scatter plot for the test set, we click Statistical Analysis->Scatter Plot->Edit Parameters.

Update the scatter plot par.csv file with the following parameters.

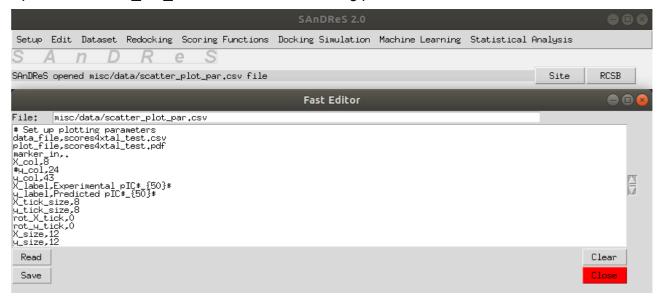


Figure 17. Fast Editor showing the file scatter plot par.csv.

Click the Save and the Close buttons.

Click Statistical Analysis->Scatter Plot->Generate. Click the Plot button. Click the Close button.

We have the following plot.

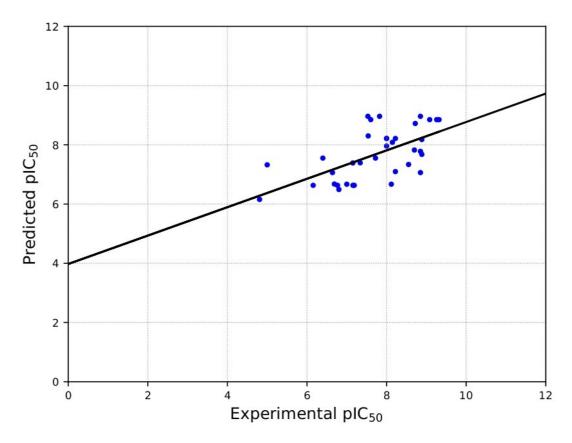


Figure 18. Scattering plot of predicted pIC₅₀ vs. experimental pIC_{i50}.

We can see the agreement between predicted and experimental values for the test set.

Now, we save our results as a zipped folder. Click *Setup->Project Directory->Backup Current Project*. Click the Yes option. After backing up the current project directory, you get the following message:

Successfully created a backup of the directory ./datasets/CDK19_IC50_AlphaFold/

You may delete or unzip this zipped folder using additional options in the Setup menu.

To finish this session, click on Setup->Exit. Click on the Yes option.

We finished Tutorial 02.

For both tutorials, we now have the .job files that could be used to predict binding for these protein systems.

8. GitHub and Wiki

You find addition information about SAnDReS available on GitHub (https://github.com/azevedolab/sandres) and the Wiki (https://github.com/azevedolab/sandres/wiki/SAnDReS).

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10. Appendix: Regression Methods

In the following page, we describe all parameters used for each regression method available in SAnDReS 2.0. The values for these parameters are in the file *misc/data/ml.csv*. Mostly, we keep the same names used in the Scikit-Learn manual. The exceptions are the parameters *rand_in+* and *cv_in**. In the following table, we provide links for complete descriptions of the parameters for each regression method.

Half of the regression methods available in SAnDReS use cross-validation (CV). We implemented the Kfold class from Scikit-Learn to perform cross-validation. The Kfold class builds an n-fold cross-validation loop and tests the generalization ability of regression. Cross-validation better estimates of how well we could generalize to predict unseen data.

Scikit-Learn (Pedregosa et al., 2011) provides some regression classes with built-in cross-validation implementation, e.g., ElasticNetCV. However, this inclusion of built-in CV is not available for all regression methods (e.g., AdaBoostRegressor). Therefore, we adopted the same CV approach (Coelho & Richert, 2015) for the regression methods in SAnDReS 2.0. The MLRegMPy package has a class (ValidationLoop) that carries out cross-validation for all CV methods.

Reference: Coelho LP, Richert W. (2015) Building Machine Learning Systems with Python. 2nd ed. Packt Publishing Ltd. Birmingham UK, 301 pp. See page 162 (Cross-validation for regression). Before applying the regression methods, SAnDReS scales the data using thefollowing methods available in Scikit-Learn: StandardScaler, MaxAbsScaler, MinMaxScaler, and RobustScaler.

Regression methods available in SAnDReS 2.0.

Method	Cross Validation	Parameters in <i>ml.in</i> file	Default values for parameters in ml.in file	Link
AdaBoostRegressor	None	Regression method,	AdaBoostRegressor,	https://scikit-
9		base estimator,	None,	learn.org/stable/modules
		n estimators,	50,	/generated/sklearn.ense
		learning rate,	1.0,	mble.AdaBoostRegress
		loss,	linear,	<u>or.html</u>
		random state	None	
AdaBoostRegressorCV	Kfold class	Regression method,	AdaBoostRegressorCV,	https://scikit-
ŭ		base estimator,	None,	learn.org/stable/modules
		n estimators,	50,	/generated/sklearn.ense
		learning rate,	1.0,	mble.AdaBoostRegress
		loss,	linear,	<u>or.html</u>
		random state,	None,	
		cv in*	5	
ARDRegression	None	Regression method,	ARDRegression,	https://scikit-
9		n iter,	1000,	learn.org/stable/modules
		tol,	1e-3,	/generated/sklearn.linear
		alpha 1,	1e-6,	model.ARDRegression.
		alpha 2,	1e-6,	html
		lambda 1,	1e-6,	- Harri
		lambda 2,	1e-6,	
		compute score,	False,	
		threshold lambda,	10000,	
		fit intercept,	True,	
		copy X,	True,	
		verbose,	False,	
		rand in ⁺	1123581321	
ARDRegressionCV	Kfold class	Regression method,	ARDRegressionCV,	https://scikit-
7 ii (B) (ogi ocolorio v	Tribia blacc	n iter,	1000,	learn.org/stable/modules
		tol,	1e-3,	/generated/sklearn.linear
		alpha 1,	1e-6,	model.ARDRegression.
		alpha_2,	1e-6,	html
		lambda 1,	1e-6,	110111
		lambda 2,	1e-6,	
		compute score,	False,	
		threshold lambda,	10000,	
		fit intercept,	True,	
		copy X,	True,	
		verbose,	False,	
		rand in ⁺ ,	1123581321,	
		cv in*	5	
		· _=		

BaggingRegressor	None	Regression	BaggingRegressor,	https://scikit-
		method,base_estimator,	None,	learn.org/stable/module
		n_estimators,	10,	s/generated/sklearn.en
		max_samples,	1.0,	semble.BaggingRegres
		max_features,	1.0,	sor.html
		bootstrap,	True,	
		bootstrap_features,	False,	
		oob_score,	False,	
		warm_start,	False,	
		n_jobs,	-1,	
		random_state,	None,	
		verbose	0	

BaggingRegressorCV	Kfold class	Regression method,	BaggingRegressorCV,	https://scikit-
		base estimator,	None,	learn.org/stable/module
		n estimators,	10,	s/generated/sklearn.en
		max samples,	1.0,	semble.BaggingRegres
		max features,	1.0,	sor.html
		bootstrap,	True,	
		bootstrap features,	False,	
		oob score,	False,	
		warm start,	False,	
		n jobs,	-1,	
		random_state,	None,	
		verbose,	0,	
		cv_in*	5	
BayesianRidge	None	Regression method,	BayesianRidge,	https://scikit-
		n iter,	1000,	learn.org/stable/modules
			1e-3,	/generated/sklearn.linear
		alpha 1,	1e-6,	model.BayesianRidge.
		alpha 2,	1e-6,	html
		lambda 1,	1e-6,	
		lambda 2,	1e-6,	
		alpha init,	None,	
		lambda init,	None,	
		compute score,	False,	
		fit intercept,	True,	
		copy X,	True,	
		verbose,	False,	
		rand_in+	1123581321	
BayesianRidgeCV	Kfold class	Regression method,	BayesianRidgeCV,	https://scikit-
		n iter,	1000,	learn.org/stable/modules
		tol,	1e-3,	/generated/sklearn.linear
		alpha 1,	1e-6,	model.BayesianRidge.
		alpha 2,	1e-6,	html
		lambda 1,	1e-6,	
		lambda 2,	1e-6,	
		alpha init,	None,	
		lambda_init,	None,	
		compute_score,	False,	
		fit intercept,	True,	
		copy X,	True,	
		verbose,	False,	
		rand_in+,	1123581321,	

		cv_in*	5	
DecisionTreeRegressor	None	Regression method,	DecisionTreeRegressor,	https://scikit-
		criterion,	squared_error,	learn.org/stable/module
		splitter,	best,	s/generated/sklearn.tre
		max_depth,	None,	e.DecisionTreeRegress
		min_samples_split,	2,	<u>or.html</u>
		<pre>min_samples_leaf,</pre>	1,	
		<pre>min_weight_fraction_leaf,</pre>	0.0,	
		max_features,	None,	
		random_state,	None,	
		<pre>max_leaf_nodes,</pre>	None,	
		<pre>min_impurity_decrease,</pre>	0.0,	
		ccp_alpha	0.0	

DecisionTreeRegressorCV	Kfold class	Regression method,	DecisionTreeRegressorCV,	https://scikit-
		criterion,	squared_error,	learn.org/stable/modules
		splitter,	best,	/generated/sklearn.tree.
		max_depth,	None,	DecisionTreeRegressor.
		min_samples_split,	2,	html
		min_samples_leaf,	1,	
		min weight fraction leaf,	0.0,	
		max features,	None,	
		random_state,	None,	
		max leaf nodes,	None,	
		min impurity decrease,	0.0,	
		ccp alpha,	0.0,	
		cv in*	5	
ElasticNet	None	Regression method,	ElasticNet,	https://scikit-
		alpha,	1.0,	learn.org/stable/modules
		11 ratio,	0.5,	/generated/sklearn.linear
		fit intercept,	True,	model.ElasticNet.html
		precompute,	False,	
		max iter,	1000,	
		copy X,	True,	
		tol,	1e-4,	
		warm start,	False,	
		positive,	False,	
		random state,	None,	
		selection,	cyclic,	
		rand in ⁺	1123581321	
ElasticNetCV	Kfold class	Regression method,	ElasticNetCV,	https://scikit-
	1	alpha,	1.0,	learn.org/stable/modules
		11 ratio,	0.5,	/generated/sklearn.linear
		fit intercept,	True,	model.ElasticNet.html
		precompute,	False,	
		max iter,	1000,	
		copy X,	True,	
		tol,	1e- 4,	
		warm start,	False,	
		positive,	False,	
		random state,	None,	
		selection,	cyclic,	
		rand in ⁺ ,	1123581321,	
		cv in*	5	

ExtraTreeRegressor	None	Regression method,	ExtraTreeRegressor,	https://scikit-
		criterion,	squared_error,	learn.org/stable/modules
		splitter,	random,	/generated/sklearn.tree.
		max_depth,	None,	ExtraTreeRegressor.htm
		min_samples_split,	2,	Ī
		min_samples_leaf,	1,	
		<pre>min_weight_fraction_leaf,</pre>	0.0,	
		max_features,	1.0,	
		random_state,	None,	
		min_impurity_decrease,	0.0,	
		max_leaf_nodes,	None,	
		ccp_alpha	0.0	

ExtraTreeRegressorCV	Kfold class	Regression method,	ExtraTreeRegressorCV,	https://scikit-
Š .		criterion,	squared error,	learn.org/stable/modules
		splitter,	random,	/generated/sklearn.tree.
		max depth,	1	ExtraTreeRegressor.htm
		min samples split,	None,	I
		min samples leaf,	2,	_
		min_weight_fraction_leaf,	1,	
		max_features,	0.0,	
		random_state,	1.0,	
		min_impurity_decrease,	None,	
		max_leaf_nodes,	0.0,	
		ccp_alpha,	None,	
		cv_in*	0.0,	
			5	
			*	
ExtraTreesRegressor	None	Regression method,	ExtraTreesRegressor,	https://scikit-
		n_estimators,	142,	learn.org/stable/module
		criterion,	squared_error,	s/generated/sklearn.en
		max_depth,	None,	semble.ExtraTreesReg
		min_samples_split,	2,	ressor.html
		min_samples_leaf,	1,	
		min_weight_fraction_leaf,	0.0,	
		max_features,	1.0,	
		max_leaf_nodes,	None,	
		min_impurity_decrease,	0.0,	
		bootstrap,	False,	
		oob_score,	False,	
		n_jobs,	-1,	
		random_state,	1123581321,	
		verbose,	0,	
		warm_start,	False,	
		ccp_alpha,	0.0,	
		max_samples	None	

ExtraTreesRegressorCV	Kfold class	Regression method,	ExtraTreesRegressorCV,	https://scikit-
		n_estimators,	142,	learn.org/stable/modules
		criterion,	squared_error,	/generated/sklearn.ense
		max_depth,	None,	mble.ExtraTreesRegres
		min_samples_split,	2,	sor.html
		min_samples_leaf,	1,	
		<pre>min_weight_fraction_leaf,</pre>	0.0,	
		max_features,	1.0,	
		max_leaf_nodes,	None,	
		min_impurity_decrease,	0.0,	
		bootstrap,	False,	
		oob_score,	False,	
		n_jobs,	-1,	
		random_state,	1123581321,	
		verbose,	0,	
		warm_start,	False,	
		ccp_alpha,	0.0,	
		max_samples,	None,	
		cv_in*	5	

GaussianProcessRegressor	None	Regression method,	GaussianProcessRegressor,	https://scikit-
_		kernel,	None,	learn.org/stable/modules
		alpha,	1e-10,	/generated/sklearn.gaus
		optimizer,	fmin_l_bfgs_b,	sian process.Gaussian
		n restarts optimizer,	0,	ProcessRegressor.html
		normalize_y,	False,	
		copy X train,	True,	
		random_state	None	
GaussianProcessRegressorCV	Kfold class	Regression method,	GaussianProcessRegressorCV,	https://scikit-
-		kernel,	None,	learn.org/stable/module
		alpha,	1e-10,	s/generated/sklearn.ga
		optimizer,	fmin l bfgs b,	ussian process.Gaussi
		n restarts optimizer,	0,	anProcessRegressor.ht
		normalize y,	False,	ml
		copy X train,	True,	_
		random_state,	None,	
		cv_in*	5	
GradientBoostingRegressor	None	Regression method,	GradientBoostingRegressor,	https://scikit-
		loss,	squared_error,	learn.org/stable/modules
		learning_rate,	0.1,	/generated/sklearn.ense
		n_estimators,	100,	mble.GradientBoostingR
		subsample,	1.0,	egressor.html
		criterion,	friedman_mse,	
		min_samples_split,	2,	
		min_samples_leaf,	1,	
		<pre>min_weight_fraction_leaf,</pre>	0.0,	
		max_depth,	3,	
		min_impurity_decrease,	0.0,	
		init,	None,	
		random_state,	None,	
		max_features,	None,	
		alpha,	0.9,	
		verbose,	0,	
		max_leaf_nodes,	None,	
		warm_start,	False,	
		validation_fraction,	0.1,	
		n_iter_no_change,	None,	
		tol,	1e-4,	
		ccp_alpha	0.0	

GradientBoostingRegressorCV	Kfold class	Regression method,	GradientBoostingRegressorCV,	https://scikit-
		loss,	squared error,	learn.org/stable/modules
		learning rate,	0.1,	/generated/sklearn.ense
		n estimators,	100,	mble.GradientBoostingR
		subsample,	1.0,	egressor.html
		criterion,	friedman mse,	
		min_samples_split,	2,	
		min_samples_leaf,	1,	
		min_weight_fraction_leaf,	0.0,	
		max_depth,	3,	
		<pre>min_impurity_decrease,</pre>	0.0,	
		init,	None,	
		random_state,	None,	
		max_features,	None,	
		alpha,	0.9,	
		verbose,	0,	
		max_leaf_nodes,	None,	
		warm_start,	False,	
		validation_fraction,	0.1,	
		n_iter_no_change,	None,	
		tol,	1e-4,	
		ccp_alpha,	0.0,	
		cv_in*	5	

HuberRegressor	None	Regression method,	HuberRegressor,	https://scikit-
		epsilon,	1.35,	learn.org/stable/modules
		max_iter,	1000,	/generated/sklearn.linea
		alpha,	0.0001,	_model.HuberRegressor
		warm_start,	False,	<u>.html</u>
		fit_intercept,	True,	
		tol,	1e-5,	
		rand in ⁺	1123581321	
HuberRegressorCV	Kfold class	Regression method,	HuberRegressorCV,	https://scikit-
		epsilon,	1.35,	learn.org/stable/modules
		max_iter,	1000,	/generated/sklearn.linear
		alpha,	0.0001,	model.HuberRegressor
		warm_start,	False,	<u>.html</u>
		fit_intercept,	True,	
		tol,	1e-5,	
		rand in ⁺ ,	1123581321,	
		cv in*	5	
KernelRidge	None	Regression method,	KernelRidge,	https://scikit-
-		alpha,	1.0,	learn.org/stable/modules
		kernel,	linear,	/generated/sklearn.kern
		gamma,	None,	el_ridge.KernelRidge.ht
		degree,	3.0,	ml
		coef0,	1.0,	
		kernel_params	None	
KernelRidgeCV	Kfold class	Regression method,	KernelRidgeCV,	https://scikit-
rtementageov	Triola diass	alpha,	1.0,	learn.org/stable/modules
		kernel,	linear,	/generated/sklearn.kern
		gamma,	None,	el ridge.KernelRidge.ht
		degree,	3.0,	ml
		coef0,	1.0,	1111
		kernel params,	None,	
		cv in*	5	
KneighborsRegressor	None	Regression method,	KNeighborsRegressor,	https://scikit-
g	1.10.1.0	n neighbors,	5,	learn.org/stable/module
		weights,	uniform,	s/generated/sklearn.nei
		algorithm,	auto,	ghbors.KNeighborsReg
		leaf size,	30,	ressor.html
		p,	2,	100001.IIIIII
		metric,	minkowski,	
		metric params,	None,	
		meeric_parame,	110110 /	

	n jobs	-1	

KneighborsRegressorCV	Kfold class	Regression method,	KNeighborsRegressorCV,	https://scikit-
		n neighbors,	5,	learn.org/stable/modules
		weights,	uniform,	/generated/sklearn.neigh
		algorithm,	auto,	bors.KNeighborsRegres
		leaf size,	30,	sor.html
		p,	2,	
		metric,	minkowski,	
		metric_params,	None,	
		n jobs,	-1,	
		cv in*	5	
Lasso	None	Regression method,	Lasso,	https://scikit-
		alpha,	0.65,	learn.org/stable/modules
		fit_intercept,	True,	/generated/sklearn.linear
		precompute,	False,	model.Lasso.html
		copy_X,	True,	
		max_iter,	1000,	
		tol,	1e-4,	
		warm_start,	False,	
		positive,	False,	
		random_state,	1123581321,	
		selection	cyclic	
LassoCV	Kfold class	Regression method,	LassoCV,	https://scikit-
		alpha,	0.65,	learn.org/stable/modules
		fit_intercept,	True,	/generated/sklearn.linear
		precompute,	False,	model.Lasso.html
		copy_X,	True,	
		max_iter,	1000,	
		tol,	1e-4,	
		warm_start,	False,	
		positive,	False,	
		random_state,	1123581321,	
		selection,	cyclic,	
		cv_in*	5	

LinearRegression	None	Regression method,	LinearRegression,	https://scikit-
	1.155	fit intercept,	True,	learn.org/stable/modules
		copy X,	True,	/generated/sklearn.linear
		n jobs,	-1,	model.LinearRegressio
		positive,	False,	n.html
		rand in ⁺	1123581321	
LinearRegressionCV	Kfold class	Regression method,	LinearRegressionCV,	https://scikit-
		fit intercept,	True,	learn.org/stable/module
		copy_X,	True,	s/generated/sklearn.lin
		n_jobs,	-1,	ear_model.LinearRegre
		positive,	False,	ssion.html
		_	1123581321,	
		rand_in ⁺ ,	· ·	
		cv_in*	5	
LinearSVR	None	Regression,	LinearSVR,	https://scikit-
		epsilon,	1e-2,	learn.org/stable/modules
		tol,	1e-8,	/generated/sklearn.svm.
		С,	1.0,	<u>LinearSVR.html</u>
		loss,	epsilon_insensitive,	
		fit_intercept,	True,	
		intercept_scaling,	1.0,	
		dual,	True,	
		verbose,	0,	
		random_state,	1123581321,	
		max iter	1000	

LinearSVRCV	Kfold class	Regression,	LinearSVRCV,	https://scikit-
		epsilon,	1e-2,	learn.org/stable/modules
		tol,	1e-8,	/generated/sklearn.svm.
		C,	1.0,	LinearSVR.html
		loss,	epsilon insensitive,	
		fit intercept,	True,	
		intercept scaling,	1.0,	
		dual,	True,	
		verbose,	0,	
		random state,	1123581321,	
		max iter,	1000,	
		cv_in*	5	
MLPRegressor	None	Regression,	MLPRegressor,	https://scikit-
_		hidden_layer_sizes,	75,	learn.org/stable/modules
		activation,	logistic,	/generated/sklearn.neur
		solver,	lbfgs,	al network.MLPRegress
		alpha,	0.01,	or.html
		batch_size,	auto,	
		learning_rate,	adaptive,	
		learning_rate_init,	0.001,	
		power_t,	0.5,	
		max_iter,	200,	
		shuffle,	True,	
		random_state,	1123581321,	
		tol,	5e−3,	
		verbose,	False,	
		warm_start,	False,	
		momentum,	0.9,	
		nesterovs_momentum,	True,	
		early_stopping,	False,	
		validation_fraction,	0.1,	
		beta_1,	0.9,	
		beta_2,	0.999,	
		epsilon,	1e-8,	
		n_iter_no_change,	10,	
		max_fun	15000	

MLPRegressorCV	Kfold class	Regression,	MLPRegressorCV,	https://scikit-
		hidden layer sizes,	75,	learn.org/stable/modules
		activation,	logistic,	/generated/sklearn.neur
		solver,	lbfgs,	al network.MLPRegress
		alpha,	0.01,	or.html
		batch_size,	auto,	
		learning_rate,	adaptive,	
		learning_rate_init,	0.001,	
		power_t,	0.5,	
		max_iter,	200,	
		shuffle,	True,	
		random_state,	1123581321,	
		tol,	5e-3,	
		verbose,	False,	
		warm_start,	False,	
		momentum,	0.9,	
		nesterovs_momentum,	True,	
		early_stopping,	False,	
		validation_fraction,	0.1,	
		beta_1,	0.9,	
		beta_2,	0.999,	
		epsilon,	1e-8,	
		n_iter_no_change,	10,	
		max_fun,	15000,	
		cv_in*	5	

NuSVR	None	Regression method,	NuSVR,	https://scikit-
		nu,	0.5,	learn.org/stable/mod
		C,	1.0,	ules/generated/sklea
		kernel,	linear,	rn.svm.NuSVR.html
		degree,	1,	
		gamma,	auto,	
		coef0,	0.0,	
		shrinking,	True,	
		tol,	0.001,	
		cache_size,	200.0,	
		verbose,	False,	
		max_iter,	-1,	
		rand_in+	1123581321	
NuSVRCV	Kfold class	Regression method,	NuSVRCV,	https://scikit-
		nu,	0.5,	learn.org/stable/mod
		С,	1.0,	ules/generated/sklea
		kernel,	linear,	rn.svm.NuSVR.html
		degree,	1,	
		gamma,	auto,	
		coef0,	0.0,	
		shrinking,	True,	
		tol,	0.001,	
		cache_size,	200.0,	
		verbose,	False,	
		max_iter,	-1,	
		rand_in+,	1123581321,	
		cv_in*	5	
PassiveAggressiveRegressor	None	Regression method,	PassiveAggressiveRegressor,	https://scikit-
		С,	1.0,	learn.org/stable/mod
		fit_intercept,	True,	ules/generated/sklea
		max_iter,	1000,	rn.linear_model.Pass
		tol,	1e-3,	iveAggressiveRegres
		early_stopping,	False,	sor.html
		validation_fraction,	0.1,	
		n_iter_no_change,	5,	
		shuffle,	True,	
		verbose,	0,	
		loss,	epsilon_insensitive,	
		epsilon,	1e-4,	
		random_state,	1123581321,	
		warm_start,	True,	

		average	True	
PassiveAggressiveRegressorCV	Kfold class	Regression method,	PassiveAggressiveRegressorCV,	https://scikit-
		С,	1.0,	learn.org/stable/mod
		fit_intercept,	True,	ules/generated/sklea
		max_iter,	1000,	rn.linear model.Pass
		tol,	1e-3,	iveAggressiveRegres
		early_stopping,	False,	sor.html
		validation_fraction,	0.1,	
		n_iter_no_change,	5,	
		shuffle,	True,	
		verbose,	0,	
		loss,	epsilon_insensitive,	
		epsilon,	1e-4,	
		random_state,	1123581321,	
		warm_start,	True,	
		average,	True,	
		cv_in*	5	

RandomForestRegressor	None	Regression method,	RandomForestRegressor,	https://scikit-
		n_estimators,	142,	learn.org/stable/modules
		criterion,	squared_error,	/generated/sklearn.ense
		max_depth,	None,	mble.RandomForestReg
		min_samples_split,	2,	ressor.html
		min_samples_leaf,	1,	
		<pre>min_weight_fraction_leaf,</pre>	0.0,	
		max_features,	1.0,	
		max_leaf_nodes,	None,	
		<pre>min_impurity_decrease,</pre>	0.0,	
		bootstrap,	True,	
		oob_score,	False,	
		n_jobs,	-1,	
		random_state,	1123581321,	
		verbose,	0,	
		warm_start,	False,	
		ccp_alpha,	0.0,	
		max_samples	None	
RandomForestRegressorCV	Kfold class	Regression method,	RandomForestRegressorCV,	https://scikit-
		n_estimators,	142,	learn.org/stable/modules
		criterion,	squared_error,	/generated/sklearn.ense
		max_depth,	None,	mble.RandomForestReg
		min_samples_split,	2,	ressor.html
		min_samples_leaf,	1,	
		<pre>min_weight_fraction_leaf,</pre>	0.0,	
		max_features,	auto,	
		max_leaf_nodes,	None,	
		min_impurity_decrease,	0.0,	
		bootstrap,	True,	
		oob_score,	False,	
		n_jobs,	-1,	
		random_state,	1123581321,	
		verbose,	0,	
		warm_start,	False,	
		ccp_alpha,	0.0,	
		max_samples,	None,	
BANGAGE		cv in*	5	
RANSACRegressor	None	Regression method,	RANSACRegressor,	https://scikit-
		base_estimator,	None,	learn.org/stable/modules
		min_samples,	None,	/generated/sklearn.linear
		residual_threshold,	None,	_model.RANSACRegres

is data valid,	None,	sor.html
is model valid,	None,	
max trials,	100,	
max skips,	np.inf,	
stop n inliers,	np.inf,	
stop score,	np.inf,	
stop_probability,	0.99,	
loss,	absolute_error,	
random state	1123581321	

RANSACRegressorCV	Kfold class	Regression method,	RANSACRegressorCV,	https://scikit-
		base estimator,	None,	learn.org/stable/module
		min samples,	None,	s/generated/sklearn.lin
		residual threshold,	None,	ear model.RANSACRe
		is data valid,	None,	gressor.html
		is model valid,	None,	
		max trials,	100,	
		max skips,	np.inf,	
		stop n inliers,	np.inf,	
		stop score,	np.inf,	
		stop probability,	0.99,	
		loss,	absolute_error,	
		random_state,	1123581321,	
		cv_in*	5	
Ridge	None	Regression method,	Ridge,	https://scikit-
		alpha,	1.0,	learn.org/1.5/modules/g
		fit_intercept,	True,	enerated/sklearn.linear
		copy_X,	True,	model.Ridge.html
		max_iter,	None,	
		tol,	1e-3,	
		solver,	auto,	
		positive,	False,	
		random_state	None	
RidgeCV	Kfold class	Regression method,	RidgeCV,	https://scikit-
		alpha,	1.0,	learn.org/1.5/modules/g
		fit_intercept,	True,	enerated/sklearn.linear_
		copy_X,	True,	model.Ridge.html
		max_iter,	None,	
		tol,	1e-3,	
		solver,	auto,	
		positive,	False,	
		random_state,	None,	
		cv_in*	5	

SGDRegressor	None	Regression method,	SGDRegressor,	https://scikit-
		loss,	squared_error,	learn.org/stable/module
		penalty,	12,	s/generated/sklearn.lin
		alpha,	0.001,	ear model.SGDRegres
		11 ratio,	0.15,	sor.html
		fit_intercept,	True,	
		max_iter,	2000000000,	
		tol,	1e-3,	
		shuffle,	True,	
		verbose,	0,	
		epsilon,	0.1,	
		random_state,	1123581321,	
		learning_rate,	invscaling,	
		eta0,	0.01,	
		power_t,	0.25,	
		early_stopping,	False,	
		validation_fraction,	0.1,	
		n_iter_no_change,	5,	
		warm_start,	False,	
		average	False	

SGDRegressorCV	Kfold class	Regression method,	SGDRegressorCV,	https://scikit-
		loss,	squared_error,	learn.org/stable/mo
		penalty,	12,	dules/generated/skl
		alpha,	0.001,	earn.linear model.
		11_ratio,	0.15,	SGDRegressor.htm
		fit_intercept,	True,	1
		max_iter,	2000000000,	
		tol,	1e-3,	
		shuffle,	True,	
		verbose,	0,	
		epsilon,	0.1,	
		random_state,	1123581321,	
		learning_rate,	invscaling,	
		eta0,	0.01,	
		power_t,	0.25,	
		early_stopping,	False,	
		validation_fraction,	0.1,	
		n_iter_no_change,	5,	
		warm_start,	False,	
		average,	False,	
		cv_in*	5	
SVR	None	Regression method,	SVR,	https://scikit-
		kernel,	linear,	learn.org/stable/mod
		degree,	1,	ules/generated/sklea
		gamma,	scale,	rn.svm.SVR.html
		coef0,	0.0,	
		tol,	1e-3,	
		C,	1.0,	
		epsilon,	0.1,	
		shrinking,	True,	
		cache_size,	200.0,	
		verbose,	False,	
		max_iter,	-1,	
2) (7.2) (rand_in+	1123581321	
SVRCV	Kfold class	Regression method,	SVRCV,	https://scikit-
		kernel,	linear,	learn.org/stable/mod
		degree,	1,	ules/generated/sklea
		gamma,	scale,	rn.svm.SVR.html
		coef0,	0.0,	
		tol,	1e-3,	
		С,	1.0,	

	epsilon,	0.1,	
	shrinking,	True,	
	cache_size,	200.0,	
	verbose,	False,	
	max_iter,	-1,	
	rand in ⁺ ,	1123581321,	
	cv_in*	5	
TheilSenRegressor None	Regression method,	TheilSenRegressor,	https://scikit-
	Regression method,	True,	learn.org/stable/mod
	fit intercept,	True,	ules/generated/sklea
	copy X,	10000,	rn.linear model.Theil
	max subpopulation,	None,	SenRegressor.html
	n subsamples,	300,	
	max iter,tol,	1e-3,	
	random state,	1123581321,	
	n jobs,	-1,	
	verbose	False	
TheilSenRegressorCV Kfold cla	Regression method,	TheilSenRegressorCV,	https://scikit-
	fit intercept,	True,	learn.org/stable/mod
	copy X,	True,	ules/generated/sklea
	max subpopulation,	10000,	rn.linear model.Theil
	n subsamples,	None,	SenRegressor.html
	max_iter,	300,	
	tol,	1e-3,	
	random_state,	1123581321,	
	n_jobs,	-1,	
	verbose,	False,	
	cv_in*		

TweedieRegressor	None	Regression method,	TweedieRegressor,	https://scikit-
ı		power,	0.0,	learn.org/stable/modules
		alpha,	1.0,	/generated/sklearn.linear
		fit_intercept,	True,	model.TweedieRegres
ı		link,	auto,	sor.html
ı		max iter,	100,	
ı		tol,	1e-4,	
ı		warm start,	False,	
ı		verbose,	0,	
		rand in ⁺	1123581321	
TweedieRegressorCV	Kfold class	Regression method,	TweedieRegressorCV,	https://scikit-
ı		power,	0.0,	learn.org/stable/modul
		alpha,	1.0,	es/generated/sklearn.li
		fit intercept,	True,	near model.TweedieR
		link,	auto,	egressor.html
ı		max iter,	100,	
ı		tol,	1e-4,	
ı		warm start,	False,	
ı		verbose,	0,	
ı		rand in ⁺ ,	1123581321,	
I		cv in*	5	
VotingRegressor	None	Regression method,	VotingRegressor,	https://scikit-
1		fit intercept,	True,	learn.org/stable/modules
I		copy X,	True,	/generated/sklearn.ense
ı		n jobs,	-1,	mble.VotingRegressor.ht
ı		n estimators,	142,	ml
I		criterion,	squared error,	_
ı		max depth,	None,	
ı		min samples split,	2,	
ı		min samples leaf,	1,	
ı		min weight fraction leaf,	0.0,	
ı		max features,	auto,	
ı		max leaf nodes,	None,	
ı		min_impurity_decrease,	0.0,	
ı		bootstrap,	True,	
ı		oob score,	False,	
ı		n jobs,	None,	
ı		random state,	1123581321,	
ı		verbose,	0,	
ı		warm start,	False,	
	i i	ccp_alpha,	/	1

		<pre>max_samples, weights, n_jobs, verbose</pre>	None, None, None, False	
VotingRegressorCV	Kfold class	Regression method, fit_intercept, copy_X, n_jobs, n_estimators, criterion, max_depth, min_samples_split, min_samples_leaf, min_weight_fraction_leaf, max_features, max_leaf_nodes, min_impurity_decrease, bootstrap, oob_score, n_jobs, random_state, verbose, warm_start, ccp_alpha, max_samples, weights, n_jobs, verbose, cv_in*	VotingRegressor, True, True, -1, 142, squared_error, None, 2, 1, 0.0, auto, None, 0.0, True, False, None, 1123581321, 0, False, 0.0, None, None, None, None, None, Selse, None, None, False, None, Selse, None, False, Selse, Sels	https://scikit- learn.org/stable/modules /generated/sklearn.ense mble.VotingRegressor.ht ml

 *cv in variable holds an integer for the number of subsets used in the cross-validation process.

*rand in holds a dummy integer seed to be used to generate a Molegro Data Modeller (MDM) format file.

#eps 0, eps 1, and n samples define an array (eps) as follows:

eps_array = np.linspace(eps_0, eps_1, num=n_samples_in)

This is applied for machine-precision regularization in the computation of the Cholesky diagonal factors.