Genome Annotation

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Objectives

- Understand steps involved in genome annotation
- Demonstrate types of data and tools that can be used in genome annotation
- Learn how to postprocess genome assemblies
- QC annotation results

We need these files

cd /scratch/Botany2020NMGWorkshop/

mkdir annotation2

cd annotation2

git clone https://github.com/bcbc-group/Botany2020NMGWorkshop.git

iget -rPT /iplant/home/shared/Botany2020NMGWorkshop/annotation/2transfer/contig_15.fasta

iget -rPT /iplant/home/shared/Botany2020NMGWorkshop/annotation/2transfer/SRR5046448_contig15_1.fastq

iget -rPT /iplant/home/shared/Botany2020NMGWorkshop/annotation/2transfer/SRR5046448_contig15_2.fastq

iget -rPT /iplant/home/shared/Botany2020NMGWorkshop/annotation/2transfer/mikado.loci.gff3

iget -rPT /iplant/home/shared/Botany2020NMGWorkshop/annotation/2transfer/scipio.yaml

iget -rPT /iplant/home/shared/Botany2020NMGWorkshop/annotation/2transfer/contig_15_masked.fasta

Iget -rPT /iplant/home/shared/Botany2020NMGWorkshop/annotation/2transfer/contig_15.maker.output/maker1.all.gff

iget -rPT /iplant/home/shared/Botany2020NMGWorkshop/embryophyta_odb9

Changing docker file storage location

```
#stop docker
sudo service docker stop
#edit daemon.json
emacs /etc/docker/daemon.json
#and add:
"graph": "/scratch/docker"
#copy current dir to new one
sudo rsync -aP /var/lib/docker/ /scratch/docker
#rename old docker dir, do no delete until you test config works
sudo mv /var/lib/docker /var/lib/docker.old
```

Goals of genome annotation

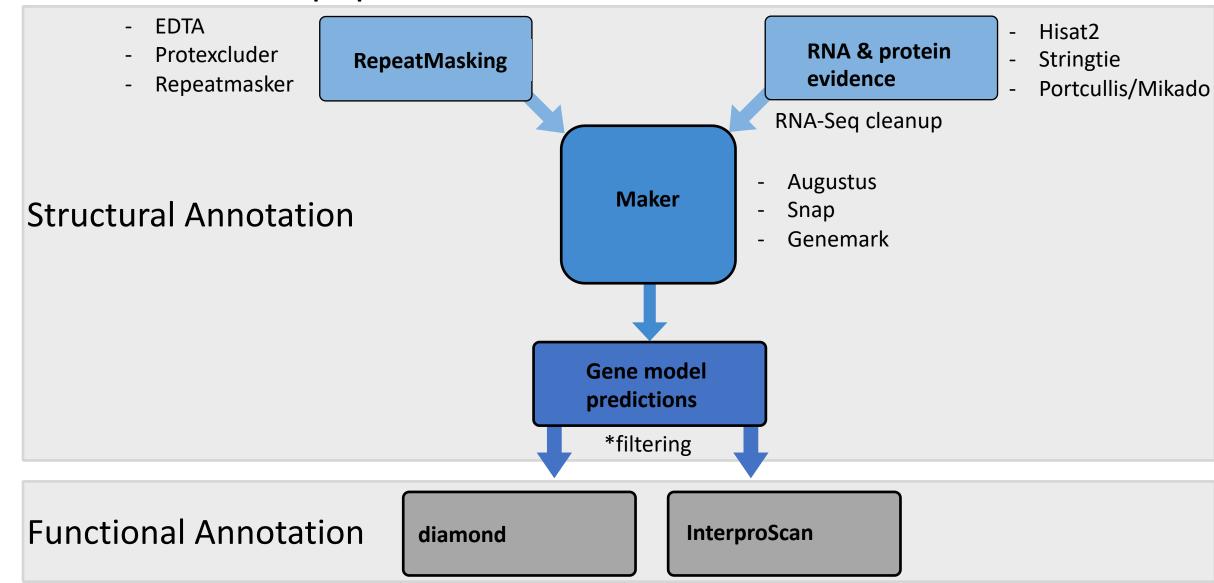
- Predict, categorize, and mask repetitive elements
- Determine gene structures as accurately as possible
- Predict possible functions of predicted genes
- Associate GO terms, domains, etc for downstream analyses

Pre-annotation QC

- Assembly quality (total length, N50, etc)
- Errors correction
- BUSCO metrics of genome

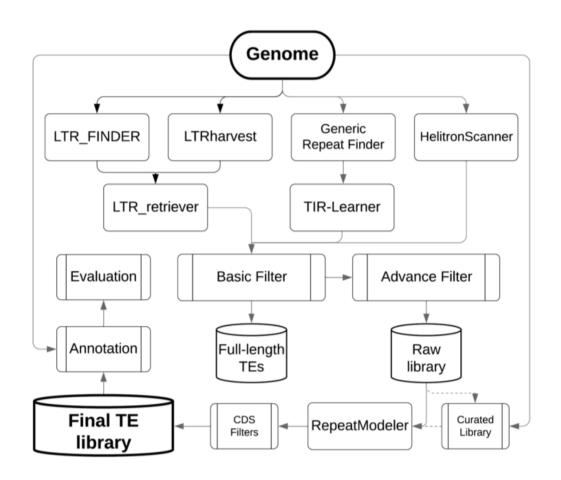
Tools for structural annotation

- EDTA https://github.com/oushujun/EDTA
- Repeatmasker http://www.repeatmasker.org/
- Braker https://github.com/Gaius-Augustus/BRAKER
- Augustus <a href="https://github.com/Gaius-Augustus/A
- Snap https://github.com/KorfLab/SNAP
- Genemark http://exon.gatech.edu/GeneMark/
- Maker https://www.yandell-lab.org/software/maker.html
- Apollo https://genomearchitect.readthedocs.io/en/latest/
- BUSCO https://gitlab.com/ezlab/busco-biocontainer



EDTA

The Extensive de novo TE Annotator (EDTA)



Tools for functional annotation

- BLAST
- Diamond
- InterProScan
- Mercator

Let's annotate our *U. gibba* FLYE assembly!

- Genome file: Ugibba_FLYE_assembly.fasta.PolcaCorrected.fa.cat.all.gz
- RNA-seq from shoots and traps: https://www.ncbi.nlm.nih.gov/sra/SRX2368915[accn]
- Proteins: uniprot_sprot_plants.fasta

 All this stuff plus some output files in /iplant/home/shared/Botany2020NMGWorkshop/annotation/

QC of FLYE *U. gibba* assembly

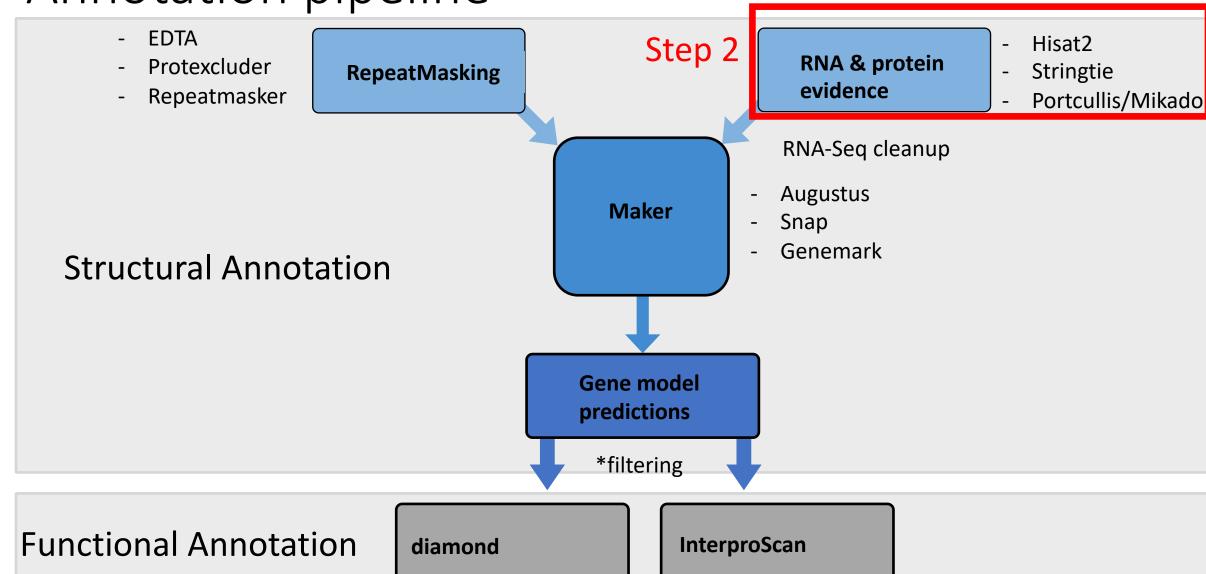
- Size =
- N50 =
- BUSCO = 93.6% complete

Annotation pipeline **EDTA** Step 2 Hisat2 Step 1 **RNA & protein** Protexcluder RepeatMasking Stringtie evidence Repeatmasker Portcullis/Mikado RNA-Seq cleanup Step 3 Augustus Maker Snap Step 5 Step 4 Genemark Structural Annotation Gene model predictions *filtering **Functional Annotation** InterproScan diamond Step 6

Annotation pipeline **EDTA** Hisat2 Step 1 **RNA & protein** Protexcluder RepeatMasking Stringtie evidence Repeatmasker Portcullis/Mikado Already performed RNA-Seq cleanup for you! Augustus Maker Snap Genemark **Structural Annotation** Gene model predictions *filtering **Functional Annotation** InterproScan diamond

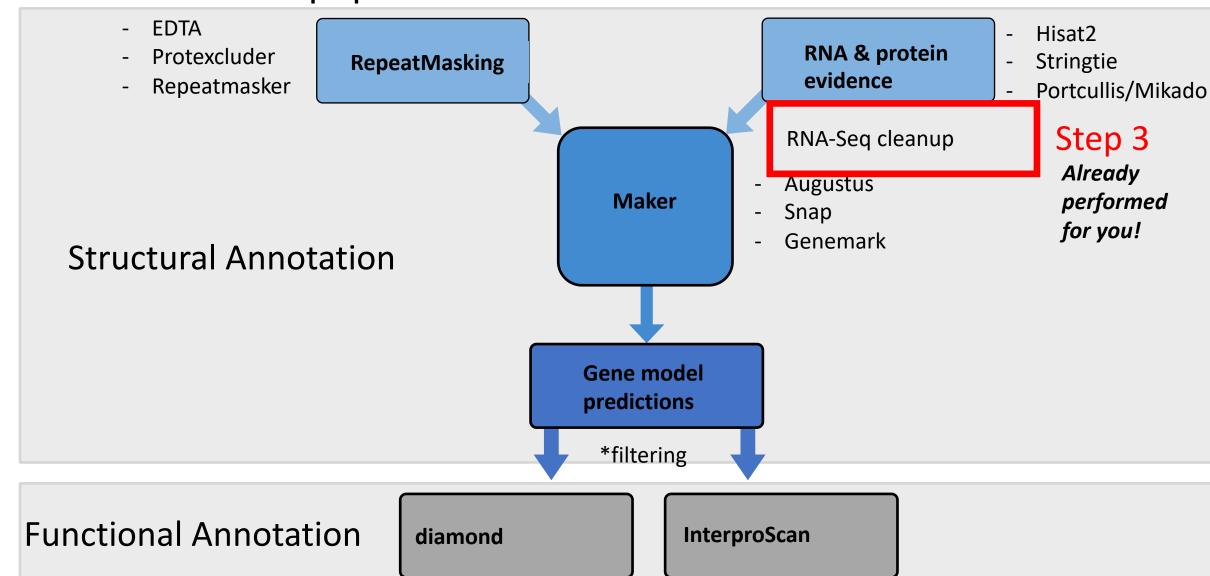
Step 1: Repeat Masking

 https://github.com/bcbcgroup/Botany2020NMGWorkshop/blob/master/annotation/1 repeat masking.sh



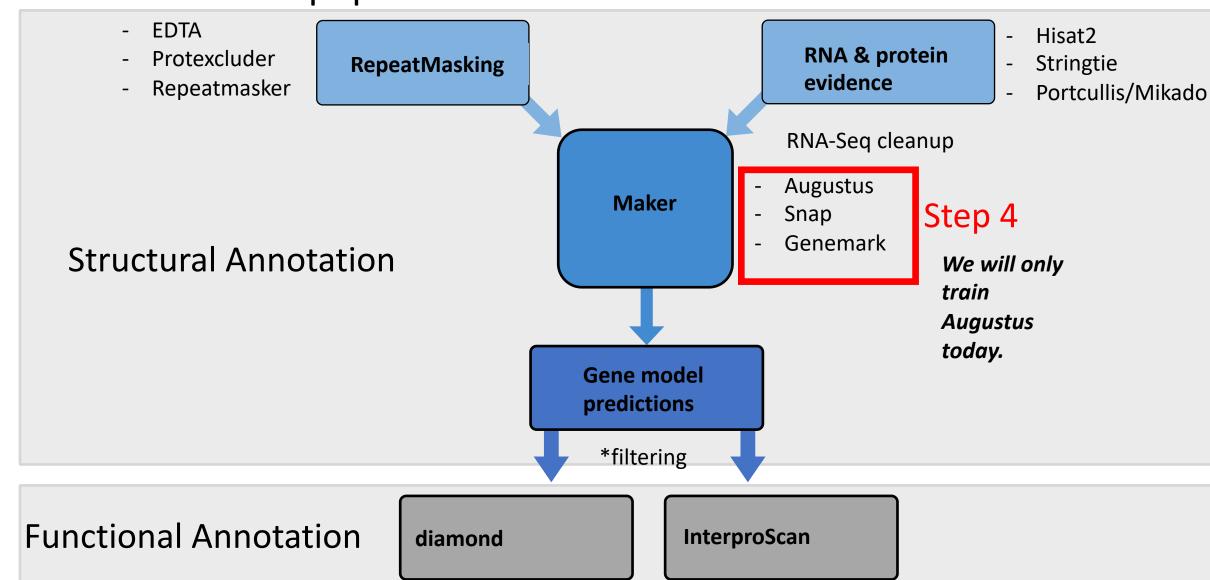
Step 2: RNA-Seq read mapping

 https://github.com/bcbcgroup/Botany2020NMGWorkshop/blob/master/annotation/2 hisat pe annot.sh



Step 3: RNA-seq cleanup

 https://github.com/bcbcgroup/Botany2020NMGWorkshop/blob/master/annotation/3 rnaseq cleanup.sh



Step 4: Training augustus and snap

- https://vcru.wisc.edu/simonlab/bioinformatics/programs/augustus/d ocs/tutorial2015/training.html
- https://vcru.wisc.edu/simonlab/bioinformatics/programs/augustus/d ocs/tutorial2015/scipio.html
- https://github.com/bcbcgroup/Botany2020NMGWorkshop/blob/master/annotation/training.s
 h

Your turn to train Augustus!

```
/opt/augustus-3.2.2/scripts/randomSplit.pl genes.gb 200 grep -c LOCUS genes.gb*

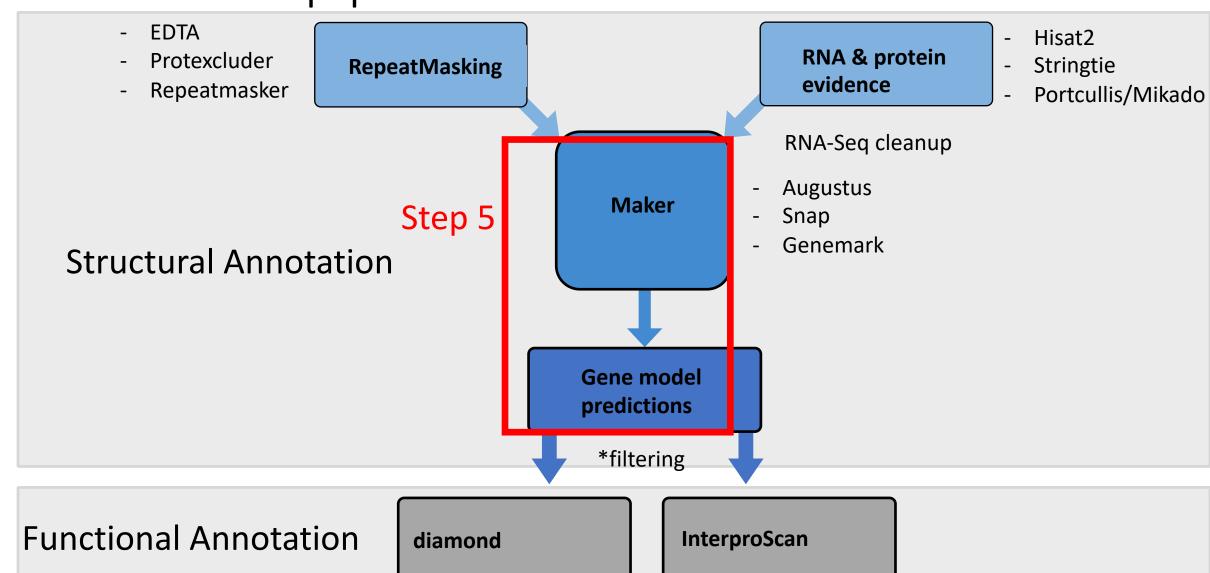
sudo chown srs57 /opt/augustus/config/species/
/opt/augustus-3.2.2/scripts/new_species.pl --species=Ugibba

etraining --species=Ugibba genes.gb.train

ls -ort $AUGUSTUS_CONFIG_PATH/species/Ugibba

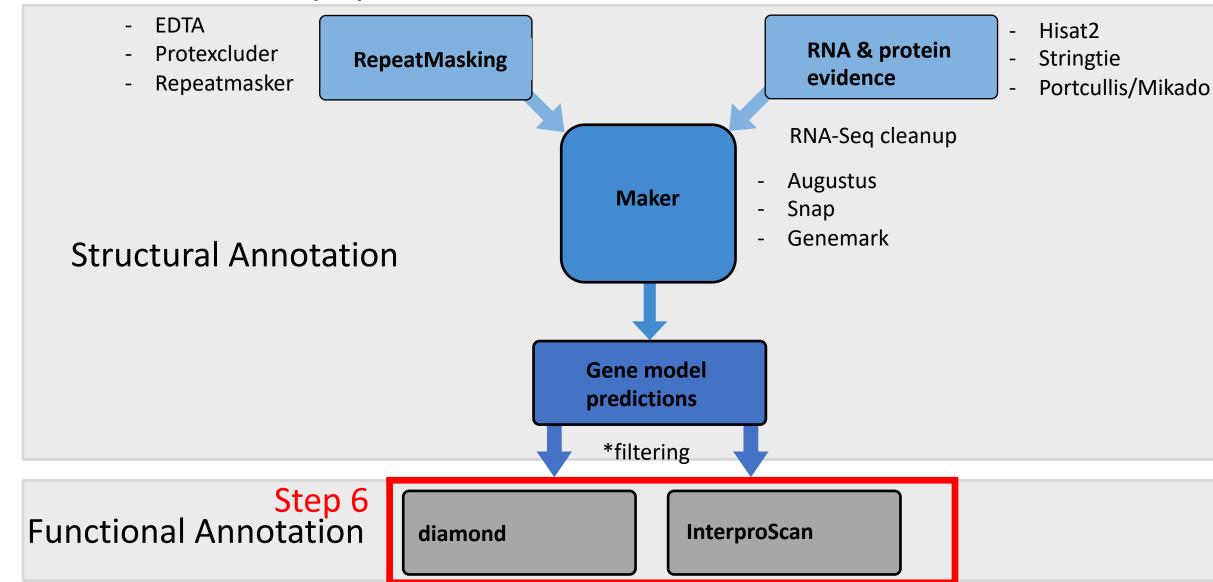
augustus --species=Ugibba genes.gb.test | tee firsttest.out
```

• These commands are also in https://github.com/bcbc-group/Botany2020NMGWorkshop/blob/master/annotation/training.sh



Step 5: Running maker

 https://github.com/bcbcgroup/Botany2020NMGWorkshop/blob/master/annotation/5 maker. sh



Postprocessing, Cleanup, and QC

- Remove Transposons
- complete genes only
- match to nr, e-20
- FPKM > 0.1
- AED > 1
- InterProScan domain
- Comparison to relative, length and number of genes
- Gene families
- BUSCO
- Change gene model names once structural annotation is completed.
- Versioning –very important
- Apollo

Step 6: Functional annotation

 https://github.com/bcbcgroup/Botany2020NMGWorkshop/blob/master/annotation/6 function annot.sh

 Maker also has several scripts for postprocessing files under: /opt/maker/bin