

Metabolomics

January 8, 2021

```
[127]: import pandas as pd
import matplotlib.pyplot as plt
import matplotlib.patches as mpatches
from sklearn.cluster import KMeans
import seaborn as sns
import numpy as np
from scipy.stats import zscore, ttest_ind, norm, hypergeom
from statsmodels.stats.multitest import multipletests
from sklearn.decomposition import PCA
%matplotlib inline

sns.set(context='talk', style='ticks', font_scale=1, rc ={'axes.spines.right':_
→False, 'axes.spines.top': False, 'pdf.fonttype':42, 'ps.fonttype':
→42}, palette='colorblind')
```

1 Figure 3 C: Targeted amino acid quantification

1.1 Define the Q-test for identification of outliers in the AA Quantification

```
[2]: q90 = [0.941, 0.765, 0.642, 0.56, 0.507, 0.468, 0.437,
0.412, 0.392, 0.376, 0.361, 0.349, 0.338, 0.329,
0.32, 0.313, 0.306, 0.3, 0.295, 0.29, 0.285, 0.281,
0.277, 0.273, 0.269, 0.266, 0.263, 0.26
]

q95 = [0.97, 0.829, 0.71, 0.625, 0.568, 0.526, 0.493, 0.466,
0.444, 0.426, 0.41, 0.396, 0.384, 0.374, 0.365, 0.356,
0.349, 0.342, 0.337, 0.331, 0.326, 0.321, 0.317, 0.312,
0.308, 0.305, 0.301, 0.29
]

q99 = [0.994, 0.926, 0.821, 0.74, 0.68, 0.634, 0.598, 0.568,
0.542, 0.522, 0.503, 0.488, 0.475, 0.463, 0.452, 0.442,
0.433, 0.425, 0.418, 0.411, 0.404, 0.399, 0.393, 0.388,
0.384, 0.38, 0.376, 0.372
]
```

```

Q90 = {n:q for n,q in zip(range(3,len(q90)+1), q90)}
Q95 = {n:q for n,q in zip(range(3,len(q95)+1), q95)}
Q99 = {n:q for n,q in zip(range(3,len(q99)+1), q99)}

def dixon_test(data, left=True, right=True, q_dict=Q95):
    """
    Keyword arguments:
        data = A ordered or unordered list of data points (int or float).
        left = Q-test of minimum value in the ordered list if True.
        right = Q-test of maximum value in the ordered list if True.
        q_dict = A dictionary of Q-values for a given confidence level,
            where the dict. keys are sample sizes N, and the associated values
            are the corresponding critical Q values. E.g.,
            {3: 0.97, 4: 0.829, 5: 0.71, 6: 0.625, ...}

    Returns a list of 2 values for the outliers, or None.
    E.g.,
        for [1,1,1] -> [None, None]
        for [5,1,1] -> [None, 5]
        for [5,1,5] -> [1, None]

    """
    assert(left or right), 'At least one of the variables, `left` or `right`,  

    ↪must be True.'
    assert(len(data) >= 3), 'At least 3 data points are required'
    assert(len(data) <= max(q_dict.keys())), 'Sample size too large'

    sdata = sorted(data)
    Q_mindiff, Q_maxdiff = (0,0), (0,0)

    if left:
        Q_min = (sdata[1] - sdata[0])
        try:
            Q_min /= (sdata[-1] - sdata[0])
        except ZeroDivisionError:
            pass
        Q_mindiff = (Q_min - q_dict[len(data)], sdata[0])

    if right:
        Q_max = abs((sdata[-2] - sdata[-1]))
        try:
            Q_max /= abs((sdata[0] - sdata[-1]))
        except ZeroDivisionError:
            pass
        Q_maxdiff = (Q_max - q_dict[len(data)], sdata[-1])

```

```

if not Q_mindiff[0] > 0 and not Q_maxdiff[0] > 0:
    outliers = [None, None]

elif Q_mindiff[0] == Q_maxdiff[0]:
    outliers = [Q_mindiff[1], Q_maxdiff[1]]

elif Q_mindiff[0] > Q_maxdiff[0]:
    outliers = [Q_mindiff[1], None]

else:
    outliers = [None, Q_maxdiff[1]]

return outliers

```

1.2 Read in AA Quant data and metadata

```

[3]: aminos = pd.read_csv(r'data/AA-Results.csv', index_col=[0], header=[0,1])
aminos = aminos.sort_index()

aminos_key = pd.read_csv(r'data/AA-MetaData.csv', index_col=[0], header=[0])

blank=aminos.loc['blank'].melt()
blank.columns=['variable', 'measure', 'value']
blank=blank[blank.measure=='Final Conc.']
blank=blank.drop('measure', axis=1)
blank['cond']='blank'
blank

```

```

[3]:

```

		variable	value	cond
12	Tryptophan (Trp)-pos Results	0.4153	blank	
13	Tryptophan (Trp)-pos Results	0.4160	blank	
14	Tryptophan (Trp)-pos Results	0.4309	blank	
15	Tryptophan (Trp)-pos Results	0.4151	blank	
16	Tryptophan (Trp)-pos Results	0.4114	blank	
...	
1171	Arginine (Arg)-pos Results	0.2365	blank	
1172	Arginine (Arg)-pos Results	0.5347	blank	
1173	Arginine (Arg)-pos Results	0.2619	blank	
1174	Arginine (Arg)-pos Results	0.2252	blank	
1175	Arginine (Arg)-pos Results	0.2366	blank	

[300 rows x 3 columns]

2 Plots results, using Q-test to filter out outliers

- Creates table df_conc, which is used to generate the subsequent figure

- Must run this code for the following segment to work (uses df_conc)

```
[4]: cols = [(x,'Final Conc.') for x in aminos.columns.levels[0].values]
fig,ax = plt.subplots(2,3,figsize=(12,12),sharex=True,sharey=True)
i=0
j=0
df_conc=pd.DataFrame()
for index,df in aminos_key.groupby(by=['Time point','Growth condition']):
    #protein =df['Protein concentration (ug/mL)']
    conc = aminos.loc[df.index.values,cols]
    conc.columns = conc.columns.droplevel(1)

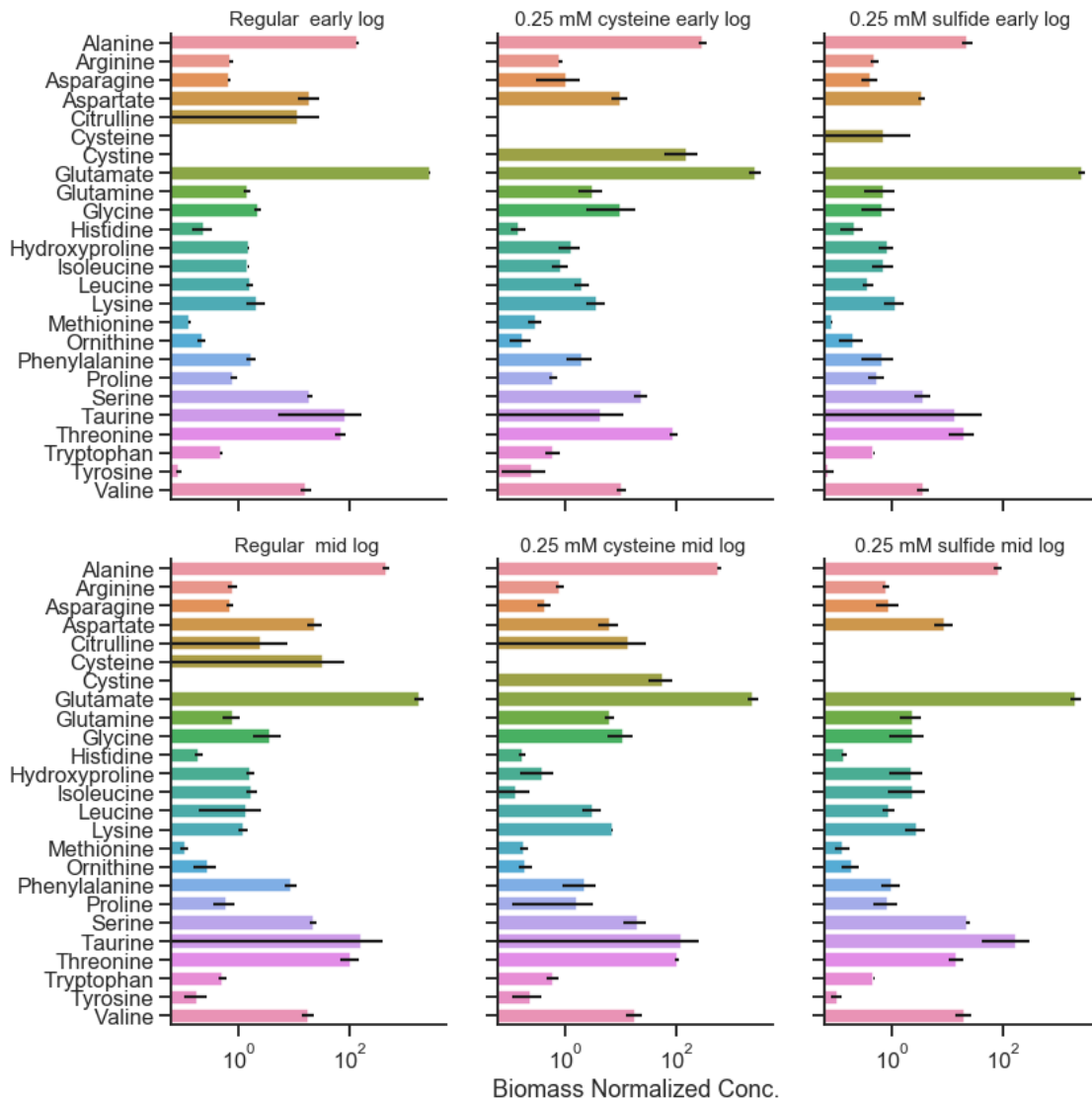
    for col in conc.columns:
        x = conc[col]
        if len(x) >= 3:
            res = np.array(dixon_test(x.values))
            res =res[res!=None]
            if len(res)>0:
                x[x==res[0]]=None
            conc.loc[:,col] = x
        cond=' '.join([index[1].split('(')[0],index[0]])[:-5]
        if '+' in cond:
            cond=cond.split('+')[1][1:]
        means = conc.mean()
        errors = conc.std()
        temp = conc.melt()
        temp['cond']=cond
        df_conc=pd.concat([df_conc,temp])
        labels = [x.split()[0] for x in conc.columns]
        sns.barplot(y=labels,x=means,xerr=errors,ax=ax[j][i%3])
        ax[j][i%3].set_title(cond,fontsize=15)
        i+=1
        if i%3 == 0 and i>0:
            j=1
        if j == 1 and i%3 == 1:
            ax[j][i%3].set_xlabel('Biomass Normalized Conc.')
    #ax[j][i%3].set_xlim(-1,50)
    ax[j][i%3].set_xscale('log')
fig.tight_layout()
sns.despine()
df_conc=pd.concat([df_conc,blank])
#plt.savefig('AminoAcid-Quant.png',bbox_inches='tight',dpi=100)
```

```
//anaconda3/lib/python3.7/site-packages/ipykernel_launcher.py:52:
RuntimeWarning: invalid value encountered in double_scalars
//anaconda3/lib/python3.7/site-packages/ipykernel_launcher.py:60:
RuntimeWarning: invalid value encountered in double_scalars
//anaconda3/lib/python3.7/site-packages/ipykernel_launcher.py:52:
```

```

RuntimeWarning: invalid value encountered in double_scalars
//anaconda3/lib/python3.7/site-packages/ipykernel_launcher.py:60:
RuntimeWarning: invalid value encountered in double_scalars
//anaconda3/lib/python3.7/site-packages/ipykernel_launcher.py:52:
RuntimeWarning: invalid value encountered in double_scalars
//anaconda3/lib/python3.7/site-packages/ipykernel_launcher.py:60:
RuntimeWarning: invalid value encountered in double_scalars
//anaconda3/lib/python3.7/site-packages/ipykernel_launcher.py:52:
RuntimeWarning: invalid value encountered in double_scalars
//anaconda3/lib/python3.7/site-packages/ipykernel_launcher.py:60:
RuntimeWarning: invalid value encountered in double_scalars
//anaconda3/lib/python3.7/site-packages/ipykernel_launcher.py:60:

```



3 Figure 3 C

- Be sure you run the code segments above, as this relies on the generation of df_conc
- Plots results with outliers removed on a log scale

```
[5]: df_conc=df_conc[(df_conc.variable!='Citruiline - pos Results')&(df_conc.
    ↪variable!='Taurine - pos Results')&(df_conc.variable!='Hydroxyproline_
    ↪(Hyp)-pos Results')]
cond_map={'Regular early log ':'Reference T1', '0.25 mM cysteine early log ':
    ↪'Cysteine T1',
    '0.25 mM sulfide early log ': 'Sulfide T1', 'Regular mid log ':
    ↪'Reference T2',
    '0.25 mM cysteine mid log ':'Cysteine T2', '0.25 mM sulfide mid log ':
    ↪'Sulfide T2','blank':'Blank'}
df_conc['labels']=df_conc.cond.map(cond_map)
rows=4
cols=4
fig=plt.figure(figsize=(cols*2.5,rows*3))
ax=plt.subplot(rows,cols,1)
i=1
order=['Arginine (Arg)-pos Results','Lysine (Lys)-pos Results','Aspartate_
    ↪(Asp)-pos Results','Serine (Ser)-pos Results',
    'Threonine (Thr)-pos Results', 'Asparagine (Asn)-pos Results','Glutamine_
    ↪(Gln)-pos Results',
    'Glycine (Gly)-pos Results', 'Proline (Pro)-pos Results',
    'Isoleucine (Ile)-pos Results', 'Leucine (Leu)-pos Results', 'Valine_
    ↪(Val)-pos Results',
    'Methionine (Met)-pos Results',
    'Alanine (Ala)-pos Results',
    'Phenylalanine (Phe)-pos Results']
for measure in order:
    df = df_conc[df_conc.variable==measure]
    g=sns.barplot(y='labels',x='value',data=df,ax=ax)
    #g.set_xticklabels(g.get_xticklabels(),rotation=90)
    g.set_xscale('log')
    g.set_ylabel('')
    g.set_xlabel('')
    g.set_title(measure.split('(')[0])
    if df['value'].max() > 500:
        g.set_xlim(10,10**4)
        #g.set_xticks(range(1,10**4))
    g.set_xlim(.01,10**4)
    g.set_xticks([10**-2,10**0,10**2,10**4])
    if (i-1)%cols != 0:
        g.set_yticklabels('')
    i+=1
    if i <= len(order):
```

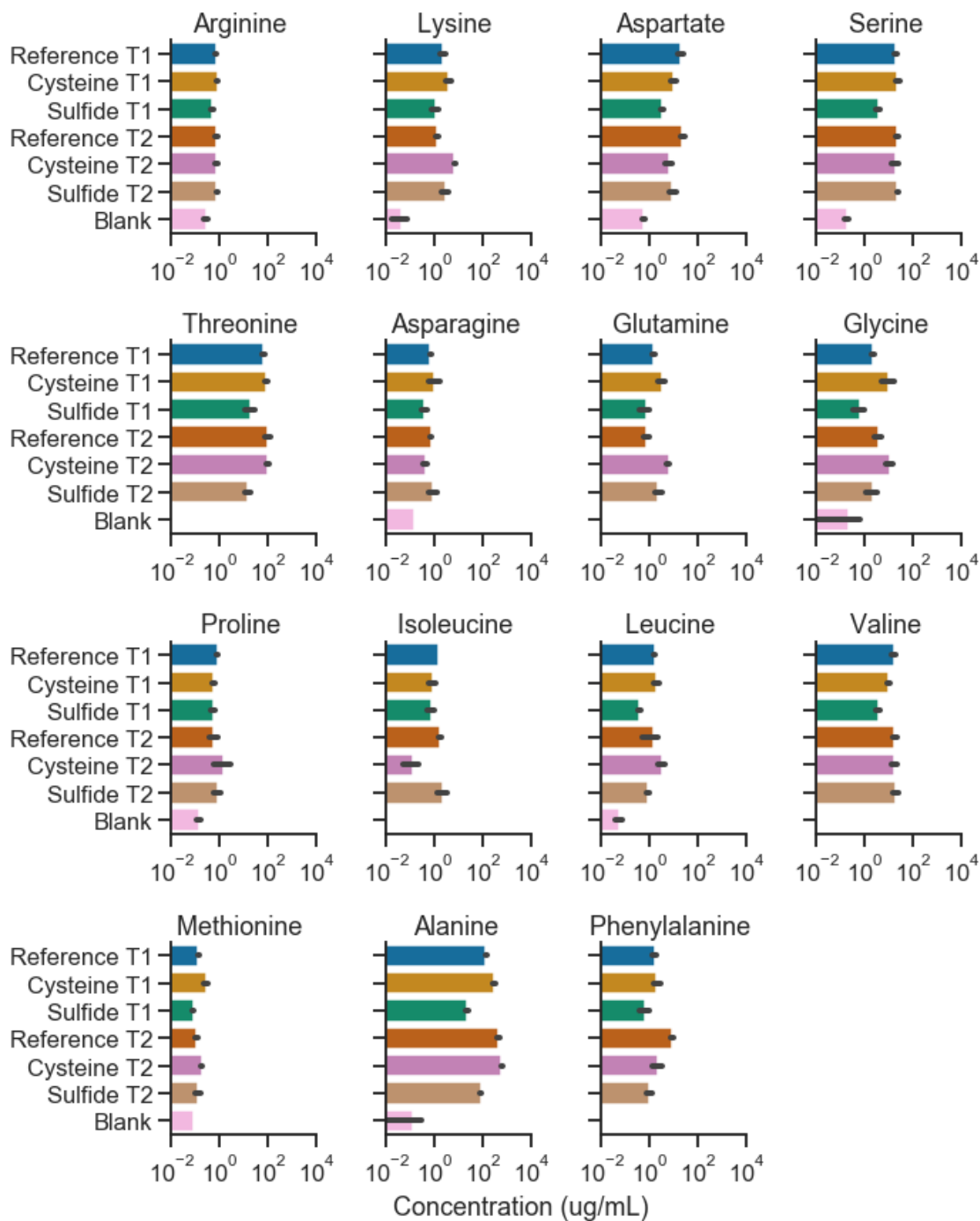
```

ax=plt.subplot(rows,cols,i)

fig.tight_layout()
fig.text(.4,0,'Concentration (ug/mL)')
#plt.savefig('2020_Targeted_aminos3.pdf',bbox_inches='tight')

```

[5]: Text(0.4, 0, 'Concentration (ug/mL)')



```

[105]: df_conc=df_conc[(df_conc.variable!='Citruiline - pos Results')&(df_conc.
    ↪variable!='Taurine - pos Results')&(df_conc.variable!='Hydroxyproline_
    ↪(Hyp)-pos Results')]
cond_map={'Regular early log ':'Reference T1', '0.25 mM cysteine early log ':
    ↪'Cysteine T1',
    '0.25 mM sulfide early log ': 'Sulfide T1', 'Regular mid log ':
    ↪'Reference T2',
    '0.25 mM cysteine mid log ':'Cysteine T2', '0.25 mM sulfide mid log ':
    ↪'Sulfide T2', 'blank':'Blank'}
df_conc['labels']=df_conc.cond.map(cond_map)
rows=4
cols=4
fig=plt.figure(figsize=(cols*3,rows*3))
ax=plt.subplot(rows,cols,1)
i=1
order=['Arginine (Arg)-pos Results','Lysine (Lys)-pos Results','Aspartate_
    ↪(Asp)-pos Results','Serine (Ser)-pos Results',
    'Threonine (Thr)-pos Results', 'Asparagine (Asn)-pos Results','Glutamine_
    ↪(Gln)-pos Results',
    'Glycine (Gly)-pos Results', 'Proline (Pro)-pos Results',
    'Isoleucine (Ile)-pos Results', 'Leucine (Leu)-pos Results', 'Valine_
    ↪(Val)-pos Results',
    'Methionine (Met)-pos Results',
    'Alanine (Ala)-pos Results',
    'Phenylalanine (Phe)-pos Results']
for measure in order:
    df = df_conc[df_conc.variable==measure]
    g=sns.boxplot(y='labels',x='value',data=df,ax=ax)
    sns.stripplot(y='labels',x='value',data=df,ax=ax,color='black',alpha=0.5)
    #g.set_xticklabels(g.get_xticklabels(),rotation=90)
    #g.set_xscale('log')
    g.set_ylabel('')
    g.set_xlabel('')
    g.set_title(measure.split('(')[0])

    #if df['value'].max() > 500:
    #    g.set_xlim(10,10**4)
    #    g.set_xticks(range(1,10**4))
    #g.set_xlim(.01,10**2)
    #g.set_xticks([10**-3,10**-2,10**0,10**2])
    if df.value.dropna().max() < 10 and df.value.dropna().max()>1:
        if df.value.dropna().max() < 6:
            if df.value.dropna().max() < 3:
                g.set_xticks([0,1,2,3])

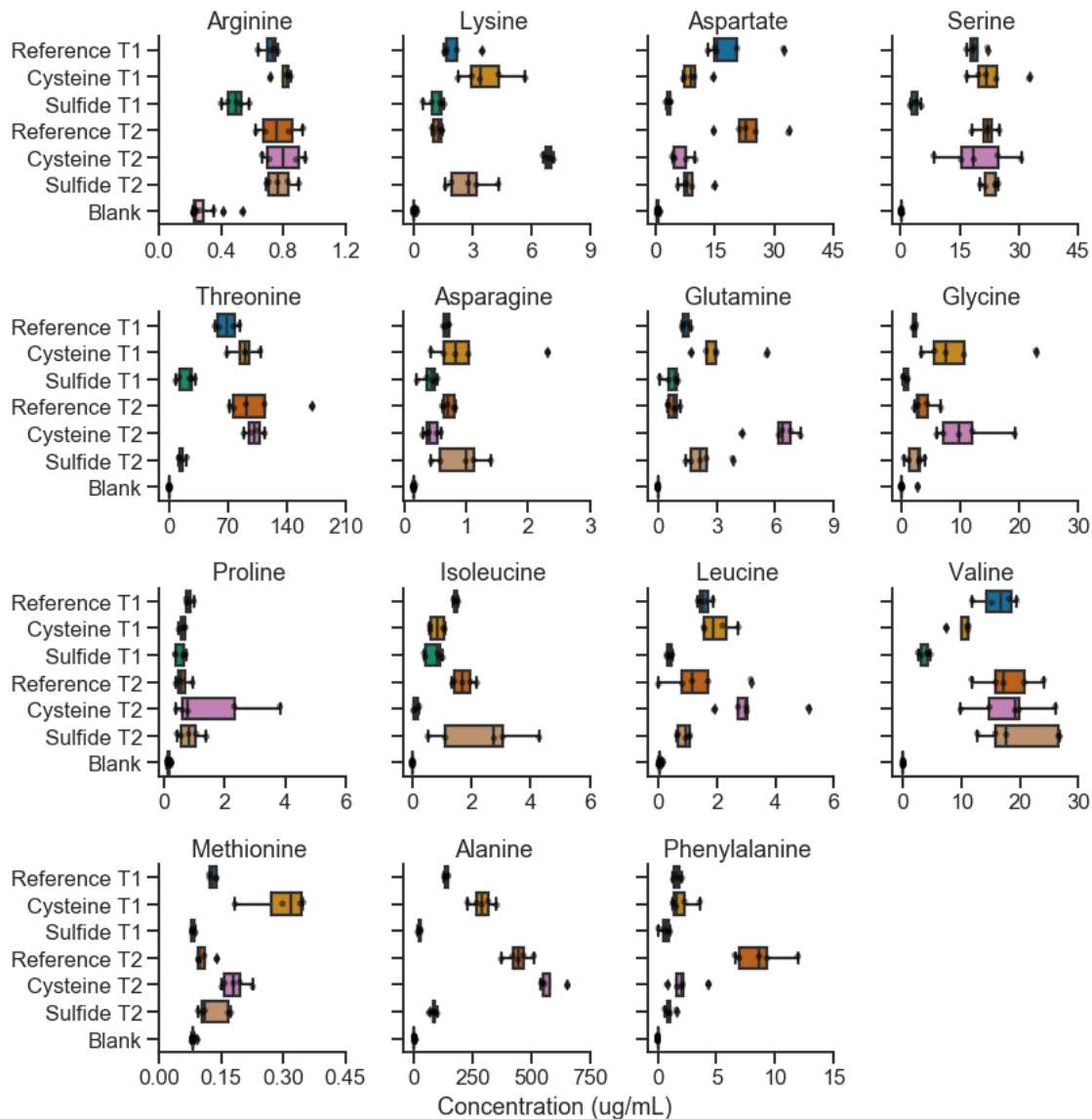
```



```

        else:
            g.set_xticks([0,2,4,6])
    else:
        g.set_xticks([0,3,6,9])
if df.value.dropna().max() > 10 and df.value.dropna().max()<100:
    if df.value.dropna().max() < 30:
        if df.value.dropna().max() < 15:
            g.set_xticks([0,5,10,15])
        else:
            g.set_xticks([0,10,20,30])
    else:
        g.set_xticks([0,15,30,45])
if df.value.dropna().max() >100:
    if df.value.dropna().max() >500:
        g.set_xticks([0,250,500,750])
    else:
        g.set_xticks([0,70,140,210])
if df.value.dropna().max() <2:
    if df.value.dropna().max() <.5:
        g.set_xticks([0,.15,.3,.45])
    else:
        g.set_xticks([0,.4,.8,1.2])
if (i-1)%cols != 0:
    g.set_yticklabels('')
i+=1
if i <= len(order):
    ax=plt.subplot(rows,cols,i)
fig.tight_layout()
fig.text(.4,0,'Concentration (ug/mL)')
plt.savefig('2020_Targeted_aminos_boxplot.pdf',bbox_inches='tight')

```



4 Figure 4 C

- Requires above code to be run first (Fig 3 C)

```
[6]: c=sns.color_palette('colorblind')
c=[c[0],c[2],c[4]]
order=['Threonine (Thr)-pos Results',
       'Isoleucine (Ile)-pos Results', 'Leucine (Leu)-pos Results', 'Valine_
       ↳(Val)-pos Results']
for measure in order:
    df = df_conc[df_conc.variable==measure].dropna()
```

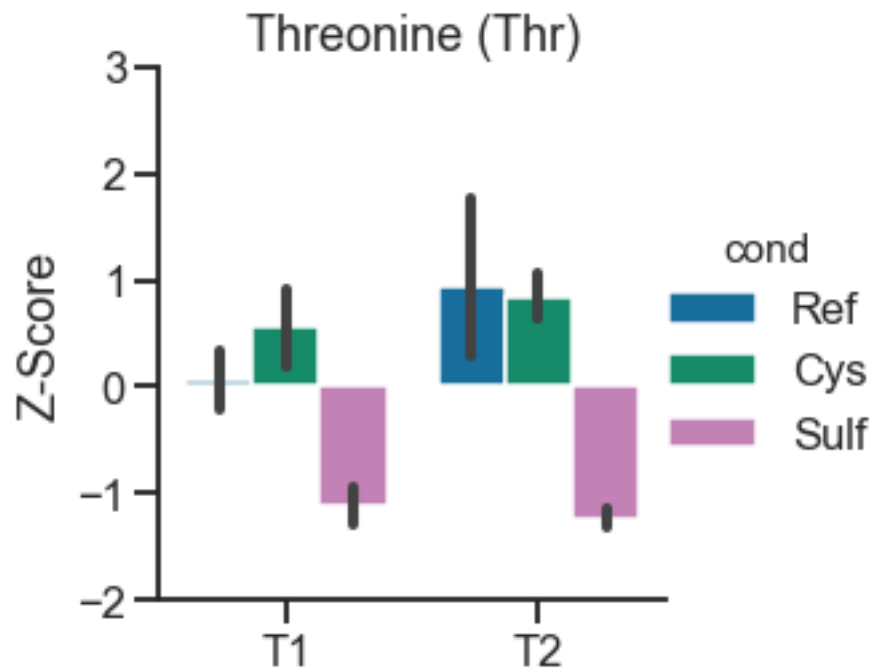
```

df['cond']=df['cond'].map(cond_map)
df=df[df['cond']!='Blank']
t=[]
cond=[]
for val in df['cond']:
    val=val.split()
    if val[0] !='Sulfide':
        cond.append(val[0][:3])
    else:
        cond.append(val[0][:4])
    t.append(val[1])
df['cond']=cond
df['t']=t

df['Z-Score']=zscore(df['value'].values)
plt.figure(figsize=(4,4))
sns.
→catplot(x='t',hue='cond',y='Z-Score',data=df,palette=c,kind='bar',height=4)
    #plt.legend(loc=2,bbox_to_anchor=(1,1))
plt.xlabel('')
plt.yticks([-2,-1,0,1,2,3])
plt.title(measure.split('-')[0])
    #plt.savefig(measure.split('-')[0]+'-zscore.pdf',bbox_inches='tight')
plt.show()
plt.clf()

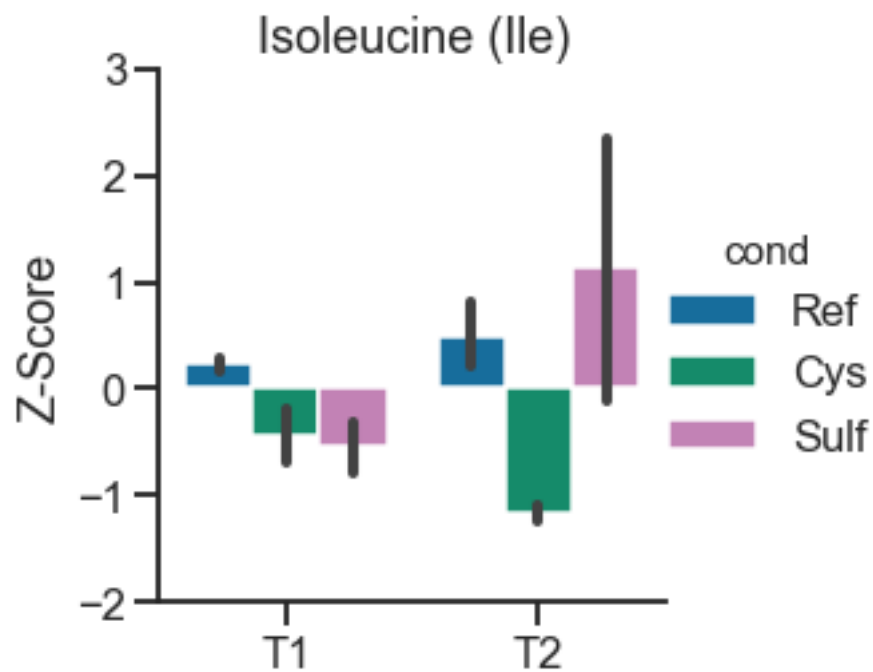
```

<Figure size 288x288 with 0 Axes>



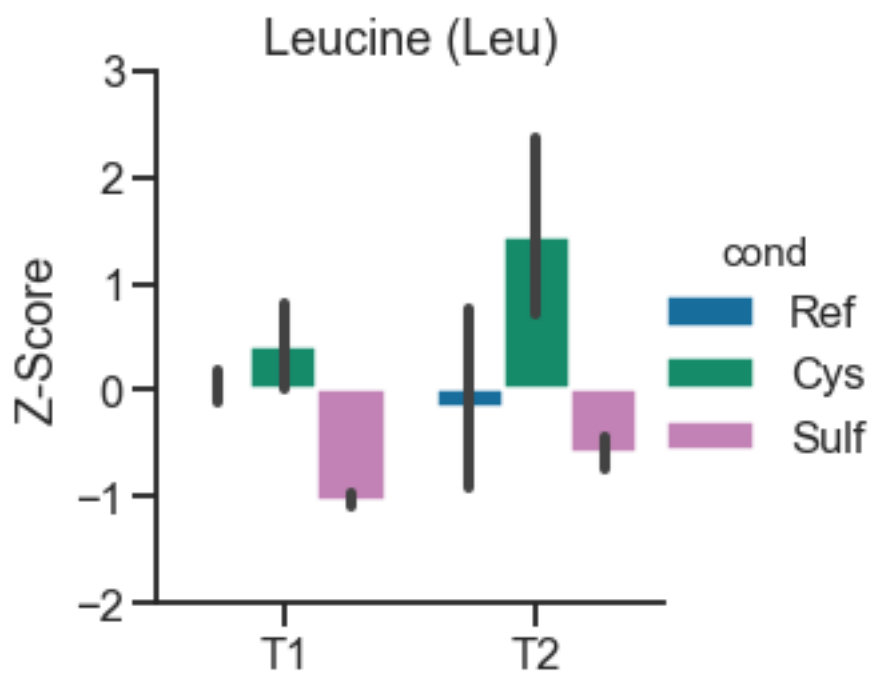
<Figure size 432x288 with 0 Axes>

<Figure size 288x288 with 0 Axes>



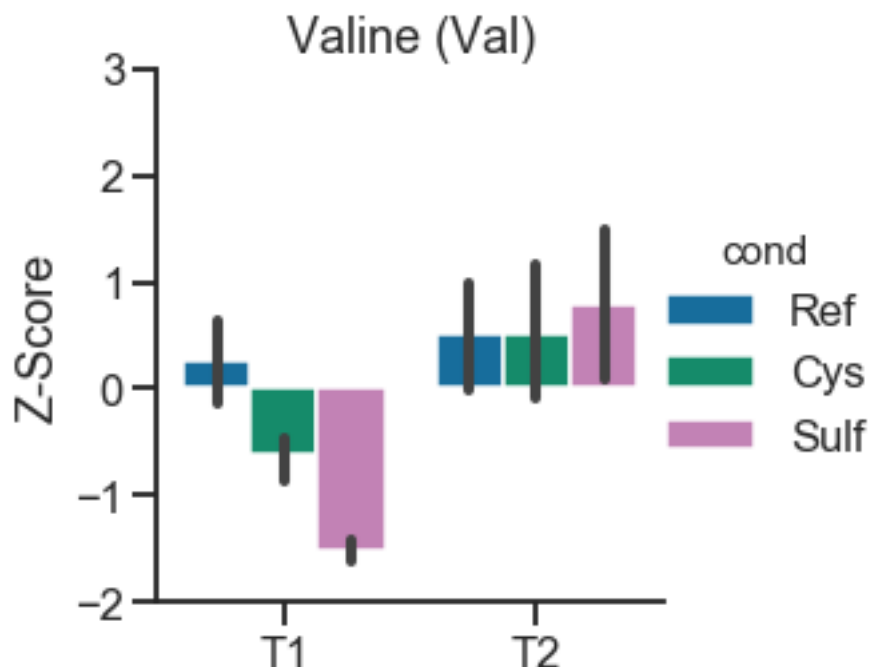
<Figure size 432x288 with 0 Axes>

<Figure size 288x288 with 0 Axes>



<Figure size 432x288 with 0 Axes>

<Figure size 288x288 with 0 Axes>



<Figure size 432x288 with 0 Axes>

5 Untargeted Metabolomics

- Results tables used below were generated by XCMS online

6 Figure 4 B

- XCMS online systems biology results
- Provides pathway dysregulation for multi sample comparison (cys vs reg vs sulf)

```
[144]: sysbio = pd.read_csv(r'data/Untargeted_Met_SysBioRes_All.tsv', sep='\t')
sysbio = sysbio[sysbio['p-values'] <= .01]
sysbio['q-values'] = multipletests(sysbio['p-values'], method='fdr_bh', alpha=0.
    ↪01)[:2][1]
sysbio['logp'] = -np.log(sysbio['p-values'])
sysbio = sysbio[sysbio['All metabolites2*'] >= 4]
sysbio['overlap'] = 100 * sysbio['Overlapping putative metabolites1'] / sysbio['All_
    ↪metabolites2*']
sysbio = sysbio.sort_values(by=['logp', 'overlap'], ascending=False)
sysbio
```

[144]:

	Pathway	Overlapping genes	\
0	isoleucine biosynthesis I (from threonine)	0	
2	valine biosynthesis	0	
3	incomplete reductive TCA cycle	0	
4	2-oxopentenoate degradation	0	
5	phosphopantothenate biosynthesis I	0	
6	respiration (anaerobic)	0	
9	glyoxylate cycle	0	
10	mixed acid fermentation	0	
11	catechol degradation to 2-oxopent-4-enoate I	0	
12	threonine degradation III (to methylglyoxal)	0	
13	pyrimidine ribonucleosides salvage I	0	
14	UDP-N-acetyl- -D-mannosaminouronate biosynthesis	0	
15	methylerythritol phosphate pathway II	0	
16	\"pyridoxal 5\"'-phosphate salvage pathway\"'	0	
17	gentisate degradation	0	
18	biotin biosynthesis from 7-keto-8-aminopelargo...	0	
21	TCA cycle IV (2-oxoglutarate decarboxylase)	0	
19	TCA cycle III (helicobacter)	0	
20	lysine biosynthesis I	0	

	Overlapping genes.1	All genes	Overlapping proteins	\
0	0%	0	0	
2	0%	0	0	
3	0%	0	0	
4	0%	0	0	
5	0%	0	0	
6	0%	0	0	
9	0%	0	0	
10	0%	0	0	
11	0%	0	0	
12	0%	0	0	
13	0%	0	0	
14	0%	0	0	
15	0%	0	0	
16	0%	0	0	
17	0%	0	0	
18	0%	0	0	
21	0%	0	0	
19	0%	0	0	
20	0%	0	0	

	Overlapping proteins.1	All proteins	Overlapping putative metabolites1	\
0	0%	0	6	
2	0%	0	3	
3	0%	0	4	
4	0%	0	3	

5	0%	0	3
6	0%	0	4
9	0%	0	3
10	0%	0	4
11	0%	0	2
12	0%	0	2
13	0%	0	2
14	0%	0	2
15	0%	0	3
16	0%	0	2
17	0%	0	2
18	0%	0	2
21	0%	0	4
19	0%	0	3
20	0%	0	3

	Overlapping putative metabolites1.1	All metabolites2*	p-values	q-values	\
0	54.5%	11	0.00068	0.00660	
2	60.0%	5	0.00120	0.00660	
3	40.0%	10	0.00120	0.00660	
4	42.9%	7	0.00200	0.00792	
5	37.5%	8	0.00260	0.00792	
6	26.7%	15	0.00300	0.00792	
9	30.0%	10	0.00430	0.00792	
10	23.5%	17	0.00440	0.00792	
11	50.0%	4	0.00540	0.00792	
12	50.0%	4	0.00540	0.00792	
13	50.0%	4	0.00540	0.00792	
14	50.0%	4	0.00540	0.00792	
15	25.0%	12	0.00700	0.00890	
16	40.0%	5	0.00790	0.00890	
17	40.0%	5	0.00790	0.00890	
18	40.0%	5	0.00790	0.00890	
21	30.8%	13	0.00890	0.00890	
19	23.1%	13	0.00890	0.00890	
20	23.1%	13	0.00890	0.00890	

	logp	overlap
0	7.293418	54.545455
2	6.725434	60.000000
3	6.725434	40.000000
4	6.214608	42.857143
5	5.952244	37.500000
6	5.809143	26.666667
9	5.449140	30.000000
10	5.426151	23.529412
11	5.221356	50.000000

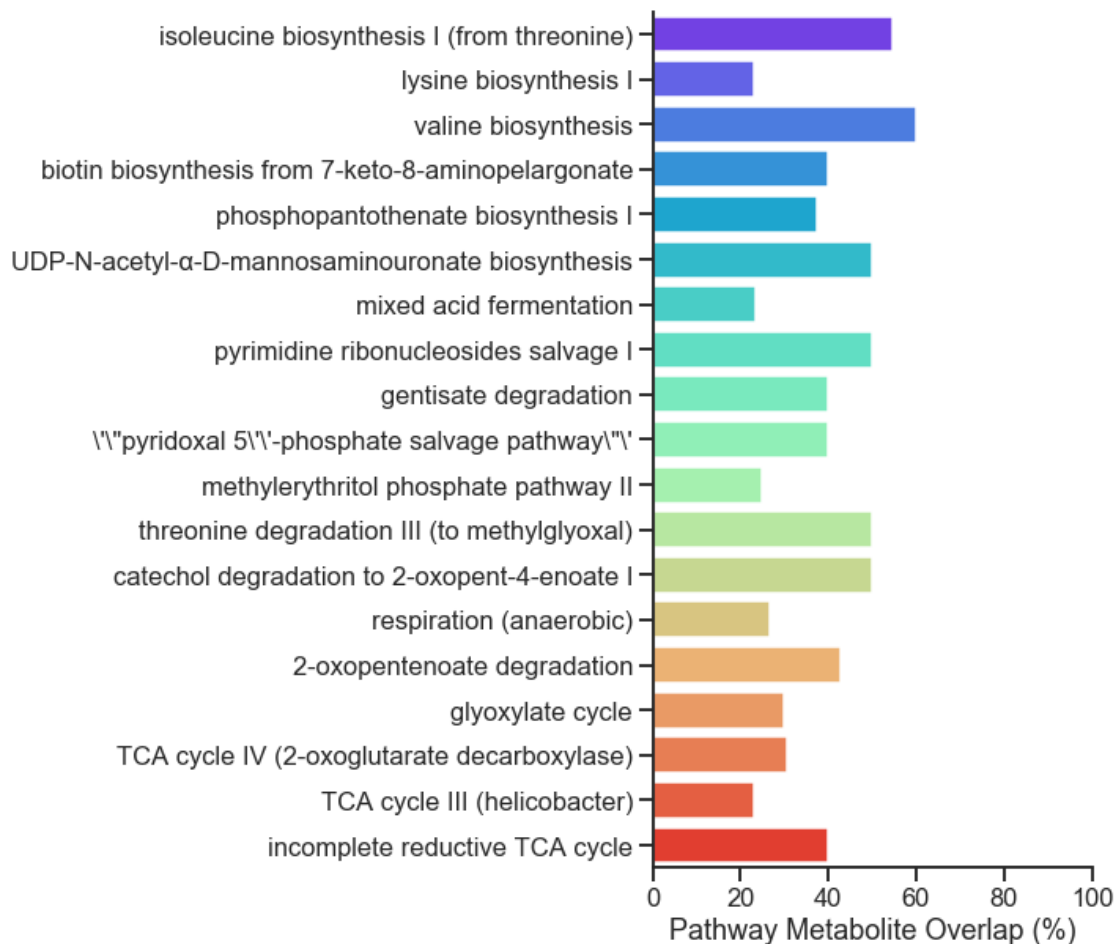
12	5.221356	50.000000
13	5.221356	50.000000
14	5.221356	50.000000
15	4.961845	25.000000
16	4.840893	40.000000
17	4.840893	40.000000
18	4.840893	40.000000
21	4.721704	30.769231
19	4.721704	23.076923
20	4.721704	23.076923

[]:

7 Names and ordering were further curated using Adobe illustrator

```
[147]: plt.figure(figsize=(5,10))
temp = sysbio[(sysbio['p-values']<=.01)&(sysbio['overlap']>=20)]
sort=[]
for p in temp['Pathway']:
    if 'lysine' in p.lower() or 'valine' in p.lower() or 'isoleucine' in p.
↳lower():
        sort.append('bcaa')
    elif 'biosynthesis' in p.lower():
        sort.append('biosynth')
    elif 'degredation' in p.lower():
        sort.append('deg')
    elif 'glyoxylate' in p.lower() or 'tca' in p.lower():
        sort.append('tca')
    elif 'fermentation' in p.lower():
        sort.append('ferm')
    else:
        sort.append('path')
temp['sort']=sort
sns.barplot(y='Pathway',x='overlap',data=temp.
↳sort_values(by='sort'),palette='rainbow')
plt.ylabel('')
plt.xlabel('Pathway Metabolite Overlap (%)')

plt.xlim(0,100)
plt.savefig('Metabolite-Pathway-Disregulation.pdf',bbox_inches='tight')
```



8 XCMS Online MultiClass Results (mummichog + feature abundances)

```
[149]: pathway = pd.read_csv(r'data/pathwayanalysis_mummichog_all.csv')
features=pd.read_csv(r'data/XCMS-MultiClass-All.csv',index_col=0)
features
```

```
[149]:
```

featureidx	name	mzmed	mzmin	mzmax	rtmed
1	M911T2_5	911.486746	911.486356	911.489983	2.049583
2	M485T23_1	484.638683	484.638013	484.639495	22.604533
3	M514T23_1	514.147566	514.147004	514.148278	22.626633
4	M515T23_1	514.649161	514.648714	514.649834	22.625600
5	M969T23	969.280946	969.279960	969.282163	22.613367
...
29983	M979T20_4	979.031523	979.026895	979.034959	19.580333

29984	M1061T20_4	1061.034120	1061.030956	1061.038579	19.574900
29985	M567T19_3	566.980125	566.974108	566.980818	19.285067
29986	M1059T20_3	1059.047693	1059.044559	1059.053117	19.578500
29987	M939T20_1	938.532292	938.528914	938.534355	19.569067

	rtmin	rtmax	npeaks	Cys_early	Cys_mid	...	\
featureidx							
1	2.049097	2.059120	3	0	0	...	
2	22.441633	22.768150	21	4	0	...	
3	22.450200	22.784867	23	5	0	...	
4	22.450200	22.751433	19	4	0	...	
5	22.441633	22.793250	24	5	0	...	
...		
29983	19.436200	19.880550	30	5	5	...	
29984	19.429500	19.682433	29	4	5	...	
29985	18.955300	19.505383	35	5	4	...	
29986	19.454600	19.634017	29	4	5	...	
29987	19.437800	19.657700	29	4	5	...	

	Sulf_4	Sulf_5	Sulf_6	Sulf_7	\
featureidx					
1	0.000000	0.000000	0.000000	0.000000	
2	2947.131112	5387.786899	10512.508700	9942.933647	
3	10228.620210	12739.203810	24752.718460	25444.025700	
4	1473.240383	3588.953005	8237.769879	8845.544354	
5	5398.305192	7712.465577	13713.700000	14791.932000	
...	
29983	74186.842800	34938.100800	36523.133400	33786.608000	
29984	66480.864000	30224.658160	32453.198770	23956.123900	
29985	37974.118000	190839.678200	180341.841400	153501.735500	
29986	119789.280000	28941.711040	36981.664000	21502.682530	
29987	38812.020000	15867.681950	20405.040330	15053.517070	

	Sulf_8	Sulf_12	Sulf_13	Sulf_14	\
featureidx					
1	0.000000	0.000000	0.000000	0.000000	
2	11833.788420	37872.27600	41237.29200	33370.244970	
3	25487.200520	65906.34182	74647.62785	67433.193000	
4	8302.467224	28255.61148	29283.38952	26005.117800	
5	14893.339120	47359.63124	54289.95368	48602.328000	
...	
29983	56593.479310	50883.76055	31963.93469	22824.688860	
29984	54586.832500	54786.08250	17157.27391	21367.899470	
29985	153928.292800	165492.47160	187036.36970	157449.180500	
29986	69536.730160	71331.79050	17133.32492	16081.842730	
29987	33345.775640	30625.96364	10903.55063	8757.898971	

	Sulf_15	Sulf_16
featureidx		
1	0.00000	0.00000
2	44785.12589	57506.10800
3	83768.10200	95294.30221
4	33901.81809	40221.23230
5	56710.08310	68136.46000
...
29983	52402.77692	100848.12300
29984	42938.66087	97891.91788
29985	171034.72650	86020.71200
29986	54714.21035	169284.58920
29987	28008.05973	56703.22411

[29987 rows x 61 columns]

9 Extract feature ID and compound mapping

```
[150]: cmp_dict={}
temp = pathway.loc[:,['overlap_features(id)','overlap_input(featureidx)']].
↳dropna()
for index in temp.index:
    names = temp.loc[index,'overlap_features(id)']
    ids =temp.loc[index,'overlap_input(featureidx)']
    for name, i in zip(names.split(';'),ids.split(';')):
        if ',' in i:
            cmp_dict[name]=[int(x) for x in i.split(',')]
        else:
            cmp_dict[name]=int(i)
cmp_dict
```

```
[150]: {'CPD-15056': 68,
'2-ACETO-2-HYDROXY-BUTYRATE': 234,
'PYRUVATE': [117, 241, 174, 197, 42, 298, 102, 34],
'1-KETO-2-METHYLVALERATE': [213, 44],
'2-KETO-3-METHYL-VALERATE': [179, 38],
'CPD-16013': 68,
'GLYCEROL-3P': [100, 303],
'GLYCEROL': 44,
'CMP': 53,
'2-KETO-ISOVALERATE': 38,
'2-ACETO-LACTATE': [161, 184, 234, 158],
'FUM': 220,
'SUC': 100,
'MAL': [220, 241],
'4-HYDROXY-2-KETOVALERATE': [161, 184, 234, 158],
```

'OXOPENTENOATE': [158, 161],
 '2-DEHYDROPANTOATE': 234,
 'L-PANTOATE': [213, 44],
 'GLYOX': 241,
 'CPD-14965': 239,
 'AMINO-OXOBUT': 27,
 'METHYL-GLYOXAL': 184,
 'UMP': 289,
 'UDP-N-ACETYL-D-GLUCOSAMINE': [115, 296],
 'UDP-MANNAC': [115, 296],
 'HYDROXY-METHYL-BUTENYL-DIP': 110,
 'PYRIDOXAL': 239,
 'PYRIDOXINE': 113,
 'DETHIOBIOTIN': 111,
 'S-ADENOSYLMETHIONINE': 242,
 'DELTA1-PIPERIDEINE-2-6-DICARBOXYLATE': [39, 21],
 'AMINO-HYDROXYMETHYL-METHYL-PYR-P': 86,
 'CHORISMATE': 188,
 'ASCORBATE': [34, 117, 197, 298, 42, 174, 102],
 'CPD-329': [34, 117, 197, 298, 42, 174, 102],
 '2-AMINOACRYLATE': [68, 262],
 'OROTATE': 247,
 'GLUCOSAMINE-1P': 171,
 '2-D-THREO-HYDROXY-3-CARBOXY-ISOCAPROATE': 38,
 '2K-4CH3-PENTANOATE': [179, 38],
 '3-CARBOXY-3-HYDROXY-ISOCAPROATE': 38,
 'ACETYL-GLU': [49, 21],
 '3OH-4P-OH-ALPHA-KETOBUTYRATE': [153, 96],
 'TMP': 210,
 'CPD-10754': [101, 233],
 'CPD-13722': 234,
 'CPD-180': 171,
 'PREPHENATE': 188,
 'HCN': 262,
 'BUTYRIC_ACID': [67, 71, 213],
 'XANTHOSINE': 254,
 'D-XYLULOSE': [161, 158, 184],
 'DCMP': 210,
 'CPD-7682': 215,
 'CPD0-1159': 312,
 'DTDP-D-GLUCOSE': 280,
 'SULFATE': 79,
 'PSEUDOURIDINE-5-P': 289,
 'ACRYLATE': 184,
 'DODECANOATE': 105}

```
[151]: rev_cmp_dict = {}
for key in cmp_dict:
    idx = cmp_dict[key]
    if type(idx) != type([]):
        idx = [idx]
    for i in idx:
        if i not in rev_cmp_dict:
            rev_cmp_dict[i]=[key]
        else:
            rev_cmp_dict[i].append(key)
rev_cmp_dict
```

```
[151]: {68: ['CPD-15056', 'CPD-16013', '2-AMINOACRYLATE'],
234: ['2-ACETO-2-HYDROXY-BUTYRATE',
      '2-ACETO-LACTATE',
      '4-HYDROXY-2-KETOVALERATE',
      '2-DEHYDROPANTOATE',
      'CPD-13722'],
117: ['PYRUVATE', 'ASCORBATE', 'CPD-329'],
241: ['PYRUVATE', 'MAL', 'GLYOX'],
174: ['PYRUVATE', 'ASCORBATE', 'CPD-329'],
197: ['PYRUVATE', 'ASCORBATE', 'CPD-329'],
42: ['PYRUVATE', 'ASCORBATE', 'CPD-329'],
298: ['PYRUVATE', 'ASCORBATE', 'CPD-329'],
102: ['PYRUVATE', 'ASCORBATE', 'CPD-329'],
34: ['PYRUVATE', 'ASCORBATE', 'CPD-329'],
213: ['1-KETO-2-METHYLVALERATE', 'L-PANTOATE', 'BUTYRIC_ACID'],
44: ['1-KETO-2-METHYLVALERATE', 'GLYCEROL', 'L-PANTOATE'],
179: ['2-KETO-3-METHYL-VALERATE', '2K-4CH3-PENTANOATE'],
38: ['2-KETO-3-METHYL-VALERATE',
      '2-KETO-ISOVALERATE',
      '2-D-THREO-HYDROXY-3-CARBOXY-ISOCAPROATE',
      '2K-4CH3-PENTANOATE',
      '3-CARBOXY-3-HYDROXY-ISOCAPROATE'],
100: ['GLYCEROL-3P', 'SUC'],
303: ['GLYCEROL-3P'],
53: ['CMP'],
161: ['2-ACETO-LACTATE',
      '4-HYDROXY-2-KETOVALERATE',
      'OXOPENTENOATE',
      'D-XYLULOSE'],
184: ['2-ACETO-LACTATE',
      '4-HYDROXY-2-KETOVALERATE',
      'METHYL-GLYOXAL',
      'D-XYLULOSE',
      'ACRYLATE'],
158: ['2-ACETO-LACTATE',
```

```

'4-HYDROXY-2-KETOVALERATE',
'OXOPENTENOATE',
'D-XYLULOSE'],
220: ['FUM', 'MAL'],
239: ['CPD-14965', 'PYRIDOXAL'],
27: ['AMINO-OXOBUT'],
289: ['UMP', 'PSEUDOURIDINE-5-P'],
115: ['UDP-N-ACETYL-D-GLUCOSAMINE', 'UDP-MANNAC'],
296: ['UDP-N-ACETYL-D-GLUCOSAMINE', 'UDP-MANNAC'],
110: ['HYDROXY-METHYL-BUTENYL-DIP'],
113: ['PYRIDOXINE'],
111: ['DETHIOBIOTIN'],
242: ['S-ADENOSYLMETHIONINE'],
39: ['DELTA1-PIPERIDEINE-2-6-DICARBOXYLATE'],
21: ['DELTA1-PIPERIDEINE-2-6-DICARBOXYLATE', 'ACETYL-GLU'],
86: ['AMINO-HYDROXYMETHYL-METHYL-PYR-P'],
188: ['CHORISMATE', 'PREPHENATE'],
262: ['2-AMINOACRYLATE', 'HCN'],
247: ['OROTATE'],
171: ['GLUCOSAMINE-1P', 'CPD-180'],
49: ['ACETYL-GLU'],
153: ['3OH-4P-OH-ALPHA-KETOBUTYRATE'],
96: ['3OH-4P-OH-ALPHA-KETOBUTYRATE'],
210: ['TMP', 'DCMP'],
101: ['CPD-10754'],
233: ['CPD-10754'],
67: ['BUTYRIC_ACID'],
71: ['BUTYRIC_ACID'],
254: ['XANTHOSINE'],
215: ['CPD-7682'],
312: ['CPD0-1159'],
280: ['DTPD-D-GLUCOSE'],
79: ['SULFATE'],
105: ['DODECANOATE']}]

```

10 Unique feature indexes mapping to a metabolite

```

[152]: idxs = []
for key in cmp_dict:
    if type(cmp_dict[key]) == type(1):
        idxs.append(cmp_dict[key])
    else:
        idxs+=cmp_dict[key]
idxs=pd.unique(idxs)
idxs

```

```
[152]: array([ 68, 234, 117, 241, 174, 197,  42, 298, 102,  34, 213,  44, 179,
           38, 100, 303,  53, 161, 184, 158, 220, 239,  27, 289, 115, 296,
          110, 113, 111, 242,  39,  21,  86, 188, 262, 247, 171,  49, 153,
           96, 210, 101, 233,  67,  71, 254, 215, 312, 280,  79, 105])
```

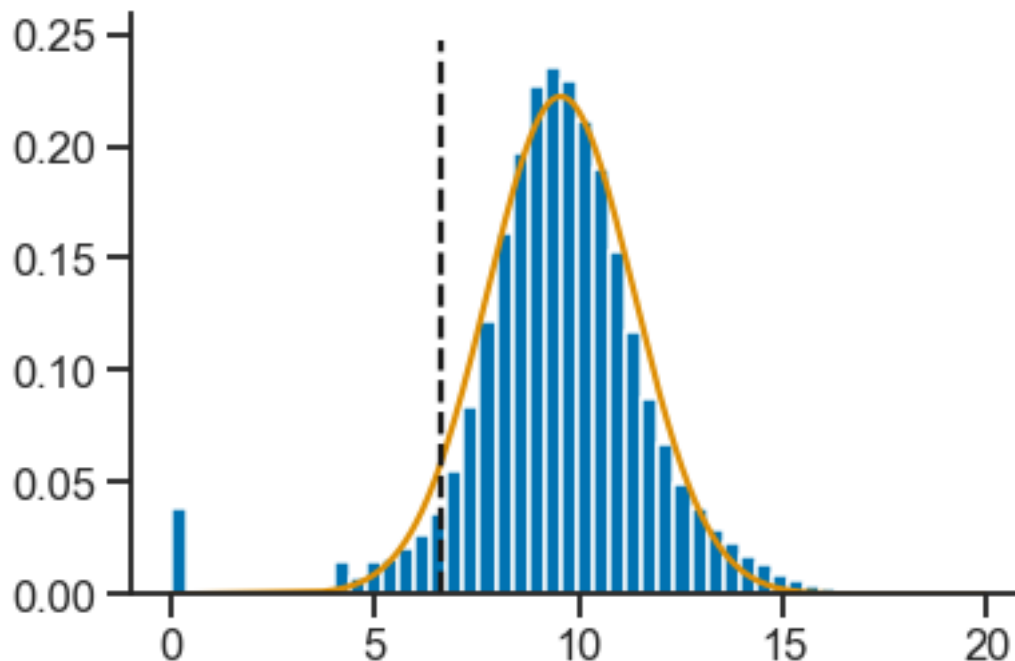
11 Feature abundance normalization

- Convert Area to Log Area
- Convert Log Area to Z-score

```
[153]: logarea = pd.DataFrame(index=features.index, columns=features.columns[31:
    ↪], data=features.iloc[:, 31:])
logarea = np.log(logarea + 1).dropna()

vals = logarea.values.ravel()
hist = plt.hist(vals, bins=50, density=True)
dist = norm(loc=np.median(vals), scale=np.std(vals)/1.25)
plt.plot(sorted(vals), dist.pdf(sorted(vals)))

keep = []
toss = []
cutoff = 0.05
x = dist.ppf(cutoff)
plt.plot((x, x), plt.ylim(), 'k--')
for i in logarea.index:
    vals = logarea.loc[i, :].values
    truth = dist.cdf(vals) > cutoff
    if len(truth[truth == True]) / len(vals) > .5:
        keep.append(i)
    else:
        toss.append(i)
logarea = logarea.loc[keep, :]
```

```
[154]: cols = ['Cys_1', 'Cys_1', 'Cys_1', 'Cys_1', 'Cys_1', 'Cys_2', 'Cys_2', 'Cys_2',
               'Cys_2', 'Cys_2', 'Reg_1', 'Reg_1', 'Reg_1', 'Reg_1',
               'Reg_2', 'Reg_2', 'Reg_2', 'Reg_2', 'Reg_2', 'Sulf_1', 'Sulf_1',
               'Sulf_1', 'Sulf_1', 'Sulf_1', 'Sulf_2', 'Sulf_2', 'Sulf_2',
               'Sulf_2', 'Sulf_2']
cols = [(x.split('_')[0],x.split('_')[1]) for x in cols]

temp = logarea.copy()
temp.columns = pd.MultiIndex.from_tuples(cols)
avglogarea = temp.groupby(level=[0,1],axis=1).median()
zscores = temp.groupby(level=[0,1],axis=1).median().T.apply(zscore).T
zscores
```

```
[154]:
```

	Cys		Reg		Sulf	
	1	2	1	2	1	2
featureidx						
2	0.139815	-2.077243	0.885792	0.776001	-0.209111	0.484747
3	-0.133942	-1.838233	1.128196	0.922728	-0.527024	0.448275
4	0.115952	-2.076387	0.906507	0.780861	-0.184705	0.457773
5	0.026579	-1.984705	1.013676	0.749986	-0.377800	0.572265
6	0.105237	-1.870644	1.060993	0.790479	-0.649394	0.563329
...
29983	1.081408	-0.302608	-0.095585	0.099708	-1.876962	1.094039
29984	1.083481	-0.375817	0.157349	0.148293	-1.942739	0.929433
29985	0.470406	1.072179	-1.822936	-0.776935	0.260781	0.796505

```
29986      1.151228 -0.782653  0.520241 -0.458928 -1.535637  1.105749
29987      1.381079 -0.660045 -0.857043  0.257514 -1.254223  1.132718
```

```
[28508 rows x 6 columns]
```

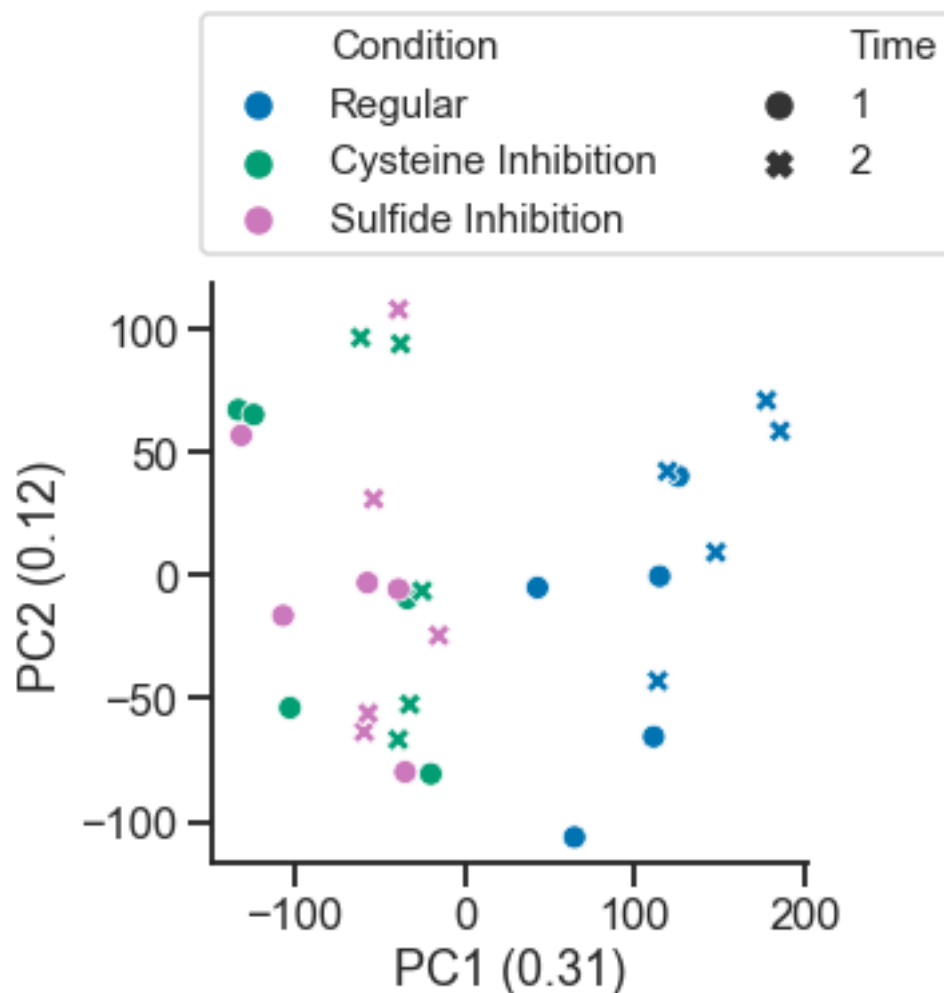
12 Supplementary Figure 4C: PCA Analysis with all features

```
[16]: colors=sns.color_palette('colorblind')
cols = ['Cys_1', 'Cys_1', 'Cys_1', 'Cys_1', 'Cys_1', 'Cys_2', 'Cys_2', 'Cys_2',
        'Cys_2', 'Cys_2', 'Reg_1', 'Reg_1', 'Reg_1', 'Reg_1', 'Reg_1',
        'Reg_2', 'Reg_2', 'Reg_2', 'Reg_2', 'Reg_2', 'Sulf_1', 'Sulf_1',
        'Sulf_1', 'Sulf_1', 'Sulf_1', 'Sulf_2', 'Sulf_2', 'Sulf_2',
        'Sulf_2', 'Sulf_2']
cols = [(x.split('_')[0],x.split('_')[1]) for x in cols]
temp = logarea.copy()
temp.columns = pd.MultiIndex.from_tuples(cols)
temp=temp[['Reg', 'Cys', 'Sulf']]
logarea2 = temp
pca_data =temp.copy().T
pca_data=pca_data.apply(zscore)
pca = PCA(n_components=2)
color=sns.color_palette('colorblind')
c=[colors[0],colors[2],colors[4]]
pca_coord = pca.fit_transform(pca_data)
pca_coord= pd.DataFrame(pca_coord,index=pca_data.index,columns=['PC1', 'PC2'])
pca_coord = pca_coord.reset_index()
pca_coord.columns=['Condition', 'Time', 'PC1', 'PC2']
labels = {'Cys':'Cysteine Inhibition', 'Reg':'Regular', 'Sulf':'Sulfide_
→Inhibition'}
pca_coord['Condition']=pca_coord['Condition'].map(labels)
print('Explained Variance... PC1:%s, PC2:%s'%tuple(pca.
→explained_variance_ratio_))
loadings = pd.DataFrame(pca.components_,index=['PC1', 'PC2'],columns=pca_data.
→columns)

plt.figure(figsize=(4,4))
sns.
→scatterplot(x='PC1',y='PC2',hue='Condition',style='Time',data=pca_coord,palette=c)
plt.legend(bbox_to_anchor=(-.05,1.5),ncol=2,loc=2,fontsize=15)
plt.xlabel('PC1 (%s)%(pca.explained_variance_ratio_[0].round(2))')
plt.ylabel('PC2 (%s)%(pca.explained_variance_ratio_[1].round(2))')
#plt.savefig('Pred_Metabolites_PCA_All.pdf',bbox_inches='tight')
```

```
Explained Variance... PC1:0.30613375327757675, PC2:0.12063218765258428
```

```
[16]: Text(0, 0.5, 'PC2 (0.12)')
```



13 Supplementary Figure 4D: PCA Analysis with features mapping to metabolites

```
[76]: colors=sns.color_palette('colorblind')
cols = ['Cys_1', 'Cys_1', 'Cys_1', 'Cys_1', 'Cys_1', 'Cys_2', 'Cys_2', 'Cys_2',
        'Cys_2', 'Cys_2', 'Reg_1', 'Reg_1', 'Reg_1', 'Reg_1', 'Reg_1',
        'Reg_2', 'Reg_2', 'Reg_2', 'Reg_2', 'Reg_2', 'Sulf_1', 'Sulf_1',
        'Sulf_1', 'Sulf_1', 'Sulf_1', 'Sulf_2', 'Sulf_2', 'Sulf_2',
        'Sulf_2', 'Sulf_2']
cols = [(x.split('_')[0],x.split('_')[1]) for x in cols]
temp = logarea.copy()
temp.columns = pd.MultiIndex.from_tuples(cols)
temp=temp[['Reg','Cys','Sulf']]
logarea2 = temp
```

```

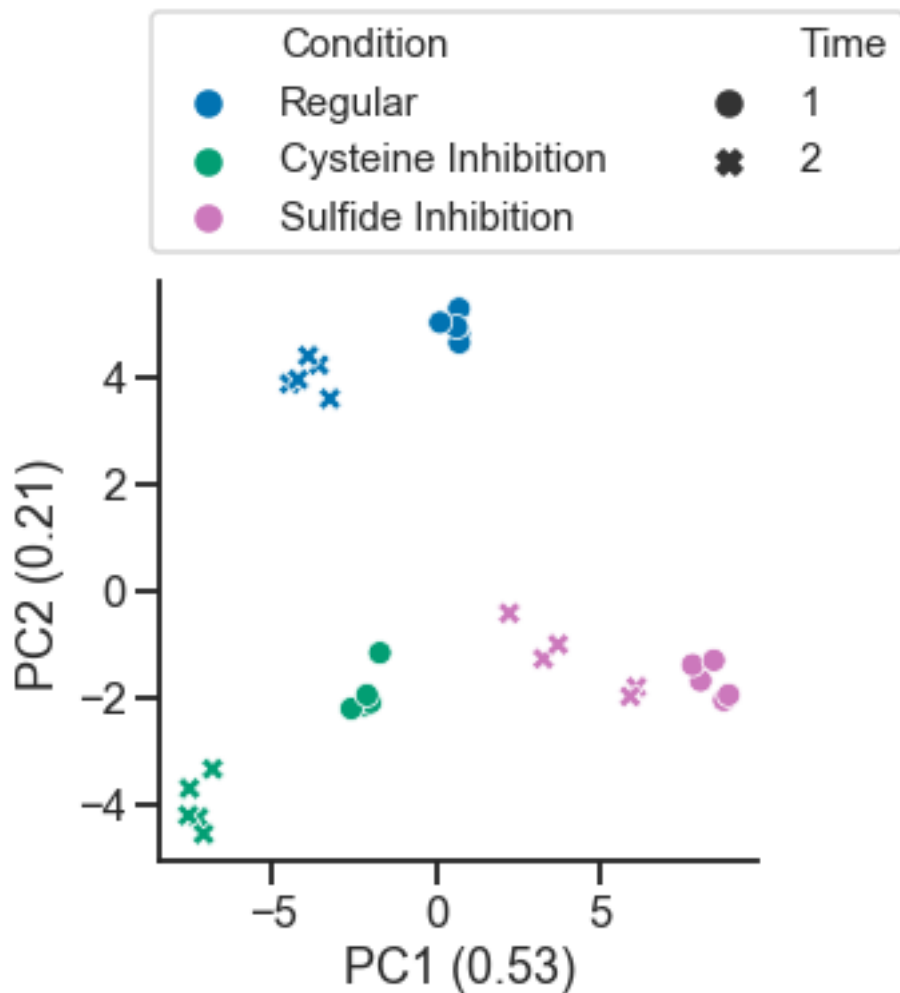
pca_data = temp.loc[idxs,:].copy().T
pca_data=pca_data.apply(zscore)
pca = PCA(n_components=2)
color=sns.color_palette('colorblind')
c=[colors[0],colors[2],colors[4]]
pca_coord = pca.fit_transform(pca_data)
pca_coord= pd.DataFrame(pca_coord,index=pca_data.index,columns=['PC1', 'PC2'])
pca_coord = pca_coord.reset_index()
pca_coord.columns=['Condition', 'Time', 'PC1', 'PC2']
labels = {'Cys': 'Cysteine Inhibition', 'Reg': 'Regular', 'Sulf': 'Sulfide_
↳Inhibition'}
pca_coord['Condition']=pca_coord['Condition'].map(labels)
print('Explained Variance... PC1:%s, PC2:%s'%tuple(pca.
↳explained_variance_ratio_))
loadings = pd.DataFrame(pca.components_,index=['PC1', 'PC2'],columns=pca_data.
↳columns)

plt.figure(figsize=(4,4))
sns.
↳scatterplot(x='PC1',y='PC2',hue='Condition',style='Time',data=pca_coord,palette=c)
plt.legend(bbox_to_anchor=(-.05,1.5),ncol=2,loc=2,fontsize=15)
plt.xlabel('PC1 (%s)%(pca.explained_variance_ratio_[0].round(2)))
plt.ylabel('PC2 (%s)%(pca.explained_variance_ratio_[1].round(2)))
#plt.savefig('Pred_Metabolites_PCA_All.pdf',bbox_inches='tight')

```

Explained Variance... PC1:0.5316343239426855, PC2:0.21420861404263722

[76]: Text(0, 0.5, 'PC2 (0.21)')



14 Obtain feature pathway mapping

```
[155]: idx=pathway.index[:282]
feature_idx=[]
pathmap={}
for i in idx:
    idx_set=pathway.loc[i,:]['overlap_input(featureidx)']
    if type(idx_set)!=type(''):
        continue
    path =pathway.loc[i,:]['pathway']
    for ID in idx_set.split(';'):
        for j in ID.split(','):
            if int(j) not in feature_idx:
                feature_idx.append(int(j))
            if int(j) not in pathmap:
```

```

        pathmap[int(j)]=[path]
    elif path not in pathmap[int(j)]:
        pathmap[int(j)].append(path)

```

pathmap

```

[155]: {68: ['isoleucine biosynthesis I (from threonine)',
            'methionine biosynthesis I',
            'cysteine biosynthesis/homocysteine degradation'],
       234: ['isoleucine biosynthesis I (from threonine)',
            'valine biosynthesis',
            '2-oxopentenoate degradation',
            'phosphopantothenate biosynthesis I',
            'androstenedione degradation'],
       117: ['isoleucine biosynthesis I (from threonine)',
            'valine biosynthesis',
            'incomplete reductive TCA cycle',
            '2-oxopentenoate degradation',
            'respiration (anaerobic)',
            'mixed acid fermentation',
            'methylerythritol phosphate pathway II',
            'gentisate degradation',
            'lysine biosynthesis I',
            'alanine biosynthesis I',
            '4-aminobenzoate biosynthesis',
            'L-ascorbate biosynthesis IV',
            'thiazole biosynthesis I (E. coli)',
            'tryptophan biosynthesis',
            "pyridoxal 5'-phosphate biosynthesis I",
            'gluconeogenesis I',
            'androstenedione degradation',
            'streptomycin biosynthesis',
            'alanine degradation IV',
            'pyruvate fermentation to acetate II',
            'D-fructuronate degradation',
            'pyruvate fermentation to lactate',
            '<i>N</i>-acetylneuraminate and <i>N</i>-acetylmannosamine degradation',
            'pyruvate fermentation to ethanol III',
            'glycolysis II (from fructose 6-phosphate)',
            'glycolysis III (from glucose)',
            'L-serine degradation',
            'L-cysteine degradation II',
            'tryptophan degradation II (via pyruvate)',
            'pyruvate decarboxylation to acetyl CoA',
            'thiazole biosynthesis II (Bacillus)'],
       241: ['isoleucine biosynthesis I (from threonine)',
            'valine biosynthesis',

```

'incomplete reductive TCA cycle',
 '2-oxopentenoate degradation',
 'respiration (anaerobic)',
 'glyoxylate cycle',
 'mixed acid fermentation',
 'methylerythritol phosphate pathway II',
 'gentisate degradation',
 'TCA cycle III (helicobacter)',
 'lysine biosynthesis I',
 'TCA cycle IV (2-oxoglutarate decarboxylase)',
 'TCA cycle V (2-oxoglutarate:ferredoxin oxidoreductase)',
 'alanine biosynthesis I',
 '4-aminobenzoate biosynthesis',
 'thiazole biosynthesis I (E. coli)',
 'tryptophan biosynthesis',
 "pyridoxal 5'-phosphate biosynthesis I",
 'gluconeogenesis I',
 'androstenedione degradation',
 'streptomycin biosynthesis',
 'alanine degradation IV',
 'pyruvate fermentation to acetate II',
 'D-fructuronate degradation',
 'pyruvate fermentation to lactate',
 '<i>N</i>-acetylneuraminate and <i>N</i>-acetylmannosamine degradation',
 'pyruvate fermentation to ethanol III',
 'glycolysis II (from fructose 6-phosphate)',
 'glycolysis III (from glucose)',
 'L-serine degradation',
 'L-cysteine degradation II',
 'tryptophan degradation II (via pyruvate)',
 'pyruvate decarboxylation to acetyl CoA',
 'thiazole biosynthesis II (Bacillus)'],
 174: ['isoleucine biosynthesis I (from threonine)',
 'valine biosynthesis',
 'incomplete reductive TCA cycle',
 '2-oxopentenoate degradation',
 'respiration (anaerobic)',
 'mixed acid fermentation',
 'methylerythritol phosphate pathway II',
 'gentisate degradation',
 'lysine biosynthesis I',
 'alanine biosynthesis I',
 '4-aminobenzoate biosynthesis',
 'L-ascorbate biosynthesis IV',
 'thiazole biosynthesis I (E. coli)',
 'tryptophan biosynthesis',
 "pyridoxal 5'-phosphate biosynthesis I",

'gluconeogenesis I',
 'androstenedione degradation',
 'streptomycin biosynthesis',
 'alanine degradation IV',
 'pyruvate fermentation to acetate II',
 'D-fructuronate degradation',
 'pyruvate fermentation to lactate',
 '<i>N</i>-acetylneuraminate and <i>N</i>-acetylmannosamine degradation',
 'pyruvate fermentation to ethanol III',
 'glycolysis II (from fructose 6-phosphate)',
 'glycolysis III (from glucose)',
 'L-serine degradation',
 'L-cysteine degradation II',
 'tryptophan degradation II (via pyruvate)',
 'pyruvate decarboxylation to acetyl CoA',
 'thiazole biosynthesis II (Bacillus)'],
 197: ['isoleucine biosynthesis I (from threonine)',
 'valine biosynthesis',
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 '2-oxopentenoate degradation',
 'respiration (anaerobic)',
 'mixed acid fermentation',
 'methylerythritol phosphate pathway II',
 'gentisate degradation',
 'lysine biosynthesis I',
 'alanine biosynthesis I',
 '4-aminobenzoate biosynthesis',
 'L-ascorbate biosynthesis IV',
 'thiazole biosynthesis I (E. coli)',
 'tryptophan biosynthesis',
 'pyridoxal 5'-phosphate biosynthesis I",
 'gluconeogenesis I',
 'androstenedione degradation',
 'streptomycin biosynthesis',
 'alanine degradation IV',
 'pyruvate fermentation to acetate II',
 'D-fructuronate degradation',
 'pyruvate fermentation to lactate',
 '<i>N</i>-acetylneuraminate and <i>N</i>-acetylmannosamine degradation',
 'pyruvate fermentation to ethanol III',
 'glycolysis II (from fructose 6-phosphate)',
 'glycolysis III (from glucose)',
 'L-serine degradation',
 'L-cysteine degradation II',
 'tryptophan degradation II (via pyruvate)',
 'pyruvate decarboxylation to acetyl CoA',
 'thiazole biosynthesis II (Bacillus)'],

42: ['isoleucine biosynthesis I (from threonine)',
 'valine biosynthesis',
 'incomplete reductive TCA cycle',
 '2-oxopentenoate degradation',
 'respiration (anaerobic)',
 'mixed acid fermentation',
 'methylerythritol phosphate pathway II',
 'gentisate degradation',
 'lysine biosynthesis I',
 'alanine biosynthesis I',
 '4-aminobenzoate biosynthesis',
 'L-ascorbate biosynthesis IV',
 'thiazole biosynthesis I (E. coli)',
 'tryptophan biosynthesis',
 'pyridoxal 5'-phosphate biosynthesis I',
 'gluconeogenesis I',
 'androstenedione degradation',
 'streptomycin biosynthesis',
 'alanine degradation IV',
 'pyruvate fermentation to acetate II',
 'D-fructuronate degradation',
 'pyruvate fermentation to lactate',
 '<i>N</i>-acetylneuraminate and <i>N</i>-acetylmannosamine degradation',
 'pyruvate fermentation to ethanol III',
 'glycolysis II (from fructose 6-phosphate)',
 'glycolysis III (from glucose)',
 'L-serine degradation',
 'L-cysteine degradation II',
 'tryptophan degradation II (via pyruvate)',
 'pyruvate decarboxylation to acetyl CoA',
 'thiazole biosynthesis II (Bacillus)'],

298: ['isoleucine biosynthesis I (from threonine)',
 'valine biosynthesis',
 'incomplete reductive TCA cycle',
 '2-oxopentenoate degradation',
 'respiration (anaerobic)',
 'mixed acid fermentation',
 'methylerythritol phosphate pathway II',
 'gentisate degradation',
 'lysine biosynthesis I',
 'alanine biosynthesis I',
 '4-aminobenzoate biosynthesis',
 'L-ascorbate biosynthesis IV',
 'thiazole biosynthesis I (E. coli)',
 'tryptophan biosynthesis',
 'pyridoxal 5'-phosphate biosynthesis I',
 'gluconeogenesis I',

'androstenedione degradation',
 'streptomycin biosynthesis',
 'alanine degradation IV',
 'pyruvate fermentation to acetate II',
 'D-fructuronate degradation',
 'pyruvate fermentation to lactate',
 '<i>N</i>-acetylneuraminate and <i>N</i>-acetylmannosamine degradation',
 'pyruvate fermentation to ethanol III',
 'glycolysis II (from fructose 6-phosphate)',
 'glycolysis III (from glucose)',
 'L-serine degradation',
 'L-cysteine degradation II',
 'tryptophan degradation II (via pyruvate)',
 'pyruvate decarboxylation to acetyl CoA',
 'thiazole biosynthesis II (Bacillus)'],
 102: ['isoleucine biosynthesis I (from threonine)',
 'valine biosynthesis',
 'incomplete reductive TCA cycle',
 '2-oxopentenoate degradation',
 'respiration (anaerobic)',
 'mixed acid fermentation',
 'methylerythritol phosphate pathway II',
 'gentisate degradation',
 'lysine biosynthesis I',
 'alanine biosynthesis I',
 '4-aminobenzoate biosynthesis',
 'L-ascorbate biosynthesis IV',
 'thiazole biosynthesis I (E. coli)',
 'tryptophan biosynthesis',
 'pyridoxal 5'-phosphate biosynthesis I',
 'gluconeogenesis I',
 'androstenedione degradation',
 'streptomycin biosynthesis',
 'alanine degradation IV',
 'pyruvate fermentation to acetate II',
 'D-fructuronate degradation',
 'pyruvate fermentation to lactate',
 '<i>N</i>-acetylneuraminate and <i>N</i>-acetylmannosamine degradation',
 'pyruvate fermentation to ethanol III',
 'glycolysis II (from fructose 6-phosphate)',
 'glycolysis III (from glucose)',
 'L-serine degradation',
 'L-cysteine degradation II',
 'tryptophan degradation II (via pyruvate)',
 'pyruvate decarboxylation to acetyl CoA',
 'thiazole biosynthesis II (Bacillus)'],
 34: ['isoleucine biosynthesis I (from threonine)',

'valine biosynthesis',
 'incomplete reductive TCA cycle',
 '2-oxopentenoate degradation',
 'respiration (anaerobic)',
 'mixed acid fermentation',
 'methylerythritol phosphate pathway II',
 'gentisate degradation',
 'lysine biosynthesis I',
 'alanine biosynthesis I',
 '4-aminobenzoate biosynthesis',
 'L-ascorbate biosynthesis IV',
 'thiazole biosynthesis I (E. coli)',
 'tryptophan biosynthesis',
 'pyridoxal 5'-phosphate biosynthesis I",
 'gluconeogenesis I',
 'androstenedione degradation',
 'streptomycin biosynthesis',
 'alanine degradation IV',
 'pyruvate fermentation to acetate II',
 'D-fructuronate degradation',
 'pyruvate fermentation to lactate',
 '<i>N</i>-acetylneuraminate and <i>N</i>-acetylmannosamine degradation',
 'pyruvate fermentation to ethanol III',
 'glycolysis II (from fructose 6-phosphate)',
 'glycolysis III (from glucose)',
 'L-serine degradation',
 'L-cysteine degradation II',
 'tryptophan degradation II (via pyruvate)',
 'pyruvate decarboxylation to acetyl CoA',
 'thiazole biosynthesis II (Bacillus)'],
 213: ['isoleucine biosynthesis I (from threonine)',
 'phosphopantothenate biosynthesis I',
 'lysine fermentation to acetate and butyrate'],
 44: ['isoleucine biosynthesis I (from threonine)',
 'cardiolipin biosynthesis I',
 'phosphopantothenate biosynthesis I',
 'glycerol degradation I',
 'triacylglycerol degradation'],
 179: ['isoleucine biosynthesis I (from threonine)',
 'leucine biosynthesis',
 'isoleucine degradation I'],
 38: ['isoleucine biosynthesis I (from threonine)',
 'valine biosynthesis',
 'phosphopantothenate biosynthesis I',
 'alanine biosynthesis I',
 'leucine biosynthesis',
 'valine degradation I',

'isoleucine degradation I'],
 100: ['cardiolipin biosynthesis I',
 'incomplete reductive TCA cycle',
 'respiration (anaerobic)',
 'glycerol degradation I',
 'aerobic respiration I (cytochrome c)',
 'glyoxylate cycle',
 'mixed acid fermentation',
 'TCA cycle III (helicobacter)',
 'lysine biosynthesis I',
 'TCA cycle IV (2-oxoglutarate decarboxylase)',
 'TCA cycle V (2-oxoglutarate:ferredoxin oxidoreductase)',
 'methionine biosynthesis I',
 'CDP-diacylglycerol biosynthesis I',
 '4-aminobutyrate degradation II',
 'ketolysis',
 'glycerophosphodiester degradation',
 '4-aminobutyrate degradation III',
 '3-oxoadipate degradation',
 '4-aminobutyrate degradation I'],
 303: ['cardiolipin biosynthesis I',
 'glycerol degradation I',
 'CDP-diacylglycerol biosynthesis I',
 'glycerophosphodiester degradation'],
 53: ['cardiolipin biosynthesis I',
 'pyrimidine ribonucleosides salvage I',
 'methylerythritol phosphate pathway II',
 'pyrimidine ribonucleotides salvage',
 'purine and pyrimidine metabolism',
 'CMP phosphorylation',
 'coenzyme A biosynthesis',
 'phosphatidylethanolamine biosynthesis I'],
 161: ['valine biosynthesis',
 '2-oxopentenoate degradation',
 'catechol degradation to 2-oxopent-4-enoate I',
 'catechol degradation to 2-oxopent-4-enoate II',
 'protocatechuate degradation III (<I>para</I>-cleavage pathway)',
 'xylose degradation I'],
 184: ['valine biosynthesis',
 '2-oxopentenoate degradation',
 'threonine degradation III (to methylglyoxal)',
 'xylose degradation I',
 'acrylonitrile degradation I'],
 158: ['valine biosynthesis',
 '2-oxopentenoate degradation',
 'catechol degradation to 2-oxopent-4-enoate I',
 'catechol degradation to 2-oxopent-4-enoate II',

'protocatechuate degradation III (<I>para</I>-cleavage pathway)',
 'xylose degradation I'],
 220: ['incomplete reductive TCA cycle',
 'respiration (anaerobic)',
 'aerobic respiration I (cytochrome c)',
 'glyoxylate cycle',
 'mixed acid fermentation',
 'gentisate degradation',
 'TCA cycle III (helicobacter)',
 'TCA cycle IV (2-oxoglutarate decarboxylase)',
 'TCA cycle V (2-oxoglutarate:ferredoxin oxidoreductase)',
 'arginine biosynthesis II (acetyl cycle)',
 'gluconeogenesis I',
 'inosine-5'-phosphate biosynthesis II",
 'adenosine ribonucleotides <i>de novo</i> biosynthesis',
 'citrulline-nitric oxide cycle',
 "inosine-5'-phosphate biosynthesis I",
 'urea cycle',
 'arginine biosynthesis IV (archaebacteria)'],
 239: ['catechol degradation to 2-oxopent-4-enoate I',
 "pyridoxal 5'-phosphate salvage pathway",
 'catechol degradation to 2-oxopent-4-enoate II',
 'protocatechuate degradation III (<I>para</I>-cleavage pathway)'],
 27: ['threonine degradation III (to methylglyoxal)',
 'threonine degradation II'],
 289: ['pyrimidine ribonucleosides salvage I',
 'UMP biosynthesis',
 'pyrimidine ribonucleotides salvage',
 'purine and pyrimidine metabolism',
 'UTP and CTP biosynthesis',
 'pseudouridine degradation',
 'pyrimidine nucleobases salvage I'],
 115: ['UDP-<i>N</i>-acetyl-α-D-mannosaminouronate biosynthesis',
 'UDP-<i>N</i>-acetyl-D-glucosamine biosynthesis I',
 'CMP-pseudamate biosynthesis',
 'anhydromuropeptides recycling',
 'mycothiol biosynthesis',
 'UDP-<i>N</i>-acetylmuramoyl-pentapeptide biosynthesis II (lysine-
 containing)',
 'UDP-<i>N</i>-acetylmuramoyl-pentapeptide biosynthesis III (<i>meso</i>-DAP-
 containing)',
 'CMP-<i>N</i>-acetylneuraminate biosynthesis II (bacteria)'],
 296: ['UDP-<i>N</i>-acetyl-α-D-mannosaminouronate biosynthesis',
 'UDP-<i>N</i>-acetyl-D-glucosamine biosynthesis I',
 'CMP-pseudamate biosynthesis',
 'anhydromuropeptides recycling',
 'mycothiol biosynthesis',

'UDP-<i>N</i>-acetylmuramoyl-pentapeptide biosynthesis II (lysine-containing)',
 'UDP-<i>N</i>-acetylmuramoyl-pentapeptide biosynthesis III (<i>meso</i>-DAP-containing)',
 'CMP-<i>N</i>-acetylneuraminate biosynthesis II (bacteria)'],
 110: ['methylerythritol phosphate pathway II'],
 113: ["pyridoxal 5'-phosphate salvage pathway"],
 111: ['biotin biosynthesis from 7-keto-8-aminopelargonate'],
 242: ['biotin biosynthesis from 7-keto-8-aminopelargonate',
 '4-amino-2-methyl-5-diphosphomethylpyrimidine biosynthesis',
 'thiazole biosynthesis I (E. coli)',
 'cyclopropane fatty acid (CFA) biosynthesis',
 'factor 420 biosynthesis',
 'menaquinol-6 biosynthesis',
 '<i>S</i>-adenosyl-L-methionine cycle II',
 'menaquinol-10 biosynthesis',
 'lipolate biosynthesis and incorporation I',
 'menaquinol-11 biosynthesis',
 'S-adenosyl-L-methionine biosynthesis',
 'heme biosynthesis from uroporphyrinogen-III II',
 'cob(II)yrinate <i>a,c</i>-diamide biosynthesis I (early cobalt insertion)',
 'chlorophyllide <i>a</i> biosynthesis III',
 'chlorophyllide <i>a</i> biosynthesis I',
 'methionine degradation I (to homocysteine)',
 'lipolate biosynthesis and incorporation III (Bacillus)',
 'menaquinol-8 biosynthesis',
 'chlorophyllide <i>a</i> biosynthesis II (anaerobic)',
 'menaquinol-9 biosynthesis',
 'cob(II)yrinate <i>a,c</i>-diamide biosynthesis II (late cobalt incorporation)'],
 39: ['lysine biosynthesis I'],
 21: ['lysine biosynthesis I',
 'arginine biosynthesis II (acetyl cycle)',
 'ornithine biosynthesis'],
 86: ['4-amino-2-methyl-5-diphosphomethylpyrimidine biosynthesis',
 'hydroxymethylpyrimidine salvage',
 'thiamin salvage IV (yeast)'],
 188: ['4-aminobenzoate biosynthesis',
 'tryptophan biosynthesis',
 'tyrosine biosynthesis I',
 'chorismate biosynthesis from 3-dehydroquinate'],
 262: ['methionine biosynthesis I',
 'taxiphyllin bioactivation',
 'thiosulfate disproportionation III (rhodanese)'],
 247: ['UMP biosynthesis'],
 171: ['UDP-<i>N</i>-acetyl-D-glucosamine biosynthesis I',
 'anhydromuropeptides recycling',

```

'streptomycin biosynthesis'],
49: ['arginine biosynthesis II (acetyl cycle)', 'ornithine biosynthesis'],
153: ["pyridoxal 5'-phosphate biosynthesis I"],
96: ["pyridoxal 5'-phosphate biosynthesis I"],
210: ['purine and pyrimidine metabolism',
'dTMP <i>de novo</i> biosynthesis (mitochondrial)',
'pyrimidine deoxyribonucleosides salvage',
'pyrimidine deoxyribonucleotides <i>de novo</i> biosynthesis I',
'pyrimidine deoxyribonucleotides <i>de novo</i> biosynthesis II',
'salvage pathways of pyrimidine deoxyribonucleotides',
'<i>N</i><sup>10</sup>-formyl-tetrahydrofolate biosynthesis',
'pyrimidine deoxyribonucleotide phosphorylation'],
101: ['CMP-pseudamate biosynthesis'],
233: ['CMP-pseudamate biosynthesis'],
67: ['lysine fermentation to acetate and butyrate'],
71: ['lysine fermentation to acetate and butyrate'],
254: ['xanthine and xanthosine salvage',
'urate biosynthesis/inosine 5'-phosphate degradation',
'purine ribonucleosides degradation'],
215: ['lysine degradation VI'],
312: ['oleate &beta;-oxidation'],
280: ['dTDP-L-rhamnose biosynthesis I'],
79: ['sulfate activation for sulfonation'],
105: ['palmitate biosynthesis II (bacteria and plants)']]

```

```

[ ]: def kmeans_cluster(df,k):
    X = df.values
    km = KMeans(n_clusters=k, random_state=0)
    return km.fit_predict(X)

```

15 Figure 3A: Heatmap of feature z-scores for features mapping to metabolites

```

[18]: df = pca_data.T.groupby(level=[0,1],axis=1).mean().loc[idxs,:].copy()
df=df[['Reg','Cys','Sulf']]
paths=['isoleucine biosynthesis I (from threonine)','valine_
→biosynthesis','glycerol degradation I','glyoxylate cycle','TCA cycle III_
→(helicobacter)','TCA cycle IV (2-oxoglutarate decarboxylase)']
colors = sns.color_palette('colorblind')
i=0
path_clr = {'None':(1,1,1)}
row_clr=[]
for p in paths:
    path_clr[p]=colors[i]
    row_clr.append(pd.Series(index=df.index,data=np.nan))
    for idx in df.index:

```

```

        if p in pathmap[idx]:
            row_clrs[i][idx]=p
        else:
            row_clrs[i][idx]='None'
    row_clrs[i]=row_clrs[i].map(path_clr)

    i+=1

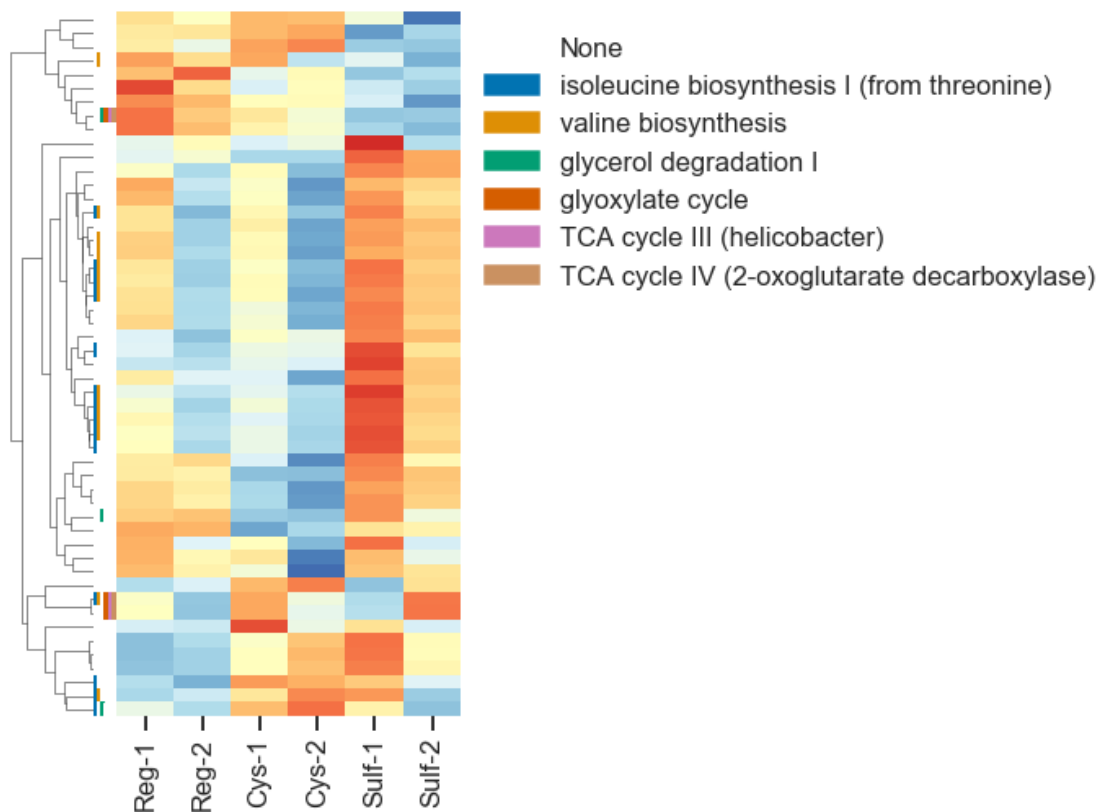
#If using plt.cm.tab20 then colors[i] must be changed to colors(i)
handles = [mpatches.Patch(color=path_clr[i], label=i) for i in path_clr.keys()]

#Creat heatmap
clust = sns.
    ↳ clustermap(df, cmap='RdYlBu', yticklabels='', figsize=(5,10), col_cluster=False, vmax=-2.
    ↳ 5, vmin=2.5, row_colors=row_clrs)
clust.ax_heatmap.set(ylabel='', xlabel='')
clust.cax.set_visible(False)

clust.ax_heatmap.
    ↳ legend(bbox_to_anchor=(1,1), loc=2, handles=handles, frameon=False)
#plt.savefig('Pred-Metabolite-Heatmap.png', bbox_inches='tight', dpi=500)

```

[18]: <matplotlib.legend.Legend at 0x1a241d7e48>



16 Figure 4B: Putative features abundances in BCAA

```
[19]: names=['CPD-15056', '1-KETO-2-METHYLVALERATE', '2-KETO-3-METHYL-VALERATE', '2-ACETO-2-HYDROXY-BUTYRATE']
index=[]
name_dict={}
for name in names:
    if type(cmp_dict[name]) == type([]):
        idx=cmp_dict[name]
    else:
        idx=[cmp_dict[name]]

    index+=idx
    for i in idx:
        if i not in name_dict:
            name_dict[i]=name
```

```

else:
    name_dict[i]=name_dict[i] + '/' + name
name_dict

```

```

[19]: {68: 'CPD-15056',
      213: '1-KETO-2-METHYLVALERATE',
      44: '1-KETO-2-METHYLVALERATE/GLYCEROL',
      179: '2-KETO-3-METHYL-VALERATE',
      38: '2-KETO-3-METHYL-VALERATE/2-KETO-ISOVALERATE',
      234: '2-ACETO-2-HYDROXY-BUTYRATE/2-ACETO-LACTATE',
      161: '2-ACETO-LACTATE',
      184: '2-ACETO-LACTATE',
      158: '2-ACETO-LACTATE',
      220: 'MAL',
      241: 'MAL',
      100: 'GLYCEROL-3P',
      303: 'GLYCEROL-3P'}

```

```

[157]: multi=[]
for cond in ['cys','reg','sulf']:
    for t in ['T1','T2']:
        for rep in range(1,6):
            multi.append((cond,t,rep))

relabund=(np.exp(logarea)-1).copy()
relabund.columns=pd.MultiIndex.from_tuples(multi)
relabund=relabund/relabund.sum(axis=0)
relabund.sum(axis=0)
relabund

```

```

[157]:

```

		cys				
		T1				
		1	2	3	4	5
featureidx						
2	0.000003	1.534291e-06	2.482685e-06	0.000003	0.000005	
3	0.000005	3.164120e-06	6.018955e-06	0.000005	0.000010	
4	0.000002	9.313779e-07	1.868791e-06	0.000002	0.000004	
5	0.000002	2.161251e-06	3.400159e-06	0.000004	0.000006	
6	0.000001	7.828048e-07	1.299537e-06	0.000001	0.000002	
...	
29983	0.000009	6.902365e-06	2.084933e-06	0.000004	0.000008	
29984	0.000008	6.814050e-06	1.628057e-06	0.000004	0.000007	
29985	0.000021	5.139379e-07	1.669532e-05	0.000026	0.000028	
29986	0.000014	1.086642e-05	1.330217e-06	0.000003	0.000009	
29987	0.000005	4.113982e-06	9.581834e-07	0.000002	0.000005	

	T2			
	1	2	3	4
featureidx				
2	4.933268e-08	2.033698e-08	2.201864e-08	2.680995e-08
3	7.142940e-07	7.961235e-07	8.298417e-07	8.040248e-07
4	2.694299e-08	9.586718e-09	5.254944e-08	3.257441e-08
5	1.839371e-07	1.307271e-07	1.542928e-07	2.095020e-07
6	1.333526e-07	1.439539e-07	1.327211e-07	1.715190e-07
...
29983	5.988915e-06	1.520856e-05	2.217725e-06	3.786984e-06
29984	5.200672e-06	1.541116e-05	1.828297e-06	3.430294e-06
29985	2.362840e-05	9.235334e-06	1.443436e-05	2.176989e-05
29986	5.687856e-06	2.549465e-05	1.635998e-06	3.668759e-06
29987	3.036996e-06	8.714433e-06	8.172636e-07	1.693315e-06

		...	sulf		
		...	T1		
	5	...	1	2	3
featureidx		...			
2	7.573280e-08	...	3.746099e-07	7.089190e-07	1.428266e-06
3	8.328074e-07	...	1.300160e-06	1.676210e-06	3.362990e-06
4	1.207442e-08	...	1.872636e-07	4.722305e-07	1.119212e-06
5	2.060366e-07	...	6.861787e-07	1.014798e-06	1.863191e-06
6	1.295445e-07	...	2.969920e-07	3.739778e-07	5.865295e-07
...
29983	1.342991e-05	...	9.429891e-06	4.597117e-06	4.962160e-06
29984	1.315275e-05	...	8.450384e-06	3.976927e-06	4.409204e-06
29985	3.881321e-05	...	4.826891e-06	2.511047e-05	2.450187e-05
29986	2.108254e-05	...	1.522642e-05	3.808118e-06	5.024457e-06
29987	7.831434e-06	...	4.933397e-06	2.087852e-06	2.772300e-06

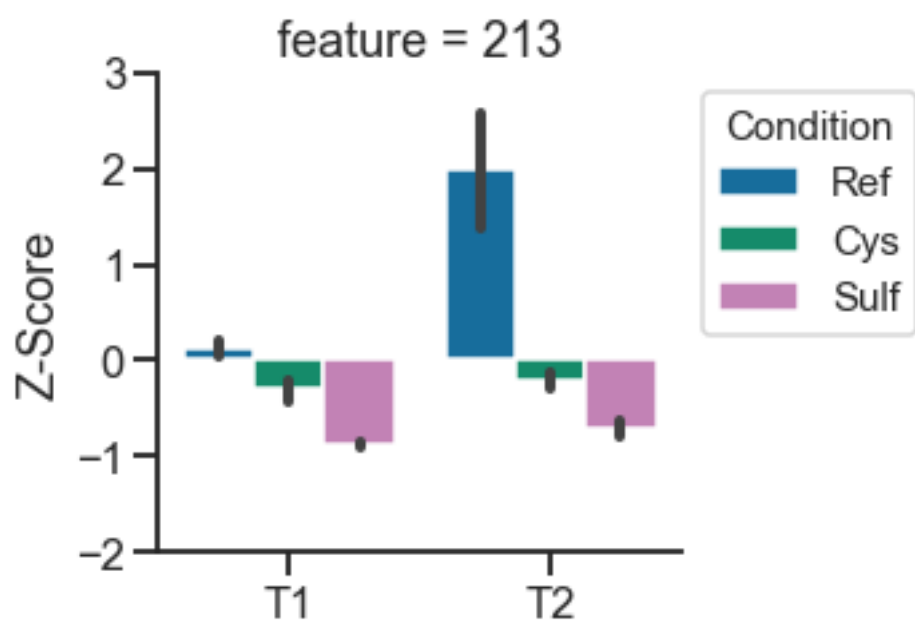
			T2		
	4	5	1	2	3
featureidx					
2	1.146211e-06	1.661544e-06	0.000005	0.000005	0.000004
3	2.933160e-06	3.578576e-06	0.000008	0.000009	0.000009
4	1.019705e-06	1.165723e-06	0.000004	0.000003	0.000003
5	1.705198e-06	2.091126e-06	0.000006	0.000006	0.000006
6	7.060791e-07	8.142010e-07	0.000002	0.000002	0.000002
...
29983	3.894884e-06	7.946109e-06	0.000006	0.000004	0.000003
29984	2.761636e-06	7.664362e-06	0.000007	0.000002	0.000003
29985	1.769551e-05	2.161258e-05	0.000021	0.000022	0.000021
29986	2.478806e-06	9.763429e-06	0.000009	0.000002	0.000002
29987	1.735353e-06	4.681973e-06	0.000004	0.000001	0.000001

	4	5
featureidx		
2	0.000006	0.000008
3	0.000010	0.000014
4	0.000004	0.000006
5	0.000007	0.000010
6	0.000002	0.000003
...
29983	0.000007	0.000014
29984	0.000005	0.000014
29985	0.000021	0.000012
29986	0.000007	0.000024
29987	0.000003	0.000008

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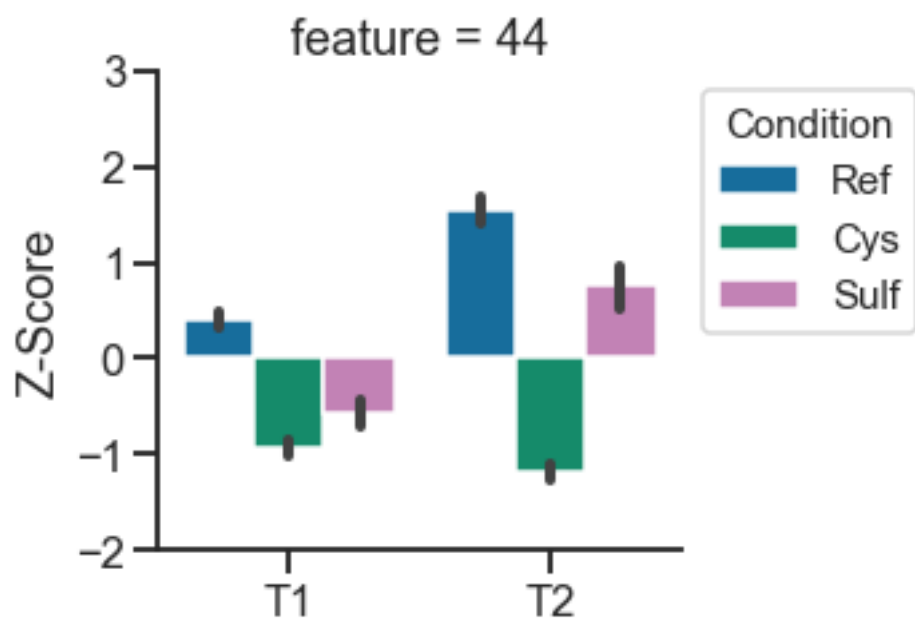
```
[21]: temp = relabund.T.apply(zscore).T.loc[index,].copy()
df = temp.unstack()
df = df.reset_index()
df['name'] = df['featureidx'].map(name_dict)
df.columns = ['cond', 't', 'rep', 'feature', 'val', 'name']
df['cond'] = df['cond'].map({'reg': 'Ref', 'cys': 'Cys', 'sulf': 'Sulf'})
c = sns.color_palette('colorblind')
c = [c[0], c[2], c[4]]
for name, df2 in df.groupby(by=['name']):
    print(name)
    g = sns.catplot(x="t", y="val",
                    hue="cond", col="feature", hue_order=['Ref', 'Cys', 'Sulf'],
                    data=df2,
    → kind="bar", palette=c, legend=False, sharey=False, height=4)
    g.axes[0][0].set_ylabel('Z-Score')
    for ax in g.axes[0]:
        ax.set_xlabel('')
        ax.set_yticks([-2, -1, 0, 1, 2, 3])
    leg = plt.legend(bbox_to_anchor=(1, 1), loc=2, title='Condition', fontsize=15)
    leg.get_title().set_fontsize('15')
    #plt.savefig('%s-box.pdf'%(+'.join(name.split('/'))), bbox_inches='tight')
    plt.show()
    plt.clf()
```

1-KETO-2-METHYLVALERATE



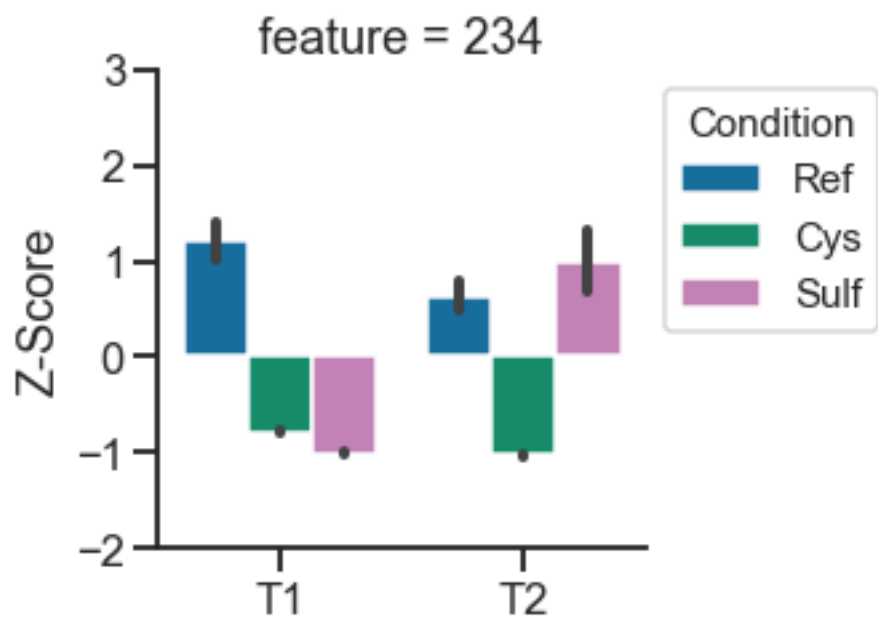
1-KETO-2-METHYLVALERATE/GLYCEROL

<Figure size 432x288 with 0 Axes>



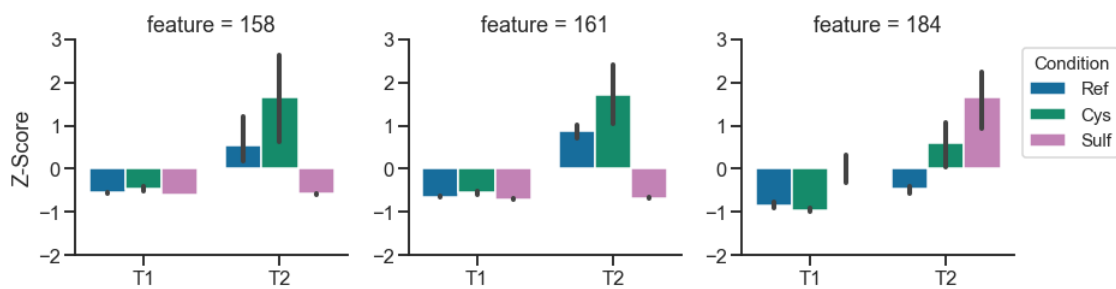
2-ACETO-2-HYDROXY-BUTYRATE/2-ACETO-LACTATE

<Figure size 432x288 with 0 Axes>



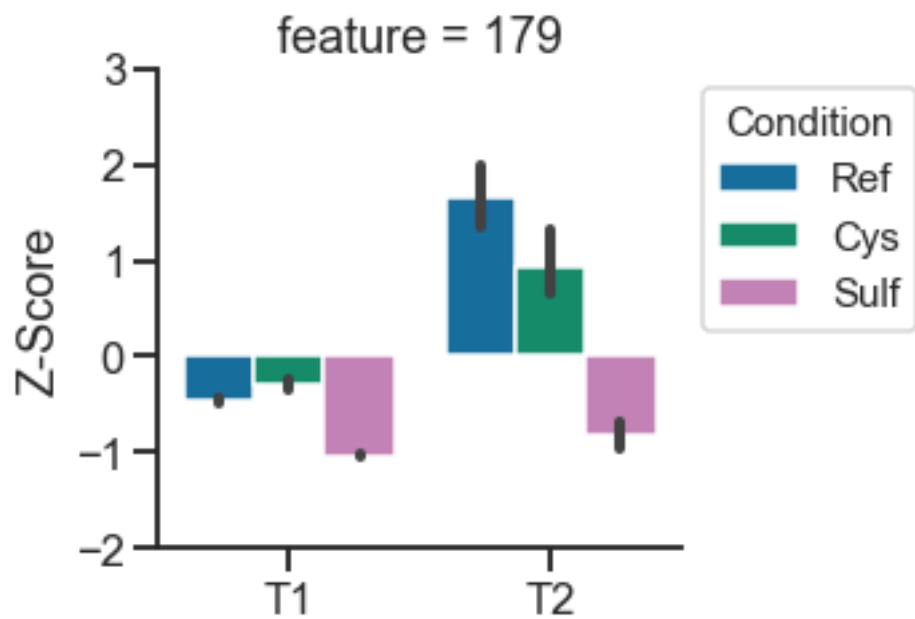
2-ACETO-LACTATE

<Figure size 432x288 with 0 Axes>



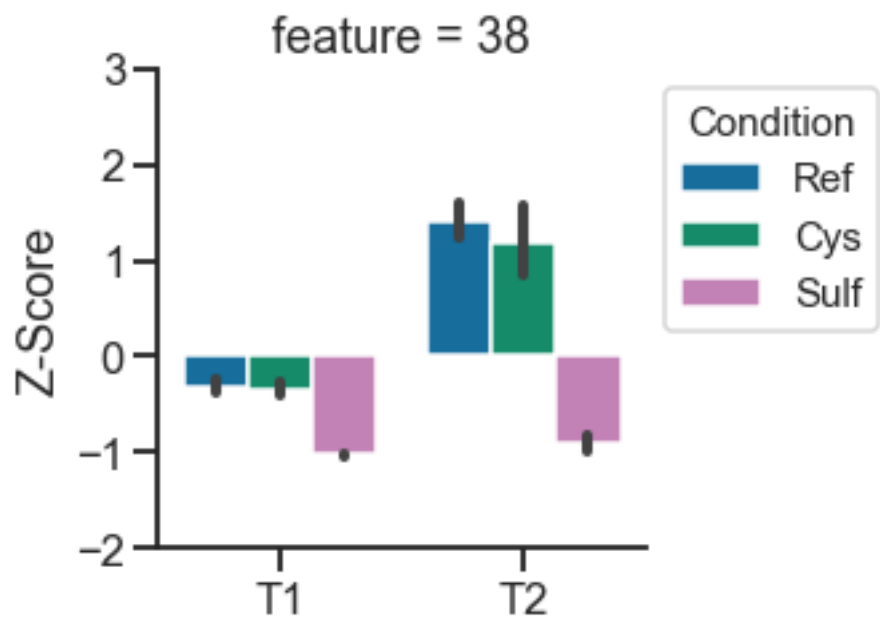
2-KETO-3-METHYL-VALERATE

<Figure size 432x288 with 0 Axes>



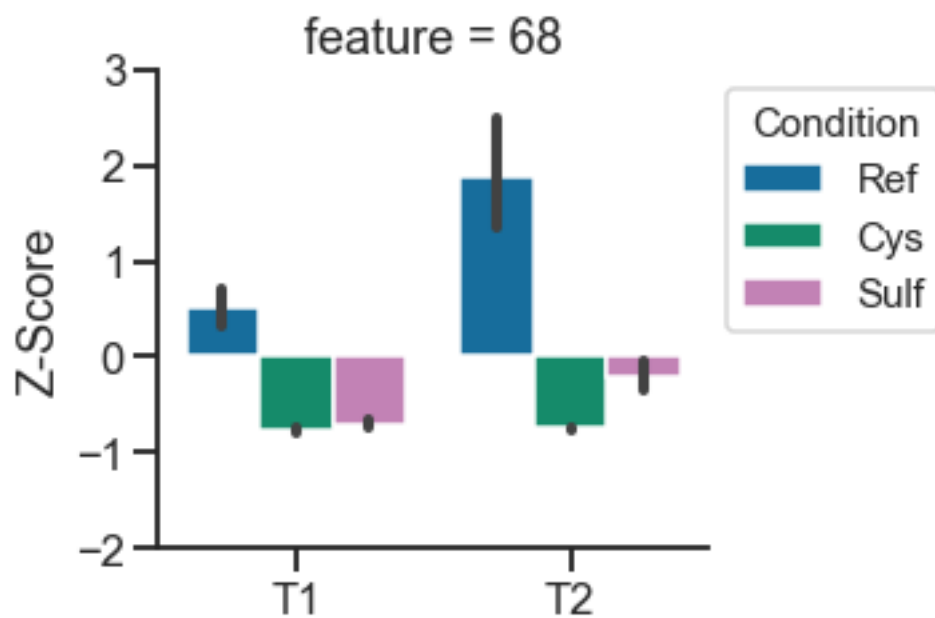
2-KETO-3-METHYL-VALERATE/2-KETO-ISOVALERATE

<Figure size 432x288 with 0 Axes>



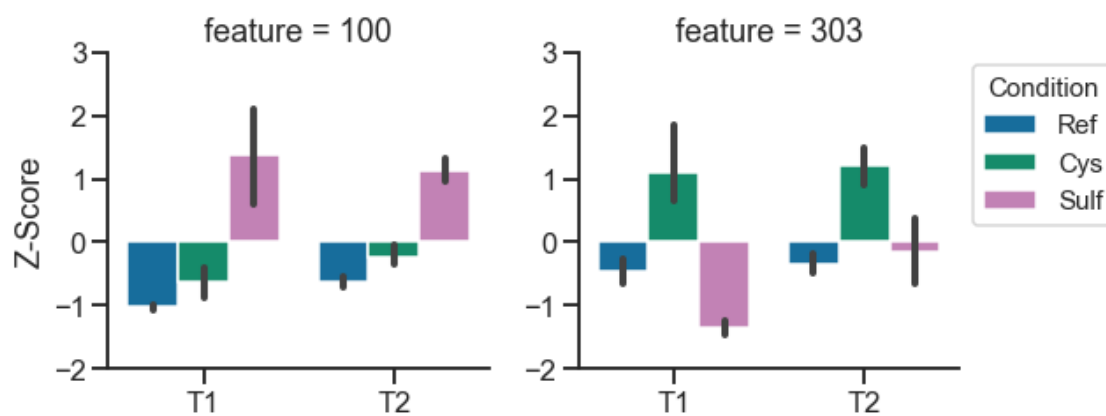
CPD-15056

<Figure size 432x288 with 0 Axes>



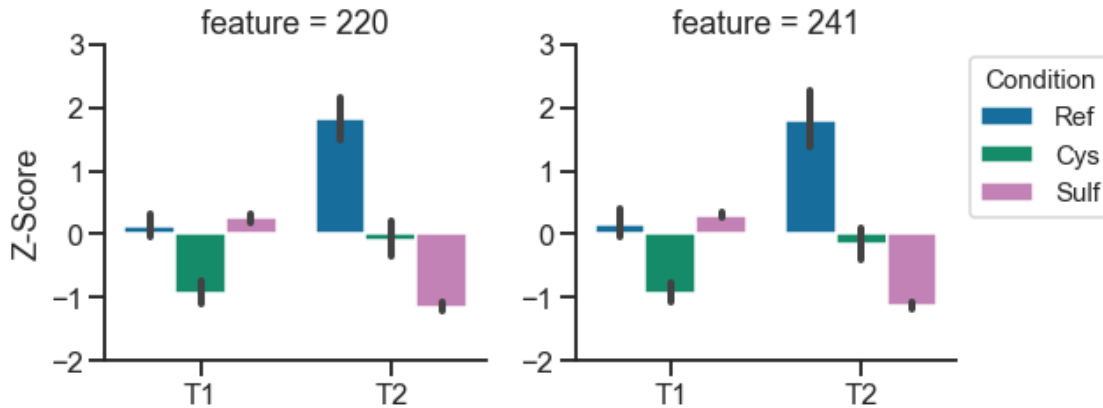
GLYCEROL-3P

<Figure size 432x288 with 0 Axes>



MAL

<Figure size 432x288 with 0 Axes>



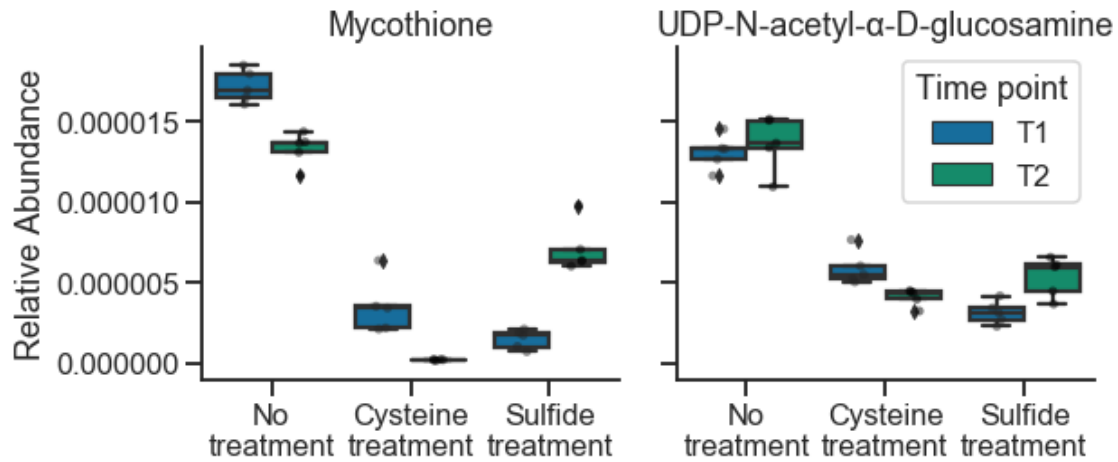
<Figure size 432x288 with 0 Axes>

17 Supplementary Figure 5: Mycothiol

```
[166]: df=pd.DataFrame(relabund.loc[[5,115, 296],:])
df=df.T.reset_index()
df.columns=['cond','time','rep','5','115','296']
fig,(ax1,ax2)=plt.subplots(1,2,figsize=(9,4),sharey=True)
c=sns.color_palette('colorblind')

g=sns.
    ↳boxplot(x='cond',y='5',hue='time',data=df,palette=[c[0],c[2],c[4]],order=['reg','cys','sulf']
sns.
    ↳stripplot(x='cond',y='5',hue='time',data=df,dodge=True,color='black',alpha=0.
    ↳5,order=['reg','cys','sulf'],ax=ax1)
g.set_ylabel('Relative Abundance')
g.set_xticklabels(['No\ntreatment','Cysteine\ntreatment','Sulfide\ntreatment'])
g.set_title('Mycothione')
g.set_xlabel('')
g.legend().set_visible(False)
g=sns.
    ↳boxplot(x='cond',y='296',hue='time',data=df,palette=[c[0],c[2],c[4]],order=['reg','cys','su
sns.
    ↳stripplot(x='cond',y='296',hue='time',data=df,dodge=True,color='black',alpha=0.
    ↳5,order=['reg','cys','sulf'],ax=ax2)
g.set_ylabel('')
g.set_xlabel('')
handles,labels=g.get_legend_handles_labels()
g.legend(handles[:3],['T1','T2'],bbox_to_anchor=(1,1),title='Time point')
g.set_xticklabels(['No\ntreatment','Cysteine\ntreatment','Sulfide\ntreatment'])
```

```
g.set_title('UDP-N-acetyl- -D-glucosamine')
fig.tight_layout()
plt.savefig('Mycothione-UPD-Relabund.pdf',bbox_inches='tight')
```



18 Supplementary Figure 4 B

```
[ ]: data=np.exp(logarea2)-1
data=data/data.sum()
cys_pvals=[]
sulf_pvals=[]
cys_sulf_pvals=[]
c=sns.color_palette('colorblind')
for i in idxs:
    df=data.loc[i].reset_index()
    plt.figure(figsize=(4,2.5))
    g=sns.boxplot(y='level_0',x=i,data=df)
    g=sns.
    ↳boxplot(y='level_0',x=i,data=df,palette=[c[0],c[2],c[4]],order=['Reg','Cys','Sulf'])
    sns.stripplot(y='level_0',x=i,data=df,color='black',alpha=0.
    ↳5,order=['Reg','Cys','Sulf'])

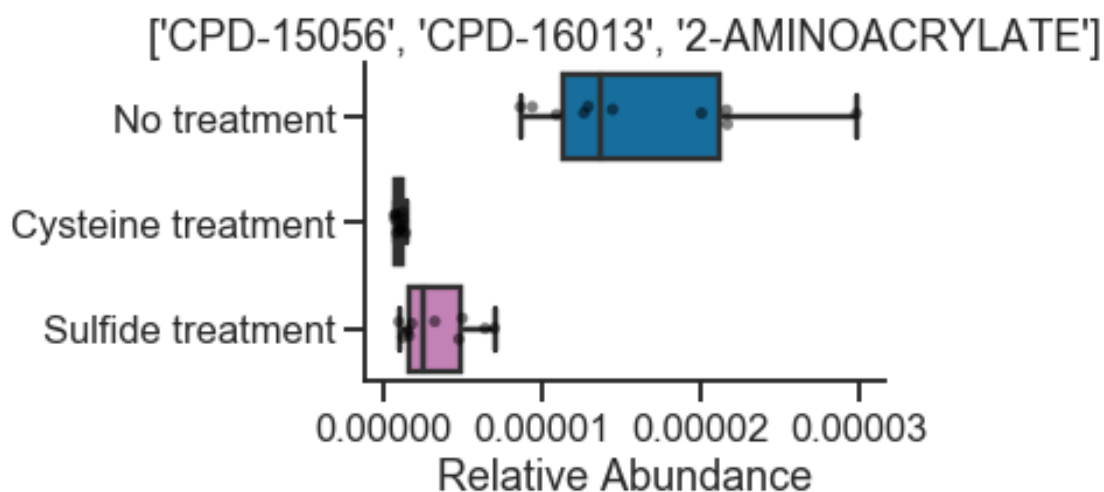
    g.set_ylabel('')
    g.set_xlabel('Relative Abundance')
    #g.set_xlim(-3.5,3.5)
    g.set_yticklabels(['No treatment','Cysteine treatment','Sulfide_
    ↳treatment'])
    reg=df[df['level_0']=='Reg'][i].values
    cys=df[df['level_0']=='Cys'][i].values
    sulf=df[df['level_0']=='Sulf'][i].values
```

```

cys_pval=ttest_ind(reg,cys,equal_var=False).pvalue
sulf_pval=ttest_ind(reg,sulf,equal_var=False).pvalue
cys_sulf_pval=ttest_ind(cys,sulf,equal_var=False).pvalue
cys_pvals.append(cys_pval)
sulf_pvals.append(sulf_pval)
cys_sulf_pvals.append(cys_sulf_pval)
g.set_title(rev_cmp_dict[i])
print(i)
#plt.savefig('metabolomics-feat-%s-boxplots.pdf'%(i),bbox_inches='tight')
plt.show()
plt.clf()
cys_pvals=multipletests(cys_pvals, method='fdr_bh',alpha=0.01)[:2][1]
sulf_pvals=multipletests(sulf_pvals, method='fdr_bh',alpha=0.01)[:2][1]
cys_sulf_pvals=multipletests(cys_sulf_pvals, method='fdr_bh',alpha=0.01)[:2][1]

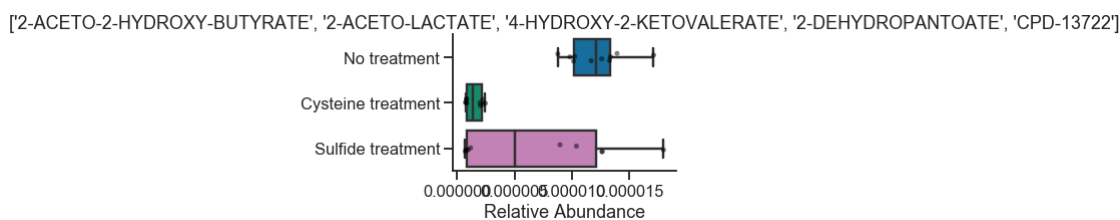
```

68



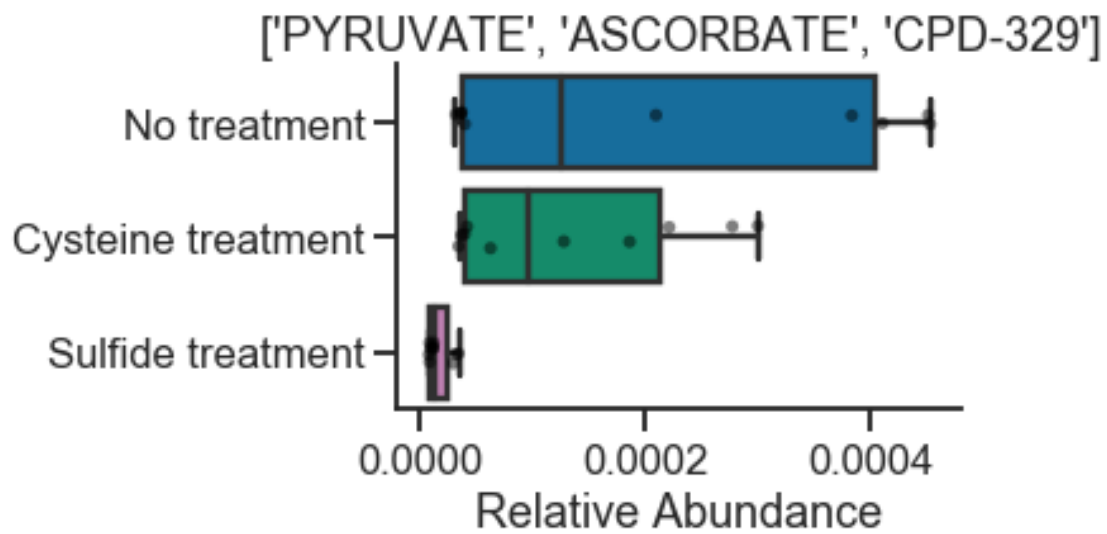
234

<Figure size 432x288 with 0 Axes>



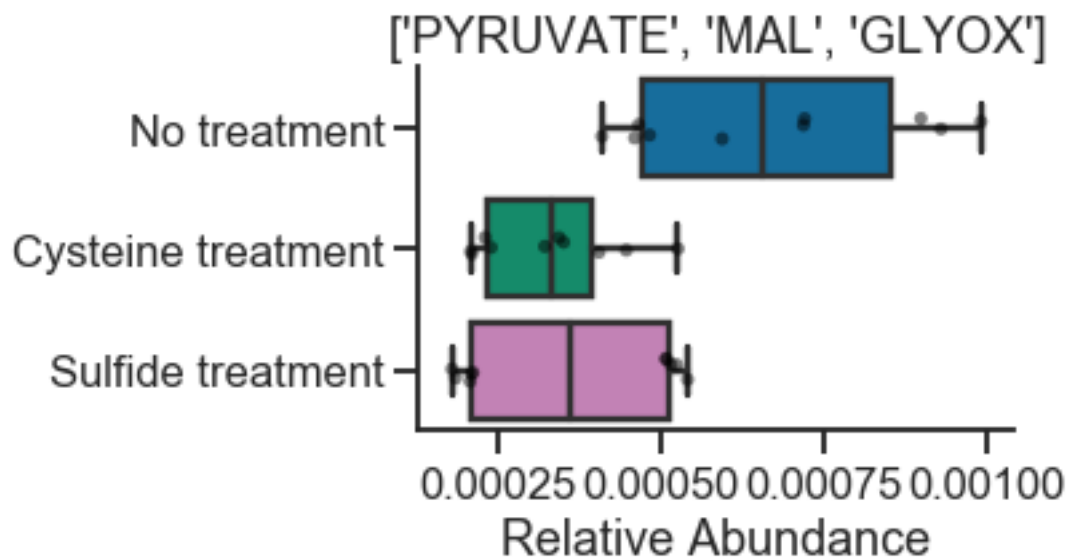
117

<Figure size 432x288 with 0 Axes>



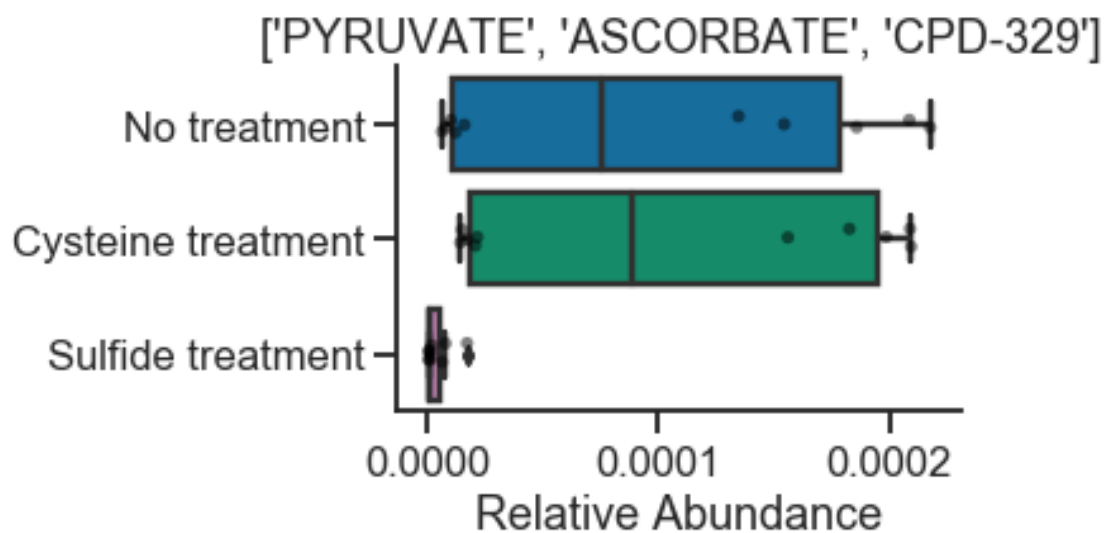
241

<Figure size 432x288 with 0 Axes>



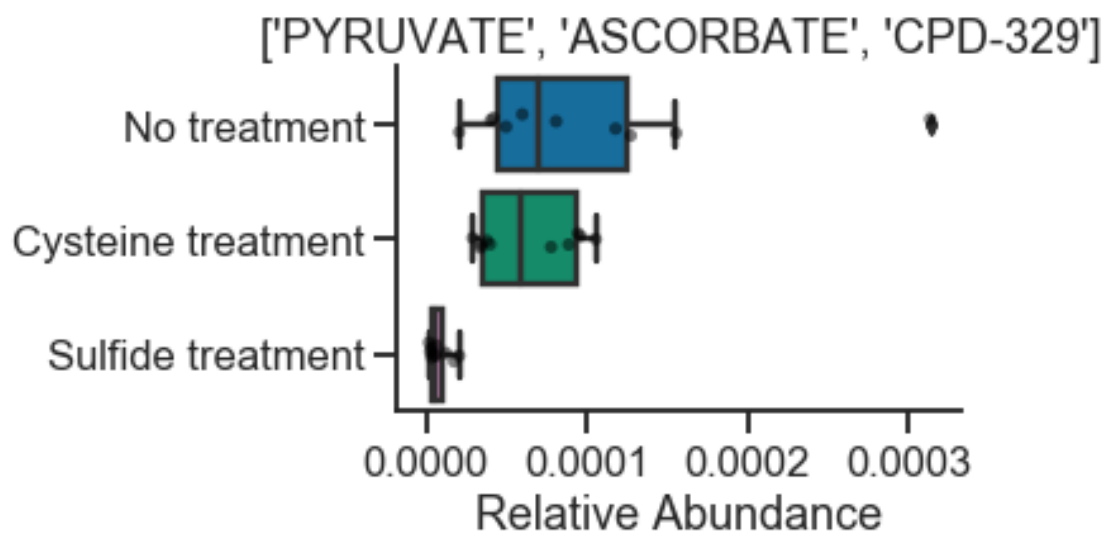
174

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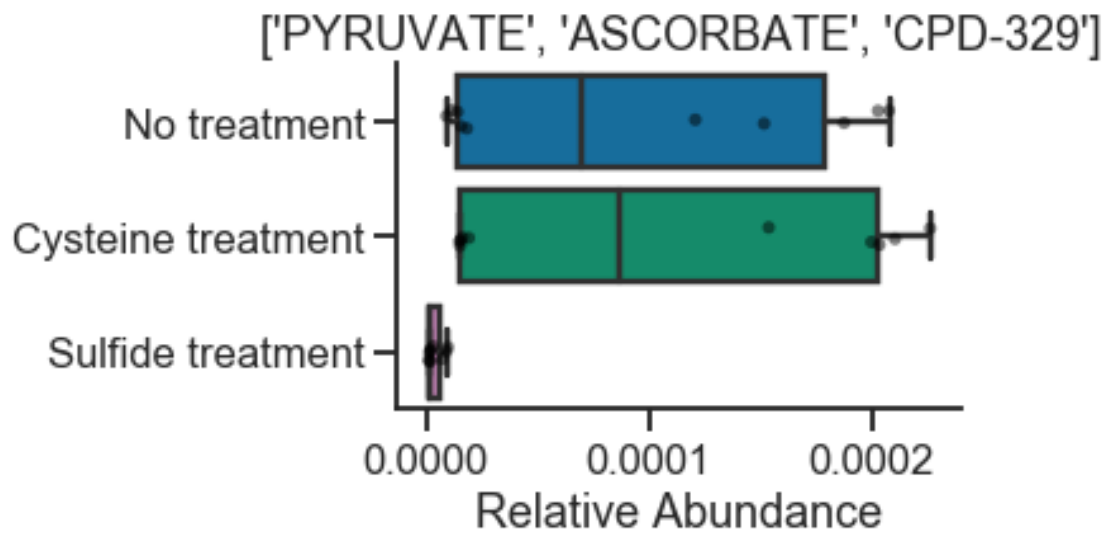
197

<Figure size 432x288 with 0 Axes>



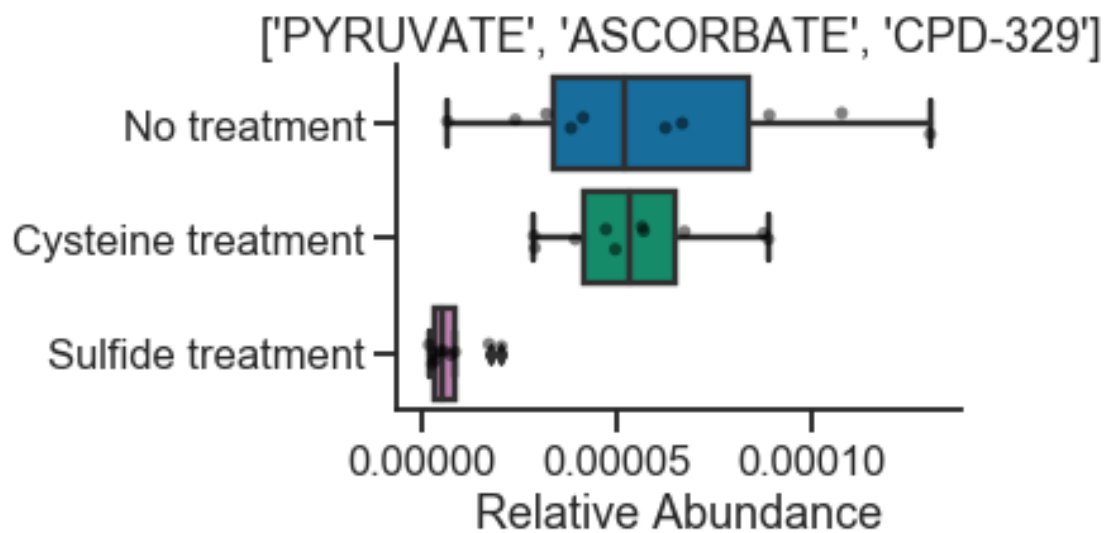
42

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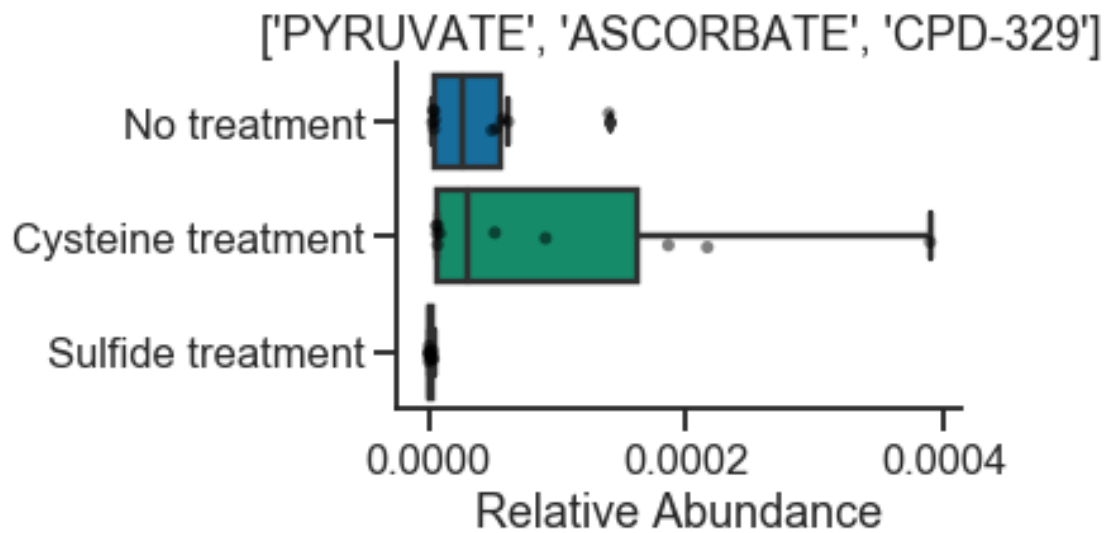
298

<Figure size 432x288 with 0 Axes>



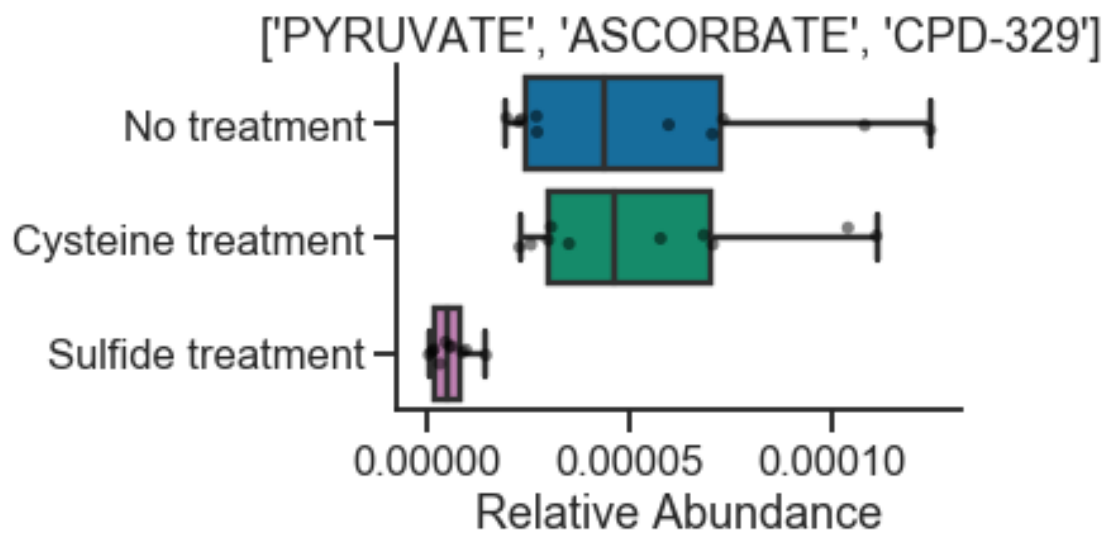
102

<Figure size 432x288 with 0 Axes>



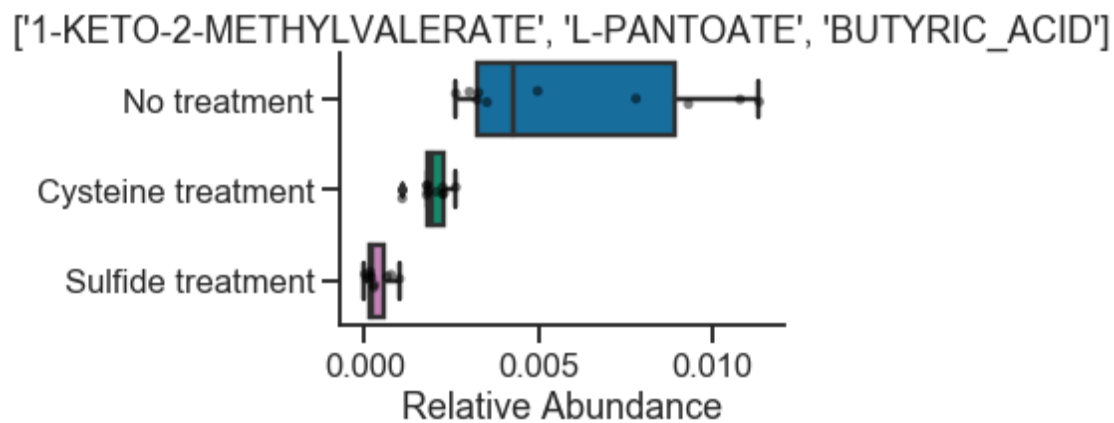
34

<Figure size 432x288 with 0 Axes>



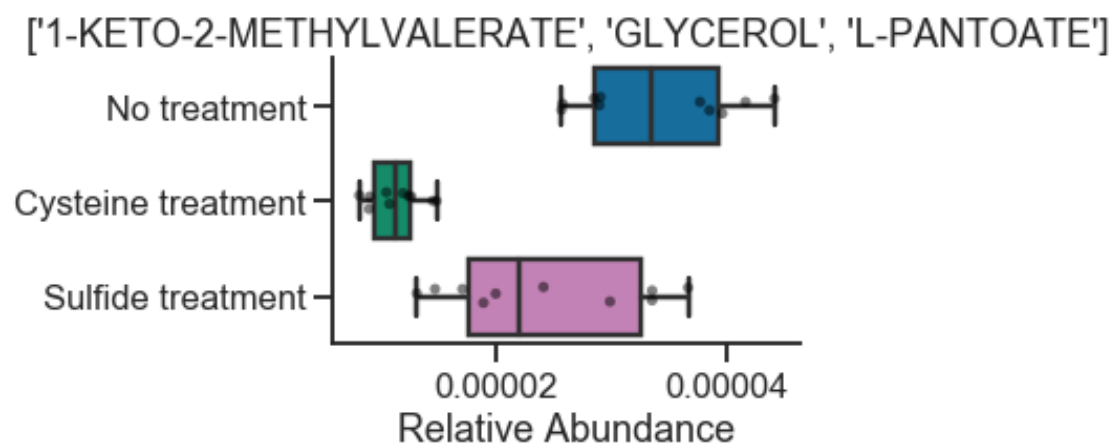
213

<Figure size 432x288 with 0 Axes>



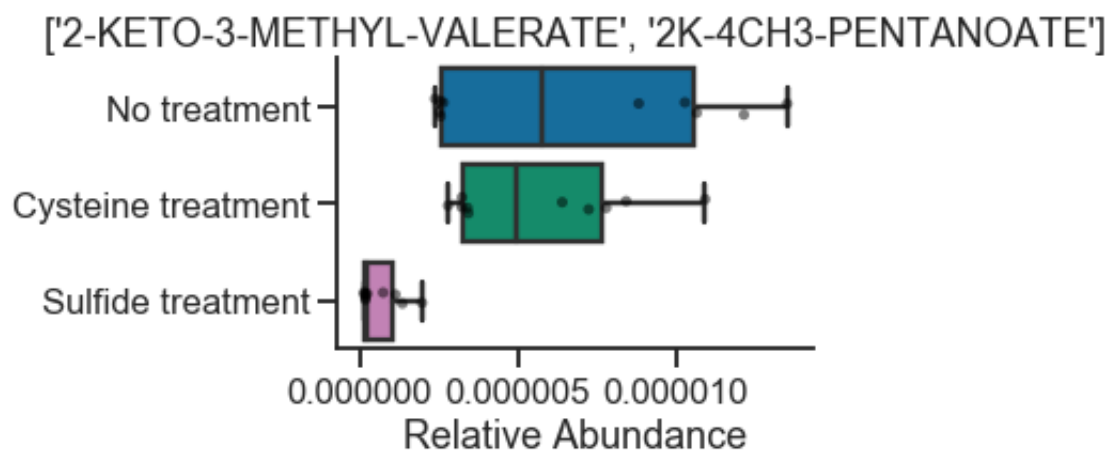
44

<Figure size 432x288 with 0 Axes>



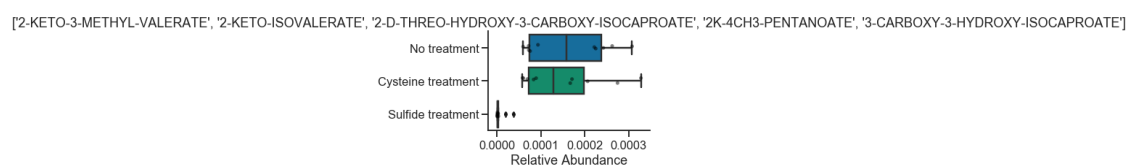
179

<Figure size 432x288 with 0 Axes>



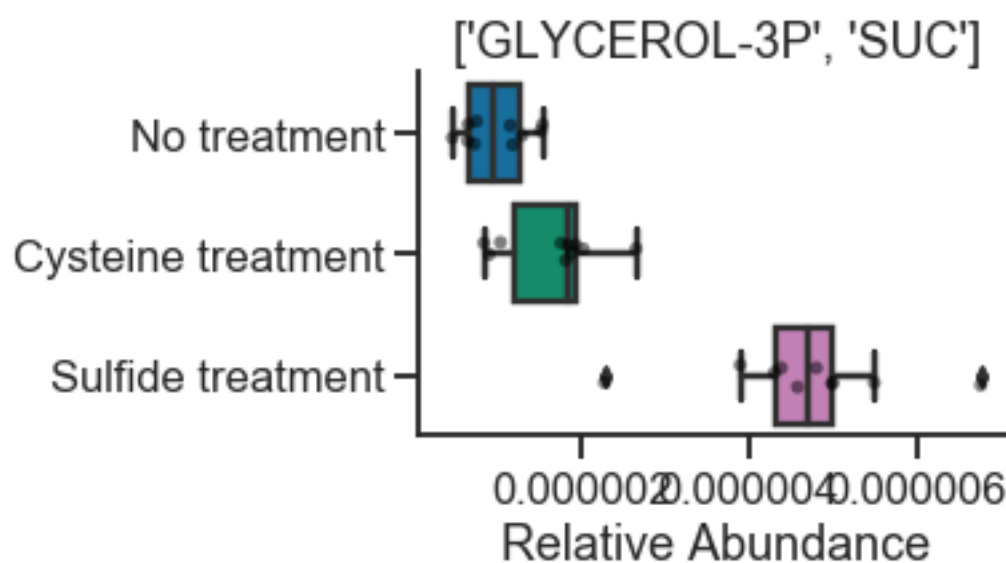
38

<Figure size 432x288 with 0 Axes>



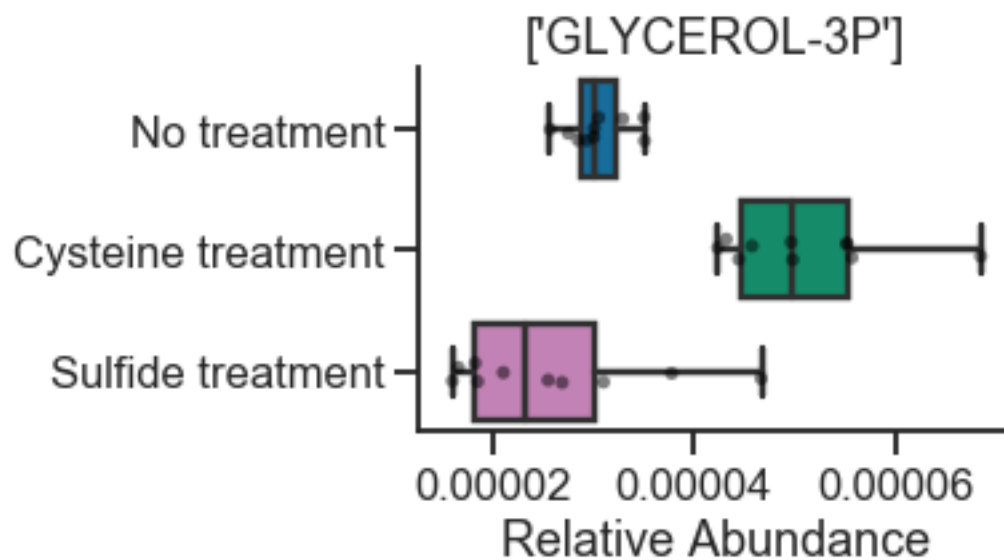
100

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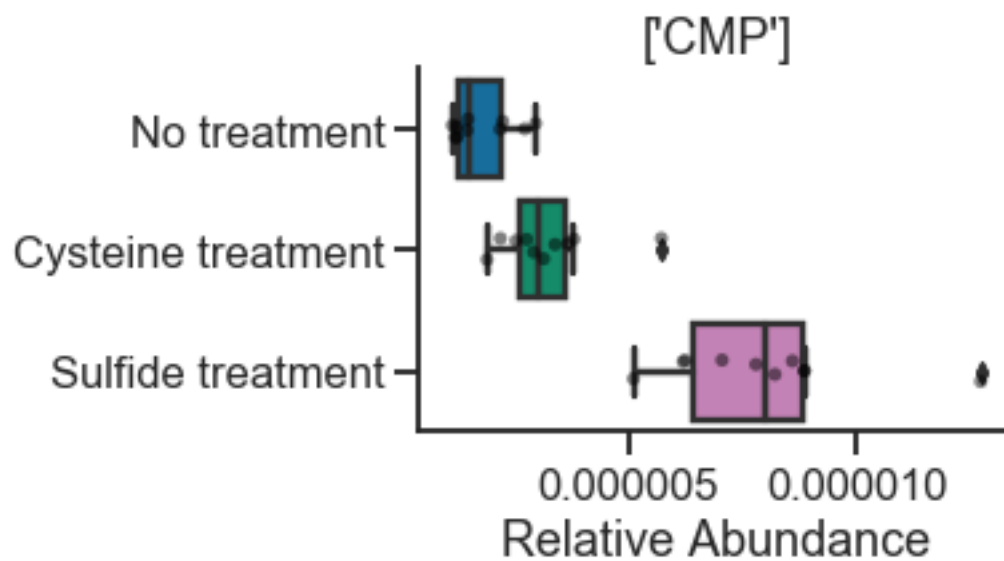
303

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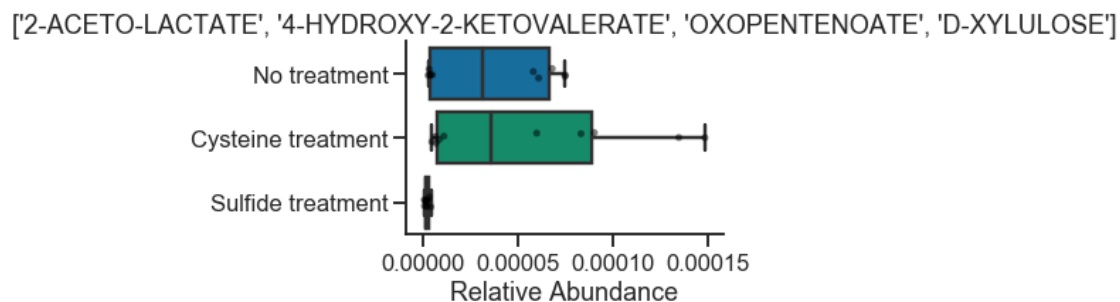
53

<Figure size 432x288 with 0 Axes>



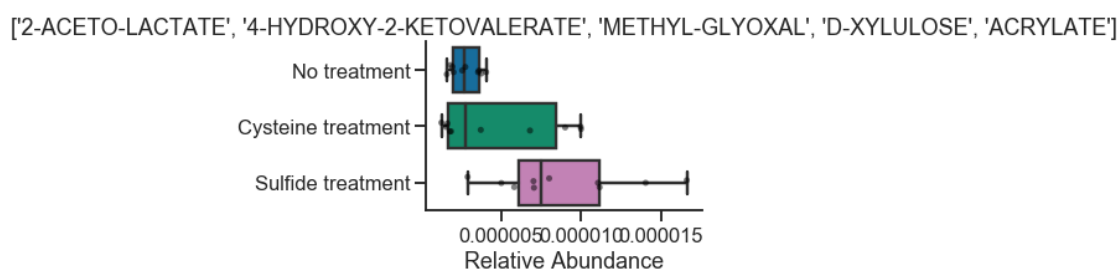
161

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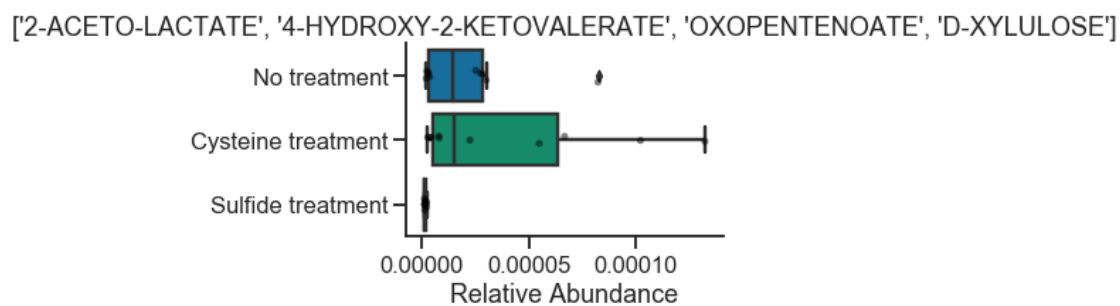
184

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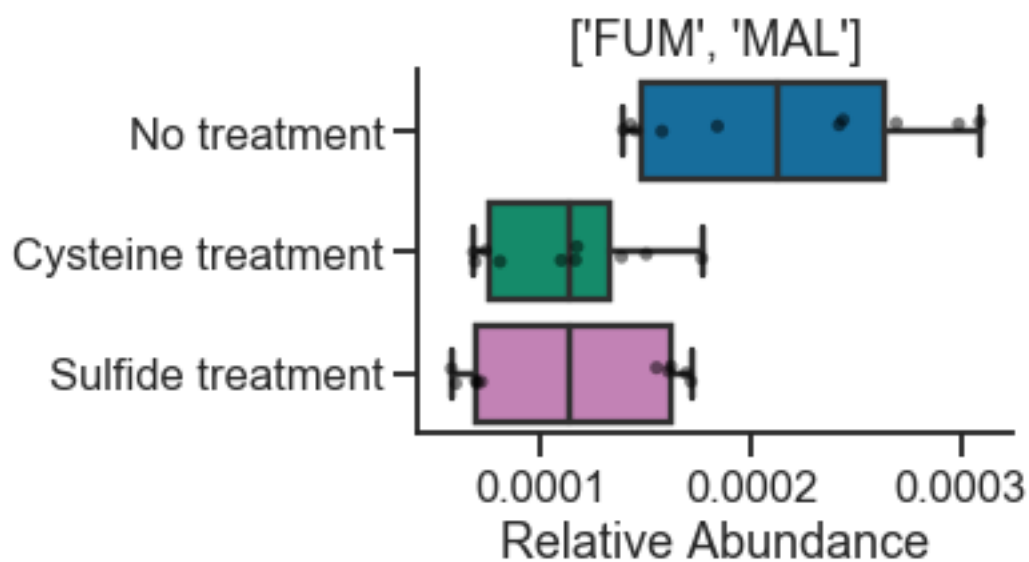
158

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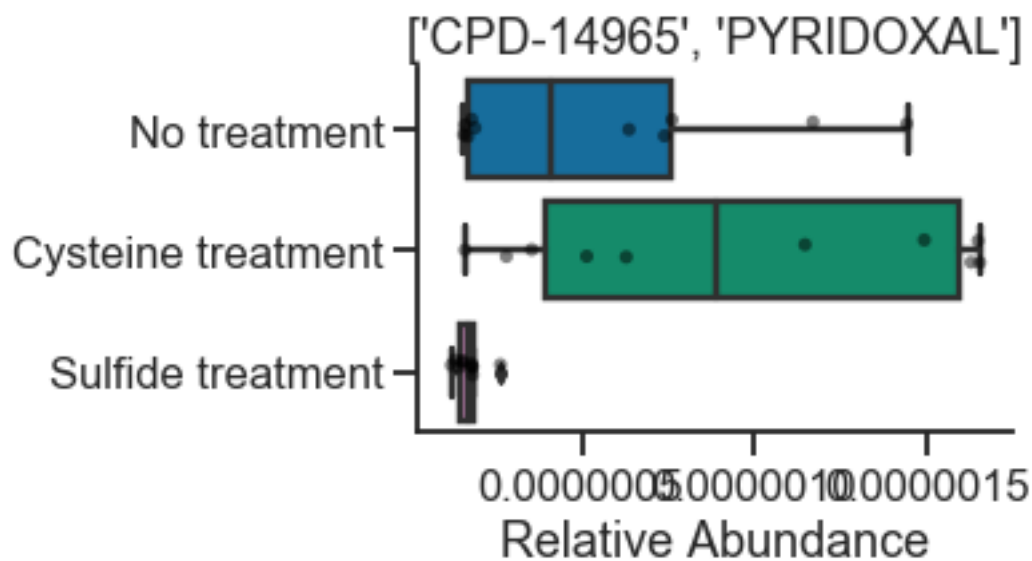
220

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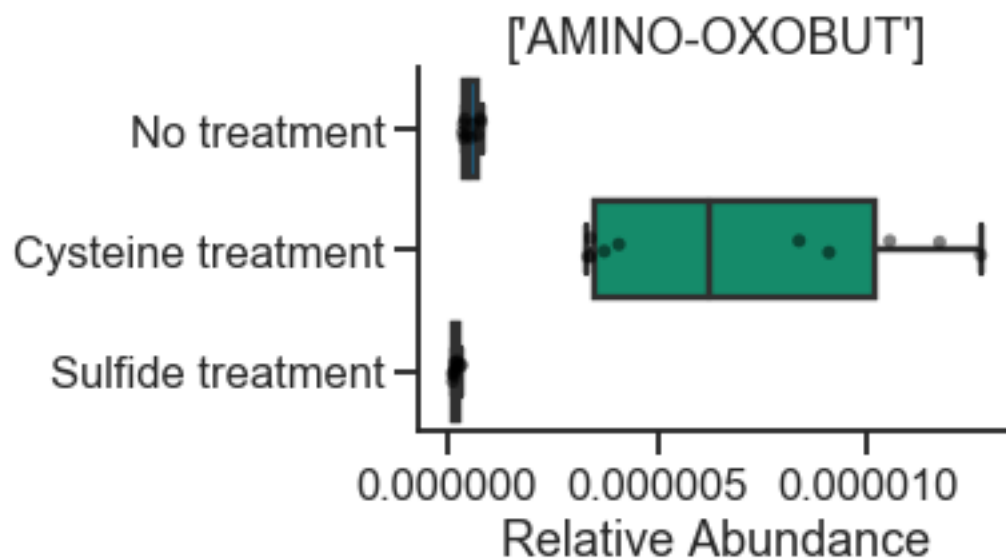
239

<Figure size 432x288 with 0 Axes>



27

<Figure size 432x288 with 0 Axes>



289

<Figure size 432x288 with 0 Axes>

[]: