

Controlled Discovery and Localization of Signals via Bayesian Linear Programming (BLiP)

Asher Spector

Stanford University, Department of Statistics

Coauthor



Lucas Janson (Harvard Statistics)

One-slide overview

Challenge: Collinearity/correlations make it challenging to perform controlled variable selection.

- Often, we can tell that *some* variables influence the outcome Y , but we don't know which ones.
- Even after fitting a model, it's unclear how to *localize* which variables may affect Y .

This talk: Given a model, how can we "localize" signal variables?

- This talk is *not* about fitting the model!
- It *is* about extracting useful information from a pre-fit model.
- For this talk, I will assume the model is Bayesian.

Empirically: Increases power 20-50% on a large-scale GWAS with ≤ 1 min of added computation!

Outline

- 1 Motivation
- 2 Methodology (BLiP)
- 3 Application to genetic fine-mapping
- 4 Advertisement for KeLP: a frequentist, knockoffs-based method (Gablitz and Sabatti, 2024)
- 5 Conclusion

Outline

- 1 Motivation
- 2 Methodology (BLiP)
- 3 Application to genetic fine-mapping
- 4 Advertisement for KeLP: a frequentist, knockoffs-based method (Gablenz and Sabatti, 2024)
- 5 Conclusion

Motivation I: genetic fine-mapping

UK Biobank dataset ($n \approx 377,000$):

- Y is disease status
- (X_1, \dots, X_p) are genetic variants ($p \approx 19,000,000$)
- Question: which features X_j influence Y ? Which are “signals?”

For simplicity, let's temporarily assume a linear model:

$$Y = X^T \beta + \epsilon \text{ with } \mathbb{E} [\epsilon | X] = 0$$

Challenge:

- (X_1, \dots, X_p) exhibit strong local correlations, e.g., $\text{Cor}(X_1, X_2) = 0.999$
- We may have no power to detect that $\beta_1 \neq 0$
- But, maybe we know (X_1, X_2) contains a signal, i.e., $\beta_{1,2} \neq 0!$

Motivation in a picture

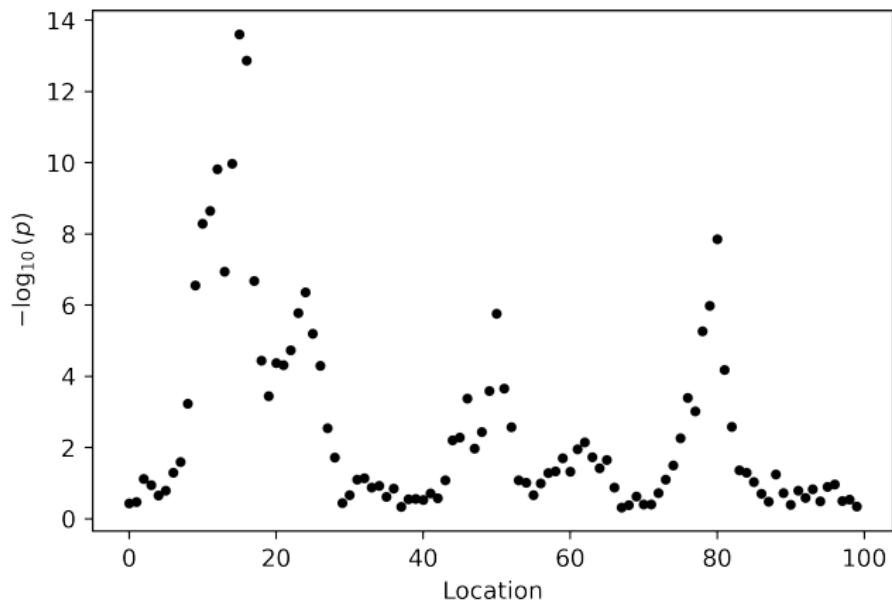


Figure: Cartoon Manhattan plot of genome; y-axis shows a measure of $\text{Corr}(Y, X_j)$ for $j = 1, \dots, 100$.

Motivation in a picture

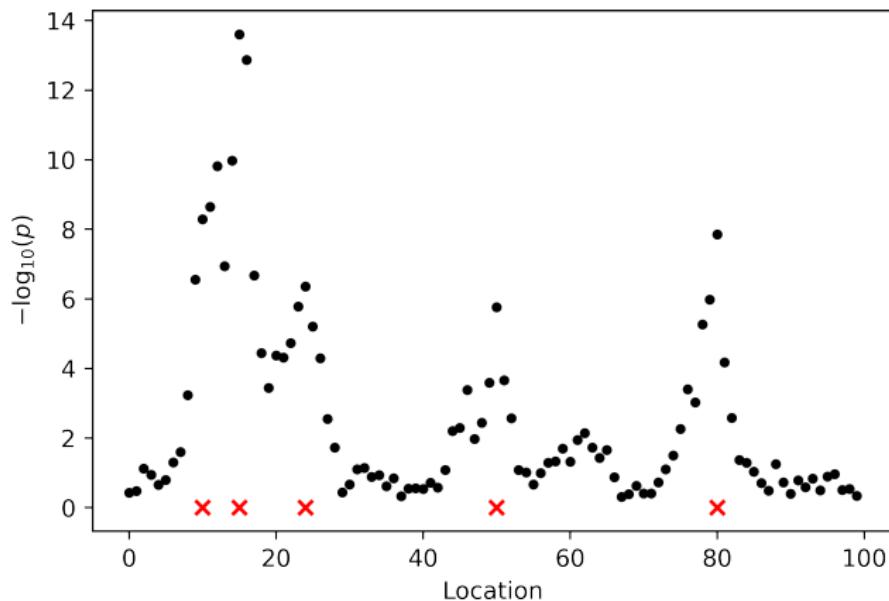


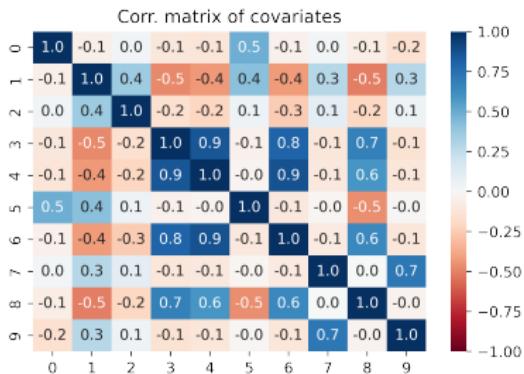
Figure: Cartoon Manhattan plot of genome; y-axis shows a measure of $\text{Corr}(Y, X_j)$ for $j = 1, \dots, 100$.

If you fit a linear model, you might find that nothing is significant!

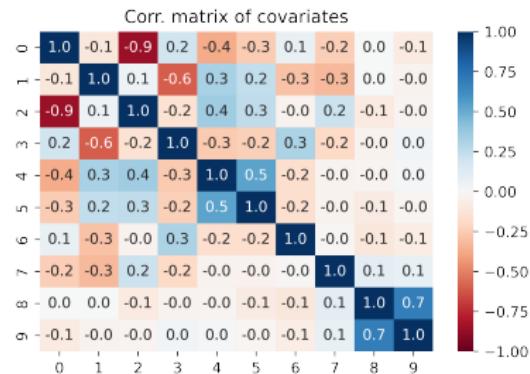
Motivation II: exploratory analysis of clinical trials

Correlated features appear in exploratory analysis of clinical trials:

- Proteomic/genomic data, even demographic data (age, baselines, etc)
- Throughout, think of X as a generic set of features
(could include treatments/interaction terms)



(a) Blum et al. (2010)



(b) Siedner et al. (2017)

Moral: for small to medium n , moderate correlations make it harder to identify treatment effect moderators / prognostic variables.

Contribution

Goals:

- Discover disjoint groups $G_1, \dots, G_R \subset \{1, \dots, p\}$ which each contain a signal
- Make R large and G_1, \dots, G_R small—both matter a lot!
- Control (e.g.) the FDR

Method: Bayesian Linear Programming (BLiP).

Input: posterior samples from any Bayesian model (e.g. Bayesian GLM/GAM)

Output: groups G_1, \dots, G_R that maximize power subject to FDR control.*

* FDR control assumes the Bayesian model is well-specified.

Contribution in a picture

Input: Samples from sparse Bayesian linear model.

Output:

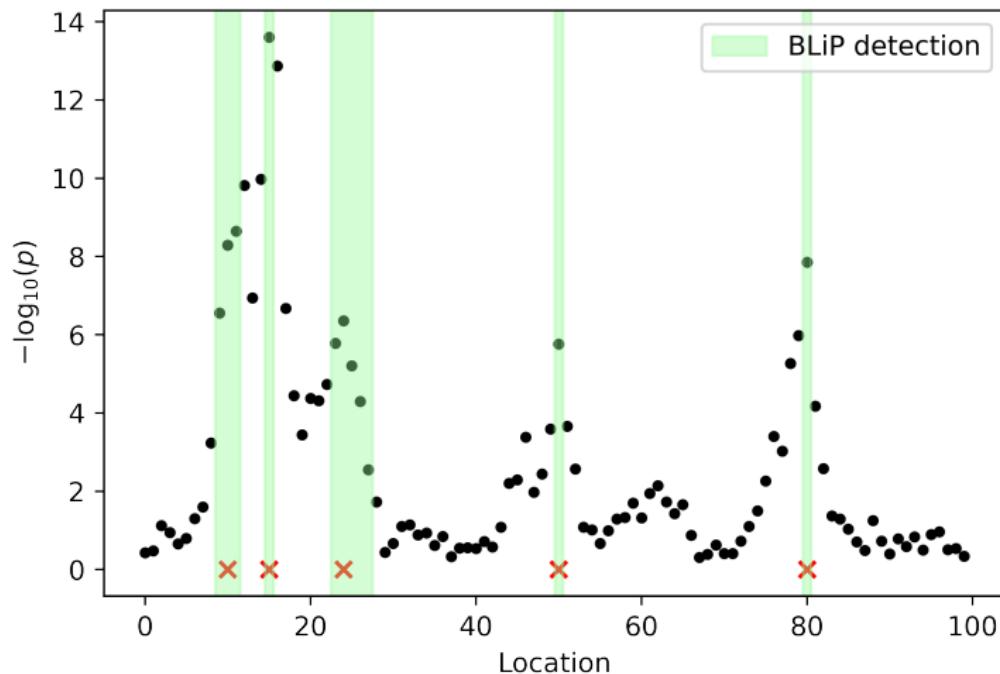


Figure: Cartoon of partial Manhattan plot of genome

Outline

- 1 Motivation
- 2 Methodology (BLiP)
- 3 Application to genetic fine-mapping
- 4 Advertisement for KeLP: a frequentist, knockoffs-based method (Gablenz and Sabatti, 2024)
- 5 Conclusion

Notation and assumptions

Notation: $X \in \mathbb{R}^p$ are features, $Y \in \mathbb{R}$ is an outcome, \mathcal{D} is dataset.

Assumption 1: The analyst specifies a Bayesian model which implies

$$\mathbb{E}[Y | X] = f_\theta(X) \text{ for } \theta \in \Theta$$

with $\theta \sim \pi$ sampled from some prior distribution.

- $S = \{j : f_\theta(X) \text{ depends on } X_j\} \subset [p]$ is the set of *signal variables*.

Assumption 2: The analyst can sample from the law of $\theta | \mathcal{D}$.

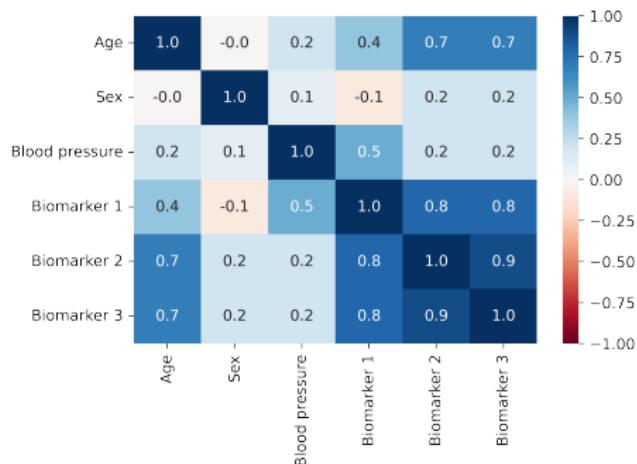
- These assumptions are not always reasonable! (see Section 5)
- But there is an enormous literature on sampling from these models
- Our question: how do we extract useful insights from these models after fitting them?

Problem statement (I)

Goal:

- Discover disjoint groups $G_1, \dots, G_R \subset \{1, \dots, p\}$ which each contain a signal
- Make R large and G_1, \dots, G_R small—both matter a lot!
- Control the (Bayesian) FDR

Emphasis: we want to pick G_1, \dots, G_R after seeing the data. E.g.:



How to group the features depends on the (unknown) signal size!

Problem statement (II)

Don't want to narrow potential discovery regions until *after* seeing data

Goal: look at the data and outputs regions G_1, \dots, G_R so as to:

$$\begin{aligned} \max & \quad \mathbb{E} [\text{Power}(G_1, \dots, G_R) \mid \text{Data}] \\ \text{s.t.} & \quad \text{FDR} := \mathbb{E} \left[\frac{\#\{G_r \text{ containing no signal}\}}{\max(1, R)} \mid \text{Data} \right] \leq q, \\ & \quad G_1, \dots, G_R \subset [p] \text{ are disjoint.} \end{aligned}$$

What does high **Power()** look like?

- As many (true) discovered regions G_r as possible
- Discovered regions G_r should be as small as possible

Existing work: no formalization of what “power” means, so cannot optimize it.

Defining resolution-adjusted power

Define a weighting function $w(G)$ that measures value of discovering a group

- Should penalize larger groups
- Canonical choice is inverse-size weighting: $w(G) = 1/|G|$
- Sum weights of true rejections to get Power():

$$\text{Power}(G_1, \dots, G_R) = \sum_{r=1}^R I_{G_r} w(G_r),$$

where $I_G = \mathbb{I}(G \cap S \neq \emptyset)$ is the indicator that G contains a signal (i.e., is a true discovery)

Remarks:

- Different w can accommodate very different scientific objections
- In practice, do we exactly know our “utility function”?
- We will see that the results are not too sensitive to precise specification

Method: Bayesian Linear Programming (I)

Method: directly solve the optimization problem:

$$\begin{aligned} \max \quad & \mathbb{E} [\text{Power}(G_1, \dots, G_R) \mid \text{Data}] \\ \text{s.t.} \quad & \text{FDR} := \mathbb{E} \left[\frac{\#\{G_r \text{ containing no signal}\}}{\max(1, R)} \mid \text{Data} \right] \leq q, \\ & G_1, \dots, G_R \subset [p] \text{ are disjoint.} \end{aligned}$$

Key observation: the power of a Bayesian method that discovers G_1, \dots, G_R is

$$\mathbb{E}[\text{Power}(G_1, \dots, G_R) \mid \text{Data}] = \mathbb{E} \left[\sum_{r=1}^R I_{G_r} w(G_r) \mid \text{Data} \right] = \sum_{G \subseteq [p]} p_G w(G) z_G,$$

- $p_G = P(G \text{ contains a signal} \mid \text{Data})$ can be computed Assumptions 1-2
- $z_G \in \{0, 1\}$ is indicator that we discover G

Method: Bayesian Linear Programming (II)

Theorem: the optimization problem is equivalent to the following integer LP:

$$\max_{\{z_G\}_{G \subseteq [p]}} \sum_G p_G w(G) z_G \quad (\text{Power})$$

$$\text{s.t.} \quad \sum_G (1 - p_G - q) z_G \leq 0 \quad (\text{FDR})$$

$$\sum_{G \subset [p]: j \in G} z_G \leq 1 \quad \forall j = 1, \dots, p \quad (\text{disjoint discoveries})$$

for decision variables $z_G \in \{0, 1\}$.

This is progress. Yet we have 2^p integer decision variables.

Method: Bayesian Linear Programming (III)

Problem: 2^p integer decision variables.

Solutions:

- ① Narrow the search space *after* looking at the data
 - Sublinear algorithm to discard $\{G : p_G \leq 0.001\}$
- ② Narrow the search space by imposing desirable structural constraints
 - E.g., ensure $|G| \leq 25$
- ③ If needed, solve the continuous relaxed LP; then round to obtain integers.

Result: Provable FDR control, verifiable near-optimality.

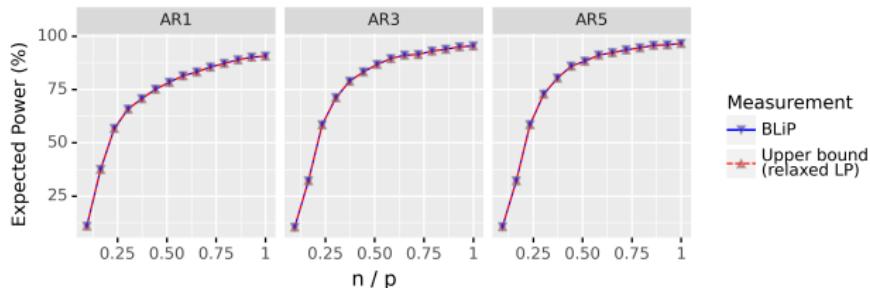


Figure: Expected power (BLiP) vs. upper bound

Putting it all together: BLiP

Input: Nearly any Bayesian model (via MCMC, variational inference) and any desired structural constraints on the discovery set

Output: disjoint discoveries which (1) verifiably nearly maximize power and (2) control the FDR.

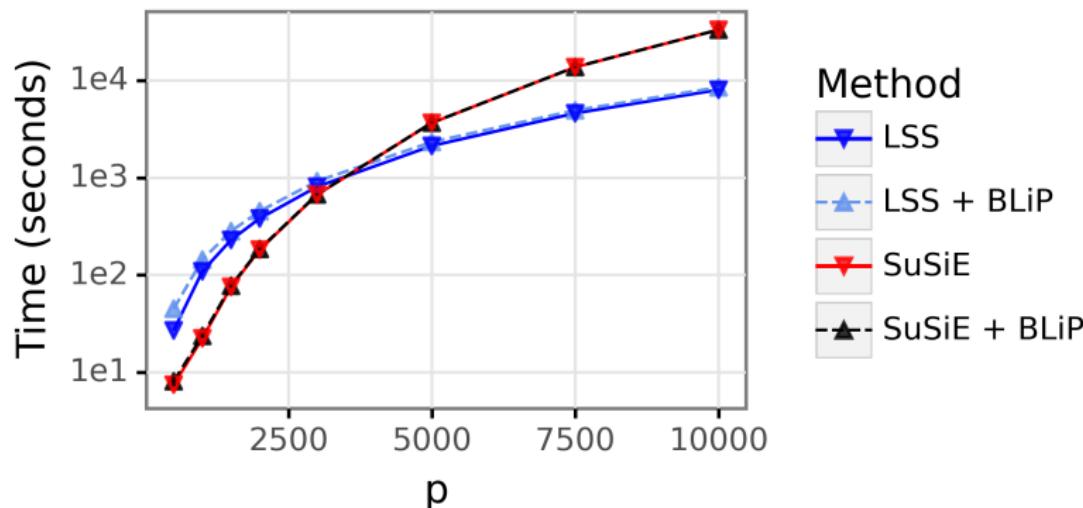


Figure: p denotes dimension of linear model being fit, with $n = p/2$

Outline

- 1 Motivation
- 2 Methodology (BLiP)
- 3 Application to genetic fine-mapping
- 4 Advertisement for KeLP: a frequentist, knockoffs-based method (Gablitz and Sabatti, 2024)
- 5 Conclusion

Fine-mapping setup

Dataset: $n \approx 337,000$, $p \approx 19,000,000$, four traits of interest.

Bayesian model: SuSiE (Wang et al., 2020)

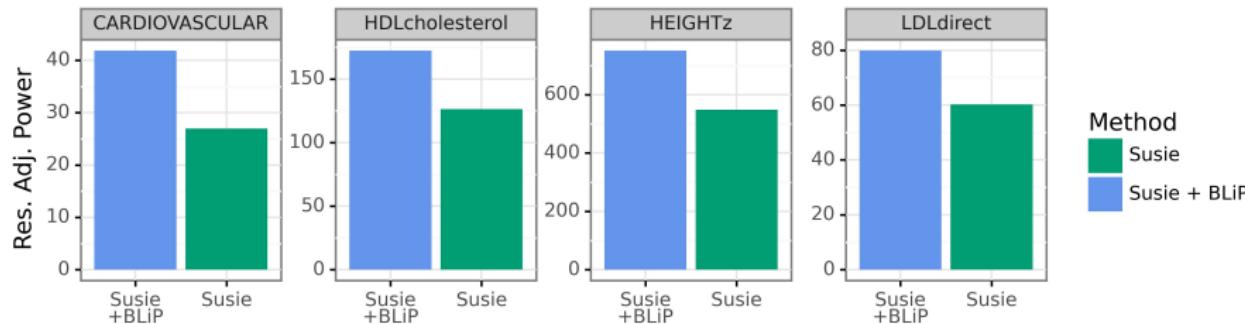
- ① SuSiE is a sparse Bayesian linear model that can be fit highly efficiently.
- ② Like BLiP, SuSiE returns regions $G_1^{\text{SuSiE}}, \dots, G_R^{\text{SuSiE}}$ of the genome.
- ③ However, SuSiE's regions are constructed heuristically.
 - Can we do better using a principled approach (BLiP)?

We run BLiP on top of a pre-fit SuSiE model from Weissbrod et al. (2019).

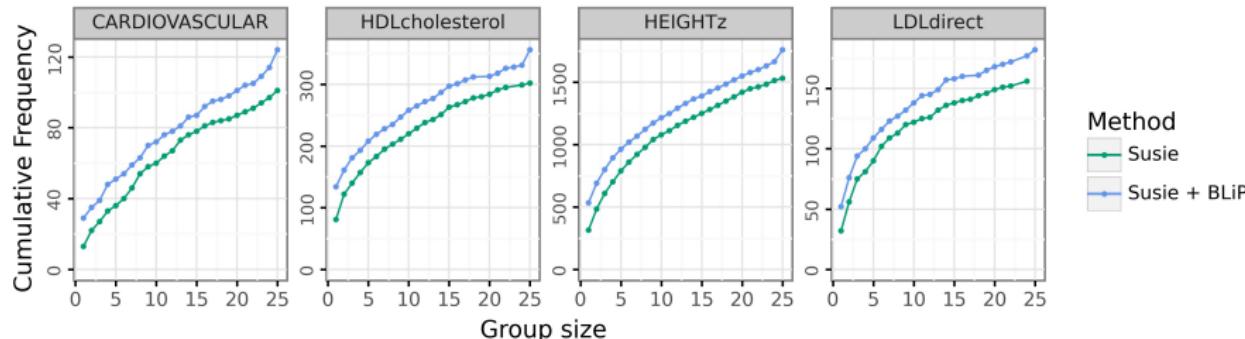
Fine-mapping results

UK Biobank data: $n \approx 337,000$, $p \approx 19,000,000$; BLiP takes < 1 min per trait

Resolution Adjusted Power on UK Biobank, N=337K



Cumulative Frequency of Discovered Group Sizes



Are our results real?

Trait	Corroboration Rate (SuSiE)	Corroboration Rate (<i>new</i>)
Height	53.5%	45.0%
HDL	57.0%	50.0%
LDL	67.3%	60.0%
Cardiovascular	82.2%	65.2%

Table: The proportion of discoveries which can be corroborated by a separate study in the NHGRI-EBI GWAS Catalog (Buniello et al., 2018).

Note the right-hand column only contains entirely new discoveries made by BLiP.

Our interpretation: this is a positive result, since all of the “low-hanging fruit” should lie in the left-hand column. Nonetheless, the numbers are comparable.

Outline

- 1 Motivation
- 2 Methodology (BLiP)
- 3 Application to genetic fine-mapping
- 4 Advertisement for KeLP: a frequentist, knockoffs-based method (Gablitz and Sabatti, 2024)
- 5 Conclusion

BLiP with knockoffs?

A weakness: BLiP assumes the Bayesian model is well-specified.

Gablenz and Sabatti (2024) also solve the BLiP optimization problem...

- ...but obtain model-free frequentist FDR guarantees.

Insights:

- Use knockoffs for model-free error control (Candes et al., 2018)
- Technical insight: use e-values to account for multiplicity (Wang and Ramdas, 2022)

TL;DR: one can perform resolution-adaptive variable selection as a frequentist.

Outline

- 1 Motivation
- 2 Methodology (BLiP)
- 3 Application to genetic fine-mapping
- 4 Advertisement for KeLP: a frequentist, knockoffs-based method (Gablenz and Sabatti, 2024)
- 5 Conclusion

Conclusion

BLiP is a powerful and efficient method for resolution-adaptive variable selection

- Provable (Bayesian) error control and verifiable near-optimality
- Substantial power gains in minutes on fine-mapping
- Software packages `pyblip` (Python) and `blipr` (R)

More in the paper:

- Applications to astronomy, change-point detection
- Potential for other signal discovery problems with spatial structure?

Paper available at: <https://arxiv.org/abs/2203.17208>

All code posted at: https://github.com/amspector100/blip_sims/

Thank you!

References

- Blum, J., Winikoff, B., Raghavan, S., Dabash, R., Ramadan, M. C., Dilbaz, B., Dao, B., Durocher, J., Yalvac, S., Diop, A., Dzuba, I. G., and Ngoc, N. T. N. (2010). Treatment of post-partum haemorrhage with sublingual misoprostol versus oxytocin in women receiving prophylactic oxytocin: a double-blind, randomised, non-inferiority trial. *Lancet*, 375(9710):217–223.
- Buniello, A., MacArthur, J. A., Cerezo, M., Harris, L. W., Hayhurst, J., Malangone, C., McMahon, A., Morales, J., Mountjoy, E., Sollis, E., Suveges, D., Vrousogou, O., Whetzel, P. L., Amode, R., Guillen, J. A., Riat, H. S., Trevanion, S. J., Hall, P., Junkins, H., Flicek, P., Burdett, T., Hindorff, L. A., Cunningham, F., and Parkinson, H. (2018). The NHGRI-EBI GWAS Catalog of published genome-wide association studies, targeted arrays and summary statistics 2019. *Nucleic Acids Research*, 47(D1):D1005–D1012.
- Candes, E., Fan, Y., Janson, L., and Lv, J. (2018). Panning for gold: Model-X knockoffs for high-dimensional controlled variable selection. *Journal of the Royal Statistical Society: Series B*, 80(3):551–577.
- Gablenz, P. and Sabatti, C. (2024). Catch me if you can: signal localization with knockoff e-values. *Journal of the Royal Statistical Society Series B: Statistical Methodology*, page qkae042.
- Siedner, M. J., Bwana, M. B., Moosa, M.-Y. S., Paul, M., Pillay, S., McCluskey, S., Atkinson, A. J. K., Marinelli, W., Marshall, P., Dill, J., ...