Neural coding and adaptation in linear-nonlinear models

Contents

1 Neural coding in retinal ganglion cells

1.1 Background

Sensory systems do a remarkable job of extracting behaviorally relevant information from the environment and communicating that information to other brain regions. The field of neural coding explores this by asking: What information do neurons encode or represent? and How do biophysical mechanisms contribute to that encoding?

We can get a handle on these questions by formulating a quantitative model that characterizes the relationship between the external environment and neural activity. These *encoding models* allow us to distill our understanding of a sensory system, highlight key computational features, tackle normative questions (e.g. optimality), and uncover biophysical limits that have shaped the evolution of neural circuits.

For this problem, you will be analyzing data recorded from a salamander retinal ganglion cell (RGC) in response to a white noise visual stimulus. You will formulate a particular encoding model known as the linear-nonlinear (LN) model [2] that predicts the response of the cell to a given stimulus, and write code to learn the parameters of this model. You will also perform an analysis known as spike-triggered covariance (STC) analysis [7], to further characterize features that this particular ganglion cell is sensitive to.

For starters, download the required files (RGC data and the Matlab template script) from the course website. All of the code you need to write will the `rgc_analysis.m` script.

1.2 Experiment details

The data you will be exploring is stored in an `hdf5` file called `rgc_data.h5` (see the template script for how to load this data into Matlab). The data consists of a 16.67 minute recording of an OFF ganglion cell from the salamander retina. The stimulus was flickering white noise bars, sampled at a frame rate of 100Hz. The `stimulus` array has dimensions (30x100000) corresponding to the pixel value of the 30 bars over 100000 frames. The `time` array contains the time of the stimulus presentation for each stimulus frame. Finally, the `spike_times` array contains the spike times of an isolated retinal ganglion cell (RGC) recorded in response to the stimulus.

1.3 Spike-triggered analysis

To analyze this cell, you will first need to compute the *spike-triggered ensemble* (STE). This is a matrix containing the stimulus that directly preceeded a particular spike, for every recorded spike. Think of the STE as a cloud of points in the high-dimensional stimulus space. As we discussed in class, the mean of this set of points is known as the *spike-triggered average*. We will also characterize this point cloud by its *covariance*, this leads us to spike-triggered covariance (STC) analysis (see [7] for more information).

Part 1 Spike-triggered analysis

1. The `rgc_analysis.m` script will loop over the set of spike times, and for each one, extract the stimulus that occurred right before that spike and store it in a matrix.

2. First, choose a length (in samples) for the temporal filter, somewhere in the 300-500ms range (remember to convert this time into samples–the sampling rate is 100Hz).

3. For the length of the filter that you chose, what is the dimensionality of the filter? (This is the product of the number of spatial dimensions and the number of temporal samples in your filter).

4. Next, you'll need to initialize the matrix that will store the spike-triggered ensemble.

5. Then, fill out the code in the for loop that loops over the set of spike times, and for each spike, store the stimulus preceeding that spike in the STE.

6. Once you have the STE, you can compute its mean and covariance to estimate the spike-triggered average and covariance, respectively. Remember to make sure that the dimensions of the covariance matrix are correct–it should be a square matrix where one side has length given by the dimensionality of the filter (not the number of spikes).

7. We will further break down the STC matrix by computing its eigendecomposition. The plotting code at the bottom of the script will visualize both the eigenvalues but also the eigenvectors. Remember, each eigenvector of the STC matrix is a spatiotemporal feature that has been unrolled as a vector.

8. The provided visualization code will generate an image of the STA, the eigenspectrum, and the STC eigenvectors.

For this part, turn in plots of the spike-triggered average, the eigenvalues of the spike-triggered covariance matrix, and the top three eigenvectors (reshaped to look like a spatiotemporal filter). This plotting code has been written for you, but make sure to add appropriate axes labels and titles to the figures. In addition, answer the following questions:

1. Describe what the spike-triggered average looks like. What does this tell you about what this ganglion cell encodes?

2. How many eigenvalues in the eigenvalue spectrum are above the noise floor? (you can estimate this by simple visual inspection of the eigenvalue spectrum, but a better approach would be to quantitatively estimate the noise distribution of eigenvalues by shuffling the data and repeating the procedure).

3. Estimate the dimensionality of the subspace of stimuli that this cell is sensitive to.

4. Describe what the eigenvectors look like. How do they compare to the STA?

5. Computing the STC requires a lot of data. How can we be sure that we have computed enough to accurately estimate the STC eigenspectrum? Can you come up with a simple way to test if we need more data (without recording more data)?

1.4 Linear-nonlinear (LN) models

In the same analysis script, you will also estimate a nonlinearity–a function that describes the threshold and amount of amplification necessary to best predict the ganglion cell response given the stimulus and the STA. To do this, you will need to do the following (again, all of this follows the template in rgc_analysis.m).

Part 2 LN modeling

1. First, you need to bin the spike times into an array that stores the number of spikes observed at a particular time.

2. There is code that discretizes the filtered stimulus (u) into a small number of bins, read through it and make sure you understand what it is doing. The discretization is used to smooth out noise in our estimate of the nonlinearity.

3. Finally, in the loop, you will need to compute the expected value of the binned spikes array conditioned on times when the filtered stimulus has a particular (discretized) value.

For this part, turn in your plot of the estimated nonlinearity (again, the plotting code is provided–you need to add axis labels and a title). In addition, answer the following:

1. What shape does the nonlinearity have? What does this imply about how the neuron responds to multiple inputs (e.g. the combination two flashes as opposed to an individual flash)?

2. Estimate (just by eye) a threshold of the nonlinearity.

3. For this stimulus, what fraction of the time is this neuron above threshold?

4. What is the dimensionality of the subspace of stimuli that a linear-nonlinear model is sensitive to?

5. To fit this linear-nonlinear model, we used a stimulus with zero mean and fixed contrast. Natural stimuli, on the other hand, have many changes in mean luminance and contrast. What does this mean for our LN model? How can the LN model deal with such stimuli?

If you are feeling rather adventurous, feel free to tackle the following (optional) problem as well:

Bonus Computing a 2-D nonlinearity

1. We computed the features this cell was sensitive to using STC analysis, but we didn't use them when creating our LN model. Let's change that.

2. The standard LN model has a single linear filter. We'll extend this to have two linear filters: the STA and the largest eigenvector (the eigenvector associated with the largest eigenvalue) of the STC matrix. This is the approach taken in Fairhall 2006 [4].

3. Now that we have two features, we have a 2-D input space. Therefore, we need a 2-D (as opposed to 1-D) nonlinearity.

4. Estimate a 2-D nonlinearity by the same ratio of histograms method, but this time, the histograms will be 2-D histograms (where the two dimensions are given by the STA and the largest STC eigenvector).

5. Plot the 2-D nonlinearity that you estimate.

6. Discuss advantages and disadvantages of this model compared to the standard 1-D LN model above.

2   Feature selectivity of single neuron models

Sensory neurons encode features of the stimulus. One basic aspect of this is the temporal pattern of the input. By virtue of their intrinsic filtering properties, different neurons may prefer different temporal patterns. Using the single compartmental models you created last week we will investigate the temporal features and the nonlinear amplification (nonlinearity) preferred by different types of neurons.

In addition, it is known that preferred sensory features may change based on the input, a process called adaptation. Many different mechanisms could accomplish changes in temporal filtering, threshold and amplification. We will investigate the adaptive effects of spiking neuron models, and look at how adaptation might arise in single neurons.

This problem, you will write your own code based off of code from last week (there is no template provided).

2.1   The spike-triggered average of the integrate-and-fire neuron

As a warm-up, we will first compute the spike-triggered average of the leaky-integrate-and-fire (LIF) neuron. That is, we will inject a random current into a simulated neuron, and compute the average current that preceeds a spike. This was computed analytically in [6], you can use the figures in that paper as a guide for what to expect.

Part 3   The STA of the LIF neuron

1. Dig up your code for the leaky integrate-and-fire neuron from last week.

2. Instead of stimulating it with a current pulse, we will stimulate it with white noise (using the `randn` command in Matlab). Keep the mean at zero, but you might have to play with the standard deviation (to do this, multiply the random current with a constant scale factor, that constant is the standard deviation). You will need to make your simulation longer as well, in order to make sure we have enough spikes (to do this, modify the `t_end` variable).

3. Now that you've got a random current simulation up and running, compute the spike-triggered average of the current that directly preceeds each spike (you only have to average the 25ms or so before a spike).

4. Generate a plot of the STA of this neuron for your report.

5. Is the filter you observe monophasic or biphasic? What does this mean about how this cell responds to current input?

6. Vary the standard deviation of the current. How does this affect the filter? Plot multiple STAs on the same figure, one for each value of the standard deviation that you choose.

7. (Optional) If you want, also compute the STC matrix for this stimulus and neural response. What does the spectrum look like? What about the eigenvectors?

2.2   Gain scaling in the hodgkin-huxley model

Next, we will investigate whether or not known single neuron mechanisms are sufficient to give rise to gain scaling. To do this, we will use the Hodgkin-Huxley (HH) simulation. For treatments on the spike-triggered average of the HH neuron, see [3, 1]. The simulation you will be running was published by Mease et. al. [5] (you should read that paper so that you know what to expect).

Again, the basic goal is we would like to understand what single neuron mechanisms could give rise to gain scaling. Specifically, we will simulate a Hodgkin-Huxley neuron in response to different current fluctuations (by varying the standard deviation of the injected current, just as we did in the integrate-and-fire neuron). We will then compute linear filters and nonlinearities (so, LN models) under these different conditions to see how they change.

Part 4   Gain scaling in the Hodgkin-Huxley neuron

1. Grab the original Hodgkin-Huxley simulation from last week (the one before we added the extra conductances).

2. Similar to Part 3, replace the current pulse with a white noise current (again, zero mean, and you'll have to play around to find a good standard deviation).

3. Compute the spike-triggered average of the Hodgkin-Huxley neuron.

4. Similar to how we did it for the data analysis problem, compute the nonlinearity that predicts the firing rate given the projection of the current onto the STA.

5. What does the STA of the Hodgkin-Huxley model look like? Is it monophasic or biphasic?

6. What about the nonlinearity? Is it similar to the nonlinearity we found in Part 2? If not, how are they different? What could cause that difference?

7. Now, compute LN models again but this time in response to current injection with different standard deviation.

8. Using the LN models across different contrasts as a guide, do you see evidence for gain scaling as a possible feature of the HH equations?

9. Turn in plots of any LN models that you fit (filters and nonlinearities).

10. Discuss the advantages and disadvantages of having individual neurons perform their own gain scaling (as opposed to it being a network phenomenon).

References

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