A quick overview of the S4 class system

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Outline

The S4 class system

- The *S4 class system* is a set of facilities provided in R for OO programming.
- Implemented in the *methods* package.
- On a fresh *R* session:
  ```
  > sessionInfo()
  ...
  attached base packages:
  [1] stats     graphics  grDevices utils     datasets
  [6] methods   base
  ```
- R also supports an older class system: the *S3 class system*.

A different world

The syntax

> `foo(x, ...)`

not:

> `x.foo(...)`

like in other OO programming languages.

The central concepts

- ▶ The core components: *classes*[1], *generic functions* and *methods*
- ▶ The glue: *method dispatch* (supports *simple* and *multiple* dispatch)

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[1] also called *formal classes*, to distinguish them from the S3 classes aka *old style classes*

The result

```
> ls('package:methods')
  [1] "addNextMethod"              "allGenerics"
  [3] "allNames"                   "Arith"
  [5] "as"                         "as<-"
  [7] "asMethodDefinition"         "assignClassDef"
...
[211] "testVirtual"                "traceOff"
[213] "traceOn"                    "tryNew"
[215] "unRematchDefinition"        "validObject"
[217] "validSlotNames"
```

- ▶ Rich, complex, can be intimidating
- ▶ The classes and methods we implement in our packages can be hard to document, especially when the class hierarchy is complicated and multiple dispatch is used

S4 in Bioconductor

- Heavily used. In BioC 3.3: 3158 classes and 22511 methods defined in 609 packages! (out of 1211 software packages)
- Top 10: 128 classes in *ChemmineOB*, 98 in *flowCore*, 79 in *IRanges*, 68 in *rsbml*, 61 in *ShortRead*, 58 in *Biostrings*, 51 in *rtracklayer*, 50 in *oligoClasses*, 45 in *flowUtils*, and 40 in *BaseSpaceR*.
- For the end user: it's mostly transparent. But when something goes wrong, error messages issued by the S4 class system can be hard to understand. Also it can be hard to find the documentation for a specific method.
- Most Bioconductor packages use only a small subset of the S4 capabilities (covers 99.99% of our needs)

Outline

Where do S4 objects come from?

From a dataset

```
> library(graph)
> data(apopGraph)
> apopGraph
A graphNEL graph with directed edges
Number of Nodes = 50
Number of Edges = 59
```

From using an object constructor function

```
> library(IRanges)
> IRanges(start=c(101, 25), end=c(110, 80))
IRanges object with 2 ranges and 0 metadata columns:
          start       end     width
      <integer> <integer> <integer>
  [1]       101       110        10
  [2]        25        80        56
```

From a coercion

```
> library(Matrix)
> m <- matrix(3:-4, nrow=2)
> as(m, "Matrix")
2 x 4 Matrix of class "dgeMatrix"
     [,1] [,2] [,3] [,4]
[1,]    3    1   -1   -3
[2,]    2    0   -2   -4
```

From using a specialized high-level constructor

```
> library(GenomicFeatures)
> makeTxDbFromUCSC("sacCer2", tablename="ensGene")

TxDb object:
# Db type: TxDb
# Supporting package: GenomicFeatures
# Data source: UCSC
# Genome: sacCer2
# Organism: Saccharomyces cerevisiae
# Taxonomy ID: 4932
# UCSC Table: ensGene
# UCSC Track: Ensembl Genes
...
```

From using a high-level I/O function

```
> library(ShortRead)
> path_to_my_data <- system.file(
+     package="ShortRead",
+     "extdata", "Data", "C1-36Firecrest", "Bustard", "GERALD")
> lane1 <- readFastq(path_to_my_data, pattern="s_1_sequence.txt")
> lane1

class: ShortReadQ
length: 256 reads; width: 36 cycles
```

Inside another object

```
> sread(lane1)

DNAStringSet object of length 256:
        width seq
    [1]    36 GGACTTTGTAGGATACCCTCGCTTTCCTTCTCCTGT
    [2]    36 GATTTCTTACCTATTAGTGGTTGAACAGCATCGGAC
    [3]    36 GCGGTGGTCTATAGTGTTATTAATATCAATTTGGGT
    [4]    36 GTTACCATGATGTTATTTCTTCATTTGGAGGTAAAA
    [5]    36 GTATGTTTCTCCTGCTTATCACCTTCTTGAAGGCTT
    ...   ... ...
  [252]    36 GTTTAGATATGAGTCACATTTTGTTCATGGTAGAGT
  [253]    36 GTTTTACAGACACCTAAAGCTACATCGTCAACGTTA
  [254]    36 GATGAACTAAGTCAACCTCAGCACTAACCTTGCGAG
```

How to manipulate S4 objects?

Low-level: getters and setters

```
> ir <- IRanges(start=c(101, 25), end=c(110, 80))
> width(ir)

[1] 10 56

> width(ir) <- width(ir) - 5
> ir

IRanges object with 2 ranges and 0 metadata columns:
          start       end     width
      <integer> <integer> <integer>
  [1]       101       105         5
  [2]        25        75        51
```

High-level: plenty of specialized methods

```
> qa1 <- qa(lane1, lane="lane1")
> class(qa1)

[1] "ShortReadQQA"
attr(,"package")
[1] "ShortRead"
```

How to find the right man page?

- `class?graphNEL` or equivalently `?`graphNEL-class`` for accessing the man page of a class
- `?qa` for accessing the man page of a generic function
- The man page for a generic might also document some or all of the methods for this generic. The *See Also:* section might give a clue. Also using `showMethods()` can be useful:

  > *showMethods("qa")*

  Function: qa (package ShortRead)
  dirPath="ShortReadQ"
  dirPath="SolexaPath"
  dirPath="character"
  dirPath="list"

- `?`qa,ShortReadQ-method`` to access the man page for a particular method (might be the same man page as for the generic)
- In doubt: `??qa` will search the man pages of all the installed packages and return the list of man pages that contain the string `qa`

Inspecting objects and discovering methods

- class() and showClass()

```
> class(lane1)
[1] "ShortReadQ"
attr(,"package")
[1] "ShortRead"
> showClass("ShortReadQ")
Class "ShortReadQ" [package "ShortRead"]

Slots:

Name:        quality        sread           id
Class: QualityScore DNAStringSet    BStringSet

Extends:
Class "ShortRead", directly
Class ".ShortReadBase", by class "ShortRead", distance 2

Known Subclasses: "AlignedRead"
```

- str() for compact display of the content of an object
- showMethods() to discover methods
- selectMethod() to see the code

Outline

Class definition and constructor

Class definition

```
> setClass("SNPLocations",
+     slots=c(
+         genome="character",   # a single string
+         snpid="character",    # a character vector of length N
+         chrom="character",    # a character vector of length N
+         pos="integer"         # an integer vector of length N
+     )
+ )
```

Constructor

```
> SNPLocations <- function(genome, snpid, chrom, pos)
+     new("SNPLocations", genome=genome, snpid=snpid, chrom=chrom, pos=pos)

> snplocs <- SNPLocations("hg19",
+             c("rs0001", "rs0002"),
+             c("chr1", "chrX"),
+             c(224033L, 1266886L))
```

Getters

Defining the `length` method

```
> setMethod("length", "SNPLocations", function(x) length(x@snpid))
> length(snplocs)  # just testing
[1] 2
```

Defining the slot getters

```
> setGeneric("genome", function(x) standardGeneric("genome"))
> setMethod("genome", "SNPLocations", function(x) x@genome)

> setGeneric("snpid", function(x) standardGeneric("snpid"))
> setMethod("snpid", "SNPLocations", function(x) x@snpid)

> setGeneric("chrom", function(x) standardGeneric("chrom"))
> setMethod("chrom", "SNPLocations", function(x) x@chrom)

> setGeneric("pos", function(x) standardGeneric("pos"))
> setMethod("pos", "SNPLocations", function(x) x@pos)

> genome(snplocs)  # just testing
[1] "hg19"
> snpid(snplocs)   # just testing
[1] "rs0001" "rs0002"
```

Defining the `show` method

```
> setMethod("show", "SNPLocations",
+     function(object)
+         cat(class(object), "instance with", length(object),
+             "SNPs on genome", genome(object), "\n")
+ )
> snplocs   # just testing
SNPLocations instance with 2 SNPs on genome hg19
```

Defining the *validity method*

```
> setValidity("SNPLocations",
+     function(object) {
+         if (!is.character(genome(object)) ||
+             length(genome(object)) != 1 || is.na(genome(object)))
+             return("'genome' slot must be a single string")
+         slot_lengths <- c(length(snpid(object)),
+                           length(chrom(object)),
+                           length(pos(object)))
+         if (length(unique(slot_lengths)) != 1)
+             return("lengths of slots 'snpid', 'chrom' and 'pos' differ")
+         TRUE
+     }
+ )
> snplocs@chrom <- LETTERS[1:3]   # a very bad idea!
> validObject(snplocs)

Error in validObject(snplocs) :
  invalid class "SNPLocations" object: lengths of slots 'snpid', 'chrom'
  and 'pos' differ
```

Defining slot setters

```
> setGeneric("chrom<-", function(x, value) standardGeneric("chrom<-"))
> setReplaceMethod("chrom", "SNPLocations",
+       function(x, value) {x@chrom <- value; validObject(x); x})

> chrom(snplocs) <- LETTERS[1:2]   # repair currently broken object

> chrom(snplocs) <- LETTERS[1:3]   # try to break it again
Error in validObject(x) :
  invalid class "SNPLocations" object: lengths of slots 'snpid', 'chrom'
  and 'pos' differ
```

Defining a coercion method

```
> setAs("SNPLocations", "data.frame",
+       function(from)
+           data.frame(snpid=snpid(from), chrom=chrom(from), pos=pos(from))
+ )

> as(snplocs, "data.frame")  # testing

    snpid chrom      pos
1 rs0001     A   224033
2 rs0002     B  1266886
```

Outline

Slot inheritance

▶ Most of the time (but not always), the child class will have additional slots:

```
> setClass("AnnotatedSNPs",
+     contains="SNPLocations",
+     slots=c(
+         geneid="character"  # a character vector of length N
+     )
+ )
```

▶ The slots from the parent class are inherited:

```
> showClass("AnnotatedSNPs")
Class "AnnotatedSNPs" [in ".GlobalEnv"]

Slots:

Name:       geneid      genome       snpid       chrom         pos
Class: character character character character     integer

Extends: "SNPLocations"
```

▶ Constructor:

```
> AnnotatedSNPs <- function(genome, snpid, chrom, pos, geneid)
+ {
+     new("AnnotatedSNPs",
+         SNPLocations(genome, snpid, chrom, pos),
+         geneid=geneid)
+ }
```

Method inheritance

- Let's create an AnnotatedSNPs object:
  ```
  > snps <- AnnotatedSNPs("hg19",
  +                 c("rs0001", "rs0002"),
  +                 c("chr1", "chrX"),
  +                 c(224033L, 1266886L),
  +                 c("AAU1", "SXW-23"))
  ```
- All the methods defined for SNPLocations objects work out-of-the-box:
  ```
  > snps
  AnnotatedSNPs instance with 2 SNPs on genome hg19
  ```
- But sometimes they don't do the right thing:
  ```
  > as(snps, "data.frame")  # the 'geneid' slot is ignored
      snpid chrom     pos
  1 rs0001  chr1  224033
  2 rs0002  chrX 1266886
  ```

- Being a SNPLocations *object* vs being a SNPLocations *instance*:

```
> is(snps, "AnnotatedSNPs")      # 'snps' is an AnnotatedSNPs object
[1] TRUE
> is(snps, "SNPLocations")       # and is also a SNPLocations object
[1] TRUE
> class(snps)                    # but is *not* a SNPLocations *instance*
[1] "AnnotatedSNPs"
attr(,"package")
[1] ".GlobalEnv"
```

- Method overriding: for example we could define a `show` method for AnnotatedSNPs objects. `callNextMethod` can be used in that context to call the method defined for the parent class from within the method for the child class.

- Automatic coercion method:

```
> as(snps, "SNPLocations")
SNPLocations instance with 2 SNPs on genome hg19
```

Incremental validity method

- The *validity method* for AnnotatedSNPs objects only needs to validate what's not already validated by the *validity method* for SNPLocations objects:

```
> setValidity("AnnotatedSNPs",
+     function(object) {
+         if (length(object@geneid) != length(object))
+             return("'geneid' slot must have the length of the object")
+         TRUE
+     }
+ )
```

- In other words: before an AnnotatedSNPs object can be considered valid, it must first be a valid SNPLocations object.

Outline

Other important S4 features

- *Virtual* classes: equivalent to *abstract* classes in Java
- Class unions (see `?setClassUnion`)
- Multiple inheritance: a powerful feature that should be used with caution. If used inappropriately, can lead to a class hierarchy that is very hard to maintain

Resources

- Man pages in the *methods* package: `?setClass`, `?showMethods`, `?selectMethod`, `?getMethod`, `?is`, `?setValidity`, `?as`
- The *Extending RangedSummarizedExperiment* section of the *SummarizedExperiment* vignette in the *SummarizedExperiment* package.
- Note: S4 is *not* covered in the *An Introduction to R* or *The R language definition* manuals[2]
- The *Writing R Extensions* manual for details about integrating S4 classes to a package
- The *R Programming for Bioinformatics* book by Robert Gentleman[3]

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[2]http://cran.fhcrc.org/manuals.html
[3]http://bioconductor.org/help/publications/books/r-programming-for-bioinformatics/