

Biomedical Data Analysis

Introducing the Problem



Biomedical Data Analysis

Assume we are concated by a bio-medical lab



- They have collected data about patients with a certain condition
- ...And they want to get a better **understanding** of the involved process

Our Dataset

This use case is based on a real-world example

...But for privacy and simplicity reasons we are going to use synthetic data

```
In [21]: data, name_map = util.generate_data(size=500, seed=42)
data
```

```
Out[21]:
```

	u0	u1	u2	u3	u4	u5	u6	u7	u8	u9	u10	u11	u12	u13	u14
0	0.0	4.052587	0.0	0.0	1.069842	-1.541433	0.984682	2.069759	-0.998354	1.615419	1.0	0.0	3.905281	1.422892	0.0
1	0.0	2.520945	1.0	0.0	-1.924131	-2.583550	4.663292	-1.633941	0.324484	0.426927	1.0	0.0	1.319270	1.771152	0.0
2	0.0	1.061444	0.0	1.0	0.288059	-0.974309	2.641967	0.823806	1.340944	1.498628	1.0	0.0	-1.072016	-0.750879	0.0
3	1.0	0.523647	1.0	1.0	1.824137	-2.841816	4.099077	-2.287757	1.230311	1.628930	1.0	1.0	1.299762	2.085999	1.0
4	0.0	2.010178	0.0	0.0	-0.050319	-2.620961	3.162254	-0.803245	-0.840076	0.507807	0.0	0.0	0.307414	-0.884796	0.0
...
495	1.0	7.434214	1.0	1.0	-1.948899	-1.283335	2.303599	0.505025	2.488334	1.713777	1.0	0.0	5.451237	0.257810	1.0
496	0.0	7.857776	1.0	0.0	0.239719	-1.181798	2.301580	-1.150514	-0.703677	2.100331	0.0	0.0	4.269326	0.760440	0.0
497	1.0	3.348010	0.0	0.0	0.147685	-2.541503	2.887376	-0.372831	1.435787	0.967976	0.0	0.0	0.576445	0.450504	0.0
498	1.0	2.784484	0.0	0.0	-2.082640	-1.224654	4.271790	-0.269379	0.876229	0.745919	1.0	1.0	0.424243	-1.446797	0.0
499	1.0	1.808553	1.0	0.0	-2.458112	-0.695131	3.231171	-2.915948	-0.078209	2.988293	1.0	1.0	-0.618186	-0.810217	1.0

500 rows × 16 columns

How do we start?



Our Dataset

Let's have a first look at the dataset

In [22]: `data.describe()`

Out [22]:

	u0	u1	u2	u3	u4	u5	u6	u7	u8	u9
count	500.000000	500.000000	500.000000	500.000000	500.000000	500.000000	500.000000	500.000000	500.000000	500.000000
mean	0.396000	1.828261	0.514000	0.330000	-0.030795	-1.420177	2.995727	-0.361947	0.449518	1.080533
std	0.489554	2.112032	0.500305	0.470684	1.440194	0.961449	1.008219	1.463672	0.980352	1.300855
min	0.000000	0.055230	0.000000	0.000000	-4.699421	-4.244599	0.033381	-5.647642	-3.000955	-2.883851
25%	0.000000	0.547481	0.000000	0.000000	-1.034566	-2.138448	2.289419	-1.295046	-0.186973	0.188749
50%	0.000000	1.127278	1.000000	0.000000	0.023120	-1.403097	3.044132	-0.320448	0.412165	1.058317
75%	1.000000	2.127061	1.000000	1.000000	0.927888	-0.836187	3.714111	0.561467	1.071871	1.974449
max	1.000000	13.486418	1.000000	1.000000	3.747794	1.706728	5.906263	4.334036	3.401382	5.526514

- There is one target binary variable Y , representing the condition under study
- All other columns represent potentially correlate variables
- We are going to refer to them as "potential correlates"



Categorical and Numerical Variables

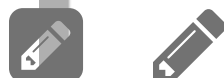
Some of the potential correlates are numeric, others are categorical

```
In [23]: # Identify numeric and categorical columns
num_cols = [c for c in data.columns[:-1] if len(data[c].unique()) > 2]
cat_cols = [c for c in data.columns[:-1] if len(data[c].unique()) == 2]
print(f'Numeric: {num_cols}')
print(f'Categorical: {cat_cols}')
```

```
Numeric: ['u1', 'u4', 'u5', 'u6', 'u7', 'u8', 'u9', 'u12', 'u13']
Categorical: ['u0', 'u2', 'u3', 'u10', 'u11', 'u14']
```

- In this synthetic dataset, all categorical variables are binary
- ...Which explains the simple filter we used to identify them

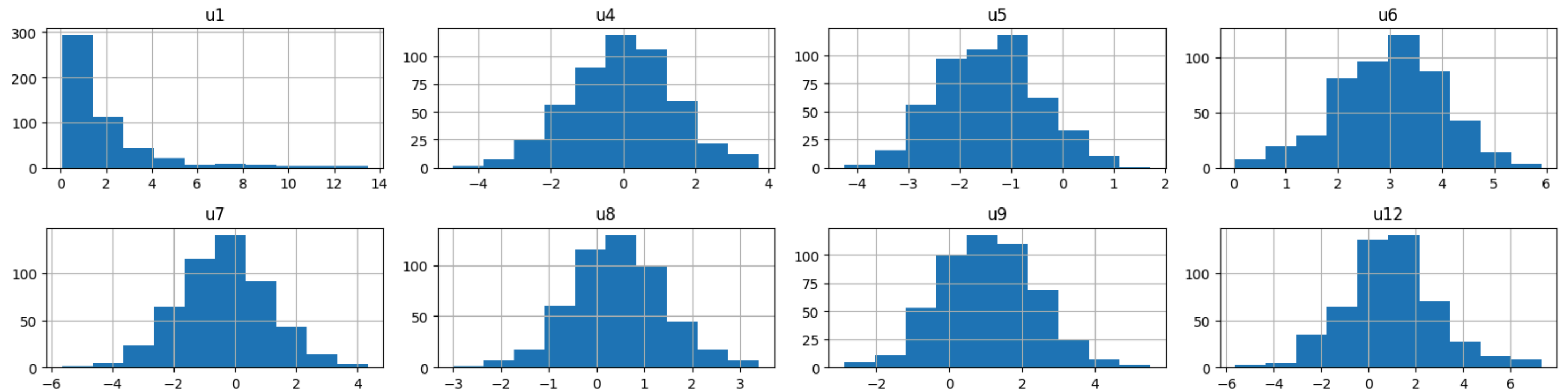
In a real world setting, you'd need to talk to a domain expert for this



Checking the Distributions

Let's check the distribution of the **numerical** candidate correlates

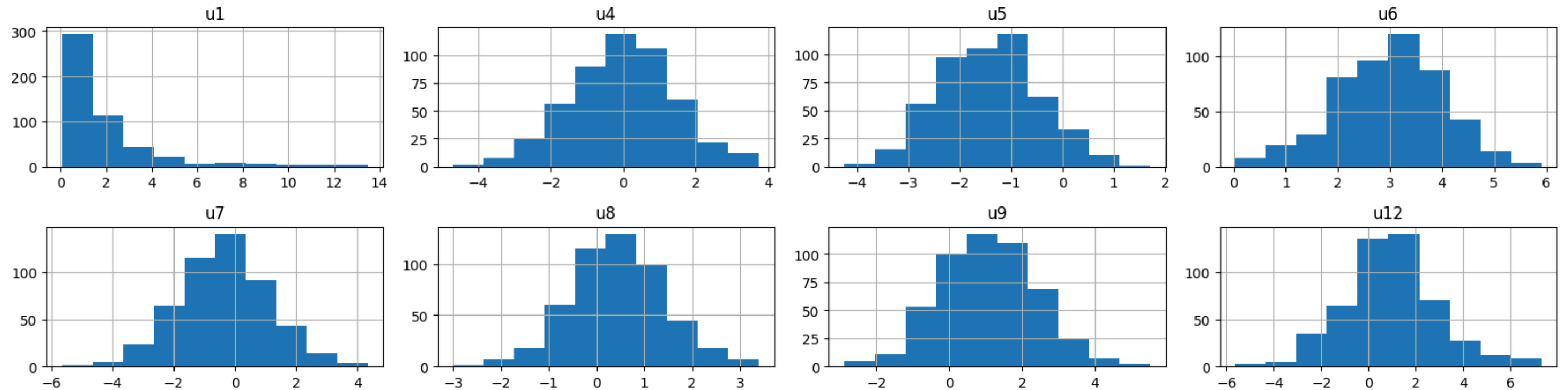
```
In [24]: _, axes = plt.subplots(nrows=2, ncols=int(np.ceil(len(num_cols)//2)), figsize=figsize)
         for ax, cname in zip(axes.ravel(), num_cols):
             data.hist(cname, ax=ax)
         plt.tight_layout()
```



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         plt.tight_layout()
```



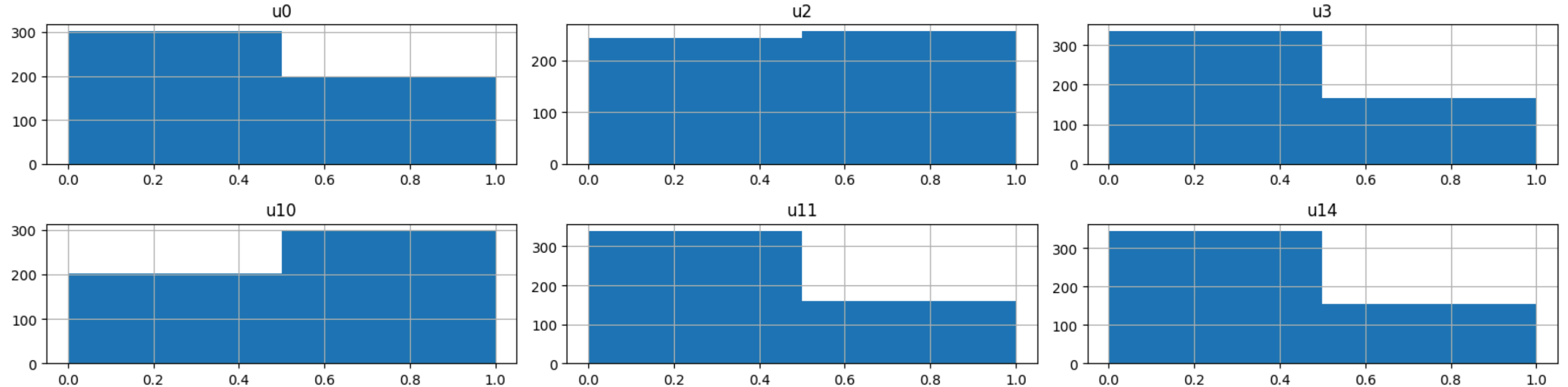
Most of them seem to follow a Normal distribution



Checking the Distributions

Let's check the distribution of the **binary** candidate correlates

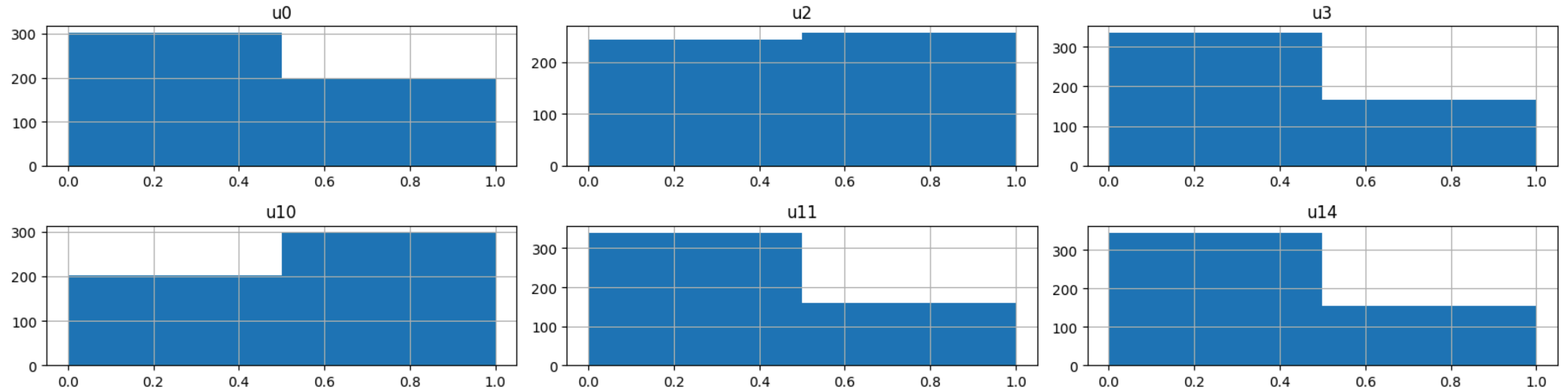
```
In [25]: _, axes = plt.subplots(nrows=2, ncols=int(np.ceil(len(cat_cols)//2)), figsize=figsize)
         for ax, cname in zip(axes.ravel(), cat_cols):
             data.hist(cname, ax=ax, bins=2)
         plt.tight_layout()
```



Checking the Distributions

Let's check the distribution of the **binary** candidate correlates

```
In [25]: _, axes = plt.subplots(nrows=2, ncols=int(np.ceil(len(cat_cols)//2)), figsize=figsize)
         for ax, cname in zip(axes.ravel(), cat_cols):
             data.hist(cname, ax=ax, bins=2)
         plt.tight_layout()
```

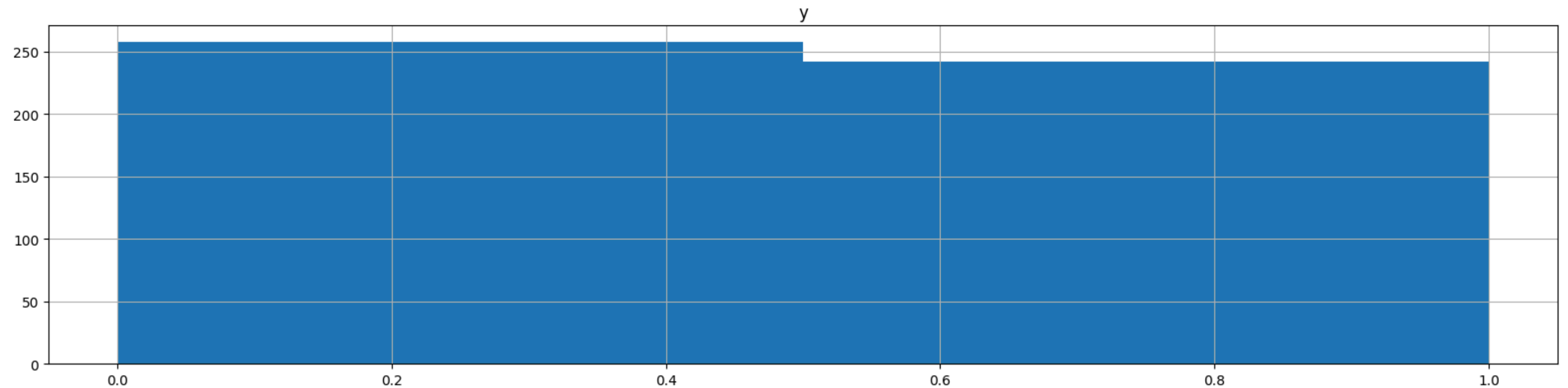


Some are well balanced, others less so

Checking the Distributions

Let's check the **target** distribution

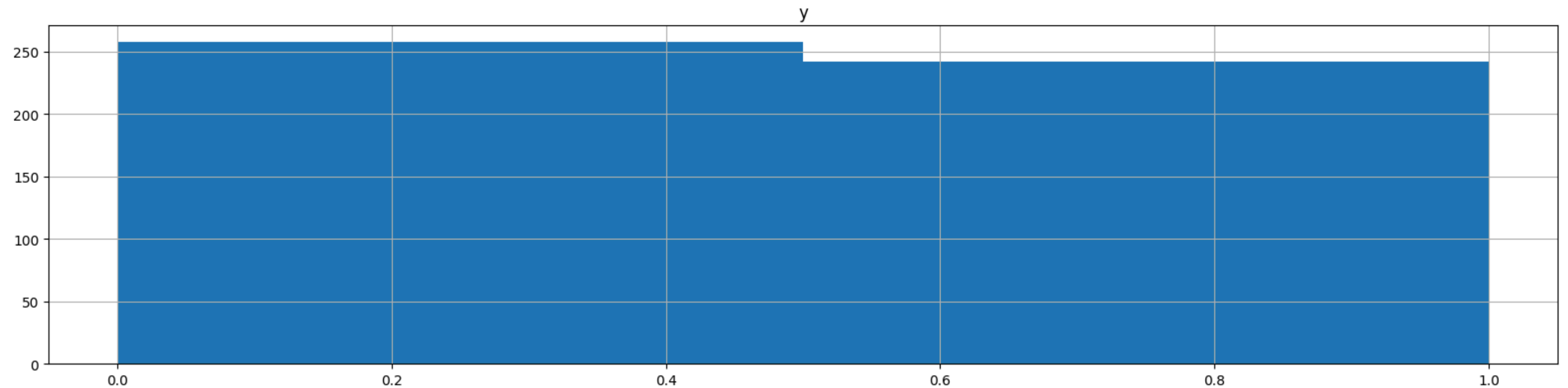
```
In [26]: data.hist('y', bins=2, figsize=figsize)  
plt.tight_layout()
```



Checking the Distributions

Let's check the **target** distribution

```
In [26]: data.hist('y', bins=2, figsize=figsize)  
plt.tight_layout()
```



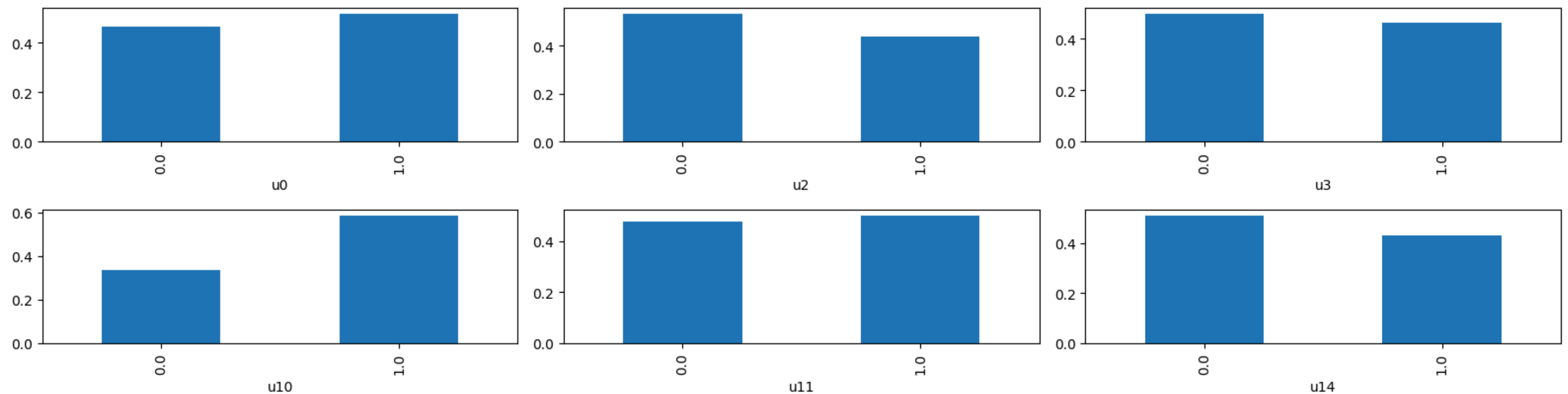
The target distribution quite balanced



Checking Univariate Dependencies

Let's check the fraction of $Y = 1$ for the **categorical** candidates

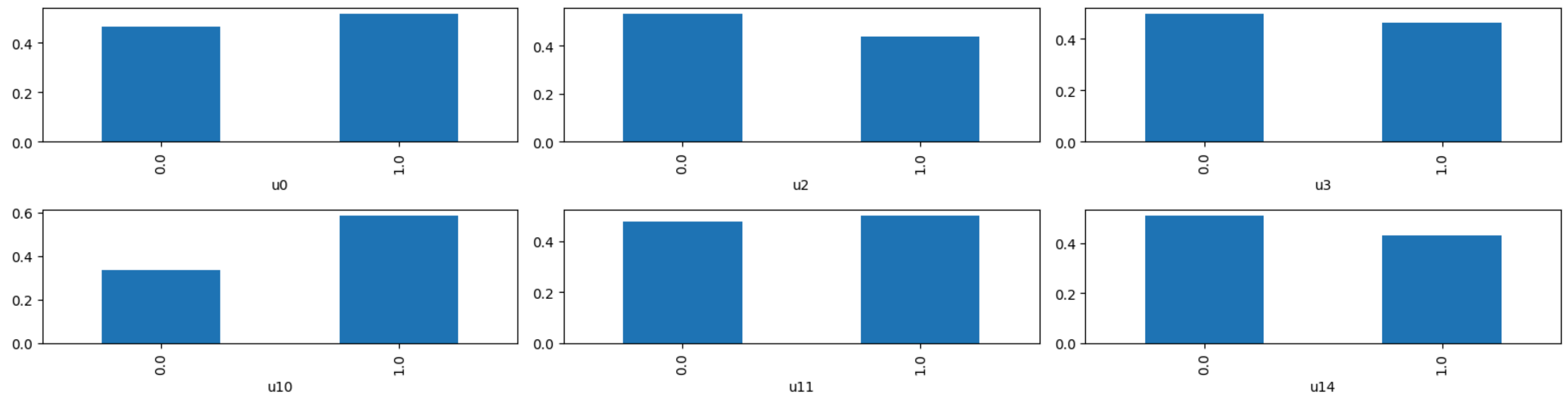
```
In [27]: _, axes = plt.subplots(nrows=2, ncols=int(np.ceil(len(cat_cols)//2)), figsize=figsize)
for ax, cname in zip(axes.ravel(), cat_cols):
    data.groupby(cname)['y'].mean().plot.bar(ax=ax)
plt.tight_layout()
```



Checking Univariate Dependencies

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```
In [27]: _, axes = plt.subplots(nrows=2, ncols=int(np.ceil(len(cat_cols)//2)), figsize=figsize)
for ax, cname in zip(axes.ravel(), cat_cols):
    data.groupby(cname)['y'].mean().plot.bar(ax=ax)
plt.tight_layout()
```

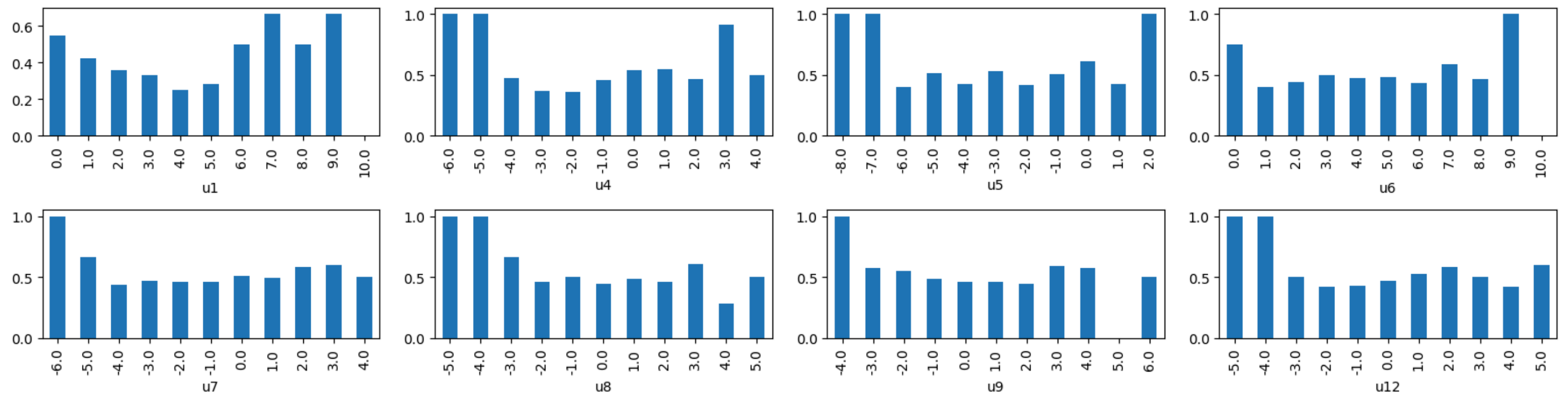


A few of them seems to have a correlation, other cases are less clear

Checking Univariate Dependencies

Let's check the fraction of $y = 1$ for the **numerical** candidates

```
In [28]: _, axes = plt.subplots(nrows=2, ncols=int(np.ceil(len(num_cols)//2)), figsize=figsize)
for ax, cname in zip(axes.ravel(), num_cols):
    bin_size = (data[cname].max() - data[cname].min()) / 10
    data['y'].groupby(data[cname] // bin_size).mean().plot.bar(ax=ax)
plt.tight_layout()
```



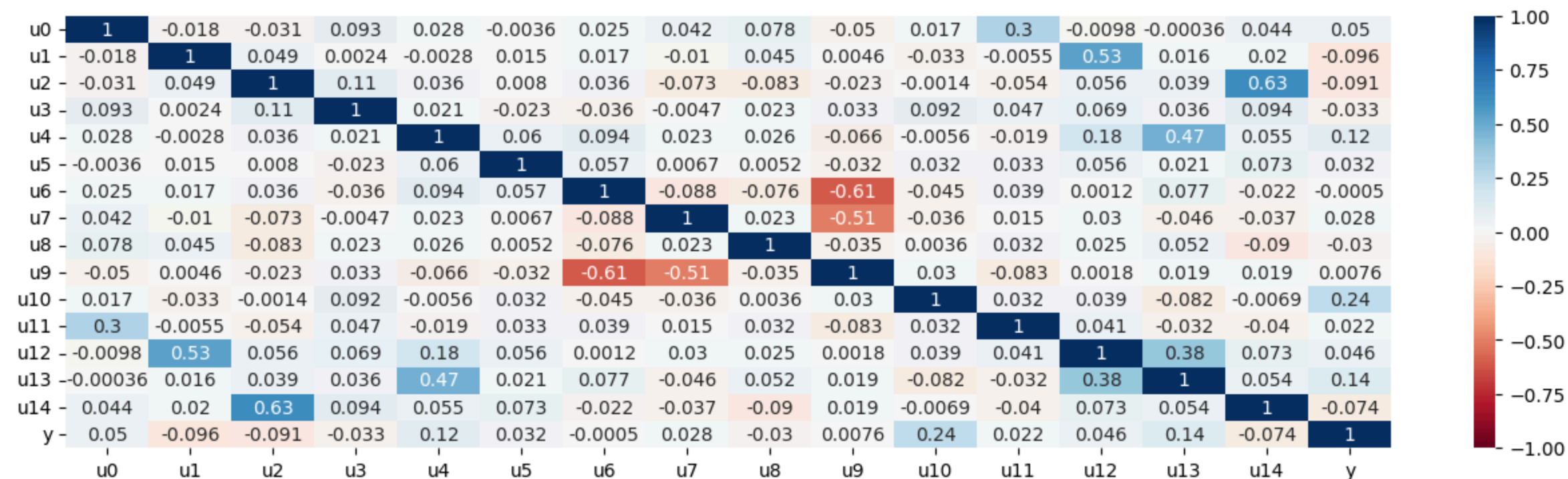
Most of them appear to have some non-linear correlation

Checking Linear Correlations

It's worth checking how all features are correlated

One way to do it is by plotting a correlation matrix (e.g. Pearson)

```
In [29]: plt.figure(figsize=figsize)
sn.heatmap(data.corr(method='pearson'), annot=True, vmin=-1, vmax=1, cmap='RdBu');
```



■ Sparse correlations in general, weak (linear) correlations for Y

**So far we have just inspected our dataset, but...
what is exactly our goal?**



Use Case Objective

Unlike in classical ML tasks, we don't have an **estimation** problem

Rather, our goal is **understanding** the process behind the data

- We want to identify the true **correlates** among our candidates
- We want to see **how** they are linked to the target y

In an ideal world, we'd like to know about **causal** relationships

...But in practice, we'll need to be happy with correlations

- Studying causality is indeed possible (a good start is Judea Pearl's book)
- ...But also very challenging, and there's no general and mature tool available

So, we'll count on the domain expert to check the correlations



Use Case Objective

Our setup also explains a quirk in the dataset

All variables except the target are called U_j , for "unknown"

- This is synthetic data, so nothing is really unknown
- In fact, the ground truth process linking Y to U is available

However, for the sake of this exercise, such process will be hidden

- We will analyze the data pretending we have no such knowledge
- At the end of our exercise we'll check the ground truth

...And we'll see how close we got to the truth!



Biomedical Data Analysis

A Baseline Approach



A Baseline Approach

Our goal is **understanding** the process behind the data

Of of many possible ways to do it consist in:

- Training an approximate model via Machine Learning
- Studying the model as a proxy for the real process

Basically, we use a ML model as an **analysis tool**

For this approach to work, we need the ML model to be **explainable**

- A few model naturally enjoy this property (e.g. linear models, simple DTs)
- Explaining other models is not obvious (e.g. Neural Networks, large ensembles)

 We will start with the simplest option: Logistic Regression

Data Preprocessing

We start with the usual data preprocessing

We will treat all candidate correlates as inputs

```
In [30]: # Input-output separation
X, y = data[data.columns[:-1]].copy(), data[data.columns[-1]].copy()
# Train-test split
X_train, X_test, y_train, y_test = train_test_split(X, y, test_size=0.2, random_state=42)
# Input standardization
scaler = StandardScaler()
X_train[num_cols] = scaler.fit_transform(X_train[num_cols])
X_test[num_cols] = scaler.transform(X_test[num_cols])
```

Even if we don't care about estimates, we need a **test set**

- This will allow us to check the model for overfitting

We also need to standardize all numeric features

- This will make the model coefficients more easily interpretable

On the Danger of Overfitting

We plan to use our model as proxy for the true process

...Which makes **overfitting is especially bad**

- Our results will strictly apply only to the model
- ...And they will be as general as the model

We will use **L1 regularization on this purpose**

Scikit learn support L1 regularizers for Logistic Regression in the form:

$$\operatorname{argmin}_{\theta} H(y, f(x, \theta)) + \frac{1}{C} \|\theta\|_1$$

- We encourage the weights to be close to 0
- ...And we attempt to sparsify the weights



Training our "Proxy" Model

We can calibrate the C parameter via cross-validation

We'll need the SAGA solver to train our model with L1 regularization

```
In [31]: base_est = LogisticRegression(penalty='l1', solver='saga')
         param_grid={'C': 1. / np.linspace(1e-1, 1e4, 100)}
         gscv = GridSearchCV(base_est, param_grid=param_grid, scoring='roc_auc')
         gscv.fit(X_train, y_train)
         lr, lr_params = gscv.best_estimator_, gscv.best_params_
```

Then we can check the performance of the refitted estimator

```
In [32]: lr_score_cv, lr_score_test = gscv.best_score_, roc_auc_score(y_test, lr.predict_proba(X_test)
         print(f'AUC score for C={lr_params["C"]:.2f}: {lr_score_cv:.2f} (cross-validation), {lr_score_test:.2f} (test)')

AUC score for C=10.00: 0.64 (cross-validation), 0.60 (test)
```

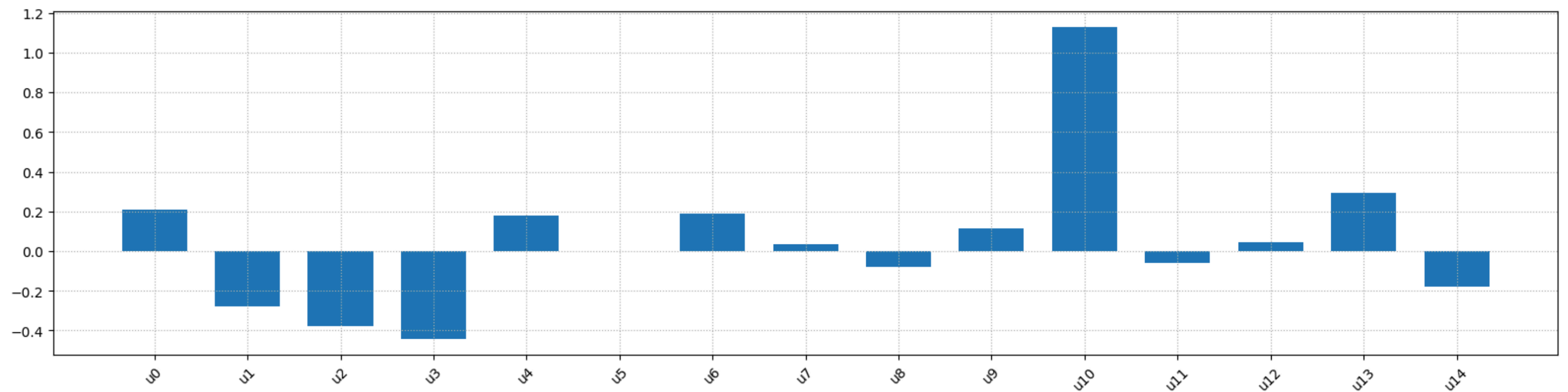
 We use the AUC score, since this is not a real classification problem



Coefficient Analysis

Finally, we can analyze the model coefficients

```
In [33]: lr_coefs = pd.Series(index=X.columns, data=lr.coef_[0])  
util.plot_bars(lr_coefs, figsize=figsize)
```



- Some variables seem to be more important than others
- The sign tells us how they are linked to the target



**This baseline approach has *many* issues
Can you spot a few ones?**



Three Key Issues with our Baseline

Issue 1: our model has **poor accuracy**

- An AUC score of 0.6 is not much above random
- ...Hence, studying our model will say little about the data



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Issue 2: our model can only capture **linear correlations**

- We can capture neither non-linear effects
- ...Nor interactions among the variables



Three Key Issues with our Baseline

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Issue 2: our model can only capture **linear correlations**

- We can capture neither non-linear effects
- ...Nor interactions among the variables

Issue 3: the coefficients are not sparse

- The L1 terms needs both to sparsify and to prevent overfitting
- ...And it cannot do both things effectively

 Additionally: it's unclear what a good level of sparsification might be

Biomedical Data Analysis

Intepretability with Non-Linear Models



Dealing with Non-Linearities

We'll start by switching to a non-linear model

By doing so:

- We can still account for non-linear correlations
- We can account for interactions among variables
- We might reach a much better accuracy
- ...And hence have a more representative proxy model



Dealing with Non-Linearities

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By doing so:

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Of course there is a price to pay

- Non-linear models are less easy to interpret
- ...And they are at a much higher risk of overfitting



Gradient Boosted Trees Model

We'll train a Gradient Boosted Trees model

We'll rely on the Extreme Gradient Boosting package (XGBoost) for this

```
In [34]: base_est = xgboost.XGBRegressor(objective='reg:logistic', tree_method='hist', importance_type='gain',
      param_grid={'max_depth': [2, 3, 4], 'n_estimators': list(range(20, 41, 5)), 'reg_lambda': np.arange(0.01, 0.1, 0.01)},
      gscv = GridSearchCV(base_est, param_grid=param_grid, scoring='roc_auc')
      gscv.fit(X, y)
      xbm, xbm_params = gscv.best_estimator_, gscv.best_params_
```

XGBoost is a library for fast, distributed, training of GBT models

It has support for **multiple loss functions**

- We are using "reg:logistic", which refers binary cross-entropy

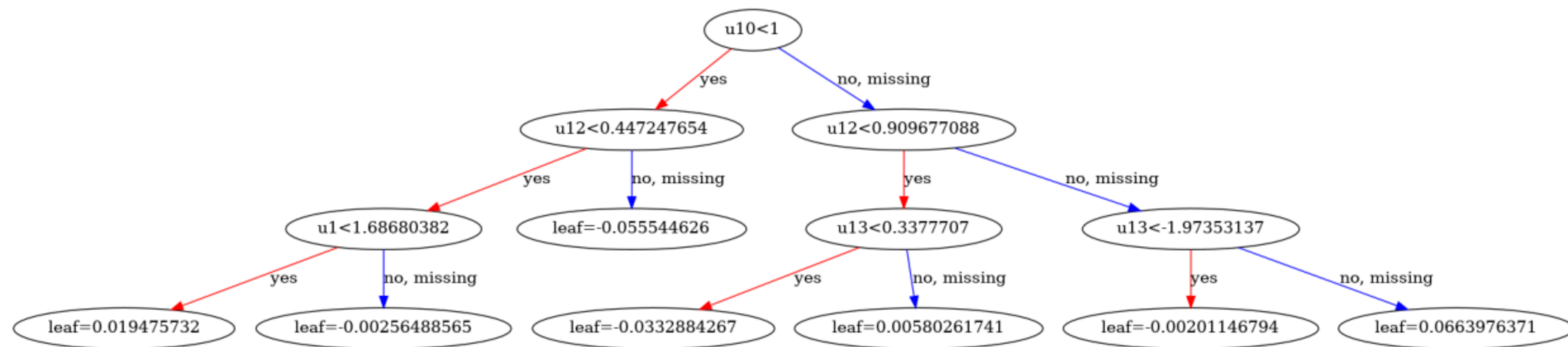
...And for **regularization** (often missing in tree-based models)

✎ The "reg_lambda" parameter refers to the weight of an L2 regularization term

Gradient Boosted Trees Model

It's easier to see how regularization work by checking a tree in the ensemble

```
In [35]: plt.figure(figsize=figsize)
xgboost.plot_tree(xbm, ax=plt.gca(), num_trees=0);
```



- Assuming T is the number of leaves and w_j is the label assigned to each leaf
- ...Then the regularization term is in the form $\sum_{k=1}^T w_j^2$

Gradient Boosted Trees Model

On our dataset, a GBT model has substantially better performance

```
In [36]: xbm_score_cv, xbm_score_test = gscv.best_score_, roc_auc_score(y_test, xbm.predict(X_test))
print(f'AUC score for {xbm_params}: {xbm_score_cv:.2f} (cross-validation), {xbm_score_test:.2f} (test)')

AUC score for {'max_depth': 3, 'n_estimators': 25, 'reg_lambda': 200.0}: 0.81 (cross-validation), 0.79 (test)
```

- The AUC score is much higher now
- There is no significant overfitting

It seems we finally have a model that we can trust



Gradient Boosted Trees Model

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```
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```

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It seems we finally have a model that we can trust

However, we know have an ensemble of many non-linear models

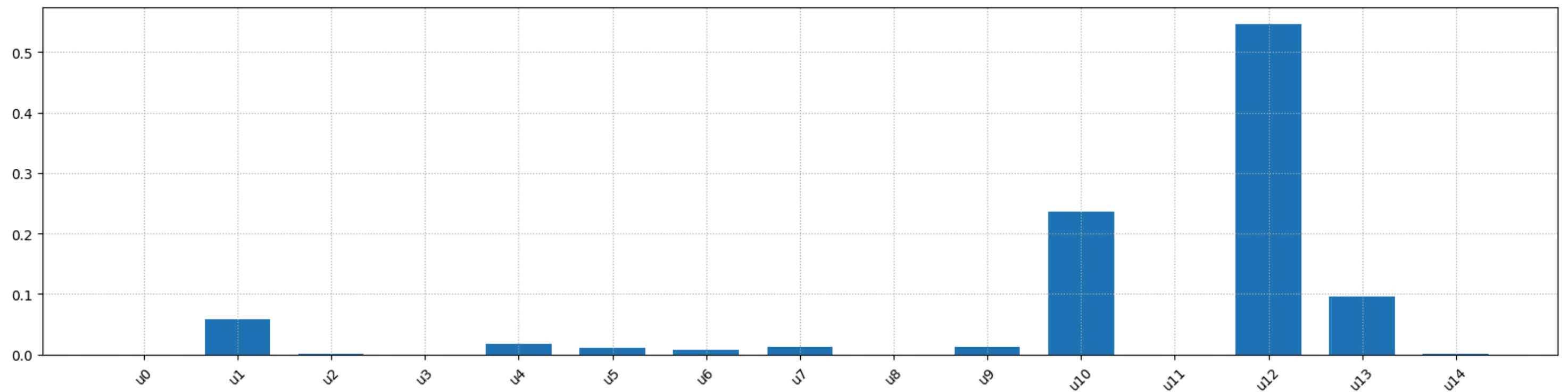
How can we make sense of that?



Feature Importances

The first option one can probably think of is using **feature importances**

```
In [37]: xbm_imp = pd.Series(index=X.columns, data=xbm.feature_importances_)
util.plot_bars(xbm_imp, figsize=figsize)
```



- The scores differ significantly from those obtained for linear regression (as expected)
- ...But what do they represent?

Which Feature Importances?

Feature importance is typically presented as this:

- For each input x_j , we sum the associated **gain** at training time
- Once training is over, we normalize the scores so that they sum up to 1

However, there are **other ways to define importance**

XGBoost supports 5 different approaches:

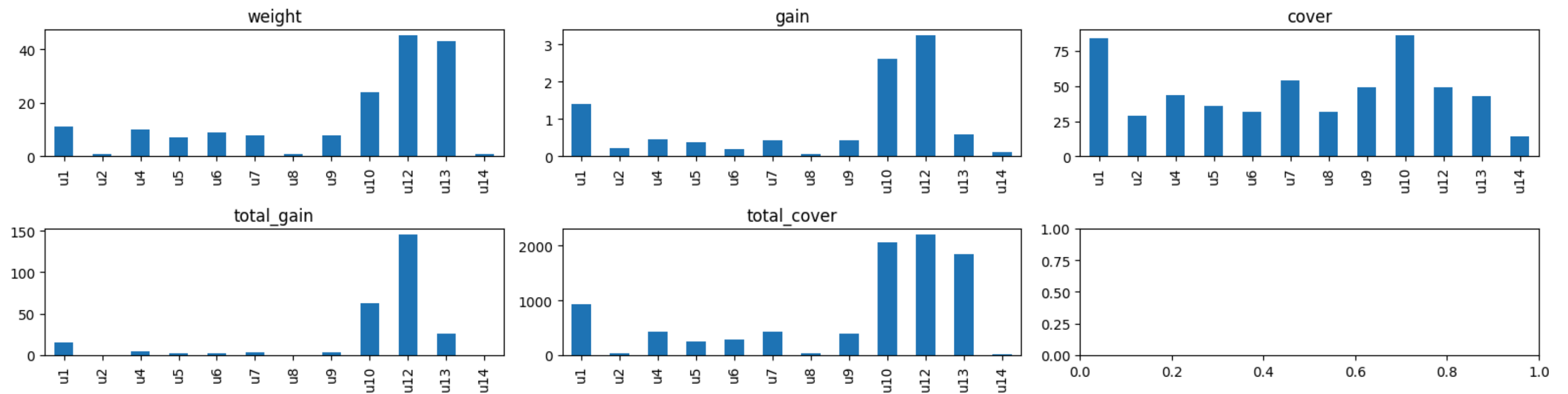
- "weight": number of times an attribute is used to split
- "gain": average gain associated to splits over an attribute
- "cover": average number of examples for which an attribute is used to decide
- "total_gain": as above, but replacing the average with a sum
- "total_cover": as above, but replacing the average with a sum



Which Feature Importances?

The values of the multiple feature importances can be quite different:

```
In [38]: _, axes = plt.subplots(nrows=2, ncols=3, figsize=figsize)
         for ax, imp_type in zip(axes.ravel(), ['weight', 'gain', 'cover', 'total_gain', 'total_cover']):
             pd.Series(xbm.get_booster().get_score(importance_type=imp_type)).plot.bar(ax=ax, title=imp_type)
         plt.tight_layout()
```



Importance and Data

Moreover, most importance scores are computed w.r.t. a dataset:

E.g. in XGBoost "gain", "cover", "total_gain", and "total_cover"

- For this reason, they are not really properties of the model
- ...But rather of the model and a reference sample

This means that the score semantic depends on the reference sample

By default, importances are computed on the training set

...Which means they are susceptible to overfitting

- The model might split on an attribute because it really is importance
- ...But also due to a spurious correlation



Permutation Importance

We can improve things by changing the way we compute importance

Given a **reference sample** $\{x_i, y_i\}_{i=1}^m$

- We can evaluate the performance of our model on the sample
- ...With that of a modified sample where the j -th input is **made** unimportant

For example, we can achieve that by **permuting the values of the input**

- This will preserve the distribution of the input
- ...But it will break all its correlations

Then, we look at the change in the model performance

- If it is small, the attribute is really unimportant
- Otherwise, the attribute is important

These scores are known as **permutation importances**



Permutation Importance

Permutation importances are robust w.r.t. spurious correlations

- We just need to repeat the process multiple times
- ...And record means and standard deviations

It's unlikely that we get a high score very often by accident

They allow us to **choose** our reference sample:

On the **training** set, the model might have overfit over the data

- The performance gap will be wider
- ...And the score will reflect how the model **is using** the data

On the **test set**, overfitting will make less of a difference

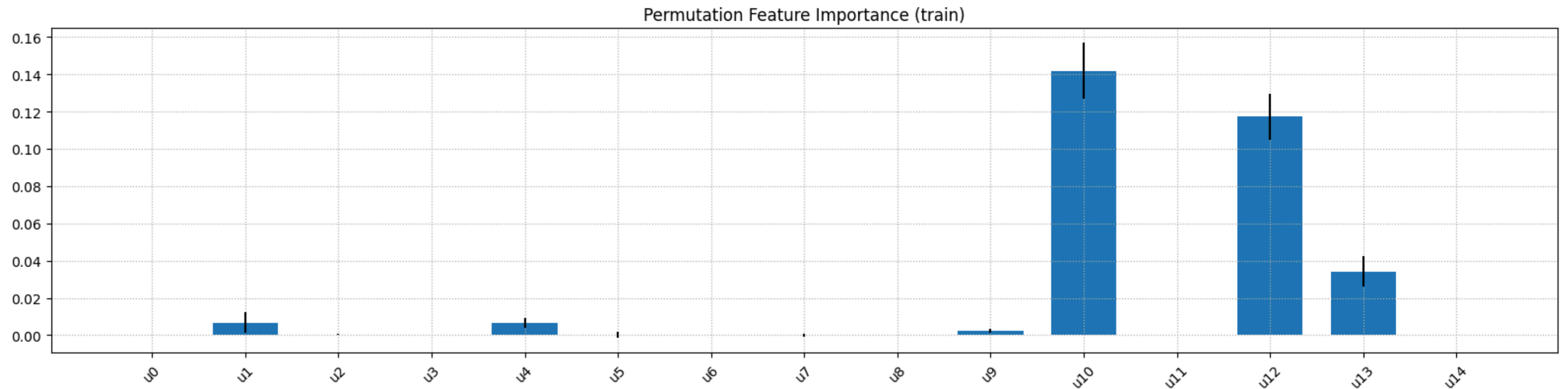
- ...And the score will reflect how correlated the attribute is with the target



Permutation Importances, on our Example

Let's check the **training** permutation importances in our case study

```
In [44]: r_train = permutation_importance(xbm, X_train, y_train, n_repeats=30, random_state=42)
xbm_p_imp = pd.Series(index=X.columns, data=r_train.importances_mean)
util.plot_bars(xbm_p_imp, figsize=figsize, std=r_train.importances_std, title='Permutation F
```



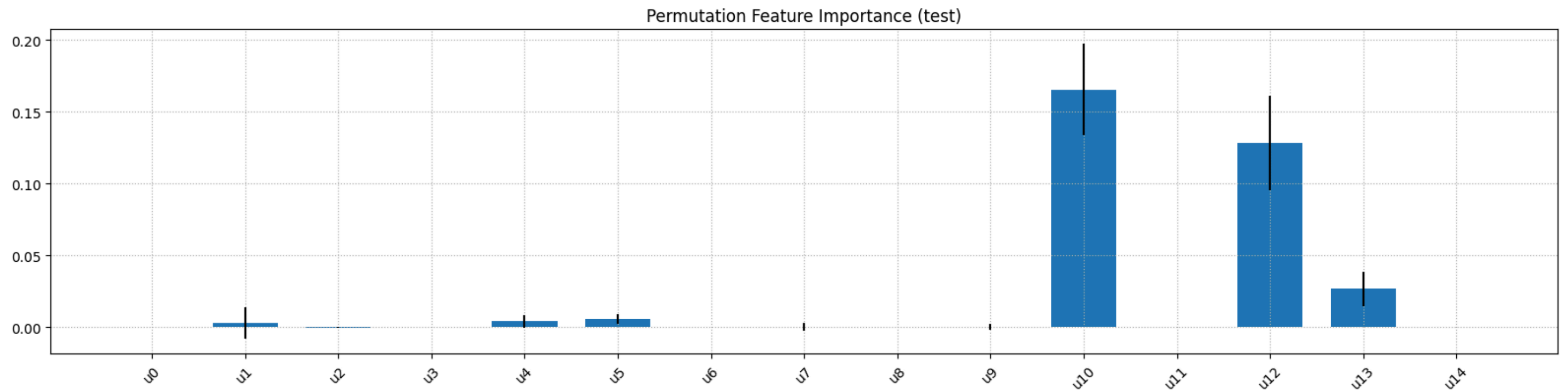
- The closely resemble those XGB "total_gain", but they are more sparse



Permutation Importances, on our Example

Let's check the **test** permutation importances in our case study

```
In [45]: r_test = permutation_importance(xbm, X_test, y_test, n_repeats=30, random_state=42)
xbm_p_imp = pd.Series(index=X.columns, data=r_test.importances_mean)
util.plot_bars(xbm_p_imp, figsize=figsize, std=r_test.importances_std, title='Permutation Fe
```



- A few low-importance features become even less relevant on the test data



Biomedical Data Analysis

Additive Feature Attribution



What we Gained, What we Lost

When we switched from Logistic Regression to GBTs we gained a lot

- A reliable proxy model
- A well defined and transparent feature importance definition
- Sparse and reliable importance scores



What we Gained, What we Lost

When we switched from Logistic Regression to GBTs we gained a lot

- A reliable proxy model
- A well defined and transparent feature importance definition
- Sparse and reliable importance scores

However, we also lost something:

With Linear Regression, we used to be able to:

- Identify the **direction** of the correlation (through the coefficient sign)
- ...And explain **individual examples**, by looking at the difference:

$$\theta^T x - \mathbb{E}_{x' \in P(X)} [\theta^T x']$$



Explaining Individual Examples

Let's look again at the last equation:

$$\theta^T x - \mathbb{E}_{x' \in P(X)} [\theta^T x']$$

- Assuming $P(X)$ is approximated by using a sample...
- ...Then $\mathbb{E}_{x' \in P(X)} [\theta^T x']$ is just the average prediction on the data

I.e. it is the prediction we could make **without access to any input value**

Therefore, the difference above represents the gap between:

- ...What we can predict given all information on one example
- ...And what we can predict with no such information

It's the collective **value of all available information**



Additive Feature Attribution

Given an example x , we can try to to the same:

$$g(z, x) = \phi_0 + \sum_{j=1}^n \phi_j(x) z_j \quad \text{with: } z_j \in \{0, 1\}$$

- Where $z_j = 1$ is the value of attribute j is known
- ... $z_j = 0$ is the value of attribute j is unknown
- ...And $\phi_j(x)$ is the **effect** of attribute j on the example x

Intuitively, we build a **linear explanation** for the model local behavior

- There are a few approaches to approximate the effect $\phi_j(x)$
- The one we'll use work by averaging the effect of unknown attributes

 The idea is based on the Game Theory notion of Shapely Value

SHAP

Using Shapely values for explanation became prominent with this paper

The work makes a number of contributions:

- It introduces the general idea of additive feature attribution
- It shows how several previous approaches fall into that category
- It show how Shapely values provide "ideal" attribution scores
- It introduces multiple techniques to approximate the values

Computing Shapely values can be very expensive:

...But the can still be approximated in the general case. We will:

- Consider limited size of example for averaging out unknown attributes
- Obtain a Shapely value approximation by training a local linear model



SHAP in Action

The authors of the SHAP paper maintain a nice Python package

...Which we are going to use to **explain** our non-linear model

```
In [47]: f = lambda x: xbm.predict(x)
explainer = shap.KernelExplainer(f, shap.sample(X_train, 100), link='logit')
shap_values = explainer(X_test)
with open(os.path.join '..', 'data', 'shap_values.pickle'), 'wb') as fp:
    pickle.dump(shap_values, fp)
```

0%| | 0/100 [00:00<?, ?it/s]

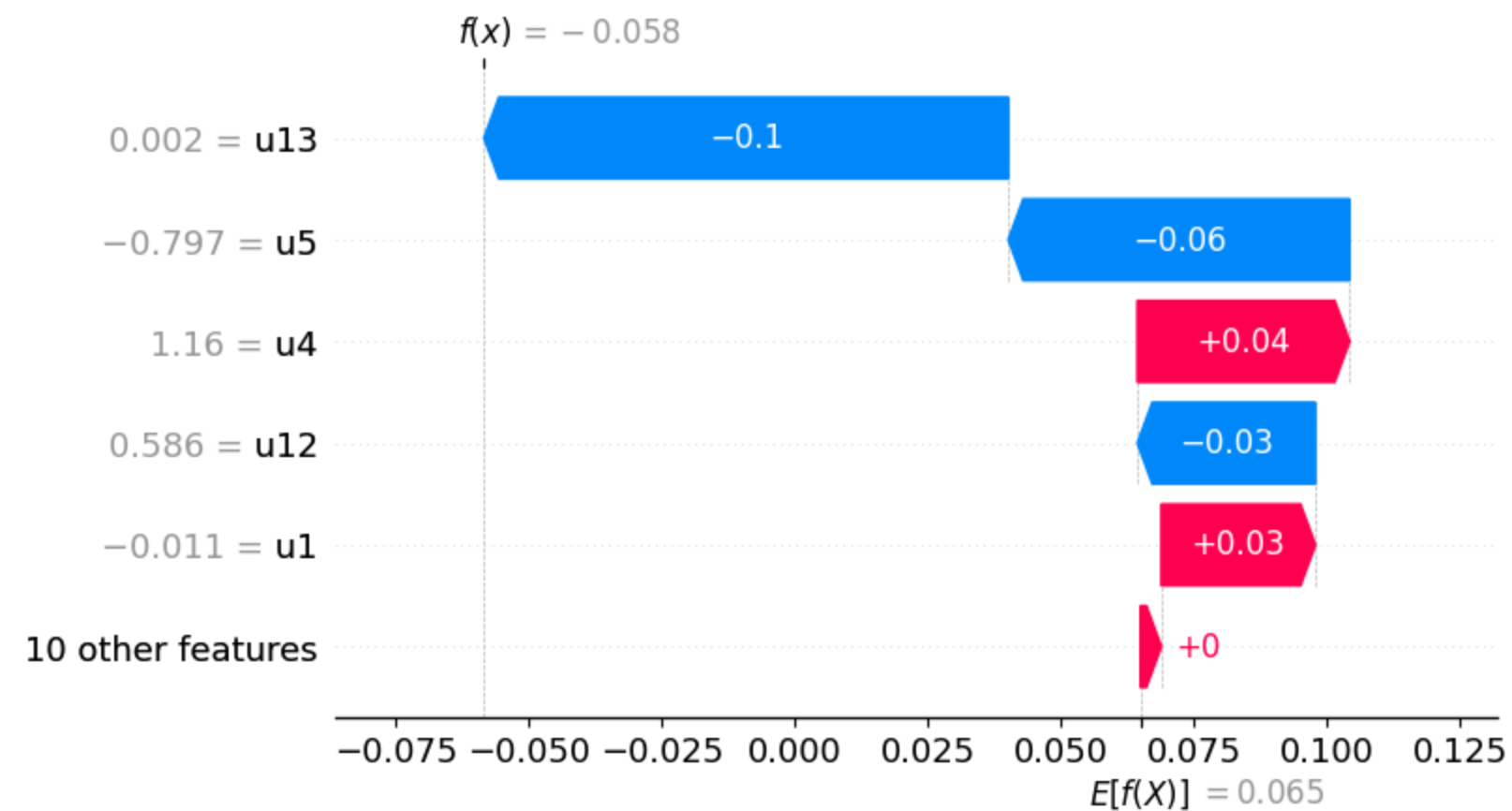
- We'll focus on the **test** data, since we want to find the true correlates
- For classifiers, it easier to explain **logits** rather than probabilities
- The process can be slow, and using a small background set is recommended
- The result contains the Shapely values, the base values, and the original data



Waterfall Plots

The SHAP library allows us to build **waterfall plots**

```
In [48]: shap.plots.waterfall(shap_values[0], max_display=6)
```



- The bars represent the Shapely values, the colors their sign

Force Plots

Waterfall plots can be "compacted" into **force plots**

Here we have again a plot for example 0:

```
In [49]: shap.initjs()  
shap.plots.force(shap_values[0])
```



Out [49] :

Visualization omitted, Javascript library not loaded!

Have you run `initjs()` in this notebook? If this notebook was from another user you must also trust this notebook (File -> Trust notebook). If you are viewing this notebook on github the Javascript has been stripped for security. If you are using JupyterLab this error is because a JupyterLab extension has not yet been written.

...And have a plot for example 99

```
In [51]: shap.plots.force(shap_values[99])
```



Out [51] :

Visualization omitted, Javascript library not loaded!

Have you run `initjs()` in this notebook? If this notebook was from another user you must also trust this notebook

Global Force Plots

Force plots can be stacked to inspect many examples at once:

```
In [52]: shap.plots.force(shap_values)
```

Out [52]:

Visualization omitted, Javascript library not loaded!

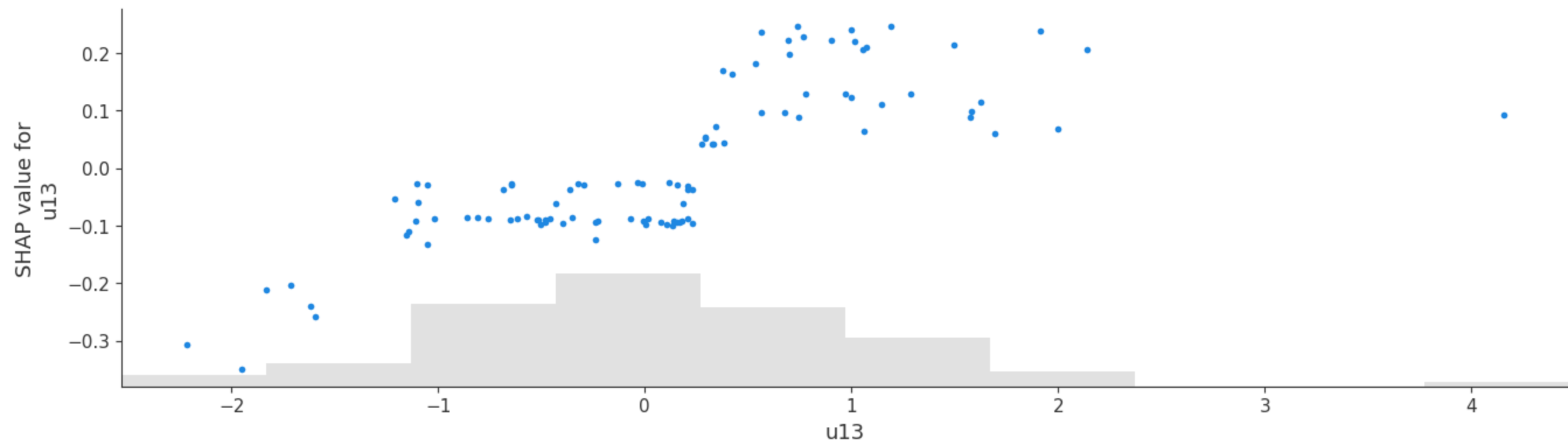
Have you run `initjs()` in this notebook? If this notebook was from another user you must also trust this notebook (File -> Trust notebook). If you are viewing this notebook on github the Javascript has been stripped for security. If you are using JupyterLab this error is because a JupyterLab extension has not yet been written.



Scatter Plots

We can use **scatter plots** to show the effect of a single feature

```
In [53]: plt.figure(figsize=figsize)
shap.plots.scatter(shap_values[:, 'u13'], ax=plt.gca())
```



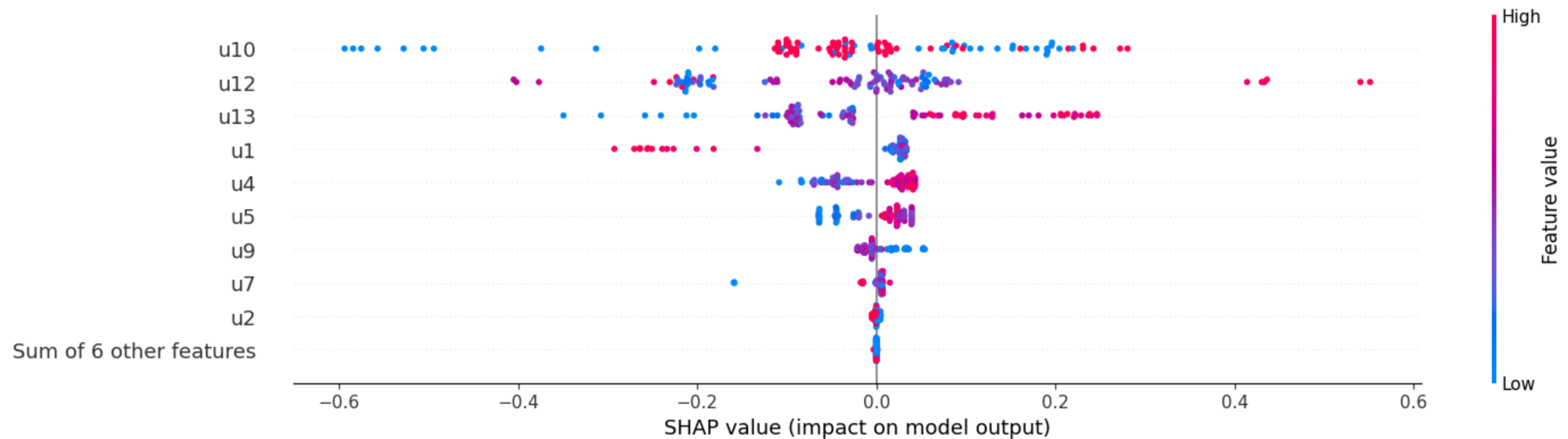
- The gray area is the histogram of the chosen feature



Beeswarm (Summary) Plot

We can stack (and color) multiple scatter plots to obtain a **beeswarm** plot:

```
In [54]: shap.plots.beeswarm(shap_values, max_display=10, plot_size=figsize)
```

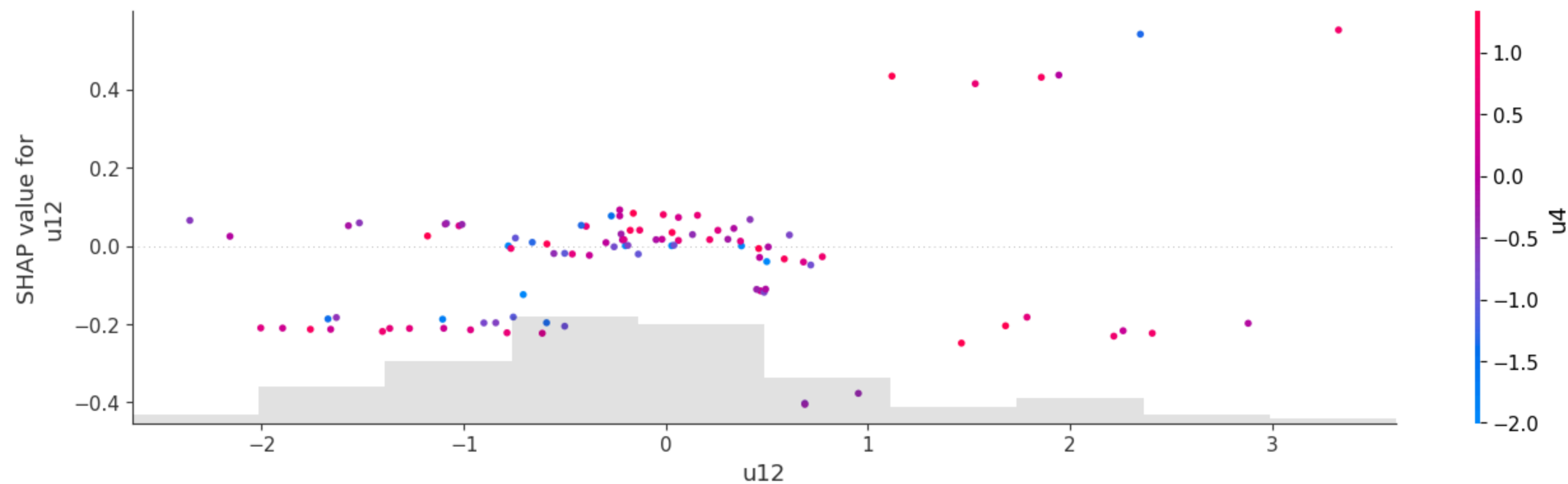


- By checking the color distribution we can indentify linear and non-linear effects

Scatter (Dependency) Plots

We can color scatter plots **by using another feature** to highlight dependency

```
In [55]: plt.figure(figsize=figsize)
shap.plots.scatter(shap_values[:, 'u12'], color=shap_values[:, 'u4'], ax=plt.gca())
```



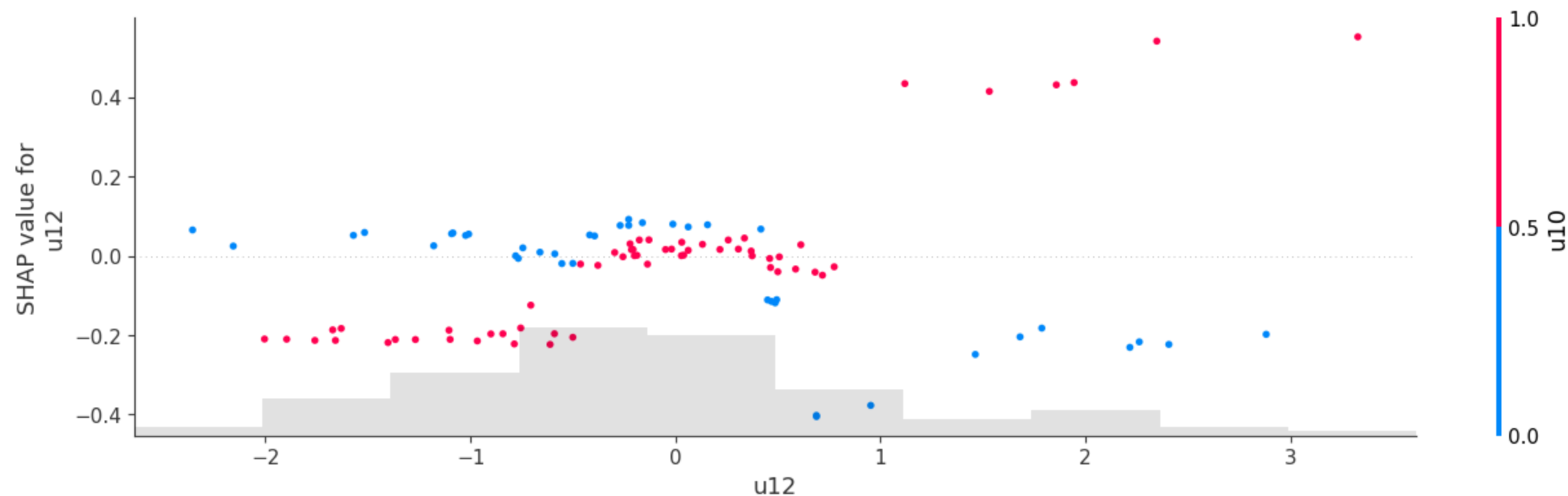
- In this case we are coloring the "u12" values by using "u4"



Scatter (Dependency) Plots

We can let the library choose the best coloring feature

```
In [56]: plt.figure(figsize=figsize)
shap.plots.scatter(shap_values[:, 'u12'], color=shap_values, ax=plt.gca())
```



- The chosen coloring feature changes how "u12" impacts the output in a noticeable way



Global Feature Analysis via SHAP

SHAP explanations can be aggregated to get global importance scores

By default, this is done by averaging absolute SHAP values:

$$\bar{\phi}_j(x) = \frac{1}{n} \sum_{i=1}^m |\phi_j(x_i)|$$

- Other aggregation functions can also be used (e.g. max)

By using aggregated SHAP scores

...We ensure that our local and global analysis have **a similar semantic**

- Permutation Importance are a perfectly viable approach
- ...And sometimes may be more appropriate than SHAP

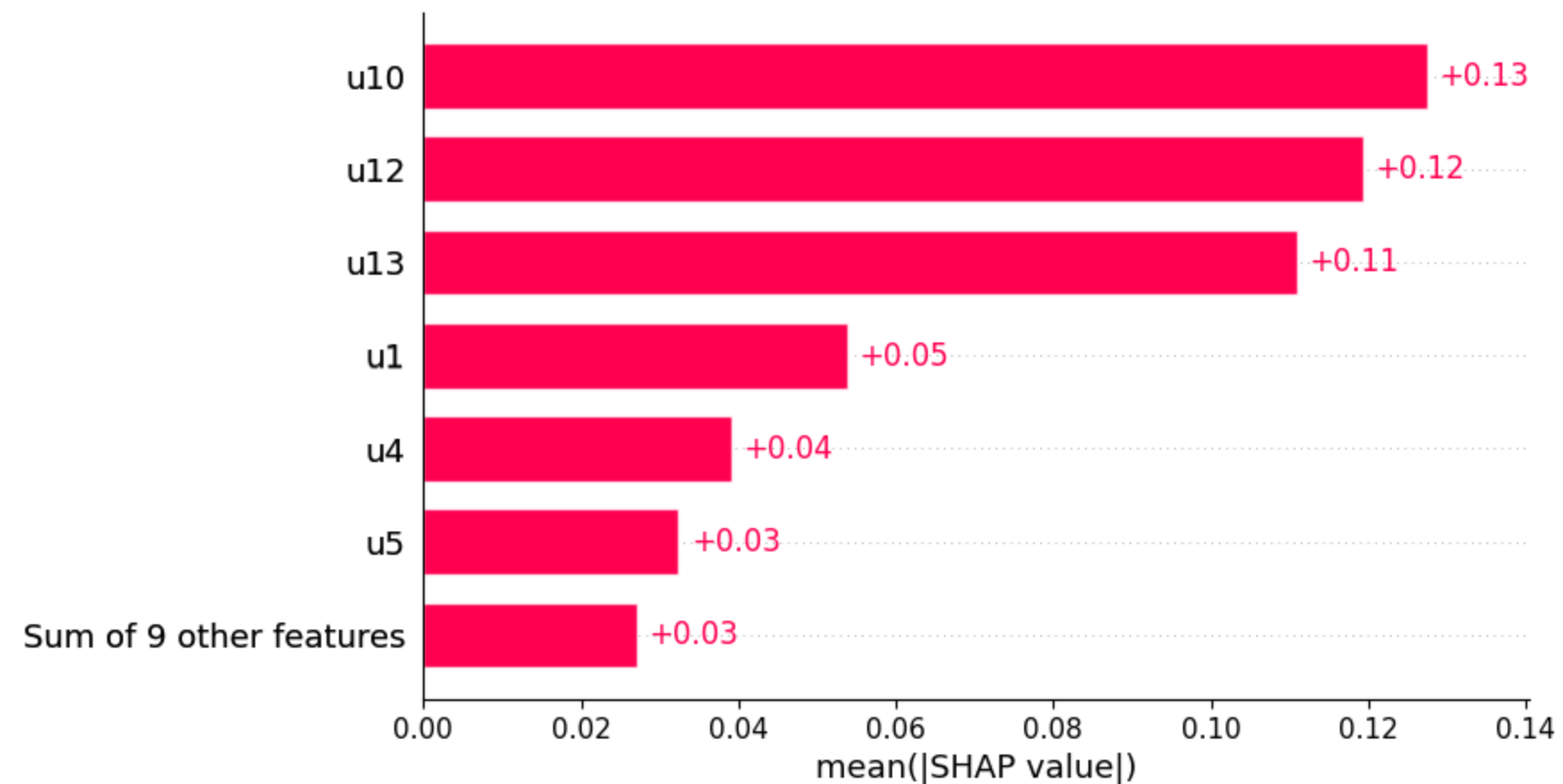
However, when doing a rigorous analysis consistency is important

Global Feature Analysis via SHAP

The SHAP library provide convenience functions to plot aggregated values

Here's how to plot mean (absolute) SHAP values:

```
In [58]: shap.plots.bar(shap_values, max_display=7)
```



Biomedical Data Analysis

Wrapping Up



What we Have Discovered

So far, we found that:

- The 4 relevant most relevant features seem to be U_1 , U_{10} , U_{12} , U_{13}

By inspecting the data we found that:

- U_{12} and U_{13} are roughly Normally distributed
- U_1 is not Normally distributed
- U_{10} is binary

By using SHAP we found that:

- U_{13} has a mostly monotonic effect
- U_{12} has a complex effect, which seems to be modulated by U_{10}

- U_1 seems to have mostly an anti-monotonic effect





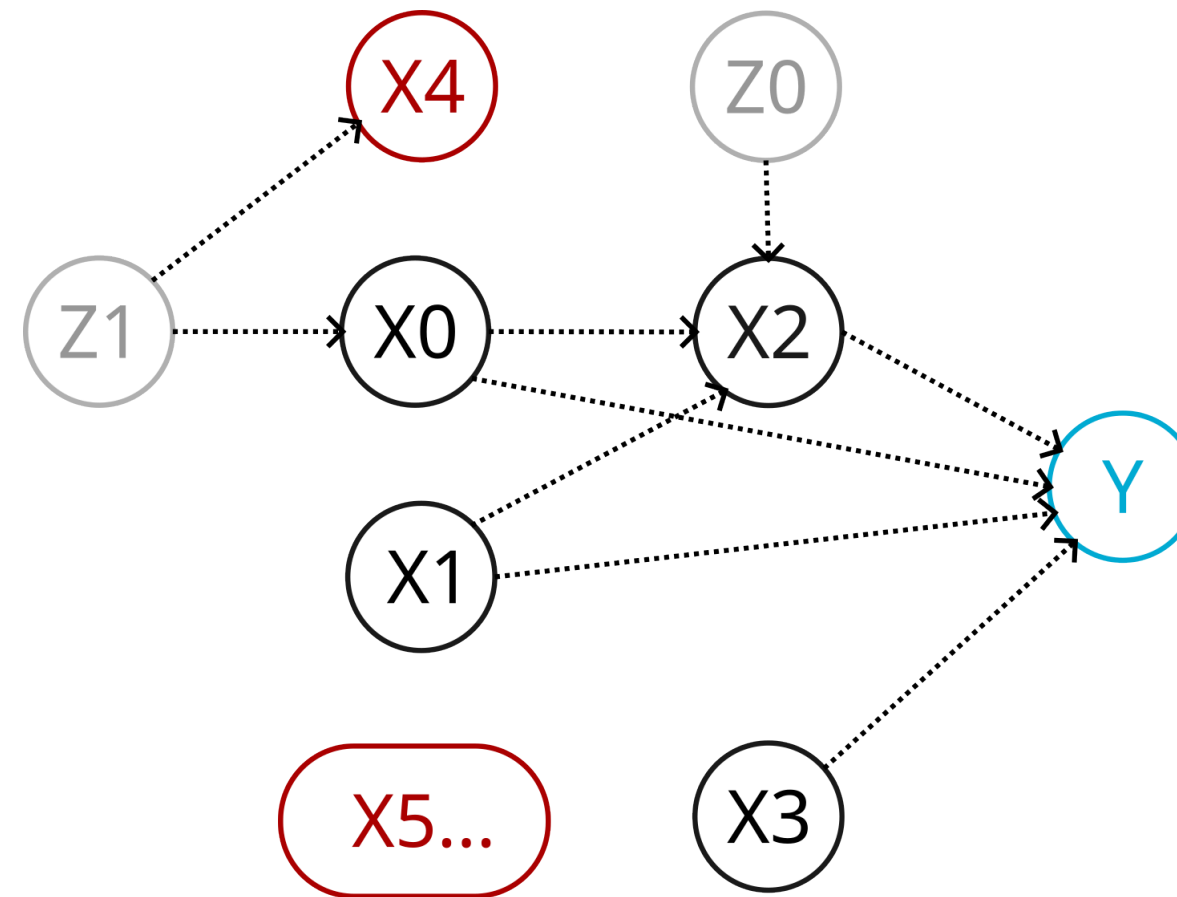
We can now inspect the ground truth process

So we can check whether our results actually hold



Checking the Ground Truth

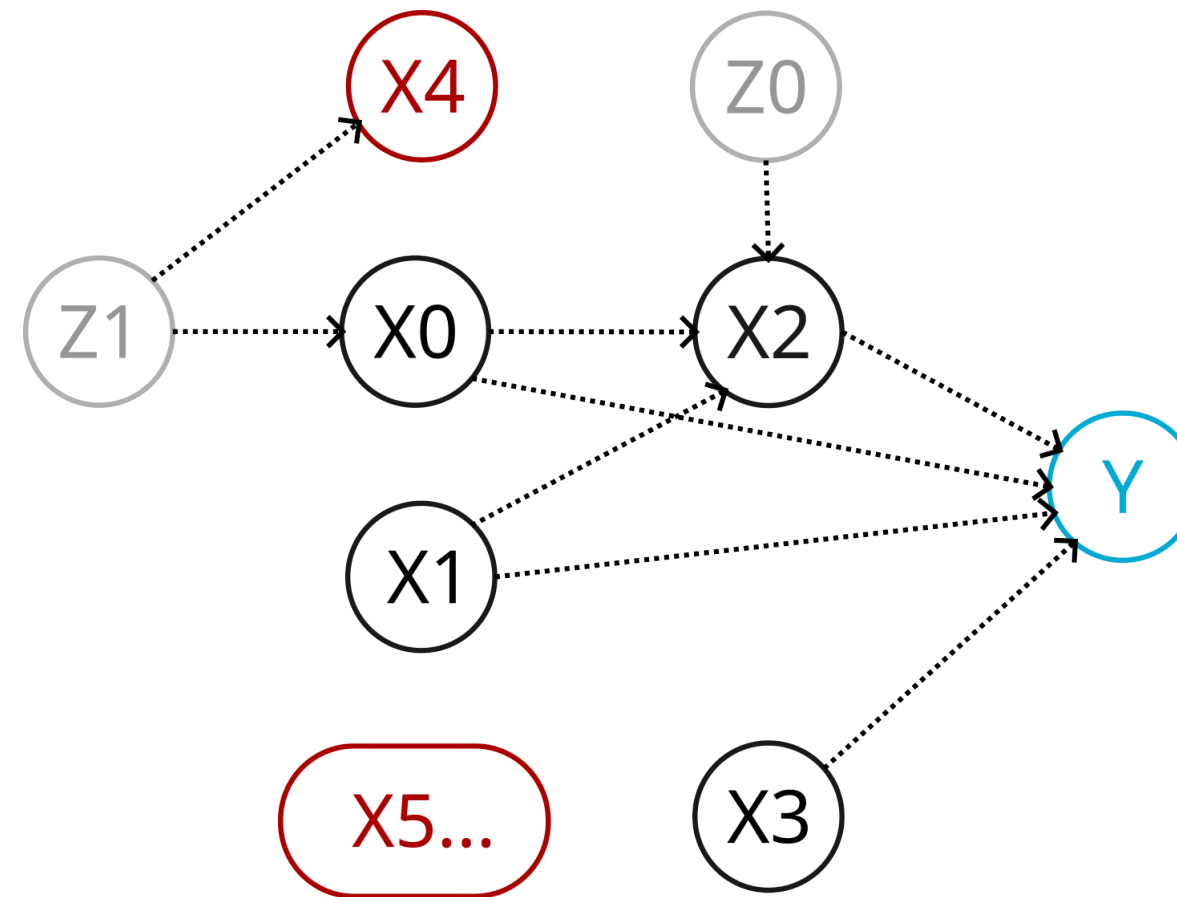
The ground-truth process is described by this causal graph:



- The Y variable (in **blue**) is the target
- The variables in **black** are those that are relevant
- The variables in **gray** are not observable, i.e. latent
- The variables in **red** are irrelevant

Checking the Ground Truth

The process was engineered to contain several classical cases

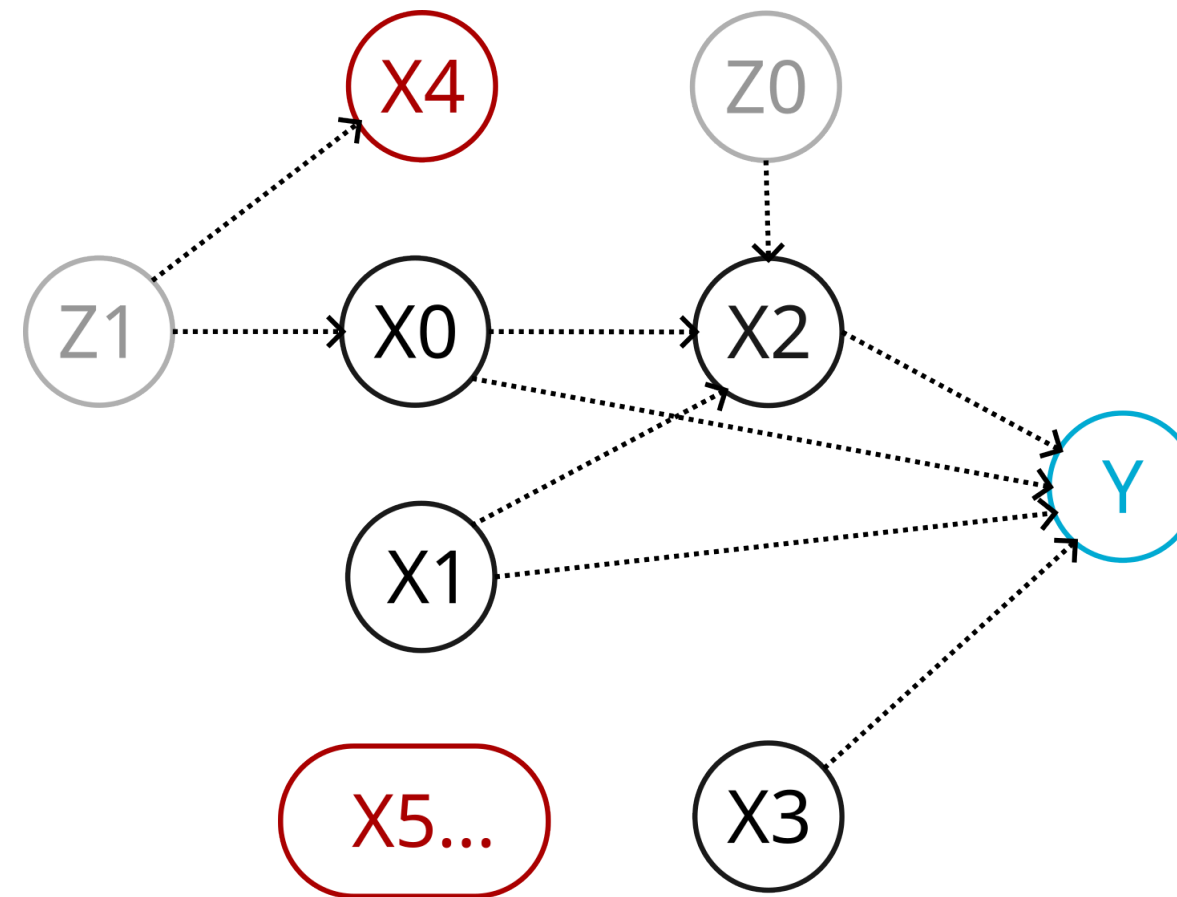


X_2 is a **mediator** between X_0 , X_1 and Y

- The variable partially hides the effect of X_0 and X_1
- If it does that completely, even Boruta cannot mark X_0 and X_1 as important
- Depending on the use case, this might be an issue

Checking the Ground Truth

The process was engineered to contain several classical cases



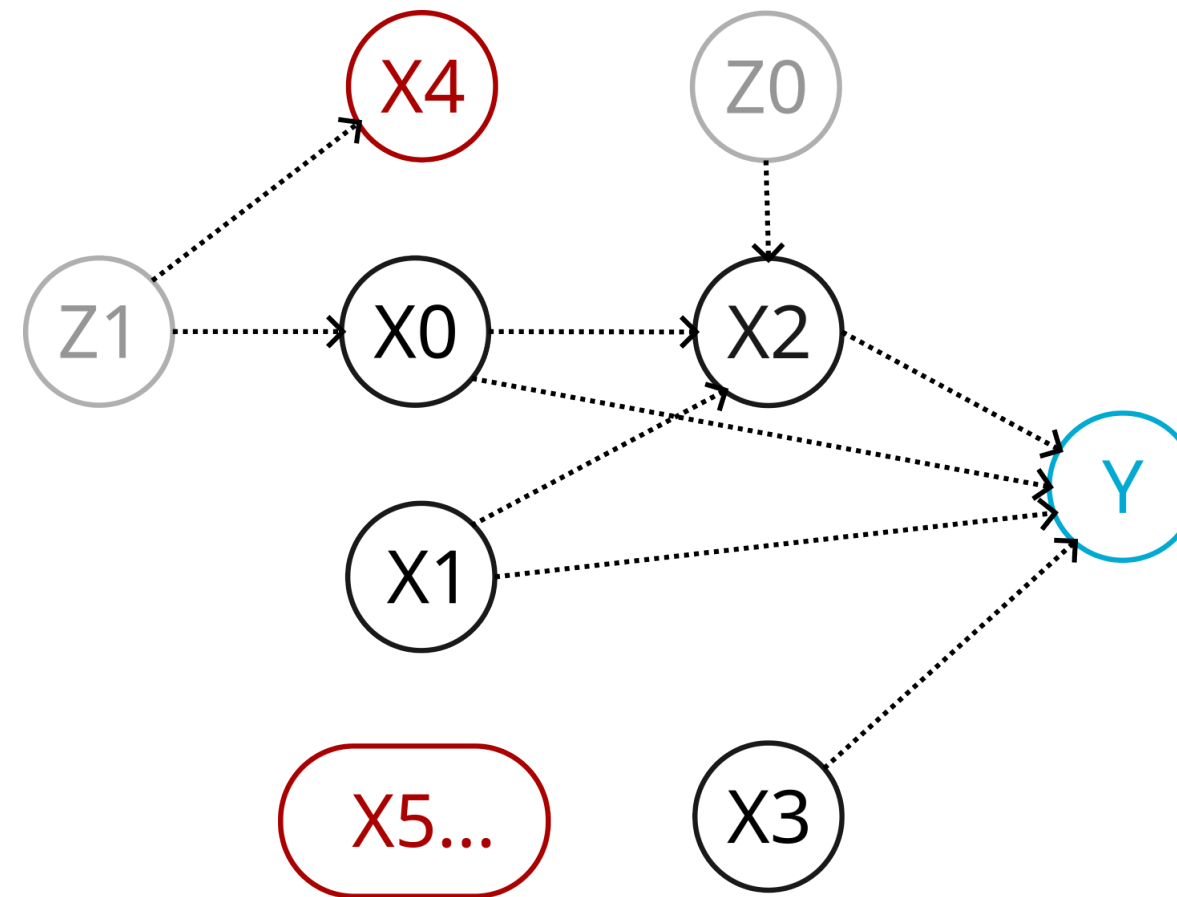
X_2 is also a complete mediator for Z_0

- ...But in this case it is a good thing!
- Z_0 is not observed, but we can account for that at least indirectly



Checking the Ground Truth

The process was engineered to contain several classical cases



Z_1 is a **confounder** and causes a correlation between X_1 and X_0

- It is totally mediated by X_1 , which is a good thing
- ...But it also causes a correlation between X_0 and X_4
- This might trick a model into considering X_4 as important

Checking the Ground Truth

In detail, the data is generated as follows:

$$Z_0 \sim \mathcal{N}(\mu = 0, \sigma = 1.5)$$

$$Z_1 \sim \mathcal{N}(\mu = 0, \sigma = 1)$$

$$X_0 \sim \mathcal{N}(\mu = 0, \sigma = 1) + Z_1$$

$$\log X_1 \sim \mathcal{N}(\mu = 0, \sigma = 1)$$

$$X_2 = 1/2(X_0 + X_1) + Z_0$$

$$X_3 \sim B(p = 0.6)$$

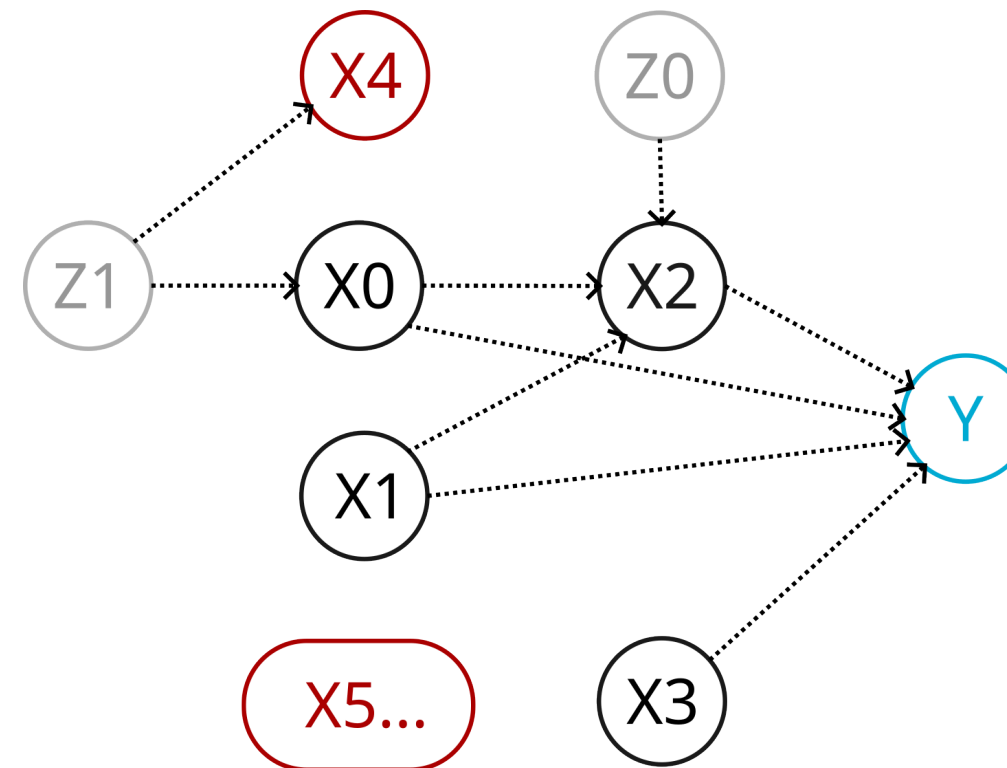
$$\text{logit}Y \sim (-1 + 2X_3)X_2 + 0.4X_0 - 0.4X_1$$

- All other variables are follow either a Normal or Bernoulli distribution
- ...And they have sparse correlations among themselves



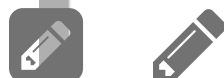
Checking the Ground Truth

Now let's check how accurate our importance estimate is:



```
In [63]: accepted = ['u1', 'u10', 'u12', 'u13']
print(f'The most relevant features are {accepted}')
print(f'...And correspond to {[name_map[f] for f in accepted]}')
```

```
The most relevant features are ['u1', 'u10', 'u12', 'u13']
...And correspond to ['X1', 'X3', 'X2', 'X0']
```



What we Have Discovered

Now, let's check our findings, with the due substitutions:

- The relevant features are $X_0, X_1, X_2, X_3 \rightarrow$ **correct!**
- All claims on distributions \rightarrow **mostly correct!**
 - X_2 is a sum of 3 variables, two of which are Normal
- X_0 has a mostly monotonic effect \rightarrow **correct!**
 - Direct monotonic effect on Y + a complex one mediated by X_2
- X_1 has a mostly anti-monotonic effect \rightarrow **correct!**
 - Direct anti-monotonic effect on Y + a complex one mediated by X_2
- X_2 has complex effect modulated by $X_3 \rightarrow$ **correct!**
 - X_3 decides the sign of the X_2 contribution



A Few Final Remarks

ML models are not just for prediction!

- They can be used for generation, anomaly detection, decision support
- ...And also as tools for a scientific analysis!

Explainability is an important topic in AI

- It is one of the main approaches to make an AI model transparent
- This critical when AI systems need to interact with human users
- ...And for some domains it is also required by existing regulations

Beware of correlated features

- Strongly correlated features (e.g mediated-mediator) may mislead algorithms

 Dealing with those is still a partially open problem!