

Problem 1: Noise-induced oscillations (Ingalls 7.8.27, 45 points)

Stochastic systems can exhibit a range of oscillatory behaviors, ranging from near-perfect periodicity to erratic cycles. To explore this behavior, consider a stochastic relaxation oscillator studied by José Vilar and colleagues (Vilar et al., 2002). The system involves an activator and a repressor. The activator enhances expression of both proteins. The repressor acts by binding the activator, forming an inert complex. A simple model of the system is:

Name and description	Reaction	Reaction propensity
R_1 (activator synthesis)	$\emptyset \longrightarrow b_A A$	$\frac{\gamma_A}{b_A} \cdot \frac{\alpha_0 + N_A/K_A}{1 + N_A/K_A}$
R_2 (repressor synthesis)	$\emptyset \longrightarrow b_R R$	$\frac{\gamma_R}{b_R} \cdot \frac{N_A/K_R}{1 + N_A/K_R}$
R_3 (activator decay)	$A \longrightarrow \emptyset$	$\delta_A N_A$
R_4 (repressor decay)	$R \longrightarrow \emptyset$	$\delta_R N_R$
R_5 (association)	$A + R \longrightarrow C$	$k_C N_A N_R$
R_6 (dissociation w/ activator decay)	$C \longrightarrow R$	$\delta_A N_C$

Here, N_A , N_R , and N_C are the molecular counts for the activator, repressor, and activator-repressor complex. The parameters b_A and b_R characterize the expression burst size. The Hill-type propensities of the synthesis reactions are not well-justified at the molecular level, but these expressions nevertheless provide a simple formulation of a stochastic relaxation oscillator.

- a) Take parameter values $\gamma_A = 250$, $b_A = 5$, $K_A = 0.5$, $\alpha_0 = 0.1$, $\delta_A = 1$, $\gamma_R = 50$, $b_R = 10$, $K_R = 1$, $k_C = 200$, and $\delta_R = 0.1$. Run a Gillespie SSA simulation of this model and verify its quasi-periodic behavior.

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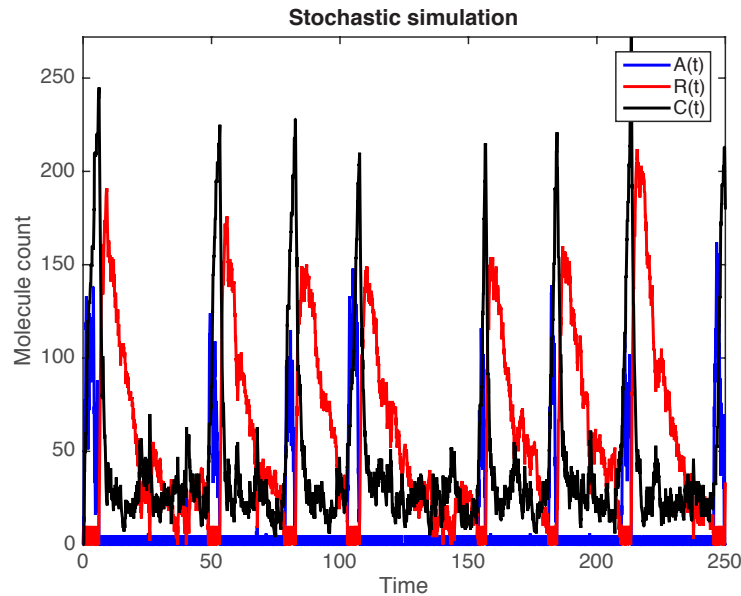
1 function [] = problemla()
2     gamma_A = 250; b_A = 5; K_A = 0.5; alpha_0 = 0.1; delta_A = 1;
3     gamma_R = 50; b_R = 10; K_R = 1; k_C = 200; delta_R = 0.1;
4     i = 1; n = 10000;
5     a = zeros(1,n); a(1) = 0;
6     r = zeros(1,n); r(1) = 0;
7     c = zeros(1,n); c(1) = 0;
8     t = zeros(1,n); t(1) = 0;
9
10    while i < n
11        rates = [ (gamma_A/b_A) * (alpha_0 + a(i)/K_A)/(1 + a(i)/K_A), ...
12                (gamma_R/b_R) * (a(i)/K_R)/(1 + a(i)/K_R), ...
13                delta_A*a(i), ...
14                delta_R*r(i), ...
15                k_C * a(i) * r(i), ...
16                delta_A * c(i) ];
17        t(i+1) = t(i)+expnrnd(1/sum(rates));
18        event = randsample(6,1,true, rates ./ sum(rates));
19        if event == 1
20            a(i+1) = a(i) + b_A; r(i+1) = r(i); c(i+1) = c(i);
21        elseif event == 2

```

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22         a(i+1) = a(i); r(i+1) = r(i) + b_R; c(i+1) = c(i);
23     elseif event == 3
24         a(i+1) = a(i) - 1; r(i+1) = r(i); c(i+1) = c(i);
25     elseif event == 4
26         a(i+1) = a(i); r(i+1) = r(i) - 1; c(i+1) = c(i);
27     elseif event == 5
28         a(i+1) = a(i) - 1; r(i+1) = r(i) - 1; c(i+1) = c(i) + 1;
29     elseif event == 6
30         a(i+1) = a(i); r(i+1) = r(i) + 1; c(i+1) = c(i) - 1;
31     end
32     i = i + 1;
33 end
34
35 % Next calculate the probability distribution at the end
36 plot(t,a,'b', 'LineWidth', 2); hold on;
37 plot(t,r,'r', 'LineWidth', 2);
38 plot(t,c,'k', 'LineWidth', 2);
39 title('Stochastic simulation')
40 xlabel('Time')
41 ylabel('Molecule count')
42 legend('A(t)', 'R(t)', 'C(t)', 'Location', 'NorthEast')
43 set(gca,'FontSize',14)
44 axis([100 250 0 max([a, r, c])])
45
46 end

```



b) The deterministic version of this model is:

$$\begin{aligned}
 \frac{da}{dt} &= \gamma_A \frac{\alpha_0 + a/K_A}{1 + a/K_A} - k_C ar - \delta_A a \\
 \frac{dr}{dt} &= \gamma_R \frac{a/K_R}{1 + a/K_R} - k_C ar + \delta_A c - \delta_R r \\
 \frac{dc}{dt} &= k_C ar - \delta_A c
 \end{aligned}$$

where a , r , and c are the concentrations of activator, repressor, and complex. Run a simulation with the same parameter values as in part (a). Does the system exhibit oscillations? How is the

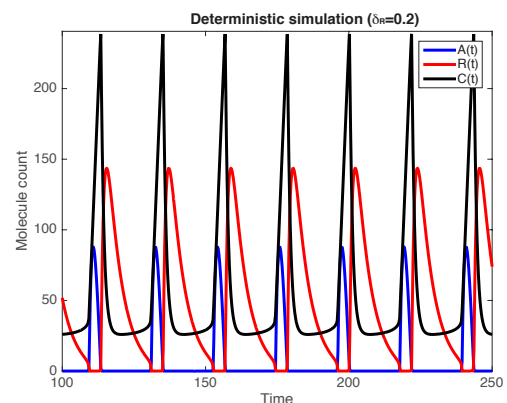
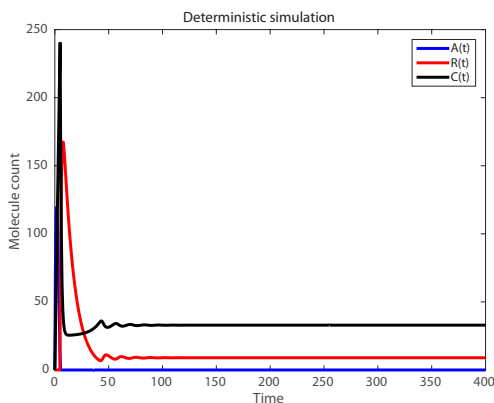
behavior different if you set $\delta_R = 0.2$?

```

1 function [] = problem1b2()
2     global gamma_A b_A K_A alpha_0 delta_A gamma_R b_R K_R k_C delta_R
3     gamma_A = 250; b_A = 5; K_A = 0.5; alpha_0 = 0.1; delta_A = 1;
4     gamma_R = 50; b_R = 10; K_R = 1; k_C = 200; delta_R = 0.2; % or delta_R = 0.1
5
6     initial_concentrations = [5, 10, 35];
7     time_interval = [0, 250];
8     [timepoints, concentrations] = ode45(@chainliddt, ...
9         time_interval, initial_concentrations);
10
11     % Next calculate the probability distribution at the end
12     plot(timepoints, concentrations(:,1), '-b', 'LineWidth', 3); hold on;
13     plot(timepoints, concentrations(:,2), '-r', 'LineWidth', 3);
14     plot(timepoints, concentrations(:,3), '-k', 'LineWidth', 3);
15     title('Deterministic simulation (\delta_R = 0.2)')
16     xlabel('Time')
17     ylabel('Molecule count')
18     legend('A(t)', 'R(t)', 'C(t)', 'Location', 'NorthEast')
19     set(gca, 'FontSize', 14)
20     axis([0 250 0 max(max(concentrations))])
21
22 end
23
24 function changes_in_concentrations = chainliddt(time, current_concentrations)
25     global gamma_A b_A K_A alpha_0 delta_A gamma_R b_R K_R k_C delta_R
26
27     A = current_concentrations(1);
28     R = current_concentrations(2);
29     C = current_concentrations(3);
30
31     change_in_A = gamma_A*(alpha_0 + A/K_A)/(1+A/K_A) - k_C*A*R - delta_A*A;
32     change_in_R = gamma_R*(A/K_R)/(1+A/K_R) - k_C*A*R + delta_A*C - delta_R*R;
33     change_in_C = k_C*A*R - delta_A*C;
34
35     changes_in_concentrations = [change_in_A; change_in_R; change_in_C];
36 end

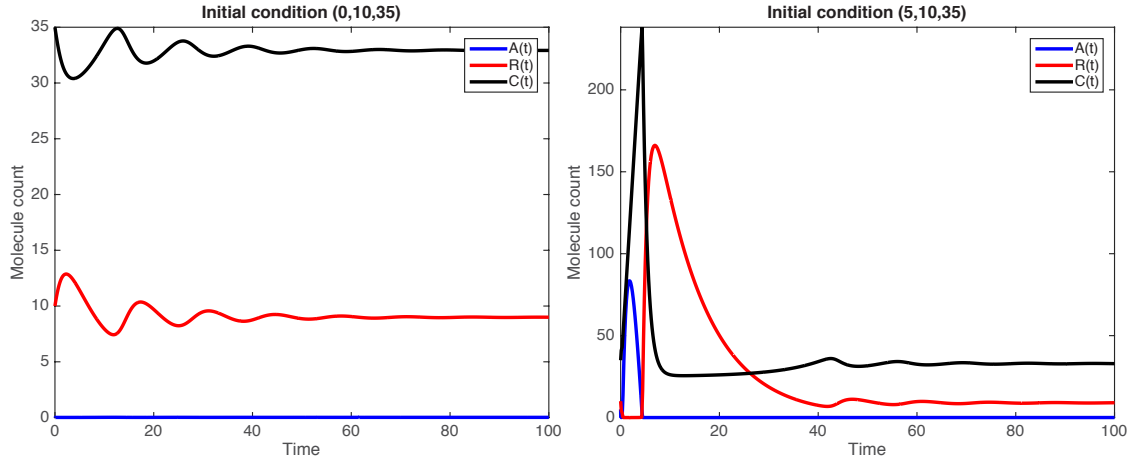
```

The system does not exhibit oscillations when $\delta_R = 0.1$ (left figure). However, when $\delta_R = 0.2$, oscillations are restored (right figure).



- c) The contrast between the behavior of the models in parts (a) and (b), for $\delta_R = 0.1$, can be explained by the excitability of this relaxation oscillato. Run two simulations of the deterministic

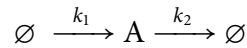
model ($\delta_R = 0.1$), one from initial conditions $(a, r, c) = (0, 10, 35)$ and another from initial conditions $(a, r, c) = (5, 10, 35)$. Verify that in the first case, the activator is quenched by the repressor, and the system remains at a low-activator steady state, whereas in the second case, this small quantity of activator is able to break free from the repressor and invoke a (single) spike in expression. Explain how noise in the activator abundance could cause repeated excitations by allowing the activator abundance to regularly cross the threshold. This is referred to as *noise-induced oscillation*.



Code is a straightforward modification of the above. The “burstiness” of expression could easily cause the activator to exceed its threshold abundance of ≤ 5 , particularly since the burst size $b_A = 5$. (Even without bursting, A will sometimes cross the activity threshold due to chance.)

Problem 2: Effect of autorepression (55 points)

Consider the open system



a) Find the master equation for this system.

The trick to this problem is to recognize that the rate of the degradation reaction $\sim nk_2$ while the rate of the synthesis reaction is constant:

$$\frac{dP(n, t)}{dt} = \begin{cases} k_2 P(1, t) - k_1 P(0, t) & : n = 0 \\ [n + 1] k_2 P(n + 1, t) + k_1 P(n - 1, t) - [k_1 + nk_2] P(n, t) & : n > 0 \end{cases}$$

b) Show that, at steady state,

$$P(N_A = n) = \frac{k_1}{k_2 n} P(N_A = n - 1, t)$$

We will show this by induction. For the base case:

$$\frac{dP(0, t)}{dt} = 0 \implies P(1, t) = \frac{k_1}{k_2} P(0, t)$$

Now suppose that the stated formula is true for some value x , i.e. $P(x, t) = \frac{k_1}{k_2 x} P(x-1, t)$. We will show that the formula holds for $x+1$:

$$\begin{aligned} \frac{dP(x, t)}{dt} = 0 &\implies P(x+1, t) = \frac{1}{k_2(x+1)} [-k_1 P(x-1, t) + (k_1 + x k_2) P(x, t)] \\ &= \frac{1}{k_2(x+1)} [-k_2 x P(x, t) + (k_1 + x k_2) P(x, t)] \\ &= \frac{k_1}{k_2(x+1)} P(x, t) \end{aligned}$$

Thus the stated formula applies for all $n \geq 0$. As the probability distribution does not depend on time at steady state, the argument t can be omitted.

- c) Use the Taylor series for e^x to derive the steady-state probability distribution:

$$P(N_A = n) = \frac{\left(\frac{k_1}{k_2}\right)^n e^{-k_1/k_2}}{n!}$$

Express our result from part (b) in terms of $P(0)$:

$$P(n) = \frac{k_1}{k_2 n} P(n-1) = \frac{\left(\frac{k_1}{k_2}\right)^2}{n(n-1)} P(n-2) = \frac{\left(\frac{k_1}{k_2}\right)^n}{n!} P(0)$$

We can find $P(0)$ using the fact that the probabilities must sum to one:

$$\begin{aligned} 1 &= P(0) \left[1 + \frac{k_1}{k_2} + \frac{k_1^2}{2k_2^2} + \dots \right] \\ &= P(0) \sum_{i=0}^{\infty} \frac{\left(\frac{k_1}{k_2}\right)^i}{i!} = e^{k_1/k_2} P(0) \implies P(0) = e^{-k_1/k_2} \end{aligned}$$

Plugging this into our result above, we obtain the desired expression:

$$P(n) = \frac{\left(\frac{k_1}{k_2}\right)^n e^{-k_1/k_2}}{n!}$$

Note that this is just a Poisson distribution with parameter $\lambda = k_1/k_2$.

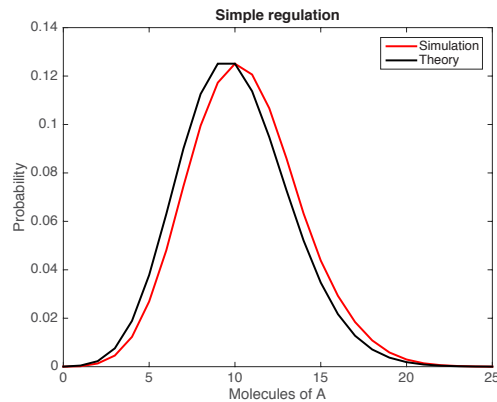
- d) Perform a Gillespie SSA simulation of the open system above with $k_1 = 10$, $k_2 = 1$. Estimate the probability distribution $P(n)$ from the timecourse $A(t)$. (Exercise your judgment in ignoring data early in the simulation and choosing an appropriate timescale for the simulation.) Plot the Poisson probability density function with $\lambda = k_1/k_2$ on the same axes for comparison.

```
1 function [] = problem1d()
2     k1 = 10; k2 = 1;
3     i = 1; n = 100000;
4     a = zeros(1,n); a(1) = 10;
5     t = zeros(1,n); t(1) = 0;
6
7     while i < n
8         rates = [k1, a(i)*k2];
```

```

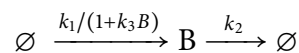
9      t(i+1) = t(i)+exprnd(1/sum(rates));
10     event = randsample(2,1,true, rates ./ sum(rates));
11     if event == 1
12         a(i+1) = a(i) + 1;
13     elseif event == 2
14         a(i+1) = a(i) - 1;
15     end
16     i = i + 1;
17 end
18
19 bins = 0:1:max(a);
20 a_binned = histc(a(n),bins)./n;
21 disp(sprintf('%d %d',length(bins),length(a_binned)));
22 plot(bins, a_binned, 'r', 'LineWidth', 2); hold on;
23 plot(bins,poisspdf(bins,k1/k2),'k', 'LineWidth', 2)
24 title('Simple regulation')
25 xlabel('Molecules of A')
26 ylabel('Probability')
27 legend('Simulation', 'Theory','Location', 'NorthEast')
28 set(gca, 'FontSize',14)
29
30 end

```



TODO: Figure out if there is a `histc()` issue.

Now consider a related molecule that exhibits hyperbolic autorepression, i.e.



- e) Perform a Gillespie SSA simulation of this system with $k_1 = 100$, $k_2 = k_3 = 1$ and estimate the probability distribution $P(n)$ from the timecourse $B(t)$. Fit a Poisson distribution to $P(n)$ and plot both on the same axes (e.g. using `poissfit()` and `poisspdf()` in MATLAB). Does the negative autoregulation system have more or less variance than expected for the simple regulation system?

```

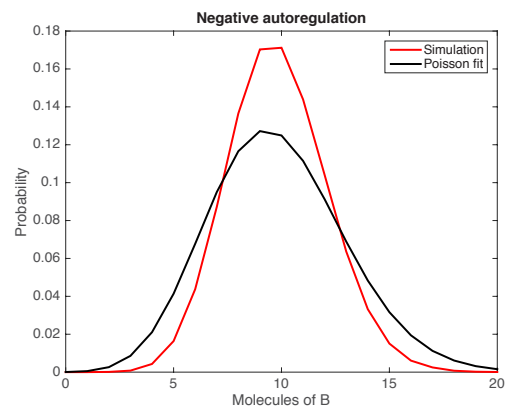
1 function [] = problem1e()
2     k1 = 100; k2 = 1; k3 = 1;
3     i = 1; n = 100000;
4     b = zeros(1,n); b(1) = 10;
5     t = zeros(1,n); t(1) = 0;
6
7     while i < n
8         rates = [k1/(1 + k3*b(i)), b(i)*k2];

```

```

9      t(i+1) = t(i)+exprnd(1/sum(rates));
10     event = randsample(2,1,true, rates ./ sum(rates));
11     if event == 1
12         b(i+1) = b(i) + 1;
13     elseif event == 2
14         b(i+1) = b(i) - 1;
15     end
16     i = i + 1;
17 end
18
19 % Next calculate the probability distribution at the end
20 bins = 0:1:max(b);
21 b_binned = histc(b,bins)./n;
22 plot(bins, b_binned, 'r', 'LineWidth', 2); hold on;
23 lambda = poissfit(b);
24 plot(bins,poisspdf(bins,lambda),'k', 'LineWidth', 2)
25 title('Negative autoregulation')
26 xlabel('Molecules of B')
27 ylabel('Probability')
28 legend('Simulation', 'Poisson fit','Location', 'NorthEast')
29 set(gca,'FontSize',14)
30
31 end

```



The simulation results for the negative autoregulation system have lower variance than the fitted Poisson distribution; this suggests that one use of negative regulation could be to limit variability.