SeqTrimNext Statistics of pre-processing

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1 Output Files

SeqTrimNext provides several files, the most interesting ones are in the following directories:

output_files

- output.less, containing an extensive information about the trimming of each sequence. It can be visualised on terminal using the command less -R.
- used_params.txt, containing the complete set of parameters used for execution of SeqTrimNext with your data
- rejected.txt, containing a list of rejected sequences together with the reason for their removal.
- initial_stats.json, containing statistics for raw sequences.
- stats.json, containing the statistics of the cleaning process.
- There is a collection of folders that gather sequences with the same MID; each folder contains a sequences file (in FASTQ format) with useful reads. There may also exists a file with reads containing low complexity regions. If you want to reconstruct a SFF with the useful segment of each pre-processed read, use sff_info file in combination with the original SFF file for the sfffile tool.

• graphs

- size_stats.png, a graph with the distribution of read lengths in raw data (see Fig. ??).
- qualities.png, a graph to inspect read qualities in raw data (see Fig. ??).
- PluginExtractInserts_insert_size.png, a graph with the distribution of read lengths after SeqTrimNext pre-processing (see Fig. 1).
- There are other graphs (mostly bar plots) that illustrate the quality of pre-processed reads. All are in PNG format.

• latex

 It is provided as a compressed file latex.zip containing all ".tex" files required to compile this document. Graphs are taken from the graph folder

2 Relevant parameters

In this section, the relevant parameters used in your experiment are shown. Full information about the parameters can be obtained from file used_params.txt





2.1 General

Plugins applied to every sequence, separated by commas. Order is important

- 1. PluginIndeterminations
- 2. PluginFindPolyAt
- 3. PluginAbAdapters
- 4. PluginUserContaminants
- 5. PluginContaminants
- 6. PluginVectors
- 7. PluginLowQuality
- 8. PluginLowComplexity
- $9. \ \ Plugin Extract Inserts$

Remove duplicated (clonal) sequences (using CD-HIT 454)

remove_clonality: false

Minimum insert size for every trimmed sequence

min_insert_size_trimmed: 30

Minimum insert size for each end of paired-end reads; true paired-ends have both single-ends longer than this value

min_insert_size_paired: 40

Seqtrim version

seqtrim_version: 2.0.67

min_sequence_size_raw:

2.2 Quality

Minimum quality value for every nucleotide

min_quality: 20

window_width:

2.3 Contaminants

Blast E-value used as cut-off when searching for contaminations

blast_evalue_contaminants: 1.0e-10

Minimum required identity (%) for a reliable contamination

blast_percent_contaminants: 85

Minimum hit size (nt) for considering a true contamination

min_contam_seq_presence: 40

Genus of input data: contaminations belonging to this genus will be ignored genus:





Is a contamination considered a source of sequence rejection? (setting to false will only trim contaminated sequences instead of rejecting the complete read)

contaminants_reject: true
Path for contaminants database
 contaminants.fasta
 cont_ribosome.fasta

3 Pre-processing statistics

Next figure is equivalent to Figure ?? but using output reads (useful sequences). The mode is expected to decrease but the shape of the plot should be similar.

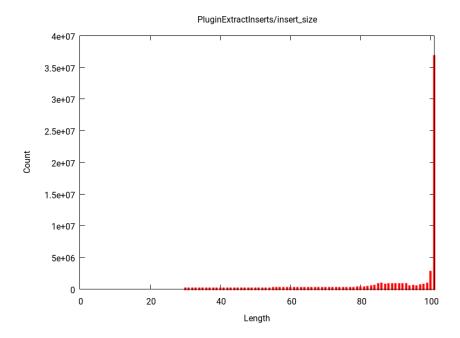


Figure 1: Size distribution of the output sequences. Short sequences (< min_insert_size_trimmed) were removed. [PluginExtractInsert_size.png]

Summary statistics of the SeqTrimNext analysis. Be careful and read all warnings that are indicating concerns about your data. In the files initial_stats.json and stats.json can be found a full statistics of your data and SeqTrimNext pre-processing





	63054976
mallest read (bp)	101
(- /	101
Mode (bp)	0
Mean (bp)	0.0
total	1599900
Rejected	4783318
complexity reads	35596
Mode (bp)	91
Mean (bp)	93.9
tnut paired reads	56636162
otai output reads	58236062
	Largest read (bp) Mode (bp) Mean (bp) total Rejected complexity reads Mode (bp)

Table 1: List of the most frequent Vectors found among your reads

Vectors	sequences
Cloning vector pAAV-MCS, complete sequence.	337451
Cloning vector pKOHPRT complete sequence.	325092
Enterobacteria phage lambda	101565
Cloning vector pVLH/hsp	40368
Cloning vector pWormgate2, complete sequence.	32309

Table 2: List of the most frequent Adapters found among your reads

Adapters	sequences
TruSeq_Universal_Adapter	331948
ABI_Solid3_Adapter_A	176880
TruSeq_Adapter_Index_4	174753
ABI_Solid3_GAPDH_Reverse_Primer	141107
Illumina_Single_End_Adapter_1	126552





Table 3: List of the most frequent Contaminants found among your reads

Contaminants	sequences
rRNA_small_subunit_Metazoa_Myotis	67005
Saccharomyces cerevisiae S288c chromosome XII, complete se-	56794
quence	
rRNA_long_subunit_Metazoa_Dasypus	46108
Podospora anserina S mat+ unordered scaffolds, whole genome	42285
shotgun sequence	
Schizosaccharomyces pombe 972h- chromosome III, complete se-	41873
quence	

Table 4: Summary of nucleotides removed in every plugin.

Plugin	Nucleotides	Percent	Warnings
Low Quality	577798730	Inf %	OK
Low Complexity	56674223	Inf $\%$	OK
Poly T	22303905	Inf $\%$	OK
Poly A	23942792	Inf $\%$	OK
Contaminants	38180703	Inf $\%$	OK
Adapters	76382458	Inf $\%$	OK
Vectors	40581720	Inf $\%$	OK
Indeterminations	202093	Inf $\%$	OK
Inserts	5472744699	Inf %	iW1

iW1 Warning!, only Inf% of nucleotides are useful

4 Rejected reads

Input sequences	63054976
Output sequences	1599900
Rejected sequences	4783318
Output paired sequences	56636162
Total output sequences	58236062
Low complexity sequences	35596





Table 5: Summary of reads removed in every plugin.

Case	Number of sequences	Percent	Warnings
	4783318	7.586~%	OK
Short inserts	2711426	4.300~%	OK
Empty Inserts	1646021	2.610~%	rdW3
Contaminants	407736	0.647~%	OK
No Valid Inserts	11323	0.018~%	OK
Low Complexity	5832	0.009~%	OK
Indeterminations	892	0.001~%	OK
Unexpected Vector	88	0.000~%	OK
Total rejected	4783318	7.586~%	OK

rdW3 Warning!, a 2.610~% of your sequences are empty (without an insert)

References

- [1] Falgueras et al. SeqTrim: a high-throughput pipeline for preprocessing any type of sequence reads. *BMC Bioinformatics* 11:38 (2010)
- [2] Weizhong Li & Adam Godzik. Cd-hit: a fast program for clustering and comparing large sets of protein or nucleotide sequences. *Bioinformatics* (2006) 22:1658-9

Thanks you for use SeqTrimNext! Send us any comment to scbi support



