Network Structure

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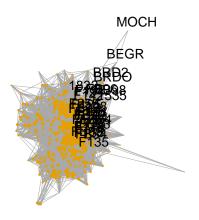
In this markdown I will:

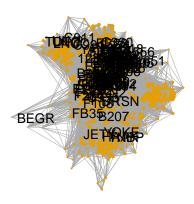
- 1. Create the network structure from the association matrix.
- 2. Evaluate local and global network metrics.
- 3. Permutate the link weights using the WalkTrap algorithm.
- 4. Evaluate and plot modularity.

PART 1: Network Structure

```
## Create social network
ig <- lapply(nxn, function (df) {</pre>
  graph_from_adjacency_matrix(
  df,
  mode = "undirected",
  weighted = TRUE,
  diag = FALSE)})
# Set the node names based on row names
row_names <- lapply(nxn, function (df) {rownames(df)})</pre>
for (i in seq_along(ig)) {
  V(ig[[i]])$name <- row_names[[i]]</pre>
## Only show IDs of HI dolphins
### subset_HI in "GLMM.R"
HI_data <- diff_raw(subset_HI(list_years))</pre>
row_names_HI <- lapply(HI_data, function (df) {</pre>
  as.vector(df$Code[(df$DiffHI == "BG" | df$DiffHI == "SD" |
                                      df$DiffHI == "FG") & df$Freq > 0])})
# Plot network
# Set up the plotting area with 1 row and 2 columns for side-by-side plots
par(mfrow=c(1, 2))
# Loop through the list of graphs and plot them side by side
for (i in 1:length(ig)) {
```

```
plot(ig[[i]],
    layout = layout_with_fr(ig[[i]]),
    edge.width = E(ig[[i]])$weight * 4,
    vertex.size = sqrt(igraph::strength(ig[[i]], vids = V(ig[[i]]), mode = c("all"), loops = TRUE) *
    vertex.frame.color = NA,
    vertex.label.family = "Helvetica",
    vertex.label = ifelse(V(ig[[i]])$name %in% row_names_HI[[i]], V(ig[[i]])$name, NA),
    vertex.label.color = "black",
    vertex.label.color = 0.8,
    vertex.label.dist = 2,
    vertex.frame.width = 0.01)
}
```





```
# Reset the plotting area to its default configuration
par(mfrow=c(1, 1))
```

PART 2: Network Metrics

Local Network Metrics

• Local clustering coefficient: Measure of the prevalence of node clusters in a network (i.e. if dolphin associates all know each other, the clustering coefficient will be high).

- Betweeness: A high betweenness means that the individual is in the communication path of other individuals, therefore, the individuals it interacts with, depend on its presence.
- Closeness: The larger the closeness centrality is for an individual, the more rapidly and easily it can influence the behavior of others.
- Degree: # Individual's associates
- Strength: Total strength of an individuals' associations

```
# Edgelist: Nodes (i & j) and edge (or link) weight
el <- readRDS("../data/el_years.RData")</pre>
# Set the node names based on row names
get_names <- function (matrix, metric) {</pre>
  row_names <- lapply(matrix, function (df) {rownames(df)})</pre>
for (i in seq_along(metric)) {
  metric[[i]][,1] <- row_names[[i]]</pre>
}
  return(metric)
# Weighted clustering coefficients
cluster <- readRDS("../data/cluster.RData")</pre>
cluster diffs <- get names(nxn, cluster)</pre>
cluster_diffs_HI <- lapply(seq_along(cluster_diffs), function(i) {</pre>
  df <- cluster_diffs[[i]]</pre>
  df_new <- as.data.frame(df[df[, 1] %in% row_names_HI[[i]], , drop = FALSE])</pre>
  return(df_new)
})
compare_cluster <- merge(</pre>
  cluster_diffs_HI[[1]][, c(1, 2)],
  cluster_diffs_HI[[2]][, c(1, 2)],
  by.x = "node",
  by.y = "node"
colnames(compare_cluster) <- c("ID", "Period.1", "Period.2")</pre>
compare_cluster[, c(2, 3)] <- sapply(compare_cluster[, c(2, 3)], as.numeric)</pre>
# Calculate differences
compare_cluster$Period.2 - compare_cluster$Period.2 - compare_cluster$Period.1
# Betweenness centrality
between <- lapply(el, function (df) {betweenness_w(df, alpha=1)})
between_diffs <- get_names(nxn, between)</pre>
between_diffs_HI <- lapply(seq_along(between_diffs), function(i) {</pre>
  df <- between_diffs[[i]]</pre>
  df_new <- as.data.frame(df[df[, 1] %in% row_names_HI[[i]], , drop = FALSE])</pre>
  return(df_new)
})
compare_between <- merge(</pre>
  between_diffs_HI[[1]],
  between_diffs_HI[[2]],
  by.x = "node",
  by.y = "node"
```

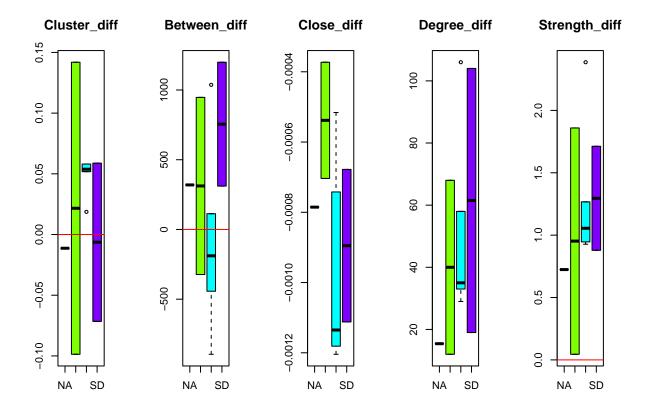
```
colnames(compare_between) <- c("ID", "Period.1", "Period.2")</pre>
compare_between[, c(2, 3)] <- sapply(compare_between[, c(2, 3)], as.numeric)</pre>
# Calculate differences
compare_between$Period.2 - compare_between$Period.1
# Closeness centrality
close <- lapply(el, function (df) {closeness_w(df, alpha=1)})</pre>
close_diffs <- get_names(nxn, close)</pre>
close_diffs_HI <- lapply(seq_along(close_diffs), function(i) {</pre>
  df <- close_diffs[[i]]</pre>
  df_new <- as.data.frame(df[df[, 1] %in% row_names_HI[[i]], , drop = FALSE])</pre>
 return(df_new)
})
compare_close <- merge(</pre>
  close_diffs_HI[[1]][, c(1, 2)],
  close_diffs_HI[[2]][, c(1, 2)],
 by.x = "node",
  by.y = "node"
colnames(compare_close) <- c("ID", "Period.1", "Period.2")</pre>
compare_close[, c(2, 3)] <- sapply(compare_close[, c(2, 3)], as.numeric)</pre>
# Calculate differences
compare close$Difference <- compare close$Period.2 - compare close$Period.1</pre>
# Degree and strength centrality
strength <- lapply(el, function (df) {degree_w(df, measure=c("degree","output"), type="out", alpha=1)})</pre>
strength_diffs <- get_names(nxn, strength)</pre>
strength_diffs_HI <- lapply(seq_along(strength_diffs), function(i) {</pre>
  df <- strength_diffs[[i]]</pre>
  df_new <- as.data.frame(df[df[, 1] %in% row_names_HI[[i]], , drop = FALSE])</pre>
 return(df_new)
})
compare_strength <- merge(</pre>
  strength_diffs_HI[[1]],
  strength_diffs_HI[[2]],
  by.x = "node",
  by.y = "node"
colnames(compare_strength) <- c("ID", "Period.1_degree", "Period.1_strength", "Period.2_degree", "Period.</pre>
compare_strength[, c(2:5)] <- sapply(compare_strength[, c(2:5)], as.numeric)</pre>
# Calculate differences
compare_strength$Difference_degree <- compare_strength$Period.2_degree - compare_strength$Period.1_degr
compare_strength$Difference_strength <- compare_strength$Period.2_strength - compare_strength$Period.1_</pre>
# Look at all of the local metrics together
## Add a column containing HI type
names_BG <- unlist(lapply(HI_data, function (df) {</pre>
  as.vector(df$Code[df$DiffHI == "BG" & df$Freq > 0])}))
names_SD <- unlist(lapply(HI_data, function (df) {</pre>
  as.vector(df$Code[df$DiffHI == "SD" & df$Freq > 0])}))
```

```
names_FG <- unlist(lapply(HI_data, function (df) {</pre>
  as.vector(df$Code[df$DiffHI == "FG" & df$Freq > 0])}))
HI_type <- ifelse(compare_cluster$ID %in% names_BG, "BG",</pre>
                                    ifelse(compare_cluster$ID %in% names_SD, "SD",
                                            ifelse(compare_cluster$ID %in% names_FG, "FG", "NA")))
local_metrics_HI <- as.data.frame(cbind(ID = compare_cluster$ID, HI_type = HI_type, Cluster_diff = comp</pre>
                                Between_diff = compare_between$Difference, Close_diff = compare_close$Di
                                Degree_diff = compare_strength$Difference_degree, Strength_diff = compar
## Add a rown to compare the averages of each metric with HI IDs
avg_metrics <- data.frame(ID = "Average", HI_type = "NA",</pre>
  Cluster_diff = mean(cluster[[2]][, 2]) - mean(cluster[[1]][, 2]),
  Between_diff = mean(between[[2]][, 2]) - mean(between[[1]][, 2]),
  Close_diff = mean(close[[2]][, 2]) - mean(close[[1]][, 2]),
  Degree_diff = mean(strength[[2]][, 2]) - mean(strength[[1]][, 2]),
  Strength_diff = mean(strength[[2]][, 3]) - mean(strength[[1]][, 3]))
local_metrics_HI <- merge(local_metrics_HI, avg_metrics, all = T)</pre>
# Make sure the metrics are numeric and HI_type is a factor
columns_to_convert <- c(3:7) # Columns to be converted to numeric</pre>
local_metrics_HI[, columns_to_convert] <- sapply(local_metrics_HI[, columns_to_convert], as.numeric)</pre>
local_metrics_HI$HI_type <- as.factor(local_metrics_HI$HI_type)</pre>
```

Plot different Metrics within HI types

```
# Set up the plotting area with 1 row and 5 columns
par(mfrow = c(1, 5))
# Get different metric values in a vector
metric value <- colnames(local metrics HI[, columns to convert])
# Define the desired order for HI types
desired_order <- c("NA", "BG", "FG", "SD")</pre>
# Loop over each metric
for (i in seq_along(columns_to_convert)) {
  # Extract the metric for the current column
  metric <- local_metrics_HI[, columns_to_convert[i]]</pre>
  # Convert HI_type to factor with the desired order
  local_metrics_HI$HI_type <- factor(local_metrics_HI$HI_type, levels = desired_order)</pre>
  # Group the metric by HI_type and order the names
  grouped_metric <- split(metric, local_metrics_HI$HI_type)</pre>
  # Set up colors based on HI type
  colors <- rainbow(length(names(grouped_metric)))</pre>
```

```
# Create the boxplot
boxplot(
    grouped_metric,
    names = names(grouped_metric),
    col = colors,
    notch = FALSE,
    main = metric_value[i],
    border = "black"
)
# Draw a horizontal line at y = 0
abline(h = 0, col = "red")
}
```



Global Network Metrics

- Size: Number of nodes.
- Density/Connectance: Proportion of realized links (observed/possible links).
- Average Path Length (geodesic): Measures the shortest distance between two random nodes then average shortest pathways between all pairs of nodes. Shows how far apart any pair of individuals will be on average.
- Geodesic path: the shortest path through the network from one node to another (1).

- Diameter: Length of the longest geodesic path (d).
- Clustering coefficient: Tendency of nodes to cluster in the network (Are the friends' friends also friends?).

```
\#' Breakdown: connectance = length(which(as.dist(orca_hwi)!=0))/(N*(N-1)/2)
## Calculate connectance for each matrix
calculate_connectance <- function(matrix) {</pre>
  N <- nrow(matrix)</pre>
  total \leftarrow N * (N - 1) / 2
  real <- sum(matrix != 0) # Count non-zero elements
  connectance <- real / total</pre>
  return(connectance)
}
connectance_list <- lapply(nxn, calculate_connectance)</pre>
connectance_list
## [[1]]
## [1] 0.5375485
##
## [[2]]
## [1] 0.4575163
# Shortest path lengths (geodesics) and diameter
# # mean shortest path
dist <- lapply(ig, function(df) {mean_distance(df)})</pre>
dist
## [[1]]
## [1] 0.004317689
## [[2]]
## [1] 0.005595755
```

PART 3: Permutate Link Weights

Walktrap algorithm breakdown with one interation

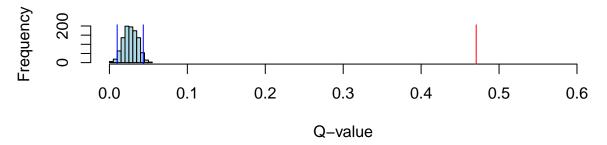
Permutate with multiple interations

```
# Run modularity permutations 1000 times for each matrix
run_mod <- function(el_list, dolphin_walk_list) {
  iter <- 1000
  randmod <- numeric(iter) # Initialize a numeric vector to store Q-values

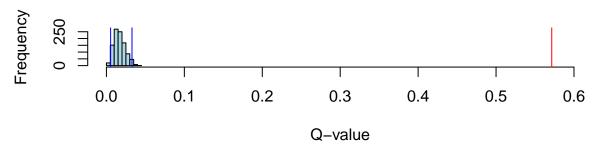
for (i in 1:iter) {
    # Save the edgelist into a new object and permutate the link weights
    auxrand <- el_list
    auxrand[, 2] <- sample(auxrand[, 2])</pre>
```

```
# Create an igraph graph from the permuted edgelist
    igrand <- graph.edgelist(as.matrix(auxrand[, 1:2]), directed = FALSE)</pre>
    E(igrand)$weight <- auxrand[, 2] # Assign link weights</pre>
    # Calculate modularity using walktrap community detection
    rand_walk <- walktrap.community(igrand)</pre>
    randmod[i] <- modularity(rand_walk) # Save Q-value into the vector</pre>
  }
  # Calculate the 95% confidence interval (two-tailed test)
  ci \leftarrow quantile(randmod, probs = c(0.025, 0.975), type = 2)
  \# Visualization of the random Q distribution
  hist(randmod, xlim = c(0, 0.6), main = "Random Q Distribution", xlab = "Q-value", ylab = "Frequency",
  # Empirical Q-value
  abline(v = modularity(dolphin_walk_list), col = "red")
  # 2.5% CI
  abline(v = ci[1], col = "blue")
  # 97.5% CI
  abline(v = ci[2], col = "blue")
  # Return a data frame with Q-value and confidence intervals
  result <- data.frame(Q = modularity(dolphin_walk_list), LowCI = ci[1], HighCI = ci[2])
  return(result)
}
# Plot null distributions for modularity
# Set up the plotting area with 2 rows and 2 column
par(mfrow=c(2, 1))
run_mod(el_list = el[[1]], dolphin_walk_list = dolphin_walk[[1]])
                        LowCI
                                  HighCI
## 2.5% 0.4706832 0.009965587 0.0434557
run_mod(el_list = el[[2]], dolphin_walk_list = dolphin_walk[[2]])
```

Random Q Distribution



Random Q Distribution



```
## Q LowCI HighCI
## 2.5% 0.5714733 0.005398006 0.03280247
```

We can reject the null hypothesis that individuals cluster at random and conclude that there is evidence that modularity is higher than what we would expect by chance.

PART 4: Modularity

• Newman's Q modularity: Stopping parameter Q removes links according to the betweenness.

```
# Create an unweighted network
system.time({
  registerDoParallel(n.cores)
  dolp_ig <- list()
  for (l in seq_along(list_years)) {
    dolp_ig[[1]] <- graph.edgelist(el[[1]][,1:2])
    # Add the edge weights to this network
    E(dolp_ig[[1]])$weight <- as.numeric(el[[1]][,3])
    # Create undirected network
    dolp_ig[[1]] <- as.undirected(dolp_ig[[1]])
}
### End parallel processing
stopImplicitCluster()
})</pre>
```

```
##
      0.09
              0.00
                      6.08
# Newman's Q modularity
newman <- lapply(dolp_ig, function (df) {cluster_leading_eigen(df, steps = -1, weights = E(df) weight,
                                start = NULL, options = arpack_defaults, callback = NULL,
                                extra = NULL, env = parent.frame())})
# Set the node names based on row names
BG <- SD <- FG <- vector("list", length = length(dolp_ig))
for (i in seq_along(dolp_ig)) {
  # Set the node names
  V(dolp_ig[[i]])$name <- rownames(nxn[[i]])</pre>
  ## Parse out what HI behavior they engage in
  BG[[i]] <- as.vector(HI_data[[i]]$Code[HI_data[[i]]$DiffHI == "BG" & HI_data[[i]]$Freq > 0])
  SD[[i]] <- as.vector(HI data[[i]]$Code[HI data[[i]]$DiffHI == "SD" & HI data[[i]]$Freq > 0])
  FG[[i]] <- as.vector(HI_data[[i]]$Code[HI_data[[i]]$DiffHI == "FG" & HI_data[[i]]$Freq > 0])
  ## Initialize label_color attribute for each node
  V(dolp_ig[[i]])$label_color <- "black"</pre>
  ## Make a different text color for each category
  V(dolp_ig[[i]])$label_color[V(dolp_ig[[i]])$name %in% BG[[i]]] <- "red"</pre>
  V(dolp_ig[[i]])$label_color[V(dolp_ig[[i]])$name %in% SD[[i]]] <- "yellow"
  V(dolp_ig[[i]])$label_color[V(dolp_ig[[i]])$name %in% FG[[i]]] <- "blue"
# Generate a vector of colors based on the number of unique memberships
for (i in seq_along(dolp_ig)) {
 V(dolp_ig[[i]])$color <- NA</pre>
  col <- rainbow(max(newman[[i]]$membership))</pre>
 for (j in 1:max(newman[[i]]$membership)){
   V(dolp_ig[[i]])$color[which(newman[[i]]$membership==j)] <- col[j]</pre>
  }
}
# Make sure the HI dolphins stand out
for (i in seq_along(dolp_ig)) {
  V(dolp_ig[[i]])$size <- ifelse(V(dolp_ig[[i]])$name %in% row_names_HI[[i]], 10, 5)
}
# Set up the plotting area with 1 row and 2 columns for side-by-side plots
par(mfrow=c(1, 2))
# Main labels for the plots
main_labels <- c("1993-2004 Network", "2005-2014 Network") # Replace with appropriate main labels
# Plot the graph with individual IDs as labels
for (i in seq_along(dolp_ig)) {
plot(dolp ig[[i]],
     layout = layout_with_fr(dolp_ig[[i]]),
```

##

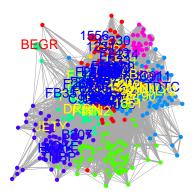
user system elapsed

```
# link weight, rescaled for better visualization
edge.width= E(dolp_ig[[i]])$weight*4,
# node size as degree (rescaled)
vertex.size= V(dolp_ig[[i]])$size,
vertex.frame.color= NA, #"black",
vertex.label.family = "Helvetica",
vertex.label=ifelse(V(dolp_ig[[i]])$name %in% row_names_HI[[i]], as.character(V(dolp_ig[[i]])$name
vertex.label.color = V(dolp_ig[[i]])$label_color,
vertex.label.cex=0.8,
vertex.label.dist=0.5,
# edge.curved=0,
vertex.frame.width=0.01)
# Add the main label above the plot
title(main = main_labels[i], line = -1)
}
```

1993-2004 Network

MGCH BEGR STBO

2005-2014 Network



Since these modules can represent functional units, I need to test which mechanisms drive the modular topology by creating null models.