## **Analysis of the imaging experiment**

mouse name: 059 task: NearFar

experimentalist: Snezana Raus-Balind

#### 1. Load the necessary object class

We use a custom-made class, ImagingSessionData, that will contain all behavioral and imaging data. We also load the matplotlib widgets to make graphis interactive under the notebook environment.

```
In [1]: from ImageAnal import *
%matplotlib widget
```

#### 2. Tell python where the data is

The required file structure is the following:

- All data should be in the data folder
- Within the data folder separate subfolders are needed for each mouse. Folder name starts with the **name** of the mouse.
- For each mouse there should be at least two folders: one for the **imaging** data and one for the **behavioral** data.
- The behavioral folder is named as MouseName\_TaskName so we need a separate folder for each different task
- The behavioral log files are in separate subfolders named by the experiment's start time within the behavioral folder e.g. 2021–02–03\_10–15–50
- The imaging folder is named as MouseName imaging
- The suite2p imaging files are also in separate folders for each experiment below the imaging folder.

```
In [2]: datapath = os.getcwd() + '/' #current working directory - look for of date_time = '2021-02-03_10-15-50' # date and time of the imaging set name = 'srb059' # mouse name task = 'NearFar' # task name

## locate the suite2p folder
suite2p_folder = datapath + 'data/' + name + '_imaging/Suite2P_4_19-
## the name and location of the imaging log file imaging_logfile_name = suite2p_folder + 'srb059_TSeries-02032021-10'
## the name and location of the trigger voltage file TRIGGER_VOLTAGE_FILENAME = suite2p_folder + 'srb059_TSeries-0203202'
```

#### 3. Load all the data - this taks ~20 secs in my computer

Python looks for the data in the specified folders. It loads the behavioral data (position,

lick and rewards) as well as the imaging data. It calculates the activity of the cells as a function of position in the different corridors and calculates their spatial tuning measures and corridor selectivity.

The name of the object that contains all the data is D1 here - Data 1.

```
In [3]: # 3. load all the data — this taks ~20 secs in my computer
        D1 = ImagingSessionData(datapath, date_time, name, task, suite2p_fo
        trigger logfile loaded
        trigger voltage signal loaded
        triggers after: 22
        n_extra_indexes 5
        candidate log indexes [0, 99, 178]
        min recorded trigger length: 0.01200000000000455
        relevant behavior located, lap time of the first frame: 870.993125
        , log reference index: 99
        slight warning - testing some late candidates failed
        suite2p data loaded
        corrected offset: 870.9851249999999 voltage_delay: 0.0080000000000
        382
        suite2p time axis loaded
        calculating dF/F and SNR...
        /Users/ubi/Projects/KOKI/VR/MiceData/ImageAnal.py:408: RuntimeWarn
        ing: invalid value encountered in true_divide
          self.dF_F[i_cell,] = (self.F[i_cell,] - baseline) / baseline
        SNR done
        dF/F calculated for cell ROI-s
        ExpStateMachineLog time interval > 1s: 10 times
            5.03966018 178.07813661 184.49403693 186.90899334 207.4688
        484
          213.4257944
                        217.7201534
                                      392.68928535 653.35621006 1802.0791
        46191
        calculating rate, reliability and Fano factor...
        calculating Skaggs spatial info...
        calculating proportion of active laps...
        calculating proportion of active laps based on dF/F ...
        calculating linear tuning specificity ...
        calculating rate, reliability and Fano factor...
        /Users/ubi/Projects/KOKI/VR/MiceData/utils.py:17: RuntimeWarning:
        invalid value encountered in true divide
          r = np.divide(r_num, r_den, out=out_vec, where=vec_nonzero)
        calculating Skaggs spatial info...
        calculating proportion of active laps...
        calculating proportion of active laps based on dF/F ...
        calculating linear tuning specificity ...
        calculating corridor selectivity ...
        calculating corridor similarity ...
```

The behavior is divided into laps (trials or runs). You can check the **number of laps** and which lap is associated with imaging data in the following way:

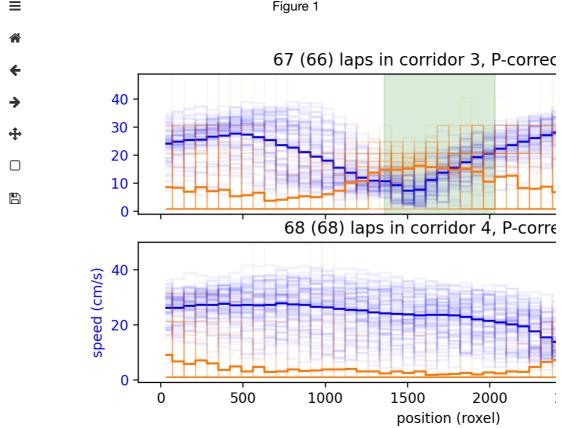
```
In [4]: print(D1.n_laps)
    print(D1.i_Laps_ImData)
```

```
178
[ 33
      34
           35
               36
                    37
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03 104
 105 106 107 108 109 110 111 112 113 114 115 116 117 118 119 120 1
 123 124 125 126 127 128 129 130 131 132 133 134 135 136 137 138 1
39 140
 141 142 143 144 145 146 147 148 149 150 151 152 153 154 155 156 1
57 158
```

So we have 178 laps and laps 33-167 contain imaging data.

#### 4. Plotting the behavioral data

You can plot the behavioral data of the session:

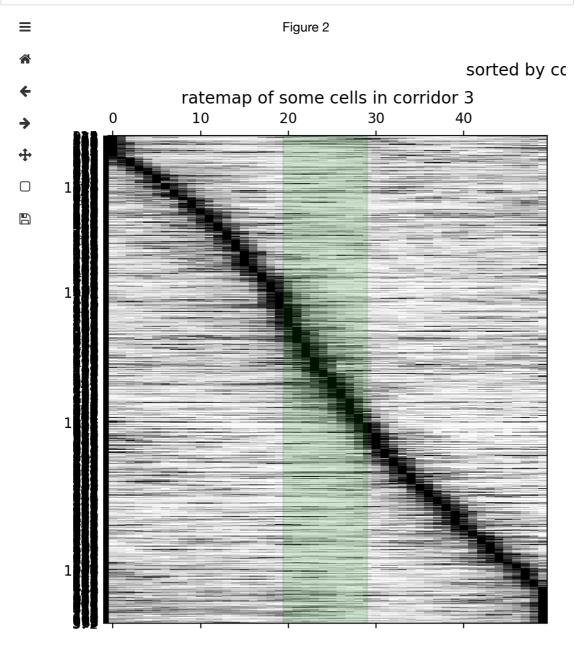


#### 5. Plot the ratemaps

First, we plot the ratemaps of some neurons. There are several options - selecting the cells, sorting and normalising the ratemaps.

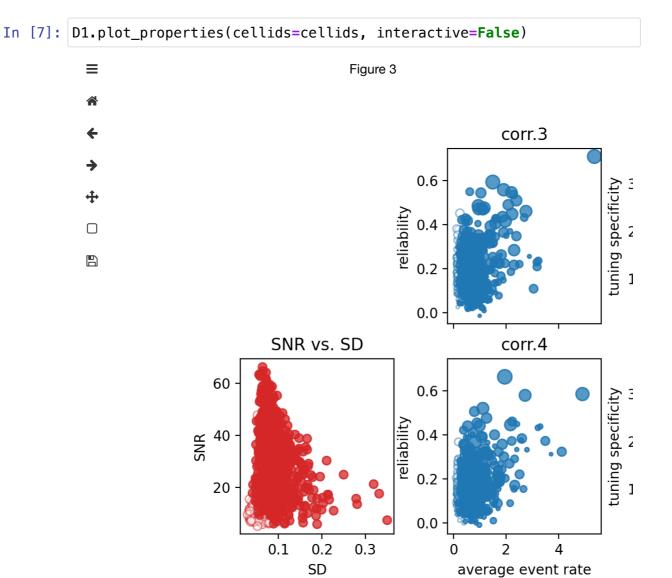
- selection: here we select all active cells (activity in at least 20% of laps), but any other selection criteria
- sorting: place fields can be sorted by either corridors
- place cells can be normalised so the peak has the same height

In [6]: cellids = np.nonzero(((D1.cell\_activelaps[0]>0.2) + (D1.cell\_activelaps[0])
D1.plot\_ratemaps(cellids = cellids, sorted=True, corridor\_sort=3, notelline)





#### 6. Plot the spatial properies of the neurons



# 7. Calculate significance of tuning by shuffling the imaging data

In [8]: D1.calc\_shuffle(cellids, 1000, 'shift', batchsize=50)

```
loading shuffling P-values from file...
        1
        selective cells: [ 3 5 15 16 23 26 29 33 34 35 36 56
        68 72 74 76 77 79
         82 84 85 87 88 89 93 95 107 115 122 125 126 141 143 144 1
        52 154
         150 160 167 166 160 177 107 107 101 105 711 717 717 721 727 721 7
In [13]: print(D1.tuned_cells)
```

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95 <b>,</b>	80,	81,	85,	86,	87,	88,	89,	90,	92,	94,
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137,	119,	120,	121,	122,	125,	128,	132,	133,	134,	136,
157,	141,	142,	143,	144,	145,	147,	148,	149,	151,	152,
170,	154,	156,	157,	158,	159,	160,	161,	162,	163,	168,
190,	171,	172,	174,	175,	177,	178,	179,	185,	187,	189,
	191,	193,	194,	195,	198,	200,	201,	204,	208,	211,
212,	213,	215,	217,	220,	221,	223,	224,	226,	229,	231,
233,	234,	235,	236,	237,	239,	240,	241,	242,	243,	249,
252,	256,	259,	264,	265,	267,	269,	274,	276,	277,	278,
279,	280,	282,	285,	286,	289,	290,	291,	292,	293,	294,
296,	297,	300,	304,	307,	308,	310,	311,	313,	314,	315,
316,	319,	320,	323,	325,	326,	327,	329,	330,	331,	334,
335,	341,	343,	347,	349,	351,	352,	353,	357,	361,	363,
365,	366,	368,	370,	371,	373,	375,	376,	377,	379,	380,
385,	388,	389,	396,	397,	399,	401,	402,	406,	409,	410,
411,	412,	415,	418,	419,	422,	427,	429,	430,	433,	434,
435,	436,	439,	442,	443,	444,	446,	447,	449,	450,	451,
454,	455,	456,	461,	463,	464,	466,	468,	472,	476,	493,
494,	496,	499,	500,	501,	502,	504,	505,	508,	509,	511,
517,	518,	519,	521,	524,	525,	526,	528,	530,	531,	532,
533,	536,	540,	541,	542,	543,	544,	545,	546,	548,	549,
550,	556,	557,	558,	559,	562,	564,	569,	570,	573,	574,
579,	580,	581,	582,	585,	586,	589,	590,	592,	595,	599,
603,	608,	612,	613,	620,	624,	627,	628,	629,	630,	632,
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```

In [14]: print(len(cellids))

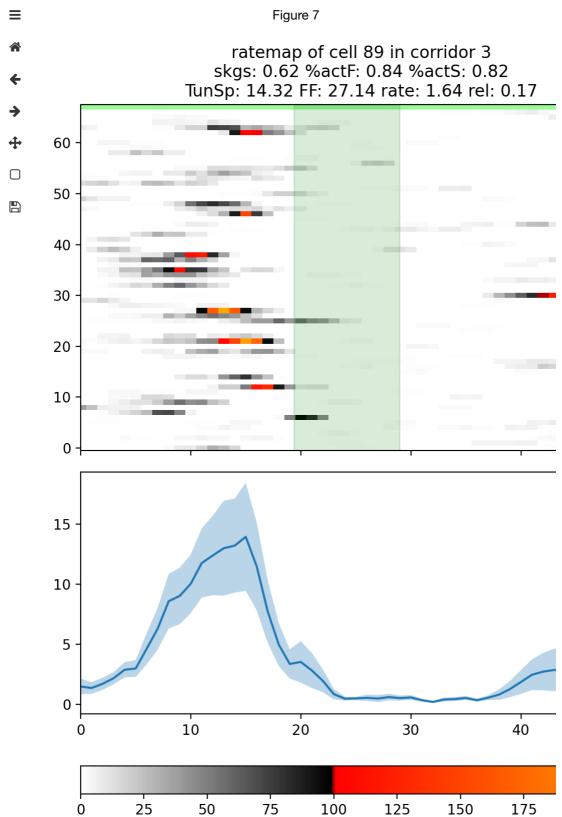
680

In [15]: print(D1.similar\_cells)

[ 13 89 106 185 221 310 313 320 327 357 409 468 521 571 818 913]

### 8. Plot the activity lap by lap

We can also plot the lap by lap activity of a selected cell. Again, there are several options, but the simplest is to plot the rate as a function of position.





```
Session parameters written into file: cell_properties_corridor_3_
N1022.csv
cell properties for corridor 3 saved into file: /Users/ubi/Proje
cts/KOKI/VR/MiceData/data/srb059_imaging/Suite2P_4_19-05-2021/anal
ysed_data/cell_properties_corridor_3_N1022.csv
Session parameters written into file: cell_properties_corridor_4_
N1022.csv
cell properties for corridor 4 saved into file: /Users/ubi/Proje
cts/KOKI/VR/MiceData/data/srb059_imaging/Suite2P_4_19-05-2021/anal
ysed_data/cell_properties_corridor_4_N1022.csv
Session parameters written into file: ratemaps corridor 3 N1022.c
sv
ratemap for corridor 3 saved into file: /Users/ubi/Projects/KOKI
/VR/MiceData/data/srb059_imaging/Suite2P_4_19-05-2021/analysed_dat
a/ratemaps_corridor_3_N1022.csv
Session parameters written into file: ratemaps_corridor_4_N1022.c
ratemap for corridor 4 saved into file:
                                          /Users/ubi/Projects/KOKI
/VR/MiceData/data/srb059_imaging/Suite2P_4_19-05-2021/analysed_dat
a/ratemaps_corridor_4_N1022.csv
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Session parameters written into file:
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lapdata saved into file: /Users/ubi/Projects/KOKI/VR/MiceData/data
/srb059_imaging/Suite2P_4_19-05-2021/analysed_data/lapdata_lap_167
N1022.csv
```

#### In []: