

Gas Chromatography- Principles and Instrumentation

CEB 4032/CFB3032: ANALYTICAL
CHEMISTRY/ANALYTICAL INSTRUMENTATION

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Chemical
Engineering

Inspiring Potential • Generating Futures

Introduction

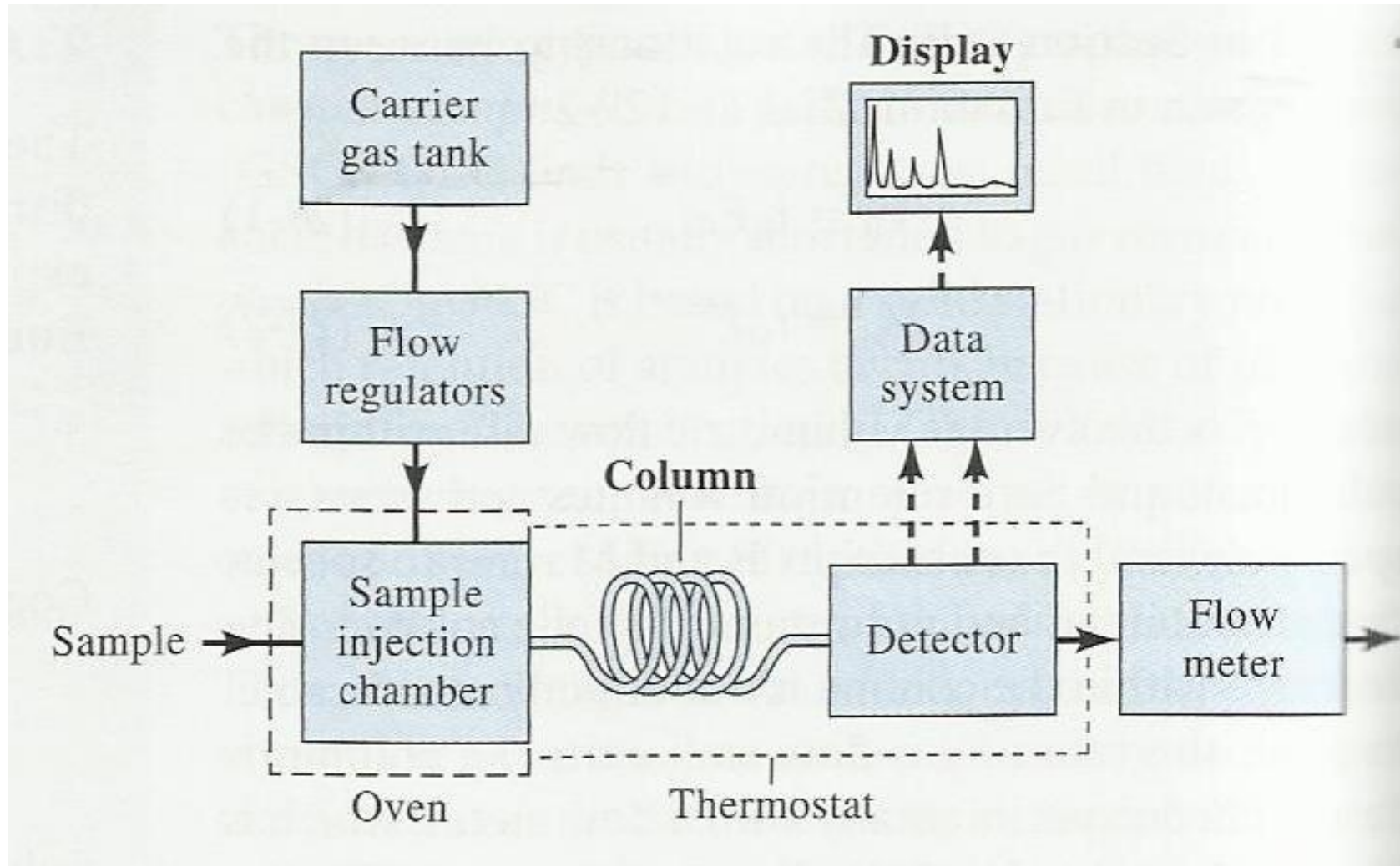
- GC is widely used for the determination of organic/volatile compounds.
- For example: the separation of benzene (bp. 80.1°C) and cyclohexane (bp. 80.8°C) is extremely simple by gas chromatography, but it is virtually impossible by conventional distillation.
- Very complex mixture can be separated by this technique.

- Two types of gas chromatography (GC):
 - i. Gas-solid (adsorption) chromatography (GSC)
 - ii. Gas-liquid (partitioning) chromatography (GLC)
- In **GSC**, analyte is adsorbed directly on solid particles of stationary phase.
- In **GLC**, the stationary phase is a **nonvolatile liquid** coated on the **inside of the column** or on a fine solid support.

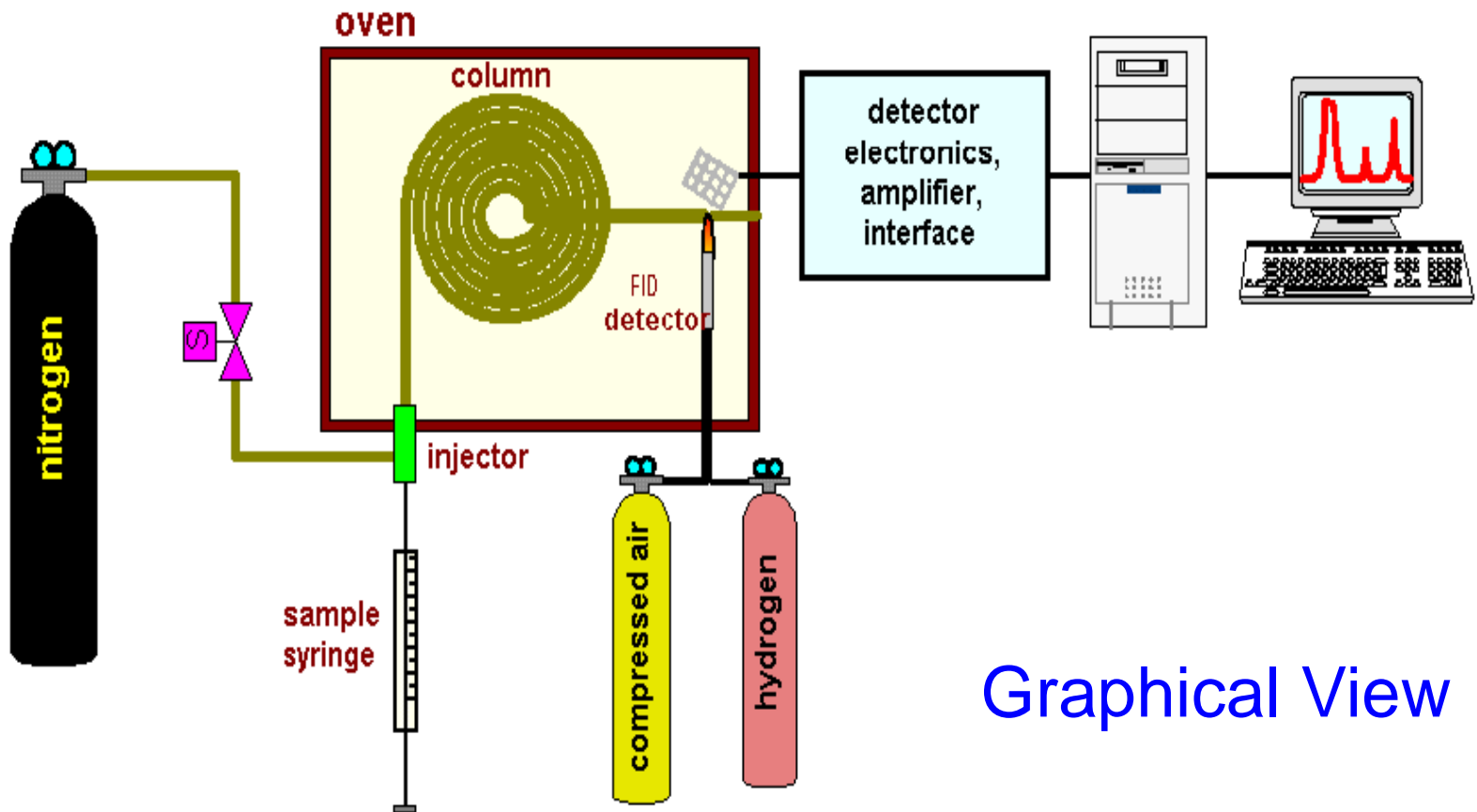
The basic components of a typical instrument for performing GC are:

- i. Carrier gas system
- ii. Sample injection system
- iii. Column
- iv. Column oven
- v. Detection system

Instrumentation of GC



Block diagram of a typical Gas Chromatography



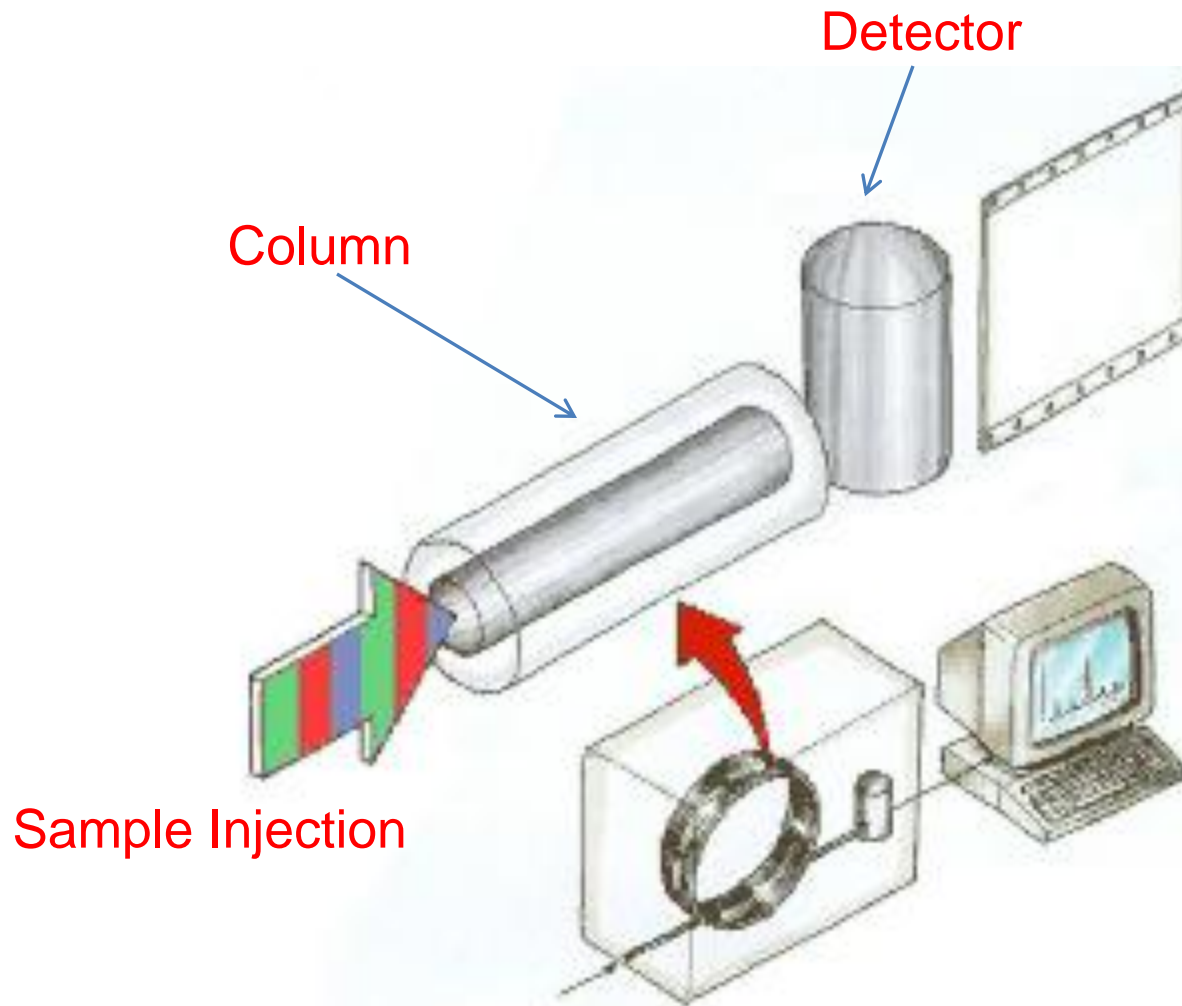
Graphical View

Principles Operation of GC

- Volatile liquid or gaseous sample is **injected** through a septum into a heated port, in which it rapidly evaporated.
- Vapor is **swept through the column (stationary phase) by carrier gas** (mobile phase → usually He, N₂ or H₂)
- The **column must be hot enough** to provide sufficient temperature for analytes to be eluted in a reasonable time.
- **Separation** occurs as the **vapor constituents equilibrate** between carrier gas and the stationary phase.

- The sample is automatically **detected** as it emerges from the column → using a variety of detectors whose **response is dependent upon the composition of the vapor.**
- The **detector is maintained at a higher temperature** than the column, so that all analytes will be gaseous.
- The **signal is fed to a recording device** where the chromatographic peaks are recorded as a **function of time → chromatogram.**

- By measuring the **retention time** and comparing this time with that of a **standard of the pure substance**, it can **identify the peak**.
- The **area under the peak** is proportional to the **concentration**, and so the **amount of the substance** can be **quantitatively** determined.



GC Operation

Modern Gas Chromatography System



These include software for recording retention time, taking peak areas and calculating concentrations.

Unleaded Gasoline

Column: DB-Petro 100

100 m × 0.25 mm I.D., 0.5 μm

J&W P/N: 122-10A6

Carrier: Helium at 25.6 cm/sec

Oven: 0°C for 15 min

0-50°C at 1°/min

50-130°C at 2°/min

130-180°C at 4°/min

180°C for 20 min

Injector: Split 1:300, 200°C

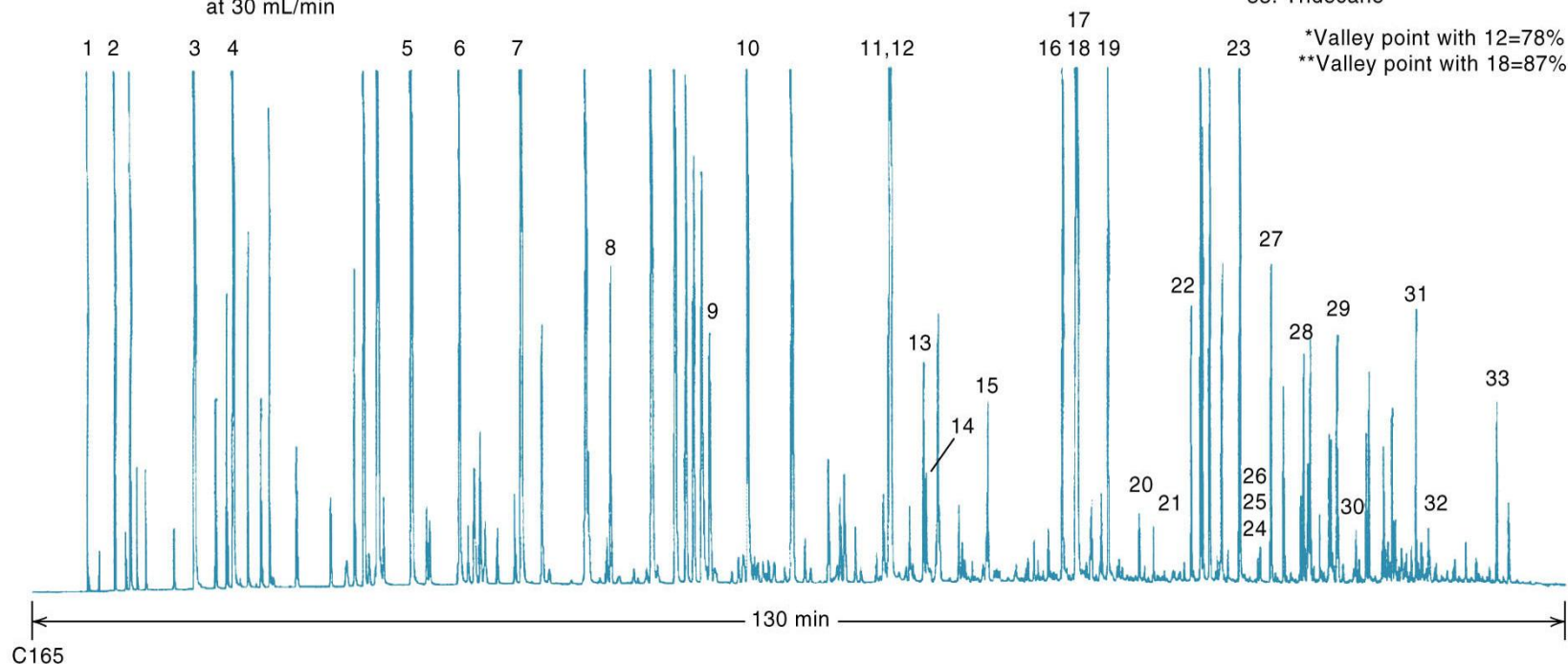
1 μL of neat sample

Detector: FID, 250°C

Nitrogen makeup gas

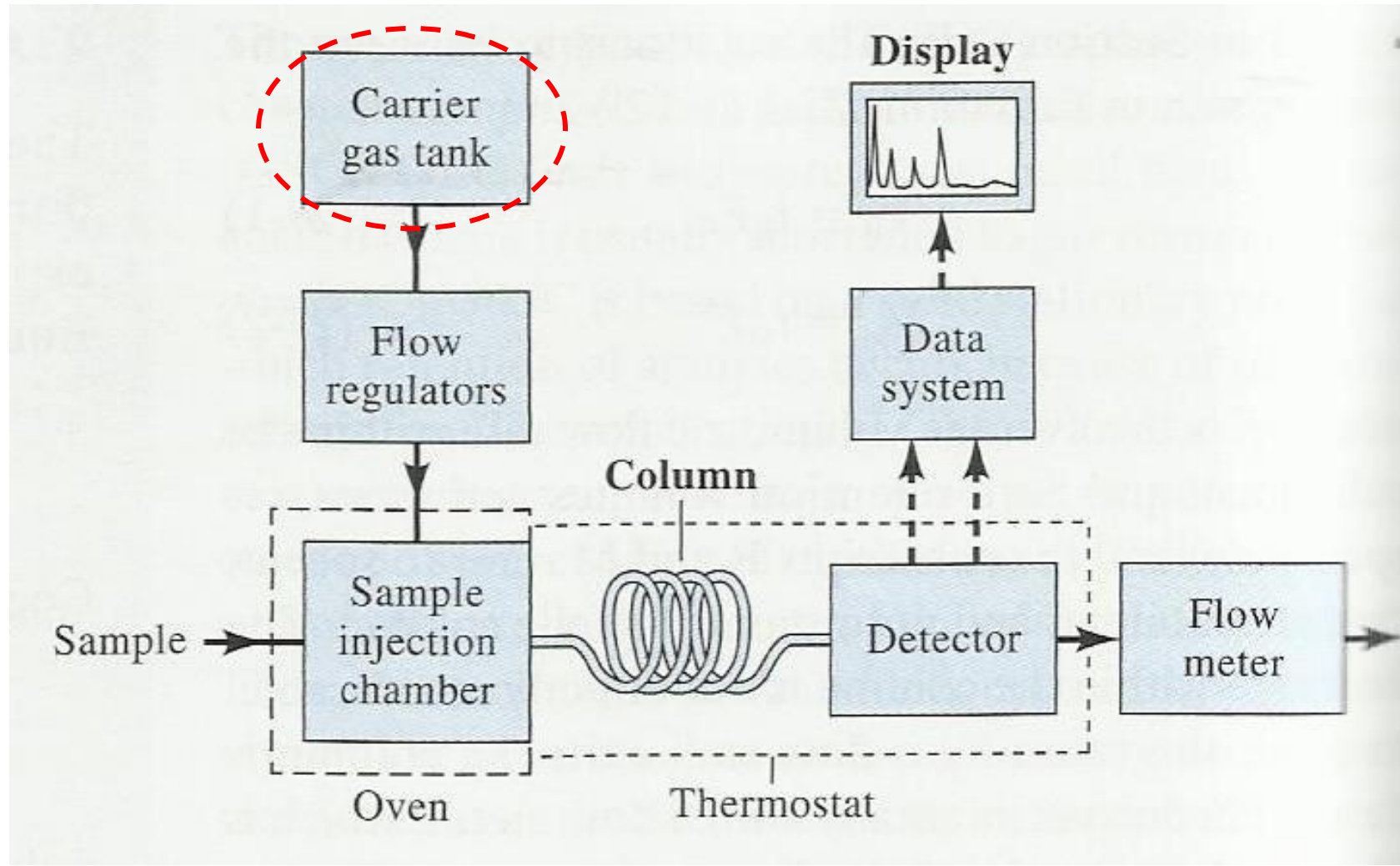
at 30 mL/min

- | | | |
|-----------------------|----------------------------|--------------------------------|
| 1. Methane | *11. Toluene | 21 Isopropylbenzene |
| 2. <i>n</i> -Butane | 12. 2,3,3-Trimethylpentane | 22. Propylbenzene |
| 3. Isopentane | 13. 2-Methylheptane | 23. 1,2,4-Trimethylbenzene |
| 4. <i>n</i> -Pentane | 14. 4-Methylheptane | 24. Isobutylbenzene |
| 5. <i>n</i> -Hexane | 15. <i>n</i> -Octane | 25. <i>sec</i> -Butylbenzene |
| 6. Methylcyclopentane | 16. Ethylbenzene | 26. <i>n</i> -Decane |
| 7. Benzene | **17. <i>m</i> -Xylene | 27. 1,2,3-Trimethylbenzene |
| 8. Cyclohexane | 18. <i>p</i> -Xylene | 28. Butylbenzene |
| 9. Isooctane | 19. <i>o</i> -Xylene | 29. <i>n</i> -Undecane |
| 10. <i>n</i> -Heptane | 20. <i>n</i> -Nonane | 30. 1,2,4,5-Tetramethylbenzene |
| | | 31. Naphthalene |
| | | 32. Dodecane |
| | | 33. Tridecane |



Typical gas chromatogram of complex mixture using a capillary column

Carrier Gas System



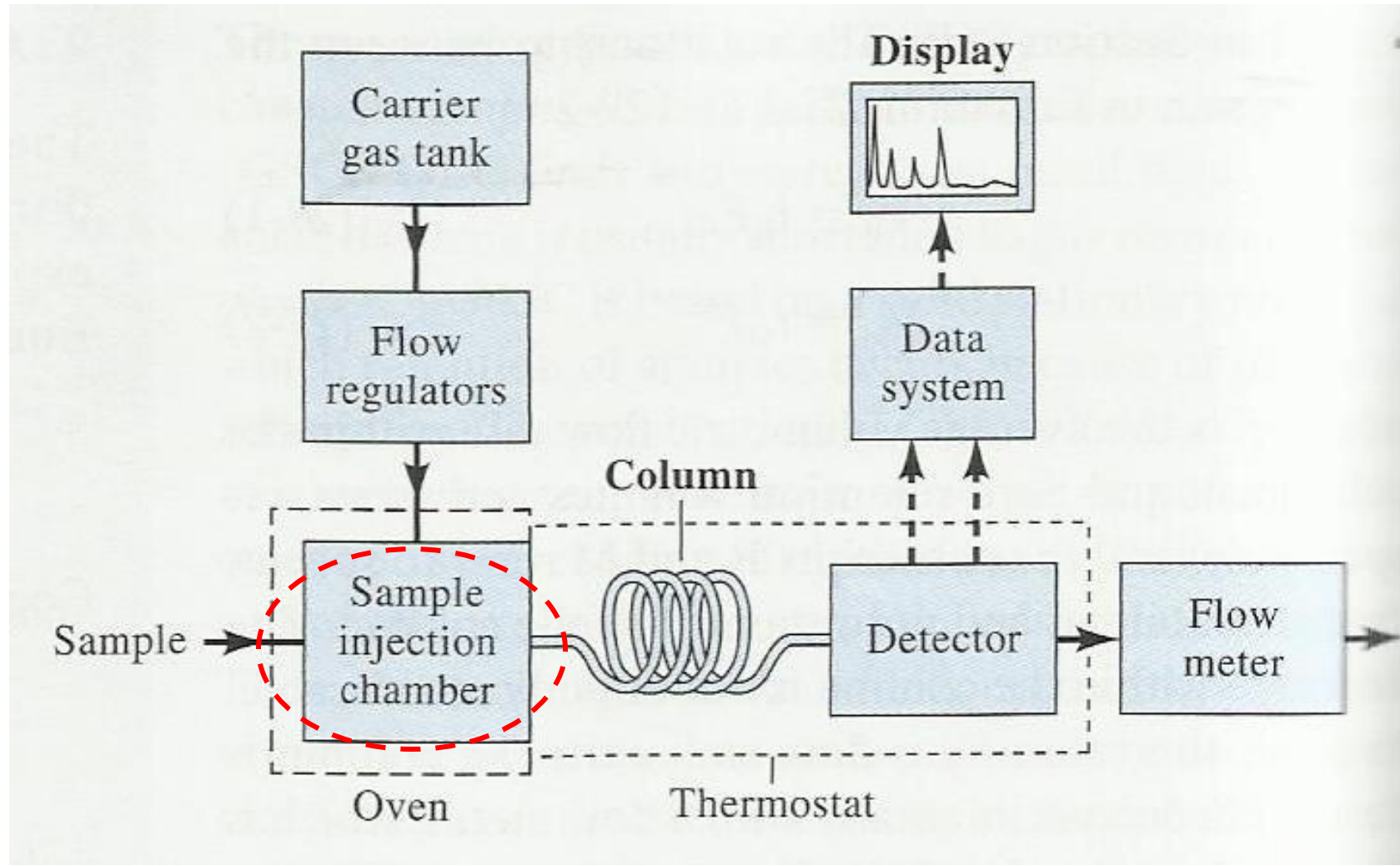
Carrier Gas System

- The **mobile-phase gas in GC** is called carrier gas and must be chemically inert.
- Example gases: helium, argon, nitrogen and hydrogen.
- The **choice of the gas** → dictated by the **type of detector**.
- These gases are available in pressurized tanks.
- Pressure regulators, gauges, and flow meters, are required to control the flow rate of the gas.

Carrier Gas System / Gas Supply



Sample Injection System

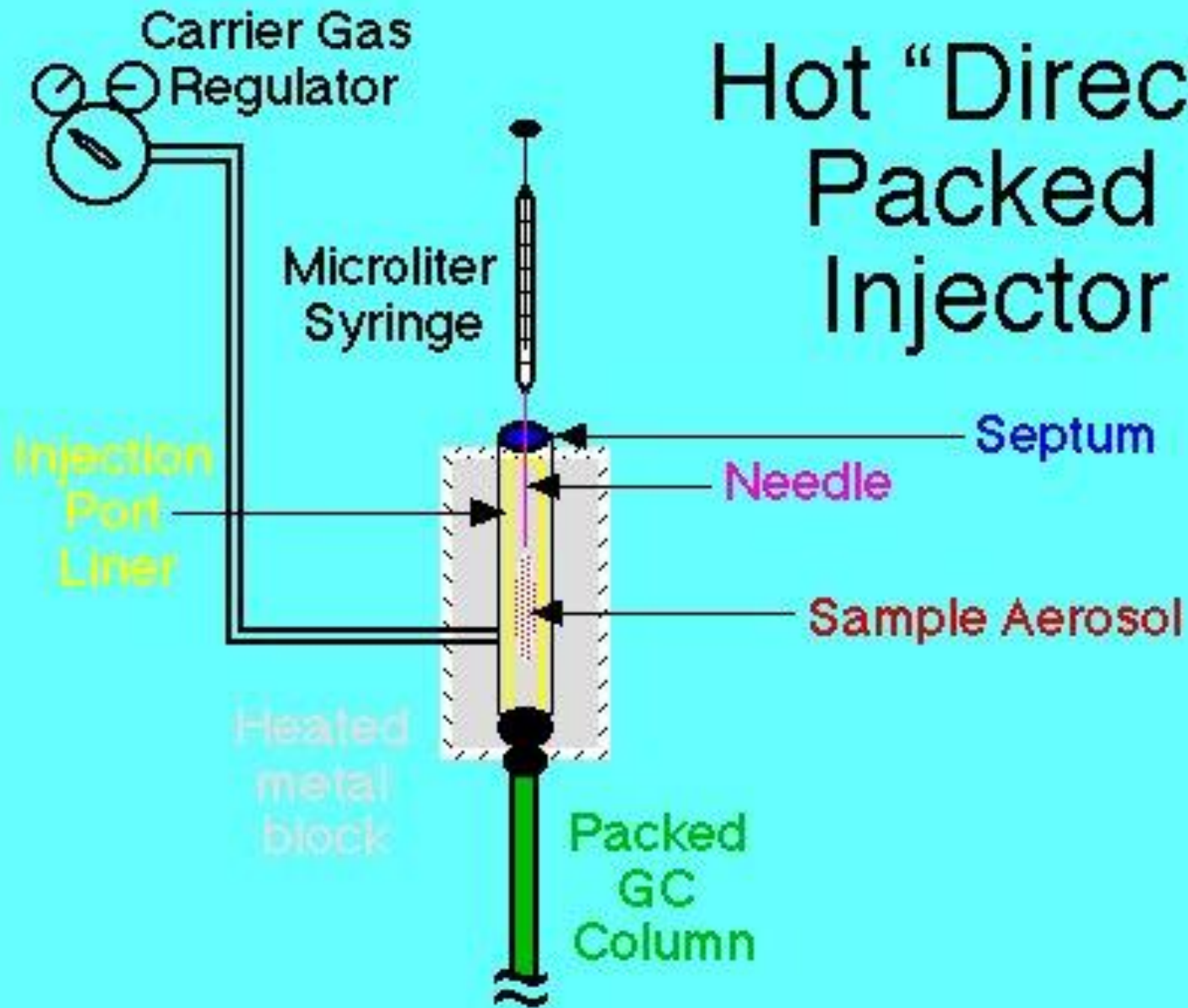


Sample Injection System

- Calibrated **microsyringes** → used to **inject samples** into a heated sample port located at the head of the column.
- For liquid samples, the **injection system** are heated to temperatures about **50 °C** above the boiling point of the **highest boiling solute**.
- The **injection port** are usually kept warmer than the column to promote **rapid vaporization** of the injected sample and **prevent sample condensation** in the detector.

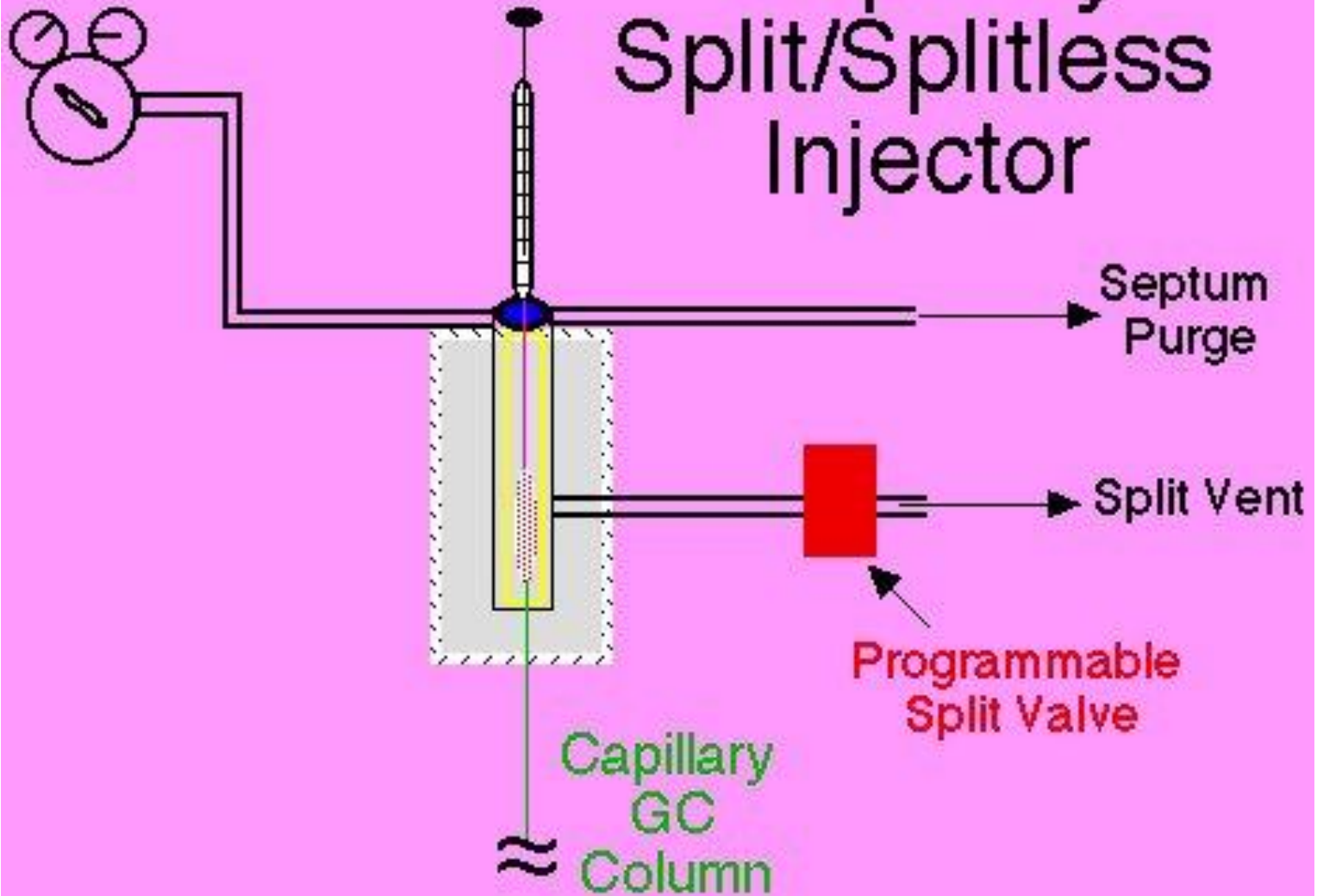
- To achieve high column efficiency, the sample must be of a suitable size.
- Slow injection or oversize samples cause band spreading and poor resolution.
- Sample size → depends on the column and sample.
- For i) packed column:
 - liquid samples of **0.1 to 10 μ l** are injected
 - for gas samples, **1-10 mL** are injected
- Gases may be injected by means of **gas-tight syringe** or through a **special gas inlet chamber** of constant volume (gas sampling valve).

Hot "Direct" Packed Injector



- For ii) capillary column → volumes of only about **1/100** size of packed column must be injected because of the lower capacity of the columns.
- **Sample splitters** are included in the gas chromatography system designed for use with **capillary columns** that deliver a **small fixed fraction** of the sample to the column, with the remainder going to the waste.

Capillary Split/Splitless Injector



Syringes



Injector Port



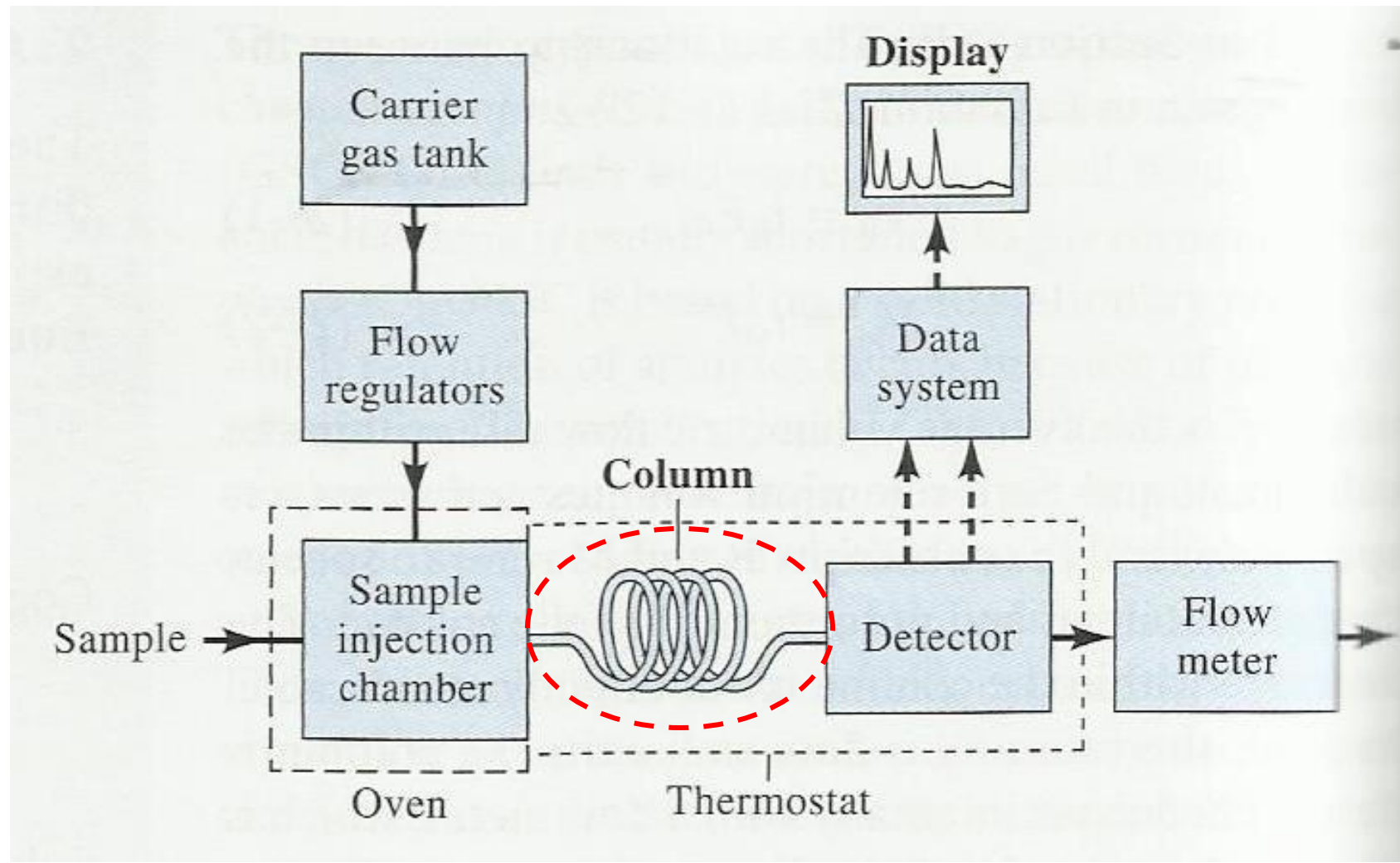
Autosampler



Injection



GC Columns



GC Columns

- Two types of columns used in GC are:
 - i. Packed column
 - ii. Capillary column (open tubular)

- Capillary columns are more commonly used today, but packed columns are still used for applications that do not require high resolution or when increased capacity is needed.

i) Packed Column

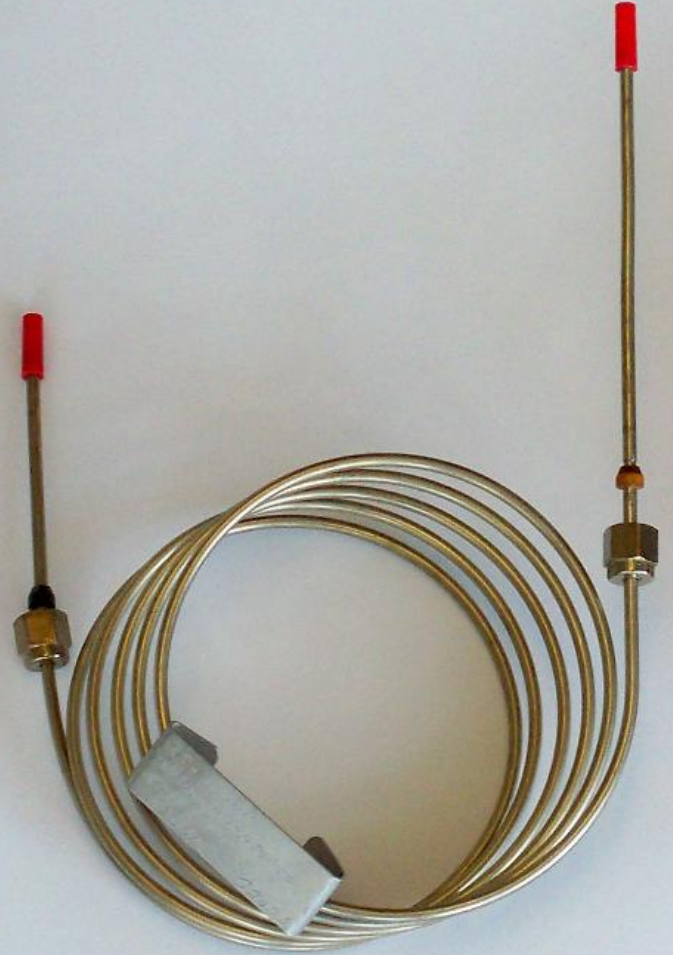
- The column contain a **fine solid support** coated with a **nonvolatile liquid stationary phase** (partitioning chromatography); or the **solid itself may be the stationary phase** (adsorption chromatography).
- useful for the separation of **small gaseous** species such as H_2 , N_2 , CO_2 , CO , O_2 , NH_2 and CH_4 and volatile hydrocarbons.

- Small particle size of fine solid **decreases the time** required for solute equilibration → **improve column efficiency**, but **higher pressure required** to force mobile phase through the column.
- Packed column are usually made of stainless steel, nickel, glass and are typically **3-6 mm in diameter** and **1-5 m in length**.

Glass Packed GC Column

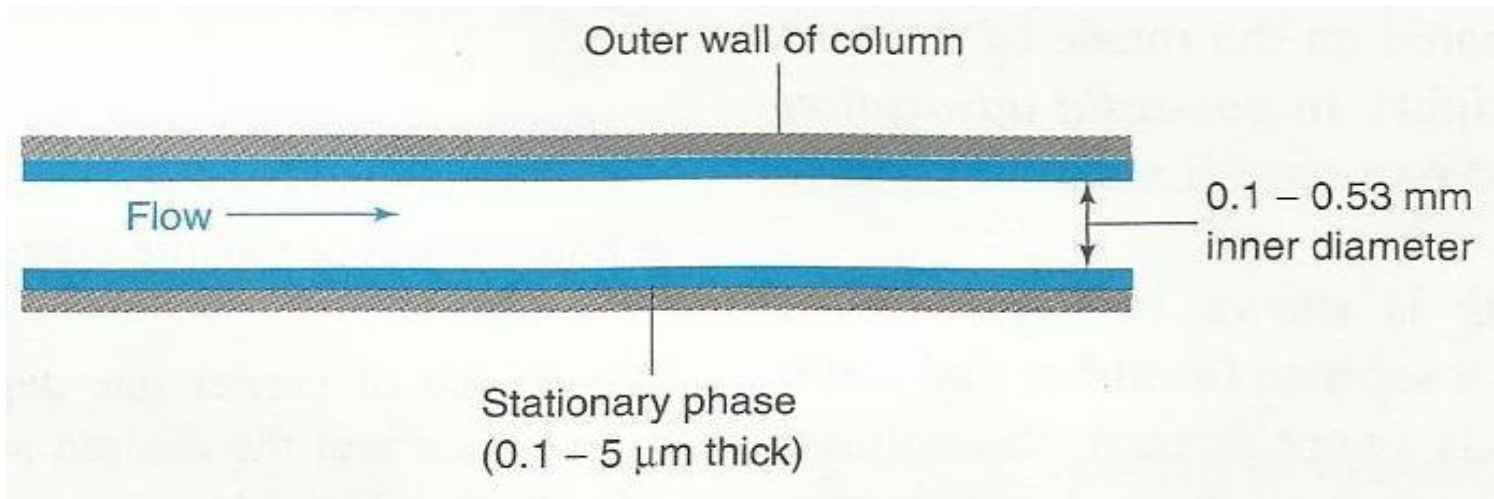


Stainless Steel GC



ii) Capillary Column (Open Tubular)

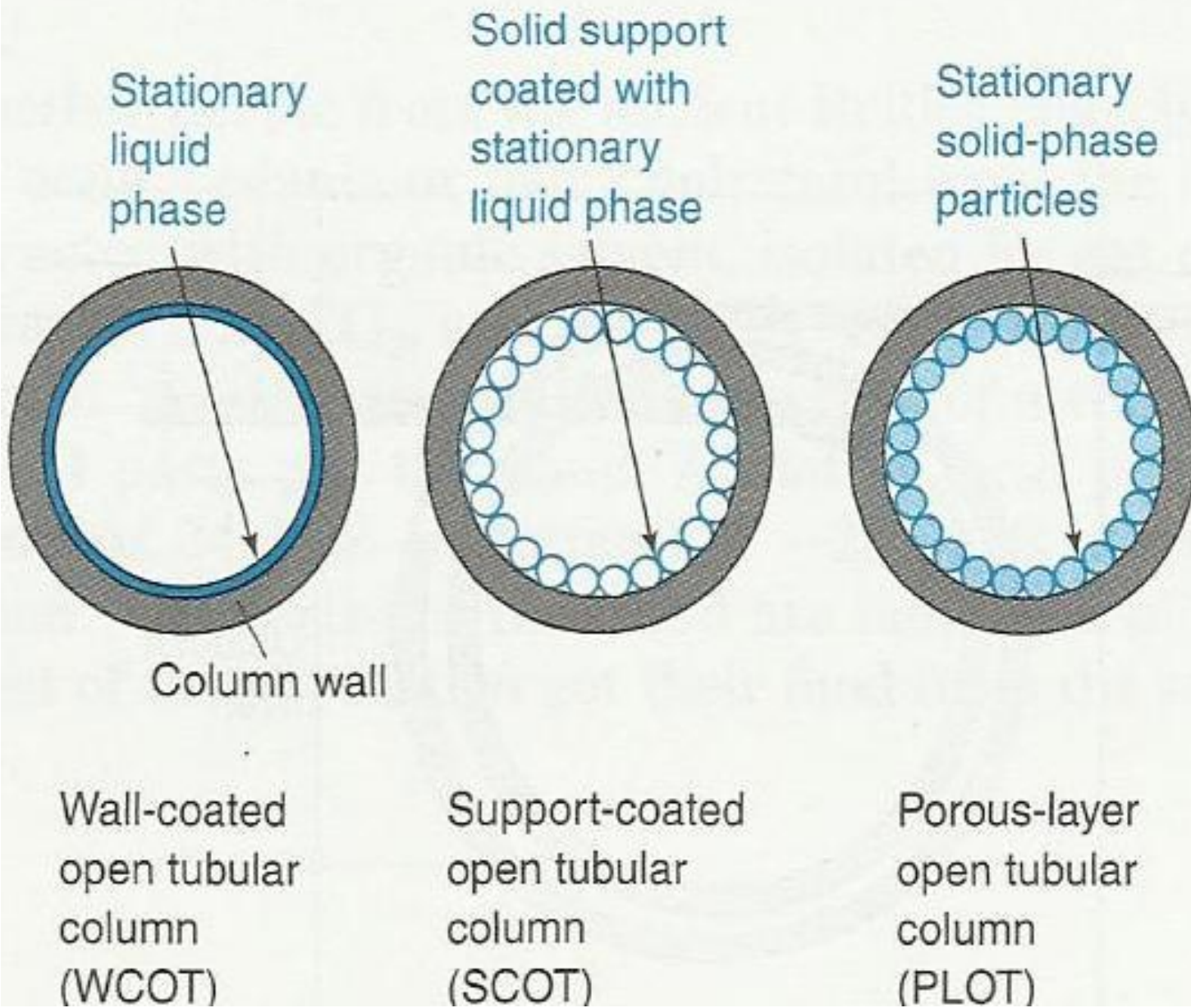
- Most widely use.
- Long, narrow open tubular columns made of fused silica (SiO_2).
- Column inner diameter = 0.10-0.53 mm and typical length = 15-100 m.



- Compared with packed columns, open tubular columns offer:
 - i. Higher resolution
 - ii. Shorter analysis time
 - iii. Greater sensitivity
 - iv. Lower sample capacity

- **Drawback:** require higher pressure to operate.

- Three types of open-tubular columns:
 - i. Wall coated open tubular column (WCOT)
 - ii. Support-coated open tubular column (SCOT)
 - iii. Porous layer tubular column (PLOT)



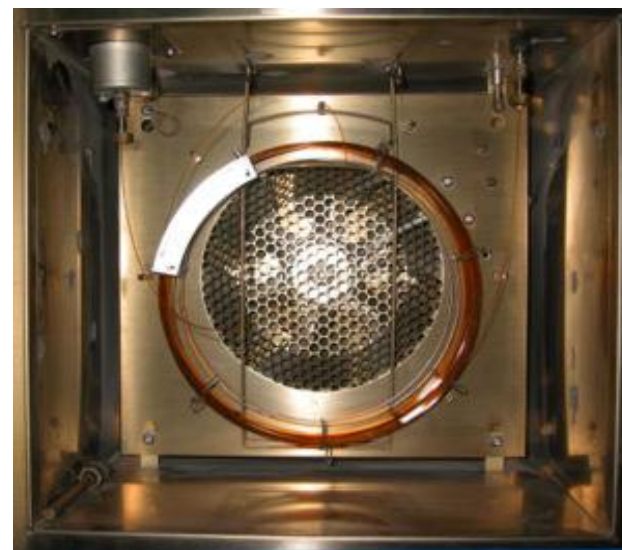


Capillary Columns

Columns placed in the Oven

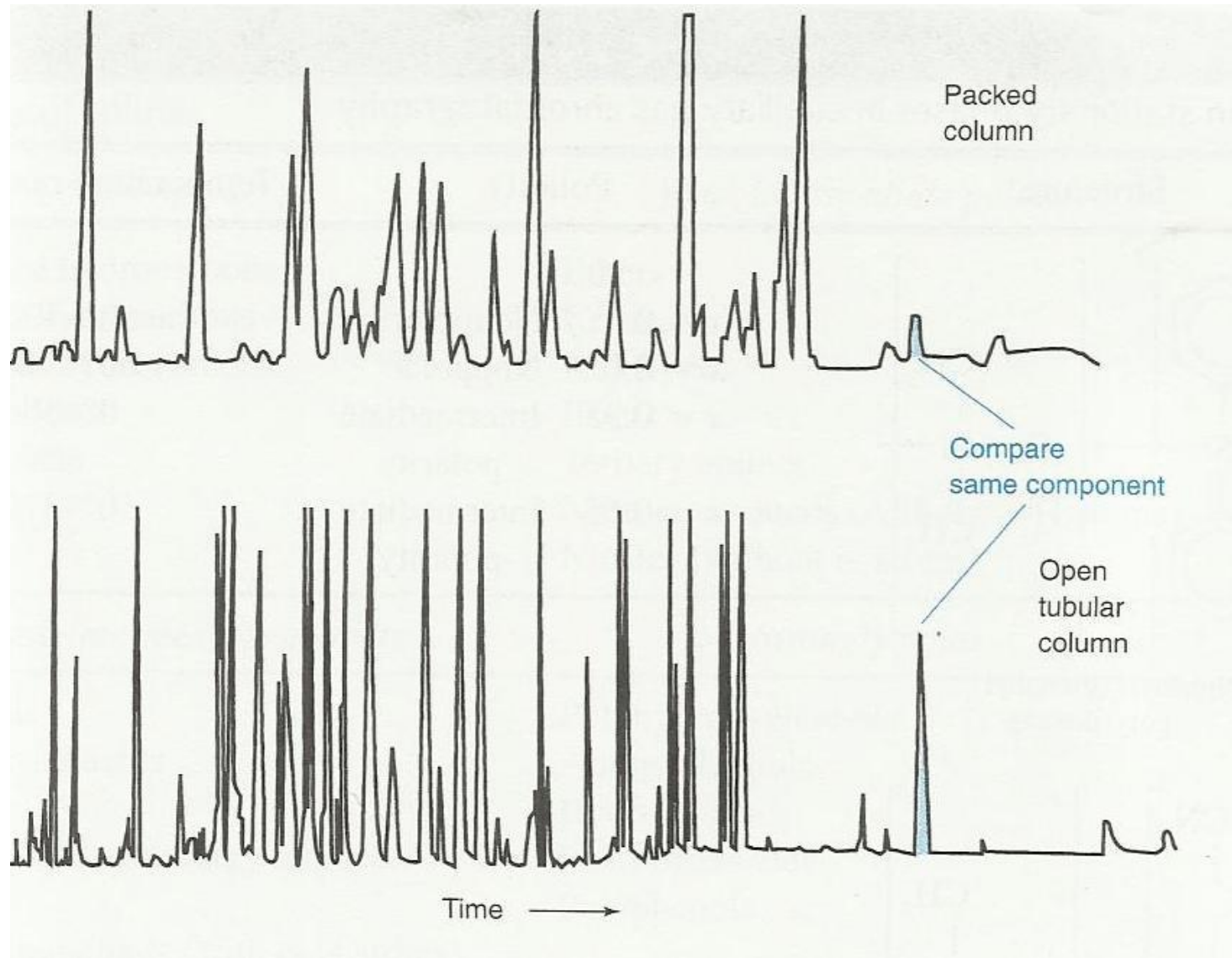


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Dual-column Configuration

Single-column Configuration



Gas Chromatographic separation of a perfume oil on a packed column and tubular column

Stationary Phases in GC columns

- The stationary phases are **high molecular weight, thermally stable polymers** that are liquids or gums.
- The **stationary phase** are selected based on their polarity on the rule “**like dissolve like**”.
- **Nonpolar columns are best for nonpolar solutes**, and vice versa.
- The most common phases are polysiloxanes and polyethylene glycols (Carbowax).

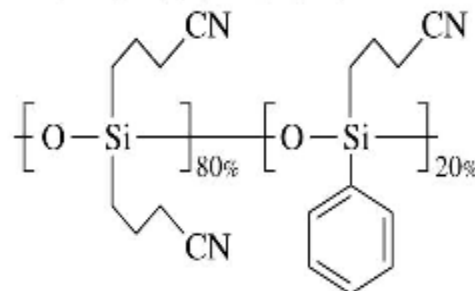
Capillary Fused Silica Stationary Phases



| Phase | Polarity | Use | Max. Temp. (°C) |
|--|--|---|-------------------|
| 100% Dimethyl polysiloxane $\left[\text{O} - \underset{\text{CH}_3}{\overset{\text{CH}_3}{\text{Si}}} - \right]_n$ | Nonpolar | Basic general-purpose phase for routine use. Hydrocarbons, polynuclear aromatics, PCBs. | 320 |
| Diphenyl, dimethyl polysiloxane $\left[\text{O} - \underset{\text{C}_6\text{H}_5}{\overset{\text{C}_6\text{H}_5}{\text{Si}}} - \right]_{x\%} \left[\text{O} - \underset{\text{CH}_3}{\overset{\text{CH}_3}{\text{Si}}} - \right]_{100-x\%}$ | 5% Low 35%, 65% Intermediate 65%, 35% Intermediate | General-purpose, good high-temperature characteristics. Pesticides. | 320 300 370 |
| 14% Cyanopropylphenyl-86% dimethylsiloxane $\left[\text{O} - \underset{\text{C}_6\text{H}_4\text{CH}_2\text{CH}_2\text{CN}}{\text{Si}} - \right]_{14\%} \left[\text{O} - \underset{\text{CH}_3}{\overset{\text{CH}_3}{\text{Si}}} - \right]_{86\%}$ | Intermediate | Separation of organochlorine pesticides listed in EPA 608 and 8081 methods. Susceptible to damage by moisture and oxygen. | 280 |



80% Biscyanopropyl–20%
cyanopropylphenyl polysiloxane

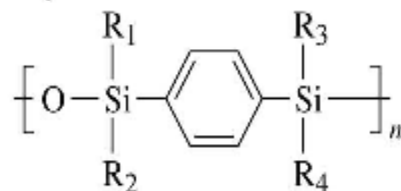


Very polar

Free acids, polysaturated fatty
acids, alcohols. Avoid polar
solvents such as water and
methanol.

275

Arylenes

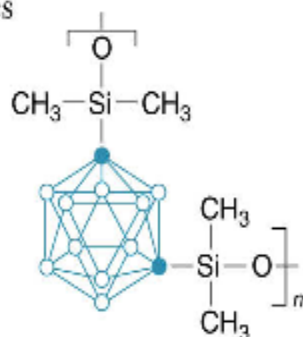


Vary R as above to
vary polarity

High temperature, low bleed

300–350

Carboranes

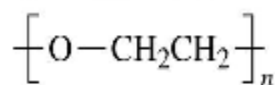


Vary R as above to
vary polarity

High temperature, low bleed

430

Poly(ethyleneglycol) (Carbowax)

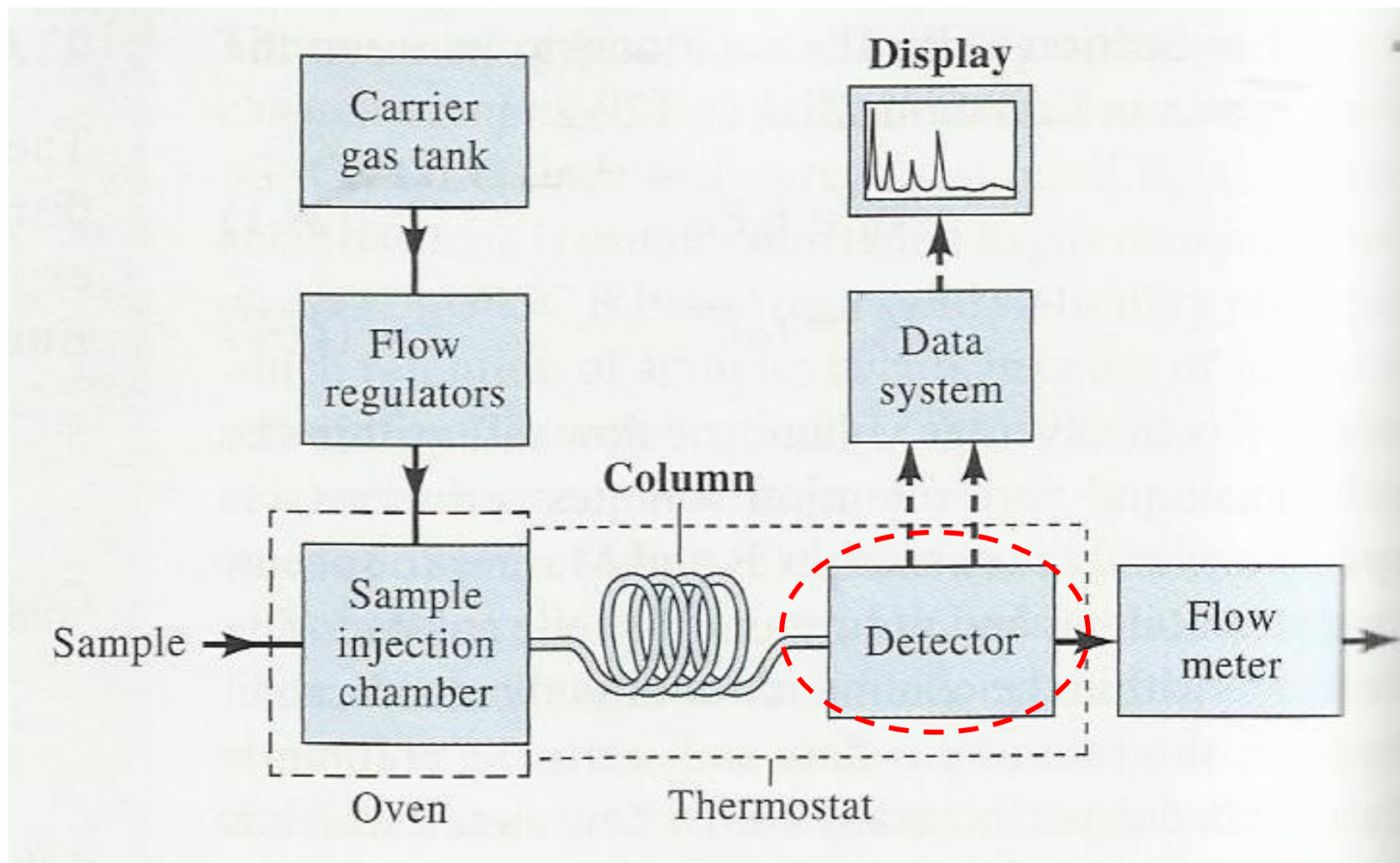


Very polar

Alcohols, aldehydes, ketones,
and separation of aromatic
isomers, e.g., xylenes

250

GC Detectors



GC Detectors

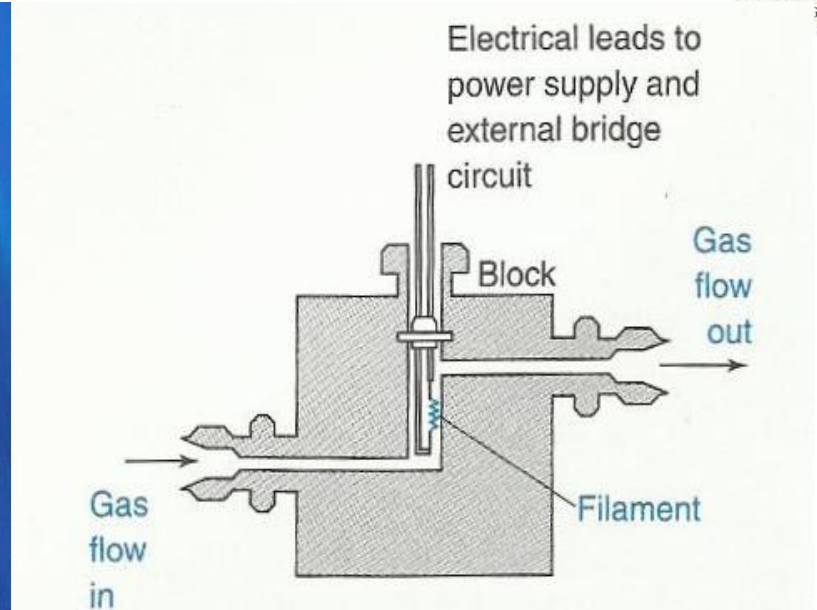
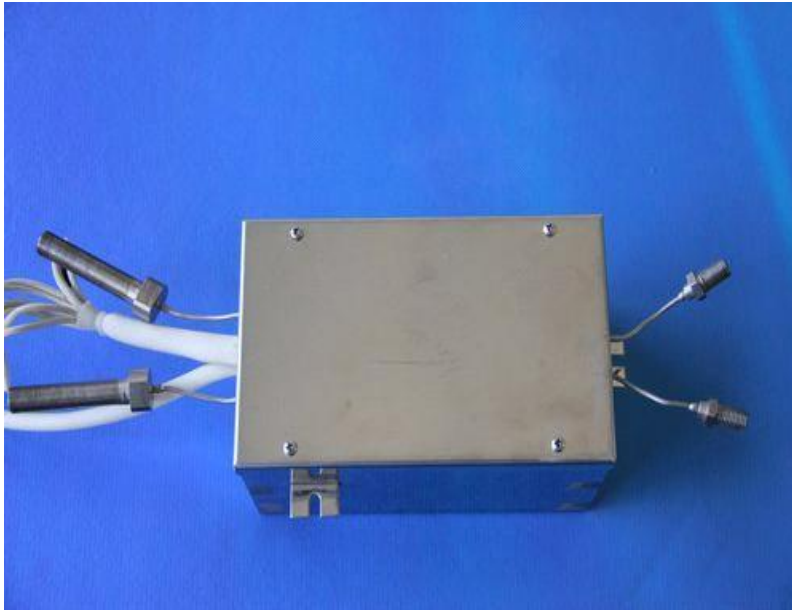
- After the components of a mixture are separated in the chromatography column, they must be **detected** at the outlet so that they can be **identified and measured**.
- A detector does **not identify what is eluted** from column → only tells us that **something is emerging**.
- Most common types of detectors:
 - Thermal Conductivity Detector (TCD)**
 - Flame Ionization Detector (FID)**

Thermal Conductivity Detector (TCD)

- The operating principle relies on the thermal conductivity of gas mixture as a **function of their composition**.
- Measures the ability of a substance to transport heat from a **hot region to cold region**.
- Helium → carrier gas commonly used in TCD → **2nd highest thermal conductivity** of the gas stream.

Thermal Conductivity at 273 K , 1 atm

| Gas | Thermal conductivity $\text{J}/(\text{K} \cdot \text{m} \cdot \text{s})$ | Molecular Weight |
|------------------------|---|---------------------|
| H_2 | 0.170 | 2 |
| He | 0.141 | 4 |
| NH_3 | 0.021 5 | 17 |
| N_2 | 0.024 3 | 28 |
| C_2H_4 | 0.017 0 | 28 |
| O_2 | 0.024 6 | 32 |
| Ar | 0.016 2 | 40 |
| C_3H_8 | 0.015 1 | 44 |
| CO_2 | 0.014 4 | 44 |
| Cl_2 | 0.007 6 | 71 |

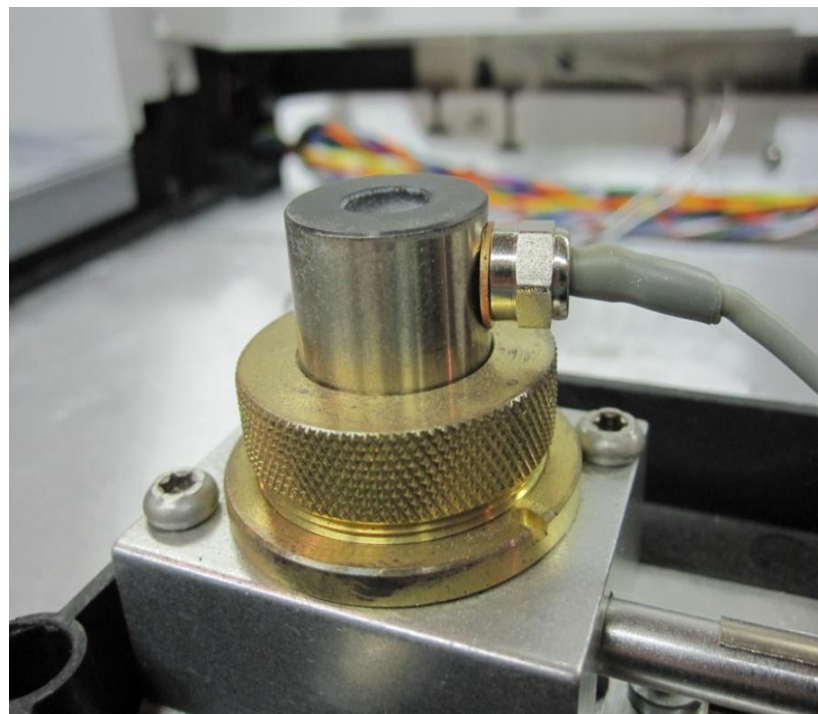
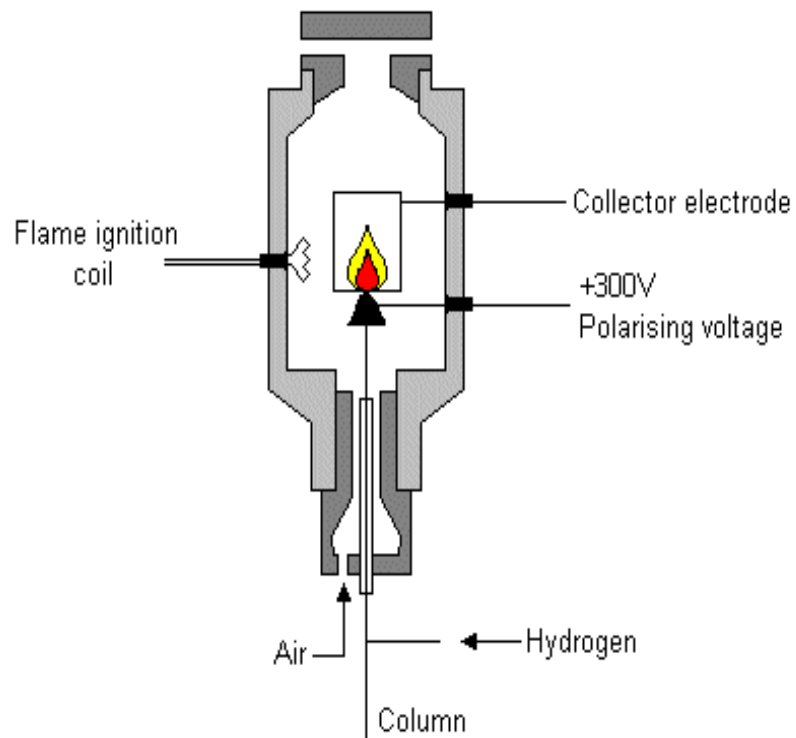


- Eluate from the column flows over a **hot filament**.
- The filament gets hotter → its electrical resistance increases, and the voltage across the filament changes.
- The **detector measures the change in voltage**.

Flame Ionization Detector (FID)

- Relies on the creation of ion/charged particles produced from the sample/compound when they are burnt by temperature in a flame.
- Eluate is burned in a mixture of H_2 and air.
- Carbon atoms produce CH radicals, which are thought to produce CHO^+ ions in the flame.
- Cations produced in the flame carry electric current from the anode flame tip to the cathode collector → detector signal.

FID Detector



- In the absence of organic solutes, the current is almost 0.
- The **detection limit is 100 times smaller** than that of the TCD.
- **FID responds to most hydrocarbons**, which constitute the vast majority of GC analytes.
- **Insensitive to nonhydrocarbon** such as H_2 , He, N_2 , O_2 , CO, CO_2 , H_2O , NH_3 , NO, H_2S and SiF_4 .

Other Detectors

- Nitrogen-Phosphorus Detector (NPD)
- Flame Photometric Detector (FPD)
- Photoionization Detector (PD)
- Sulfur Chemiluminescence Detector (SCD)
- Nitrogen Chemiluminescence Detector (NCD)
- Electron Capture Detector (ECD)



Comparison of Gas-Chromatographic Detectors



| Detector | Application | Sensitivity Range | Linearity | Remarks |
|----------------------|---|--|---|---|
| Thermal conductivity | General, responds to all substances | Fair, 5–100 ng, 10 ppm–100% | Good, except thermistors at higher temperatures | Sensitive to temperature and flow changes; concentration sensitive |
| Flame ionization | All organic substances; some oxygenated products respond poorly. Good for hydrocarbons | Very good, 10–100 pg, 10 ppb–99% | Excellent, up to 10^6 | Requires very stable gas flow; response for water is 10^4 – 10^6 times weaker than for hydrocarbons; mass-sensitive |
| Flame photometric | Sulfur compounds (393 nm), phosphorus compounds (526 nm) | Very good, 10 pg S, 1 pg P | Excellent | |
| Flame thermionic | All nitrogen- and phosphorus-containing substances | Excellent, 0.1–10 pg, 100 ppt–0.1% | Excellent | Needs recoating of sodium salts on screen; mass sensitive |
| Electron capture | All substances that have affinity to capture electrons; no response for aliphatic and naphthenic hydrocarbons | Excellent for halogen-containing substances, 0.05–1 pg, 50 ppt–1 ppm | Poor | Very sensitive to impurities and temperature changes; quantitative analysis complicated; concentration sensitive |

Characteristics of the Ideal Detector

- Adequate sensitivity.
- Good **stability and reproducibility**.
- A **linear response to solutes** that extends over several order of magnitude.
- A temperature range from room temperature to at least 400 °C.
- A **short response time** and **independent flow rate**.
- **High reliability** and **ease of use**.
- **Non destructive**.

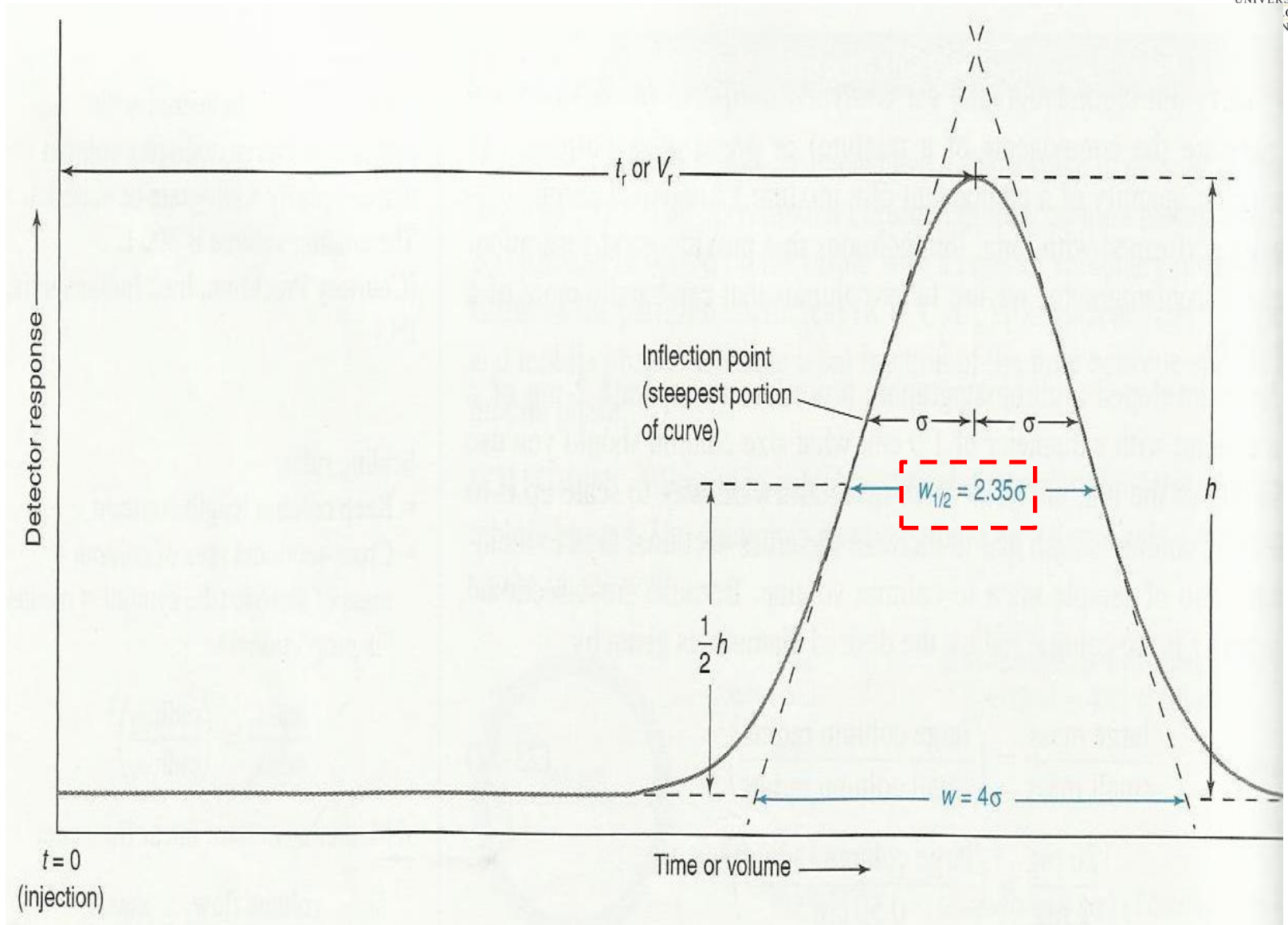
Quantitative Analyses based on GC Chromatogram

1) Analyses based on Peak Areas (or Peak Height)

- The **area of a peak is proportional to the concentration** of that component.
- In most of the computer-controlled system, the peak area is measured automatically by the computer.
- If the peak area must be measured by hand, and if the peak has a **Gaussian shape**, then the **area** is:

$$\text{Area of Gaussian peak} = 1.064 \times \text{peak height} \times w_{1/2}$$

Recall:



2a) Analyses based on Internal Standard Method (single point)

- After measuring the response factor with standard mixture, the quantity of unknown is:

$$\frac{A_x}{[X]} = F \left(\frac{A_s}{[S]} \right)$$

A_x = area of analyte signal

A_s = area of internal standard

$[X]$ = concentration of analyte

$[S]$ = concentration of standard

F = response factor

Example

When **1.06 mM** of **pentanol** and **1.53 mM** of **hexanol** were separated by gas chromatography, they gave relative peak areas of **922** and **1570** units, respectively.

When **0.570 mM** of **pentanol** was added to an **unknown containing hexanol**, the relative chromatographic peak areas were **843:816** (pentanol:hexanol).

How much hexanol did the unknown contain?

Solution

$$\text{Equation: } \frac{A_X}{[X]} = F \left(\frac{A_S}{[S]} \right)$$

S = [pentanol]

X = [hexanol]

For the standard mixture,

$$\frac{A_X}{[X]} = F \left(\frac{A_S}{[S]} \right)$$

$$\frac{1570}{1.53} = F \left(\frac{922}{1.06} \right)$$

$$F = 1.18$$

For an unknown mixture,

$$\frac{A_x}{[X]} = F \left(\frac{A_s}{[S]} \right)$$

$$\frac{816}{[X]} = 1.18 \left(\frac{843}{0.571} \right)$$

$$[x] = 0.468 \text{ mM}$$

Exercise

- A standard solution containing 6.30×10^{-8} M iodoacetone and 2.00×10^{-7} M p-dichlorobenzene (an internal standard) gave peak areas of 395 and 787, respectively, in a gas chromatogram. A 3.00-mL unknown solution of iodoacetone was treated with 0.100 mL of 1.60×10^{-5} M p-dichlorobenzene and the mixture was diluted to 10.00 mL. Gas chromatography gave peaks areas of 633 and 522 for iodoacetone and p-dichlorobenzene, respectively. Find the concentration of iodoacetone in the 3.00 mL of original solution.

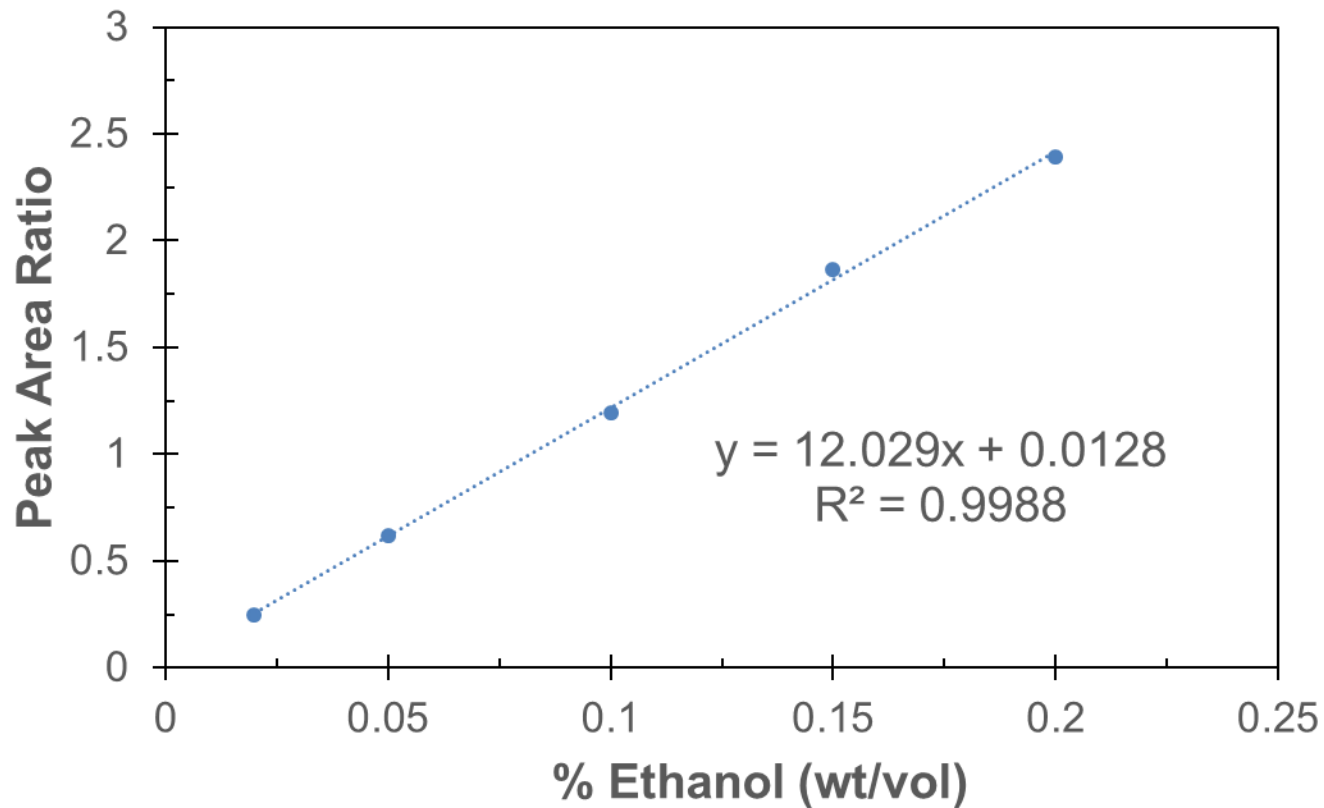
2b) Analyses based on Internal Standard Method (calibration curve)

A 5.00 mL blood sample from a suspect is spiked with 0.500 mL of aqueous 1% propanol internal standard. A 10.0-microliter portion of the mixture is injected into the GC, and the peak areas are recorded. Standards are treated in the same way. The following results were obtained:

| % Ethanol (wt/vol) | Peak Area Ethanol | Peak Area Propanol |
|--------------------|-------------------|--------------------|
| 0.0200 | 114 | 457 |
| 0.0500 | 278 | 449 |
| 0.100 | 561 | 471 |
| 0.150 | 845 | 453 |
| 0.200 | 1070 | 447 |
| Unknown | 782 | 455 |

Construct a calibration curve of the ratio of the Ethanol/Propanol area vs ethanol concentration and calculate the unknown concentration.

| % Ethanol (wt/vol) | Peak Area Ethanol | Peak Area Propanol | Peak Ratio |
|-----------------------|----------------------|-----------------------|------------|
| 0.0200 | 114 | 457 | 0.249 |
| 0.0500 | 278 | 449 | 0.620 |
| 0.100 | 561 | 471 | 1.19 |
| 0.150 | 845 | 453 | 1.87 |
| 0.200 | 1070 | 447 | 2.39 |
| Unknown | 782 | 455 | 1.72 |



Correlation, $y = 12.029 x + 0.0128$, by substitution of $y = 1.72$ into the equation, concentration of unknown, $x = 0.142 \text{ \% wt/vol}$

3) Area Normalization method

- Complete elution of all the sample constituents is necessary.
- The area of each peak is then measured and **corrected for differences in detector response** to the different eluates.
- The correction involves **dividing the area by the response factor**.
- The concentration of the analyze is found from the **ratio of its corrected area to the total corrected area** of all peaks.

Example

For a chromatogram containing three peaks, the relative areas were found to be 16.4, 45.2 and 30.2 in order of increasing retention time. Calculate the percentage of each compound if the relative detector responses were 0.600, 0.780 and 0.880, respectively.

Solution

| Compound | Relative Peak Area | Relative detector Response | Corrected Area(peak area/detector response) | % Compound |
|--------------------------|--------------------|----------------------------|---|------------|
| 1 | 16.4 | 0.600 | 27.33 | 22.9 |
| 2 | 45.2 | 0.780 | 57.95 | 48.4 |
| 3 | 30.2 | 0.880 | 34.32 | 28.7 |
| Corrected area summation | | | 119.6 | |

Temperature Selection

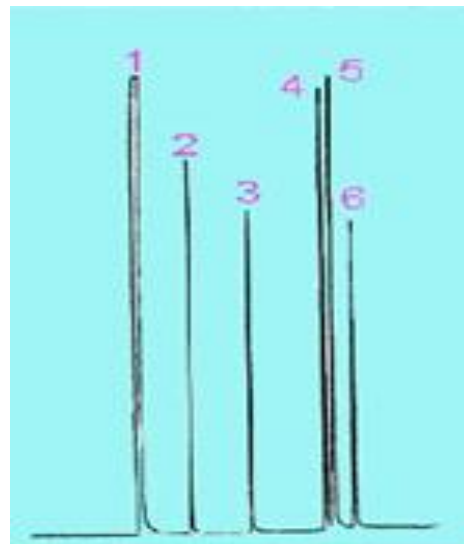
- The **proper temperature selection** in gas chromatography → depends on several factors.
- The injection temperature should be
 1. relatively **high**
 2. consistent with **thermal stability** of the sample
 3. give **faster rate of vaporization** to get the sample into the column in **small volume**.
 4. **decrease spreading** and **increase resolution** results.
- **Too high** an injection temperature → tend to **degrade the rubber septum** and cause **dirtying** of the injection port.

- The column temperature is **compromise** between
 1. **speed**
 2. **sensitivity**
 3. **resolution**
- At high column temperature → sample components spend most of their time in the **gas phase** → **eluted quickly** → poor resolution.
- At low temperature → sample spend more time in the **stationary phase** and **elute slowly** → **resolution increased** but **sensitivity is decreased** due to increased **spreading of the peaks**.

- The detector temperature must be high enough → prevent condensation of the sample components.
- Separations can be facilitated by temperature programming → the compounds eluted with more difficulty can be eluted in a reasonable time without forcing the others from the column too quickly.

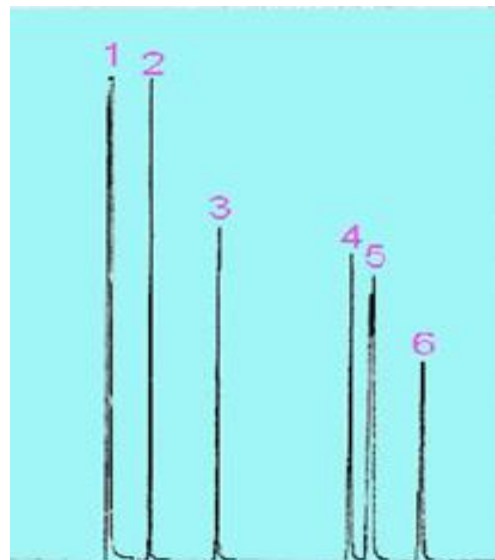
Example: GC analysis results for the same mixture using different temperature-programmed

Temperature
Program:
50 °C - 100 °C,
10 ° C/min

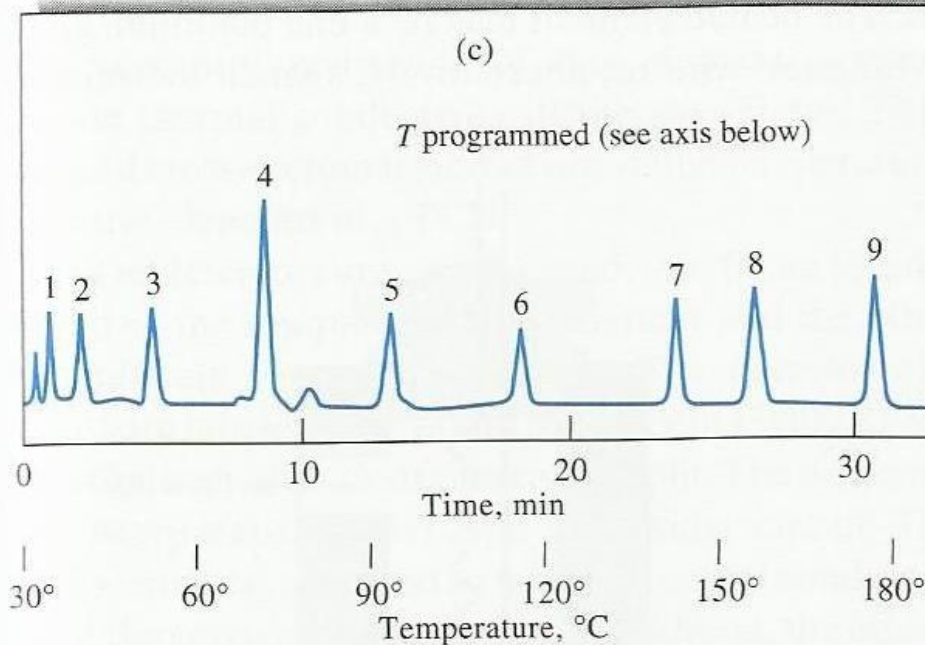
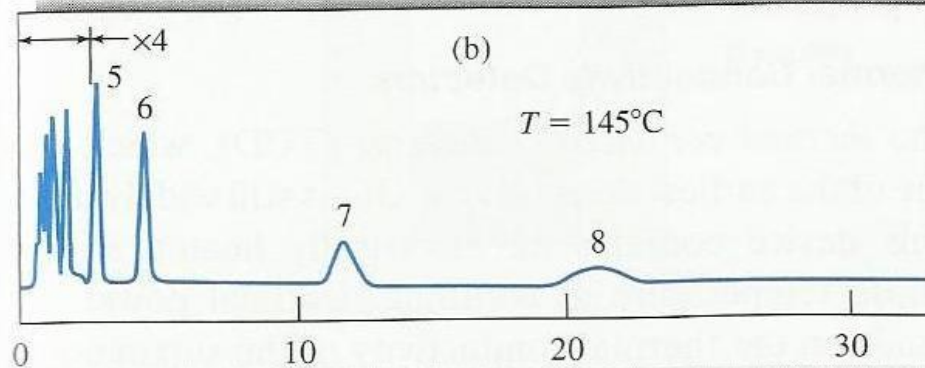
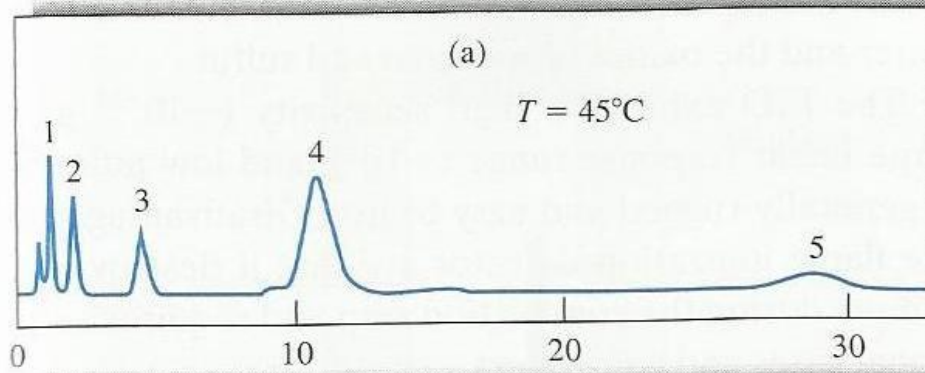


| Peak# | Compound | Retention Time |
|-------|----------------------------|----------------|
| 1 | Methanol | 2.244 min |
| 2 | Benzene | 3.260 min |
| 3 | Toluene | 4.455 min |
| 4 | Ethyl Benzene | 5.941 min |
| 5 | Para-xylene Meta-xylene | 6.115 min |
| 6 | Ortho-xylene | 6.524 min |

Temperature
Program:
60°C - isothermal



| Peak# | Compound | Retention Time |
|-------|----------------------------|------------------------|
| 1 | Methanol | 2.245 min |
| 2 | Benzene | 3.176 min |
| 3 | Toluene | 4.616 min |
| 4 | Ethyl Benzene | 7.486 min |
| 5 | Para-xylene Meta-xylene | 7.895 min 7.957 min |
| 6 | Ortho-xylene | 9.050 min |



Effect of temperature on gas chromatography:

- (a) Isothermal at 45°C
- (b) Isothermal at 145°C
- (c) Programmed at 30°C to 180°C

Method Development in Gas Chromatography

- Order of decisions:
 1. **Goal of analysis**
 - what is require?
 - qualitative or quantitative?
 - need high resolution?
 2. Sample preparation
 3. Choosing the **detector**
 4. Choosing the **column**
 5. Choosing the **injection method**
 - split/splitless/on-column injection

End of GC topic