ManyLLMs: Data Cleaning

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1 Overview

This script accompanies the analyses presented in the manuscript. Two versions are provided:

- ullet The .Rmd file can be opened and edited in ${f RStudio}$.
- A rendered **PDF** is available for reference, generated by knitting this file.

We provide full R code to ensure the analyses are transparent and reproducible. Where necessary, code is explained or unpacked in-text. This document includes:

- 1. Visualizations of key variable distributions (used in main and supplementary materials)
- 2. Justifications for major analysis decisions

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2 Step 1: Data Preparation

In this step, we prepare the environment and load data from three experimental conditions.

First, we clear the workspace using rm(list = ls(all.names = TRUE)) and load the necessary R libraries such as dplyr, ggplot2, forcats, and patchwork for data manipulation and visualization.

We then read in three .csv files corresponding to the three group conditions:

- Single-agent condition: stored in the variable single
- Pair-agent condition: stored in nob_2
- Triad-agent condition: stored in nob_3

Each dataset receives an additional column, experiment, which labels its group condition.

These datasets are then **merged** using bind_rows() into one unified data frame: combined_df

This combined dataset will be filtered, cleaned, and analyzed in the next steps.

```
rm(list = ls(all.names = TRUE))
setwd("/Users/nita/Downloads")
# 1. Read each file and add an experiment label
# Single Agent (Solo)
       <- read.csv("single_no_message_f.csv",</pre>
                                                  header = TRUE, stringsAsFactors = TRUE) %>%
  mutate(experiment = "Single")
#Pairs (groups)
        <- read.csv("nob_2_no_message_f.csv",</pre>
                                                   header = TRUE, stringsAsFactors = TRUE) %>%
  mutate(experiment = "NoB_2")
#Triad (groups)
        <- read.csv("nob_3_no_message_f.csv",</pre>
nob_3
                                                    header = TRUE, stringsAsFactors = TRUE) %>%
  mutate(experiment = "NoB_3")
# 3. Combine
combined_df <- bind_rows(single,nob_2,nob_3)</pre>
```

We remove the column onboarding from combined df as it is not needed for further analysis.

Next, we filter the dataset to retain only the observations from step -1 (pre-condition) and step 7 (post-condition), which correspond to the *single* and *group* conditions. After filtering, we apply droplevels() to clean up unused factor levels.

To focus our analysis on relevant moral judgments, we also remove any **non-moral scenarios** by filtering out rows where the type column is "Non-Moral".

Finally, we simplify verbose model names by recoding them using fct_recode() and storing the cleaned names in a new column:

model short

This step ensures the dataset is concise and easier to interpret in subsequent analysis and visualization.

```
# 4. Drop the unwanted columns
combined_df <- combined_df %>% select(-onboarding)

# 5. Select only Step -1 and 7 opinion (Single vs. Groups) no discussion
combined_df= filter(combined_df, step == -1 | step ==7) %>% droplevels()

# 6. delete the non-moral scenarios
combined_df= filter(combined_df, type!="Non-Moral") %>% droplevels
```

Next, we **standardize** the dataset names by recoding "oxford_utilitarianism_scale" to a shorter label: "oxford".

We also recode values in the type column to group similar scenario categories under clearer, more interpretable labels — for example:

```
"Factual-Killing-Utilitarian" becomes "Killing-Util" and "Factual-Saving-Deontological" becomes "Saving-Deon".
```

To enhance readability, we also **shorten long model names** by assigning simplified names to a new variable: model_short.

For items belonging to the "CNI" dataset, we create a new column cni_type by mapping each example_index to a predefined list of CNI scenario codes.

From these codes, we extract meaningful suffixes such as "acinc", "ininc", "incon", or "accon" and update the type column accordingly.

This finalizes the dataset's semantic structure, aligning it with the categories used throughout the manuscript.

```
# 7. Make llm names shorter
combined_df <- combined_df %>%
  mutate(
   model_short = fct_recode(model,
      "GPT4.1" = "gpt-4.1",
      "Lamma3.3" = "llama3.3",
      "Qwen2.5"= "qwen2.5:32b-instruct",
     "Qwen3" = "qwen3:32b",
      "QWQ" = "qwq",
      "Gemma3" = "gemma3:27b"
    )
)
# 7 changing the levles of dataset
combined_df <- combined_df %>%
  mutate(dataset = recode(dataset, "oxford_utilitarianism_scale" = "oxford"))
# 8. changing the type:
library(forcats)
combined_df$type <- fct_recode(combined_df$type,</pre>
  "Action" = "action",
  "Killing-Util" = "Factual-Killing-Utilitarian",
  "Other-Deon" = "Factual-Other-Deontological",
  "Other-Util" = "Factual-Other-Utilitarian",
```

```
"Saving-Deon" = "Factual-Saving-Deontological",
  "Beneficence" = "Impartial Beneficence",
 "Harm" = "Instrumental Harm",
"Omission" = "omission",
"CNI" = "N/A"
  # "Impersonal" and "Personal" remain unchanged
#fix cni types
cni_labels <- c(</pre>
  "dlacinc", "d6ininc", "d5ininc", "d4incon", "d3ininc", "d2ininc",
  "d1ininc", "d2acinc", "d6accon", "d5accon", "d4acinc", "d3incon",
  "d2incon", "d6incon", "d1incon", "d3accon", "d5acinc", "d4accon",
  "d2accon", "d6acinc", "d3acinc", "d4ininc", "d5incon", "d1accon"
)
combined_df <- combined_df %>%
  mutate(example_index = as.character(example_index)) %>%
  mutate(
    cni_type = case_when(
      type == "CNI" ~ factor(
        example_index,
        levels = as.character(seq_along(cni_labels)),
       labels = cni_labels
      ),
      TRUE ~ NA_character_
    )
  )
combined_df <- combined_df %>%
  mutate(
    type = ifelse(
      type == "CNI",
      str_extract(cni_type, "acinc|ininc|incon|accon"),
      as.character(type)
    ),
    type = factor(type, levels = c("acinc", "ininc", "incon", "accon",
                                    "Action", "Killing-Util", "Other-Deon",
                                    "Other-Util", "Saving-Deon", "Beneficence",
                                    "Harm", "Impersonal", "Personal", "Omission"))
```

Finally, we address a known **misalignment** in the greene dataset: any example_index values greater than 41 are off by one.

We correct this by subtracting 1 from those values.

The corrected example_index column is then converted into a factor named item to prepare it for grouped analysis.

As a sanity check, we verify that all expected item indices from 1 to 9 are present.

This concludes the *initial data preparation step*, ensuring that the dataset is **clean**, **consistent**, and fully ready for downstream analysis.

```
#Fixiing issues with Green
library(dplyr)
combined_df$example_index=as.numeric(combined_df$example_index)
# Step 1: Apply correction ONLY to greene where index > 41
combined_df <- combined_df %>%
  mutate(
    example_index = case_when(
      dataset == "greene" & example_index > 41 ~ example_index - 1,
      TRUE ~ example index
   )
  )
combined_df$item=as.factor(combined_df$example_index)
combined df$stepf=as.factor(combined df$step)
expected_indices <- 1:max(combined_df$example_index)</pre>
missing_indices <- setdiff(expected_indices, combined_df$example_index)
expected_indices <- 1:max(combined_df$example_index)</pre>
missing_indices <- setdiff(expected_indices, combined_df$example_index)
if (length(missing_indices) == 0) {
  cat(" All indices from 1 to", max(combined_df$example_index), "are present.\n")
} else {
  cat(" Missing indices:", paste(missing indices, collapse = ", "), "\n")
}
```

All indices from 1 to 64 are present.

There are 0 missing indices between 1 and 64.

3 Step 2: Cleaning and Formatting

Note that the dataset has already been *pre-processed*, as described in the **Supplementary Materials**. Here, we check for any **additional mismatches or missing entries** that may affect downstream analysis.

In this step, we clean the combined_df dataset by identifying and removing problematic values in the opinion column (also referred to as the *utilitarian score*).

We begin by converting opinion into two formats:

- as character: opinion_char
- as numeric: opinion num

```
# Calculate total invalid entries
total_rows <- nrow(combined_df)
total_missing <- sum(invalid_all, na.rm = TRUE)</pre>
```

We then define several **invalid conditions**:

- is_na detects missing values
- is_text identifies non-numeric entries (e.g., text or symbols)
- out_of_range flags numeric values outside the valid 1-7 Likert scale

All three are combined into a master logical flag: invalid_all.

We then compute and print their counts.

The **total number of invalid entries** is stored in total_missing, which equals r total_missing observations. These include any missing, misformatted, or out-of-range values in the data.

The total number of invalid entries is stored in total_missing, which equals 289 observations.

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We print a concise summary of issues found in the opinion column of combined_df:

- NA values are missing responses.
- Text values are improperly formatted (e.g., strings instead of numbers).
- Out-of-range values fall outside the valid Likert scale (1–7).
- All of these are grouped under invalid entries and counted as a share of the total.

This summary helps assess the integrity of the data before applying filters or imputation.

```
# 3. Print counts
# Summary counts for opinion cleaning
total_rows <- nrow(combined_df)
na_count <- sum(is_na)
text_count <- sum(is_text)
range_count <- sum(out_of_range)
invalid_total <- sum(invalid_all)</pre>
```

Table 1: Table: Data Quality Summary

Metric	Count
Total rows	51090
NA values	0
Non-numeric (text)	39
Out-of-range (not 1–7)	250
Total invalid entries	289

```
# 4. Show the percentage of missing values

total_rows <- nrow(combined_df)
total_missing <- sum(invalid_all, na.rm = TRUE)</pre>
```