**A Meta-analysis of Longevity Estimates of Mosquito Vectors of Disease**

Ben Lambert1,2, Ace North1 & H. Charles J. Godfray1

1 Department of Zoology, University of Oxford, South Parks Road, Oxford OX1 3PS, United Kingdom

Corresponding author:

Phone: 01865 271176

2 Present address: MRC Centre for Outbreak Analysis and Modelling, Infectious Disease Epidemiology, Imperial College London, London W2 1PG, UK.

**Abstract**

Mosquitoes are responsible for more human deaths than any other organism, yet we still know relatively little about their ecology. Mosquito lifespan is a key determinant of transmission strength for the diseases they vector, but the field experiments used to determine this quantity – mark-release-recapture (MRR) studies and wild-caught dissection of female mosquitoes – produce estimates with high uncertainty. In this paper, we use Bayesian hierarchical models to analyse a previously-published database of 232 MRR experiments and compile then analyse a database of 131 dissection studies to produce the first ever species-and genus-level estimates of mosquito lifespan. Notably, for the major African malaria vector *Anopheles gambiae s.l.*, we estimate lifespans of 4.5 days (mean estimate; 25%-75% CI: 3.8-5.1 days for unfed female mosquitoes from the MRR analysis) and 9.5 days (mean estimate; 25%-75% CI: 5.2-11.0 days from the dissection analysis); and a lifespan of 4.3 days (mean estimate; 25%-75% CI: 3.6-4.8 days, only present in the MRR database) for the predominantly East-African vector *A. funestus s.l*. We estimate LBLs of 7.0 days (mean estimate; 25%-75% CI: 4.5-8.5 days from the MRR analysis) and 5.0 days (mean estimate; 25%-75% CI: 3.5-5.1 days from the dissection analysis) for *Aedes aegypti*; and 12.1 days (mean estimate; 25%-75% CI: 10.0-13.7 days fromthe MRR analysis) for *Ae. albopictus* – the predominant vectors of dengue fever, chikungunya and Zika. Our estimates indicate that there is significant variation in lifespan across species, with most variation explained by diﬀerences between genera. In correspondence with laboratory studies, we estimate that female mosquitoes outlive males by 1.2 days on average (mean estimate; 25%-75% CI: 0.3-1.6 days). By fitting a range of survival models to the data, we determine that relatively few species within our databases indicate evidence of senescence. Our analysis applies a common framework to the analysis of databases of MRR and dissection-based experiments, allowing us to produce robust estimates of lower bounds on lifespan. It also enables us to critically appraise each field method, highlighting a need for alternative field methods for measuring this important mosquito characteristic.

**Author summary**

Mosquitoes transmit some of the most important diseases aﬄicting humans, with malaria alone killing between 0.4-1.2 million people annually, chiefly children in low-income countries. The transmission strength of these diseases depends critically on the duration of mosquito lifespans and some of the most successful disease control interventions, including insecticide-treated bednets, explicitly target reductions in mosquito longevity. In this study, we conduct meta-analyses of two important classes of field experiments used to estimate wild mosquito lifespan: mark-release-recapture studies, where mosquitoes are marked with dye then released with the number of marked mosquitoes caught monitored over time; and experiments involving dissection of wild-caught females, whose reproductive anatomy is used as a biological clock to determine physiological age. In both analyses, we estimate that most mosquito species live less than 10 days on average, which suggests that relatively few mosquitoes live sufficiently long to transmit disease. We find evidence of variation in mosquito mortality across species, with the estimates of lifespan obtained from each method largely corresponding for the few species with data from both experiments. Finally, by fitting a range of survival models to the data, we conclude that, for most species, mosquitoes do not experience strong age-related increases in mortality.

**Author contributions**

HCJG, AN and BL were involved in conceptualising this study. BL was responsible for data curation and the formal analysis of the data. BL and AN developed the statistical methodology and conducted the investigation. All authors were involved in drafting the original manuscript and revising it.

**Keywords**

mosquitoes, mortality, meta-analysis, senescence, mark-release-recapture, vector-borne disease, Bayesian, hierarchical model

**Introduction**

Some of the most important infectious diseases aﬄicting humans are transmitted by mosquitoes (Gates, 2014), including pathogens, such as the causative agent of malaria, that have been associated with humans throughout our evolutionary history (Carter and Mendis, 2002) as well recently emergent infections, such as the Zika virus (World Health Organisation, 2016). Most mosquito species have a “gonotrophic cycle” involving successive episodes of vertebrate blood feeding, egg maturation and oviposition (Silver, 2007). In order for a mosquito to transmit a pathogen it must feed on an infectious person and live long enough to complete at least one gonotrophic cycle and feed on an uninfected and susceptible individual. Adult lifespan is thus a critical determinant of the ability of a mosquito population to allow the persistence of an indirectly transmitted infection (Macdonald, 1957). Lifespan can of course be straightforwardly assessed in the laboratory, but it is generally accepted that measurements under relatively benign laboratory conditions have limited relevance in the field, and much eﬀort has been directed at estimating this parameter in the vector’s natural environment (Clements and Paterson, 1981; Guerra et al., 2014). Most work has focused on assessing average daily mortality rates, and the simplest assumption is that these do not vary with mosquito age – in this case, longevity is simply the reciprocal of mortality. Testing this assumption and discovering whether mosquitoes senesce or show other types of age-dependent mortality has also been studied in the field (Clements and Paterson, 1981; Harrington et al., 2008; Hugo et al., 2014).

There are two main strategies to estimate mosquito mortality rates and longevity. The first is through mark-release-recapture (MRR) experiments, a technique that is widely applied to estimate these parameters in many types of animal. As applied to mosquitoes, insects are caught in the field or reared in the laboratory and then marked, typically with fluorescent dust. The mosquitoes are then released into the field and then eﬀorts are made to recapture them, for example using human baits or light traps, usually over an extended period of time. Mortality rates can be statistically estimated from the numbers of recaptures given certain assumptions (Silver, 2007). The main challenges with MRR is ensuring the marking technique does not aﬀect recapture probability and distinguishing mortality from mosquitoes dispersing out of range of being recaptured. Also, releasing insects that can transmit disease (especially if this increases ambient population levels) raises important ethical issues.

The second approach is specific to female mosquitoes and makes use of their gonotrophic cycle and involves two distinct dissection-based techniques. The simplest and most widely used approach is based on the observation that the appearance of the fine tracheoles incasing ovaries changes irreversibly when ovaries first develop (Detinova, 1945). The proportion of parous individuals – those individuals that have borne offspring – can be determined by dissecting field-caught specimens and, by making assumptions of the duration of gonotrophic cycles, yields estimates of lifespan. In honour of the entomologist who first made this observation, this approach is known as Detinova’s method. The crude dissection technique needed to apply this method means it has been widely adopted, but its simplicity means it provides limited information about mortality. The next approach requires more sophisticated dissection and, rather than producing a dichotomous determination of reproductive status, yields a count of the number of reproductive cycles a mosquito has undergone. The mosquito ovary is made up of ovarioles, each of which typically produces one egg every gonotrophic cycle. After the egg passes into the oviduct, the distended ovariole does not completely recover its previous form but a discrete dilation remains which can be detected by dissecting the female reproductive organs (Polovodova, 1949). A skilled dissector can determine the number of such dilations, so providing richer data on longevity. After the scientist first observing these changes, this approach is known as Polovodova’s method. The challenges of this method include the amount of time and expertise it takes to collect data and lack of consensus regarding the type of oogenesis producing observable dilations, complicating interpretation of data. Both dissection approaches are specific to females and require conversions between physiological and chronological time (though the distribution of the number of gonotrophic cycles wild-caught mosquitoes have gone through is of direct epidemiological relevance).

An issue with all methods is that they require logistically diﬃcult and expensive field campaigns. There is thus value in conducting a meta-analysis of existing data to explore consistency across studies, to identify correlates of lifespan and to learn lessons for further studies. Here, we apply a common statistical methodology to analyse data from 232 MRR, 1490 observations of parity obtained through Detinova’s method, and 131 studies which used Polovodova’s method to determine physiological lifespan. For MRR, we make use of a valuable database of 394 mosquito studies assembled by Guerra et al. (2014). For parity observations of Detinova’s type, we use a valuable database of anopheline malaria vectors assembled by Massey et al. (2016) comprising bionomic data assembled from 877 references. We extracted data from studies using Polovodova’s method ourselves via a literature search. We concentrated on the three major genera of mosquito vectors, *Anopheles,* *Aedes* (in its traditional sense) and *Culex*, which constitute the majority of thedata.

MRR estimates the length of time a mosquito remains alive and is still in the area available for recapture, meaning that estimates of lifespan using this data are likely biased downwards. In dissections of females, the majority of ovarioles have fewer dilations than the number of gonotrophic cycles an individual has experienced, also meaning that estimates derived from these data likely understand true physiological age (Hugo et al., 2008). It is unclear which of these methods leads to lower estimates but in both cases we term our estimates lower bounds on lifespan, which we shall refer to as LBL.

**Results**

**Chronological longevity estimated from MRR studies**

*Variation in LBL across MRR studies*

To begin, we estimated LBL independently for each available MRR time-series (Figure 1; Methods). The estimates varied substantially both within and among species, though a majority were less than ten days (187 of 236 time-series estimates, based on posterior medians). In comparison, mosquito longevity in laboratory conditions is typically found to exceed 30 days (e.g. Styler et al., 2007). Our estimates ranged from 0.7 days from a study of *Anopheles* *annulipes walker* (a species predominantly found in Australasia) to 38.3 days from a study of *Aedes aegypti* (all estimates are posterior medians). It is likely that the very short longevity estimates reflect dispersalout of the recapture zone or a violation of the assumptions of our analyses, and we thus advise caution in their interpretation (see Discussion). There are multiple data sets for the most important vector species such as *Anopheles gambiae s.l.* (malaria)*, Aedes aegypti* and *albopictus* (yellow fever, dengue and Zika viruses) and *Culex tarsalis* (West Nile Fever, Western Encephalitis) all of which show considerable variation. For example, there are 54 estimates of LBL for *Ae. aegypti* which range from 2.2 days to 38.3 days with a mean of 8.3 days and coeﬃcient of variation of 0.7 (all estimates are posterior medians). To help make sense of the variation both within and among species, we next consider the following four potentially confounding factors: (i) mosquito sex, (ii) whether or not the mosquitoes are fed before release, (iii) the spatial extent of the recapture zone, and (iv) the average temperature during the MRR study.

*Mosquito sex*

The MRR studies included male-only and female-only releases, and mixed releases of both sexes which we used to estimate male and female LBL at the genus level (Fig. 2 – POSSIBLY GO INTO SUPPLEMENT). There was a consistent trend for females to live longer than males for each genus, with the diﬀerence largest for *Aedes* (2.5 days; posterior median; fraction of pairwise posterior samples of females versus males where diﬀerence was less than zero, p<0.01), followed by *Anopheles* (2.0 days; p=0.17) and *Culex* (0.3 days; p=0.34). Overall, female mosquitoes were estimated to live 0.8 days longer than males (posterior median estimate; p=0.10).

*Feeding before release*

The MRR data includes information on whether mosquitoes were pre-fed with sugar, blood, both, or alternatively unfed, which we used to determine the effects of feeding on LBL at the genus level and across all studies (Fig. S4). Since there were insufficient data on males that were fed with sugar versus unfed, we estimated a pooled effect of sugar-feeding on the log scale (see SOM). We estimate that female mosquitoes fed on sugar pre-release lived on average for 0.6 days longer than those that were not fed (posterior median; fraction of pairwise posterior samples of females versus males where diﬀerence was less than zero, p=0.15; Fig. S4; a pattern that was consistent across the genera) versus 0.5 days for males (p=0.15). The effect of blood-feeding on female mosquitoes was less marked (0.1 days; p=0.44).

*The spatial extent of the recapture zone*

Following a release of marked mosquitoes, the rate of their recapture typically reduces in time because some mosquitoes die, and also because some disperse out of the recapture area. These factors are indistinguishable in spatially-averaged recapture data which is why our estimates are lower bounds on lifespan. If dispersal out of the recapture area commonly reduces the LBL below the true lifespan then we should expect a positive correlation between the spatial extent of the recapture zone and LBL. We found no such pattern (Fig. S2), although there was a positive correlation between LBL and trap density (Fig. S3).

*Ambient temperature*

To assess whether temperature affects LBL we used weather records to calculate average temperatures at the MRR sites (see Methods). Using both linear and quadratic temperature terms in regressions, we found no significant relationship between study-site temperature and LBL (overall or within genus; Fig. S5). This result held if, instead of pooling results from all time series, we considered the four species with the most data individually (*Ae. aegypti*, *Cx. tarsalis*, *A. gambiae s.l.* and *A. culicifacies s.l.*; Fig. S6).

*Species and genus-specific variation in LBL*

We computed species (or species complex) and genus specific estimates to subsume the variation within these taxonomic groupings. Additionally, we estimated LBLs by pooling data across all of the studies. To ensure fair comparison, we present estimates for females that were not fed blood or sugar before release (Fig. 3At the species level, the longest estimate was 26.9 days for the species *Aedes simpsoni s.l.* (an African vector of yellow fever). The smallest estimate was 1.1 days for *Anopheles* *subpictus s.l.*(an Asian malaria vector)which is unfeasibly short and almost certainly reflects dispersalout of the recapture zone or a violation of the assumptions of our analyses. There were also differences in longevity at the genera level, with *Culex* estimated to have the shortest longevity (2.9 days) and *Aedes* the longest (8.1 days). *Anopheles* were estimated to live on average 6.8 days while the average across all the available data covering the three genera was 6.0 days. The diﬀerences between genera were significant (ANOVA on median LBL controlling for sex and pre-release feeding: F2,229 = 12.4, *p* <0.01; Kruskal Wallace: *, p <* 0*.*01). Measures of model fit indicated that after the eﬀect of genus is accounted for, the incorporation of a species term provides little predictive power (Fig. S1; in part explained by the latter model over-fitting the data where there are few time series per species).

**Reproductive longevity estimated from dissection studies**

Dissection allows the number of completed gonotrophic cycles to be counted and from this the mean number of cycles before death was estimated. Across the 131 studies, 95% of the individual time series estimates were less than 3 gonotrophic cycles (Fig. S7) and, overall, the mean number of cycles completed in a lifetime was 1.3 (posterior mean; Fig. 4; Table S2). The estimated greatest number of cycles was for *Anopheles sergentii* (3.0 cycles; posterior mean) which is adapted to desert conditions (it is known as the “oasis vector” of malaria; Sinka et al., 2010) and may have evolved greater longevity. The major African malaria vector *A. gambiae* *s.l.* was estimated to be the second longest living (2.4 cycles; posterior mean).The smallest estimated mean number of gonotrophic cycles was for *Anopheles* *bellator* (0.6 cycles; posterior mean) which transmits malaria in Brazil’s AtlanticForest. There were significant diﬀerences in estimated lifetime gonotrophic cycles amongst species (ANOVA: F24,106 =2.2, *p* <0.01; the non-parametric Kruskal-Wallace: , *p*<0.01).

The estimated lifetime gonotrophic cycles for the diﬀerent genera were *Anopheles,* 1.6; *Culex,* 1.2; *Mansonia*, 1.1; and *Aedes* 0.8 (Fig. 4; Table S2) and the diﬀerences between the genera were significant (ANOVA: F3,127 =3.4, *p* =0.02; the non-parametric Kruskal Wallace: , *p*<0.01).

**Comparison of longevity estimates from MRR and dissection studies**

Using the data collected from a literature search, we estimated that the first gonotrophic cycle duration had a mean of 4.3 days (std. error: 0.4 days) and, for subsequent cycles, the mean was 3.9 days (std. error: 0.4 days; see SOM). To compare the two methods, we converted numbers of gonotrophic cycles (physiological age) into lifespan (chronological age) as described in the SOM using these estimates of gonotrophic cycle duration. Table S3 provides posterior summaries of chronological lifespan for the species and genera in the dissection dataset (see also Fig. S10). For 12 species, we had enough data from both species to make a comparison, and there was a positive correlation (not statistically significant; Pearson correlation *ρ* = 0*.*34, *n* = 12, *p* = 0*.*27) between the two measures (Fig. 5), and in only one case – for *A. darlingi* - there was a significant diﬀerence in the time-series level LBLs (Table S4).

**Evidence for age-dependent mortality**

The survival model upon which the above analyses are based is the single-parameter exponential model which assumes an age-invariant mortality hazard. We also fitted five multi-parameter models that allow, in diﬀerent ways, mortality to vary with age. We did this to maximise our chance of detecting age-varying mortality (though aware of the risks of false positives with multiple estimations). In Fig. 6, we compare the predictive performance of the six models for describing lifespan in MRR studies of 33 species (see SOM for details). We categorised the evidence for age-dependent mortality in each species according to the performance of the five age-dependent models versus the exponential: ‘+’ indicated that all age-dependent models outperformed the exponential; ‘?’ indicated that the exponential outperformed one or more age-dependent models; and ‘-’ indicated that the exponential performed at least as well as all other models. Overall, we estimated that there were 8 ‘+’ species, where age-dependent mortality fit the data better; 11 ‘?’ species where the evidence was mixed; and 14 species where constant mortality models performed at least as well. The species where age-dependent mortality best fit the data included *Ae. Aegypti*, the main vector of dengue fever, Zika and chikungunya. These studies also tended to include multiple release MRR studies which, on average, were conducted over a longer period of time than the others, which may be why we 76failed to detect age-dependence in the latter (Fig S11).

In Fig. 7, we compare the predictive performance of the six models for describing lifespan in dissection studies of 25 species, and categorise the evidence in the same way as for the MRR analysis. By our metric, we determined that there were only two species with evidence for age-dependent mortality (*A. gambiae s.l.* and *A. minimus*).

Overall, we conclude that the evidence for age dependence from MRR and dissection studies is mixed. From the two groups of species that were identified as exhibiting senescence from each meta-analysis, there was no overlap. Using the Gompertz model only, we compared the predictive accuracy versus the exponential for the 12 species with data from both analyses (Fig. S12) and only in one case (*A. subpictus s.l.*) there was consensus that the age-dependent model provided a better fit to the data. It is possible that some mosquito species do not live long enough in the wild to experience physiological decline. In support of this, a Spearman’s rank correlation test indicated that there was positive correlation between the ranked estimated LBLs of the species and the ranked mean predictive accuracy of age-dependent models for the MRR analysis (*ρ*=0.19, p=0.01), however this was not significant for the dissection analysis (*ρ*=0.07, p=0.43).

**Estimates of the fraction mosquitoes capable of transmitting disease**

We can use the LBL estimates from our analysis to estimate the fraction of mosquitoes that live beyond a certain age. In order to transmit a disease, a mosquito must live longer than the length of the intrinsic incubation period (the time taken for a pathogen ingested in one blood meal to be ready to be transmitted during a future feeding event). This is a lower bound as it does not include the waiting time to find a host after feeding or egg maturation. In Fig. 8, we plot the fraction of the mosquito population that pass this threshold using estimates from both MRR and dissection studies for vector species (see SOM for references used to identify species as vectors) and their most significant diseases.

For malaria, estimates of the minimum fraction of the population that can transmit the disease vary from <0.1% for *A. subpictus* (posterior median; from the MRR analysis, as noted above likely to be due to the LBL substantially underestimating lifespan) to 52% (posterior median) for the drought-adapted and long-lived *A sergentii.* The proportions surviving long enough to become infectious for *A. gambiae s.l.* were 10% (from MRRs) and 27% (from dissection studies); and for *A.* *funestus s.l.*, 9% (from MRRs). Using the individual time series estimates, thereevidence for a diﬀerence in EIP between the species (Kruskal-Wallis used due to non-normality of data; MRR: , *p* <0.01; dissection: = 38*.*9, p<0.01).

*Ae. aegypti* and *Ae. albopictus* are the main vectors of dengue, chikungunyaand Zika viruses. Because of their short intrinsic incubation periods a greater fraction of mosquito potentially live long enough to transmit diseases (Fig. 8), rising to a maximum of 84% for *Ae. albopictus* transmitting chikungunya.

**Discussion**

In this study, we applied a Bayesian hierarchical framework to the analysis of a database of mark-release- recapture experiments and another for mosquito dissection studies to estimate mosquito lifespan. By applying a single framework, this allows us to eﬀectively synthesise information from the disparate experiments which, individually, estimate lifespan with considerable uncertainty. Due to the assumptions required to analyse the field data, our estimates represent lower bounds on lifespan (LBL). Across both meta-analyses, the estimated LBLs were mostly less than 10 days, hinting that only a small proportion of mosquitoes may live long enough to transmit disease. We determined that LBL varies across species and genera, although most variance is explained by genus. The MRR analysis includes experiments conducted on each sex individually, and we estimate that, on average, males live shorter lives than females. Pre-release feeding with sugar also lengthens lifespan across all three genera, although this eﬀect is less marked than the sex diﬀerences. In contrast to a number of lab-based experiments (Yang et al., 2009; Brady et al., 2013), temperature was not determined to significantly impact lifespan. By fitting a range of survival models to the data in both meta-analyses, we could assess evidence for age-dependent mortality. Overall, we conclude that the evidence is mixed: in the MRR experiments, in 8 of 33 species we found evidence for mosquito senescence, whereas in only 2 of 25 species included in the dissection analysis were better fit by a model incorporating an increasing risk of mortality with age.

MRR experiments are known to produce downwardly-biased estimates of lifespan. Lab experiments have demonstrated that marking can negatively impact survival

(Verhulst, Loonen, and Takken, 2013; Dickens and Brant, 2014) resulting in artificially depressed survival. MRR studies typically cannot diﬀerentiate between a mosquito dying and dispersal from the study area meaning that lifespan will be underestimated. In this study, we found a positive correlation between lifespan estimates and the density of traps, indicating that better trapping coverage likely raises estimates towards their real value. We conducted an *in silico* Monte Carlo study to determine how accurately we could estimate mosquito lifespan given study parameters in an ideal MRR experiment, where the assumptions of no emigration and harmless marking are fully satisfied (see SOM for full details). This work indicated that for many of the experiments, the short study lengths or typical numbers of mosquitoes released, results in considerable uncertainty in lifespan estimates (Fig. S12). This indicates that statistical power can be substantially increased by pooling data across experiments as we did using a Bayesian hierarchical model.

The key assumptions of dissection based methods to determine chronological age are: (i) physiological age can be accurately determined by dissection of female specimens (unlike MRR, this method can only be applied to one sex), (ii) the relationship between physiological and chronological age is known, (iii) the population being sampled is in equilibrium (recruitment matches mortality) and (iv) individual mosquitoes can be randomly sampled from the population. The reliability and accuracy of dissection has been questioned. The objections include the impracticality of dissecting more than a small proportion of ovarioles (Hoc and Wilkes, 1995), particularly in African vector species (Gillies and Wilkes, 1965), the related issue of locating ovarioles whose count of dilations represents true physiological age (Fox and Brust, 1994), and the variation in numbers of ovariolar dilations for mosquitoes of the same, known, physiological age (Kay, 1979; Russell, 1986; Hugo et al., 2008). Indeed there is considerable uncertainty concerning the fundamental question of how dilations in ovarioles form in the first place. Whilst the ‘Old School’ of thought (a term coined by Fox and Brust, 1994) headed by Polovodana (Polovodova, 1949) and Detinova (Detinova, 1962) considers dilations to result from normal oogenesis, a ‘New School’ headed by Lange and Hoc (Lange and Hoc, 1981) has challenged this assertion. The New School believe that only abortive oogenesis results in follicular dilations because normal oogenesis destroys the sack-like structures (Fox and Brust, 1994). This means that Polovodana’s method requires dissecting large numbers of ovarioles to uncover those with the most dilations, where abortive oogenesis has occurred in each gonotrophic cycle. They deem these ovarioles ‘diagnostic’ since only in these cases the number of dilations equals the number of gonotrophic cycles that have occurred. As a mosquito ages, the number of diagnostic ovarioles diminishes, since the random occurrence of normal oogenesis in a particular ovariole means its dilation count does not equal the number of gonotrophic cycles undertaken. This increased diﬃculty of finding diagnostic ovarioles as a mosquito ages would elevate the chance of age ‘hypodiagnosis’ for older specimens (Fox and Brust, 1994), and likely biases lifespan estimates downwards. The diﬃculty of locating diagnostic ovarioles has been investigated using lab populations of *Culex* and *Aedes* mosquitoes by Hugo et al. (2008), who conclude that only a small percentage of ovarioles are diagnostic. The exchange rate between physiological age and chronological age is the duration of gonotrophic cycles. Two methods are commonly used to estimate the duration of gonotrophic cycles: MRR studies (see, for example, Gillies and Wilkes, 1965), where marked mosquitoes are recaptured and dissected to determine the number of gonotrophic cycles occurring since release; and laboratory-based observations of colonies of (typically) wild-caught females, or their progeny (see, for example, Afrane et al., 2005). Whilst it is unclear how each method could bias estimated gonotrophic cycle duration, in our analysis, laboratory-based studies indicated a longer gonotrophic cycle (Fig. S9). The distributions we used to convert physiological age into calendar age were calculated by pooling data across both approaches, to incorporate uncertainty from both experimental procedures. It is possible, however, that this aggregate approach may induce biases in estimates and an approach more entrenched in experimental knowledge would fare better. If a population of mosquitoes is shrinking, this leads to a relative under-abundance of young mosquitoes, and a flattening of the survival curve, resulting in over-estimates of lifespan. For stable populations, periods when shrinking occurs must result in equal changes in the population size compared to those when it expands. If mosquito collections occur with equal frequency in each of these two modes, then aggregating the data across all sampling times and estimating a single model, as we do here, should yield an approximately unbiased estimate of lifespan. The additional uncertainty of a fluctuating population size, however, could lead us to understate the uncertainty in estimates. Field entomologists have challenged the assumption of random sampling the mosquito population, although there are conflicting opinions as to whether this results in a relative paucity (Gillies and Wilkes, 1965) or abundance (Clements and Paterson, 1981) of nulliparous individuals. In our database, there are cases where there was an obvious deficit of nulliparous individuals, which has previously been ascribed to the diﬀering distribution of resting females between indoor and outdoor traps (Detinova, 1962; Clements and Paterson, 1981). We chose to not include those counts of nulliparous individuals in our analysis where their number was less than 90% of the uniparous. Whilst we see no obvious diﬀerences in lifespan according to collection method (data not shown) or location, it is possible that the assumption of random sampling is violated, although the directionality of the bias induced by this is unclear. Overall, the assumptions underpinning estimates from dissection studies indicate that our estimates represent lower bounds on lifespan. The alternative dissection-based approach of Detinova Detinova, 1962, based on dichotomous categorisation of female mosquito specimens as ‘parous’ or ‘unparous’ relies on fewer assumptions, and is widely used. Further work examining parity rates in field specimens may be fruitful although, in principle, it oﬀers less information on the age structure of a population than Polovodova’s approach.

By applying a common method to analysing all studies in our databases, it is possible that we may have missed patterns of mortality that would have been evident from using a more bespoke approach. As our *in silico* analysis of MRR experiments indicates, however, the overdispersed data from single experiments results in high measurement error (Fig. S12). By applying diﬀerent methods to each study, this could lead us to falsely detect patterns when none are present, and we prefer a pooled approach.

The diﬀerent nature of the assumptions of each of the two methods means they oﬀer complimentary information on mosquito survival. We also note that Polovodova’s dissection-based studies require specialised expertise which will often be unavailable, whereas MRR methods can more readily be used. Furthermore, most if not all dissection methods that have been used previously are only applicable to female mosquitoes, whereas MRR can be applied to either sex and can additionally be used to determine other ecological parameters (for example, population size and dispersal). Although dissection data gives detailed of age-structure, we thus foresee a continued reliance on MRR experiments in field entomological experiments. Eﬀorts to use both approaches concurrently will be particularly useful and will allow quantification of the biases induced by the assumptions of each. Similarly, MRR experiments releasing large numbers of marked mosquitoes and recording spatiotemporally-disaggregated captures of wild and re-caught marked mosquitoes will continue be useful in estimating lifespan and dispersal.

To compare estimates of lifespan derived from MRR with those from dissection-based methods, we display the estimates of lifespan from those ten species occurring in both databases in a single plot (Fig. 5). In is reassuring that there is correlation between estimates from both approaches, although the small sample size likely hindered our ability to determine statistical significance. In both cases, we estimate that *A. sergentii* was amongst the longest lived of the anopheline species with an LBL of 12.4 days (mean estimate; 25%-75% CI: 5.9-13.8 days) from the MRR analysis and 11.9 days (mean estimate; 25%-50% CI: 7.6-14.0 days) from the analysis of dissection studies. This species is a vector of malaria in the Sahara (Sinka et al., 2010), where to act as a disease vector it must persevere through these hard conditions. It is reasonable to hypothesise that this species should live longer than those in environments where the potential for blood-feeding and oviposition is greater. The species with the greatest discrepancy in the estimates was *A. gambiae s.l.*, where we estimated LBLs of 4.5 days (mean estimate; 25%-75% CI: 3.8-5.1 days for unfed female) from the MRR analysis and 9.5 days (mean estimate; 25%-75% CI: 5.2-11.0) from the dissection analysis. Across genera, the greatest discrepancy in estimates was for *Aedes*, where the estimates from the MRR studies (8.1 days) are considerably longer than those of dissection-based studies (3.5 days). This was followed by *Culex* (a posterior mean of 2.9 days from the MRR versus 4.9 days from thedissection analysis) with the smallest discrepancy for *Anopheles* (6.8 versus 6.4 days). Across all studies we estimate from the MRR analysis that mean mosquito lifespan is 6.0 days versus 5.5 days from the dissection-based studies. Some of the diﬀerences in these group-level estimates between the two approaches is likely due to environmental and genetic diﬀerences between mosquitoes in the experiments that were analysed in each meta-analysis. However, we believe that

part of the discrepancy can be explained by the methodological diﬀerences in approaches. We speculate that diﬀerences in dispersal rate can explain some of the discrepancy. Both *Anopheles* and *Culex* mosquitoes are generally thought to fly farther during their lifetimes than *Aedes* [Charles, do you have a reference here?], meaning that the estimates from MRR-based approaches will be most downwardly-biased for these genera. This is supported by our results since the dissection-based estimates (themselves not reliant on assumptions about dispersal) are similar or exceed the MRR estimates for *Anopheles* and *Culex* mosquitoes, but not for *Aedes*.

It is widely believed mosquitoes live artificially long under the benign conditions of the laboratory. We find it informative to consider estimates of lifespan derived from observations of such populations as they constitute an upper bound on the lifespan of wild populations. Also, since the numbers of mosquitoes involved in large cage experiments often numbers in the thousands, these estimates have lower uncertainty than those from field experiments although are typically conducted on highly inbred mosquito strains. Styer et al., 2007, using colonies of 45,054 female and 55,997 male *Ae. aegypti*, determined that females lived nearly twice as long as males; the median lifespan was estimated as 31.69 ± 0.06 days for females and 16.39 ± 0.03 days for males. A similar study by Dawes et al., 2009 with a lab colony of over 1000 female *A. stephensi* found similar estimates for median lifespan (31-42 days). These estimates are many multiples of the average estimates that result from our analysis of field data which, as discussed, represent lower bound estimates. Without an unbiased method to measure mosquito lifespan, however, it is diﬃcult to quantify and explain the gap that exists between field and laboratory lifespans. The development of additional methods to estimate mosquito age, such as ‘Near-Infrared Spectroscopy’ (Mayagaya et al., 2009; Sikulu et al., 2011; Lambert et al., 2018) if they are proven to work in the field, may be of considerable worth here.

We conducted a power analysis of MRR experiments to determine whether typical experimental characteristics could detect senescence. Here we calculated the power of a maximum likelihood estimator of the ‘senescence parameter’ *β* of the Gompertz survival function (see Table SM3) for case study populations with three diﬀerent levels of senescence (Fig. S11A). This analysis indicated that power to detect senescence strongly depends on study length (Fig. S11B) but is insensitive to release size (Fig. S11C). Clements and Patterson (1981) conducted a meta-analysis of MRR and dissection-based field experiments and found evidence of an increasing risk of mortality hazard with age that is similar in magnitude to that of the ‘mild’ case considered above. For this case, detecting senescence with a power of 80% requires a study length of at least 18 days. Since the median study duration for experiments included in our analysis was 10 days (Table SM2) this could partly explain our failure to detect senescence at the species level. A number of experiments have found evidence of age-dependence in laboratory populations (Styer et al., 2007; Dawes et al., 2009). However, the artificially benign environment of the laboratory means mosquitoes live considerably longer than in the wild, where they may die because of exogenous

factors, before the eﬀects of physiological decline have had time to manifest. Field experiments have also found evidence for age-dependent mortality. Harrington et al. (2008) conducted a field experiment where mosquitoes reared under laboratory conditions were marked and released at diﬀerent ages. Analysis of the resultant MRR time-series indicated that mosquito mortality increases with age at release. It is possible, however, that this field experiment suﬀers from the same biases as laboratory-based approaches, because the released mosquitoes were often of ages considerably higher (up to 20 days) than typical estimates of wild mosquito lifespan.

As ethical concerns of contributing to disease burden are more often considered, it is now less common for MRR experiments to release female mosquitoes versus males than historically (Fig. SM2). Our analysis indicates that females outlive male mosquitoes by approximately 1.2 days (Fig. 3), meaning that diﬀerences between the sexes may exist for other ecological parameters determinable by MRR. This suggests that continued field entomological work on contained releases of mosquitoes in semi-field sites or large microcosms may be a valuable source of information on female mosquito ecology.

Our estimates of LBL indicate that mosquitoes that were sugar-fed prior to release lived on average 0.7 days longer than those that were unfed (Fig. S4) suggesting the potential value of this underappreciated aspect of the mosquito ecology to the insects. It may also partly explain the recent successes in the use of Attractive Toxic Sugar Baits as a vector control intervention (Müller, Kravchenko, and Schlein, 2008; Müller, Junnila, and Schlein, 2010; Müller et al., 2010a; Müller et al., 2010b; Beier et al., 2012). More research is needed, however, to identify the sugar-feeding frequency and food sources for wild populations.

There is evidence mainly from laboratory studies that temperature modulates mosquito ecology and behaviour (Yang et al., 2009; Brady et al., 2013; Murdock et al., 2012; Beck-Johnson et al., 2013). The locations and times of year over which the MRR studies were conducted encompassed a large range of average air temperatures, from approximately 10 oC to 35 oC and, within this, we determined no relationship between lifespan and temperature across all time series (Fig. S5) or, for any of the species with the most data (Fig. S6). It is possible that by considering a raw average of air temperature across the month, this ignored, more complex, interactions between temperature and lifespan. It is also possible that by ignoring the eﬀects of rainfall (the historical data on rainfall is less likely to be reliable for a given location), that this masked a more complex interaction between longevity and temperature. The observed laboratory relationship between lifespan and temperature, however, may not be as robust in the field if mosquitoes adjust their behaviours (such as, by seeking shade) in reaction to changes in temperature. More work exploring the relationship between mosquito ecology and temperature in semi-field experiments may be useful in probing these interactions further. In this work, we have used modern statistical methods to synthesise precious field data conducted by entomologists past and present, to produce lower bound estimates of mosquito lifespan. The importance of vector mortality for disease transmission has long been recognised, however, since even before 1957, when George Macdonald formulated the now famous Ross-Macdonald equation of R0 for malaria. Indeed, the recent declines in malaria prevalence in Sub-Saharan Africa were likely due to upscaling of interventions (insecticide-treated bednets and indoor residual spraying) that aim to reduce mosquito lifespan (Bhatt et al., 2015). Worryingly, resistance to pyrethroids, the only class of insecticide used in current insecticide-treated bednets and likely the only product to come to market in the near future, has been determined to be widespread and increasing in intensity across Sub-Saharan Africa (World Health Organization, 2018). This alarming trend highlights the need for continued MRR and dissection-based studies to monitor the eﬀectiveness of bednets and determine whether more expensive alternatives, such as nets incorporating piperonyl butoxide be deployed. It also emphasises the need for investment in new tools for real time monitoring of mosquito populations. In recent years, considerable funding has been allocated to molecular and genomic research into mosquitoes that strengthens existing interventions and suggest novel control strategies. Without commensurate funding allocated to applied vector ecology, our lack of knowledge in this area threatens our opportunity to capitalise on molecular advances and potentially hinders our ability to control of mosquito-borne disease.

**Methods**

In recent years many important vectors of disease have been shown to be complexes of closely related species, biotypes or forms that cannot be distinguished morphologically (for example, the morphospecies *Anopheles gambiae sensu lato* is now separated into the widespread *gambiae, coluzzii, arabiensis* and a number of more local species). As the majority of studies analysed here took place before molecular techniques allowed these taxa to be separated we work here chiefly with morphospecies.

**Mark-release-recapture**

Data from MRR experiments in the Guerra et al. (2014) database were examined and those with fewer than six recaptures and species with only a single MRR study were excluded for the hierarchical analysis. Of the 232 data sets, 179 involved only females, 35 males, and 18 both sex releases. For 102 data sets the age of the released mosquitoes was known (the average age of released mosquitoes was 4.0 days) while in the other cases it was unknown or unrecorded; in these cases we assumed the mosquitoes were newly emerged at the time of release and return to this assumption later. See Table SM1 for a summary of other data characteristics.

We analysed all MRR experiments within the same statistical framework (for full details see the Supplementary Online Material (SOM)). In the simplest case *NR* mosquitoes are released on day zero and the probability that they remain inthe recapture area until day *t* is *S*(*t*) when they are recaptured with probability *ψ*. We model the number of mosquitoes recaptured on day *t* using a negativebinomial sampling model with mean (*NR* − *Y* (*t* − 1)) *S* (*t*) *ψ*, where *Y* (*t* − 1) is cumulative captures before day *t*, and shape parameter *κ*. The negative binomial has been used previously in analyses of mosquito count data (Service, 1971; Nedelman, 1983) because of its ability to represent temporal over-dispersion in recaptures most likely caused by variable weather. A slight modification was required for studies with multiple releases (see SOM).

The simplest model for *S*(*t*) assumes there is a constant probability (*λ*) that a mosquito dies or leaves the recapture area so that the numbers remaining after time *t* are given by the exponential distribution, exp(−*λt*). We utilised this form extensively but in testing for senescence used five other models where *λ*(*t*) varies with time so that,

Details of the five models (Gompertz, Weibull, Gompertz-Makeham, Logistic and Logistic-Makeham), which vary in their ability to detect diﬀerent forms of age-dependent mortality, are given in the SOM. Using multiple diﬀerent types of models increased our chances of detecting senescence though also increases the likelihood of false positives.

Parameters were estimated using Bayesian techniques with relatively uninformative priors for *κ* and the parameters of *λ*(*t*), but assuming a prior for *ψ* indicating a low recapture probability (bounded in part by knowledge of the maximum daily recapture rates; see SOM). We used a Bayesian hierarchical model to estimate distributions of lifespan at the species and the genus levels, and across the complete data set. This procedure assumes that there is a distribution of lifespan parameters for each species from which those governing individual MRR time series are sampled, and similarly a distribution at the genus level from which those for individual species are derived (rather akin to random eﬀects in classical statistics). Within this framework we can also allow the parameters for individual time series to be influenced by co-variates such as diﬀerences in experimental methodology. As in the estimation of the parameters of the individual experiments, relative uninformative priors were set for the parameters of the hierarchical models except for *ψ* where again a distribution representing low recapture probabilities was assumed. Posterior distributions were derived using Markov Chain Monte Carlo (MCMC) methods with convergence assessed using the statistic (Gelman and Rubin, 1992). The predictive power of the modelwas assessed using *K* -fold cross validation which tests the ability of the model fitted to part of the data to predict the rest using multiple different partitions. Further details of the prior specification, fitting and validation through posterior predictive checks (Lambert, 2018) are given in the SOM.

Two studies of *Anopheles balabacensis* reported capture rates increasing with time, presumably reflecting a violation of our assumption of constant recapture probabilities. We omitted this species from the analysis.

The Guerra et al., (2014) database included the latitude and longitude of each study along with the date when the study began. We used this information to find estimates of the air temperature for each study using the European Centre for Medium Range Weather Forecasts’ ERA Interim Daily historical database. For each study we calculated the mean monthly temperature across a spatial area of (latitude ± 1 degree, longitude ± 1 degree), for the month at which each study was carried out. The records for this database begin in 1979, which pre-dates the study date for 65 of our 232 MRR time-series. For these time-series, we chose to estimate the air temperature by an average of the corresponding monthly temperatures over the years 1979-89.

**Dissection**

Studies using dissection to estimate mosquito longevity were located in literature databases using relevant keyword, citation and author searches, and by checking previous studies cited by the papers located (see SOM). The list of studies located with associated metadata is available as a Supplementary Online File.

Most dissection studies recorded the distribution of the number of gonotrophic cycles in mosquito samples collected over a specific period of time. Overall, we found 568 physiological age cross-sections at recorded distinct times in 72 published articles. Our statistical approach relies on steady recruitment to the adult mosquito population. To guard against the eﬀect of fluctuating population sizes on our analysis, we aggregated the data at a given location across cross-sections taken at diﬀerent times. We further omitted time series with fewer than 100 mosquitoes and for species with only one data set leaving 131 studies of mosquitoes in the genera *Anopheles, Aedes*, *Culex* and *Mansonia*.

The dissection data which we use provides measures of the age distribution of mosquitoes within each investigated population. By assuming that the population sizes were approximately fixed throughout the period of investigation, this allows us to estimate mean lifespan using a statistical model of mortality incorporating the probability of mosquito capture. We modelled the number of mosquitoes found by dissection to be of age *a* using the negative binomial distribution with mean Ψ*S*(*a*) and shape parameter *κ*, where Ψ is the product of the recruitment rate of adult mosquitoes, which we assume is constant over time, and the probability of being captured for dissection, and *S* (*a*) is the probability of surviving until age *a*. We used the number of females that have yet to lay eggs (nulliparous) to estimate the recruitment rate as described further in the SOM. Initial examination revealed that in some data sets the number of nulliparous females was anomalously low, something that has been noticed before (Gillies and Wilkes, 1965). As some

studies have suggested that the first gonotrophic cycle tends to be longer than the subsequent ones, this is probably due to diﬀerences in capture probability. In data sets where the fraction of nulliparous females was less than 90% the uniparous (completed on gonotrophic cycle) we excluded the nulliparous observation. Data was analysed using a Bayesian framework as with the MRR data with minor diﬀerences in the specification of the priors (see SOM).

To compare lifespan estimates from dissection and MRR studies we need to convert physiological age (the number of gonotrophic cycles) into chronological age. Using a literature search and a review by Silver (2007) we found 79 estimates in 42 published articles. Most estimates were obtained by dissecting females recaptured in MRR studies or by observations in the laboratory, the latter tending to give longer durations. Studies diﬀered greatly in how (if at all) they represented uncertainty in their estimate of the duration of the gonotrophic cycle. Where confidence limits were given we treated these as the relevant quantiles of a normal distribution, where a range was stated (e.g. “4-6 days”) we interpreted the bounds as the 2.5% and 97.5% quantiles of a normal distribution, and where a single figure was quoted we assumed this was the mean this distribution. Using the quantiles of the normal distribution, we estimated its mean and standard deviation by regression (see SOM). Initially we calculated distributions of gonotrophic cycle lengths at the species and then genus levels, but because of the paucity of data for many species and the lack of significant diﬀerences we aggregated the data into a single distribution. We converted physiological age to chronological age by sampling from this distribution to obtain a particular gonotrophic cycle length for each mosquito (we also explored sampling from this distribution to obtain the duration of *each* gonotrophic cycle which increased the uncertainty in lifespan estimate but did not aﬀect any of the conclusions).

**Acknowledgements**

The authors would like to thank the following for useful conversations throughout the course of this work: Austin Burt, Mike Bonsall, Thomas Churcher, Steve Lindsay and Ellie Sherrard-Smith.

**References**

1. Y. A. Afrane et al. “Eﬀects of microclimatic changes caused by land use and land cover on duration of gonotrophic cycles of *Anopheles gambiae* (Diptera: Culicidae) in western Kenya highlands”. *Journal of Medical* *Entomology* 42.6 (2005), pp. 974–980.
2. L. M. Beck-Johnson et al. “The eﬀect of temperature on Anopheles mosquito population dynamics and the potential for malaria transmission”. *PLOS one* 8.11 (2013), e79276.

1. J. C. Beier et al. “Attractive toxic sugar bait (ATSB) methods decimate populations of *Anopheles* malaria vectors in arid environments regardless of the local availability of favoured sugar-source blossoms”. *Malaria* *Journal* 11.1 (2012), p. 31.
2. S. Bhatt et al. “The eﬀect of malaria control on *Plasmodium falciparum* in Africa between 2000 and 2015”. *Nature* 526.7572 (2015), pp. 207–211.
3. O. J. Brady et al. “Modelling adult Aedes aegypti and Aedes albopictus survival at diﬀerent temperatures in laboratory and field settings”. *Parasites & vectors* 6.1 (2013), p. 351.
4. R. Carter and K. N. Mendis. “Evolutionary and historical aspects of the burden of malaria”. In: *Clinical Microbiology Reviews* 15.4 (2002), pp. 564– 594.
5. A. Clements and G. Paterson. “The analysis of mortality and survival rates in wild populations of mosquitoes”. *Journal of Applied Ecology* (1981), pp. 373–399.
6. E. J. Dawes et al. “*Anopheles* mortality is both age-and *Plasmodium*-density dependent: implications for malaria transmission”. *Malaria* *Journal* 8.1 (2009), p. 228.
7. T. S. Detinova et al. “Age grouping methods in Diptera of medical impor-tance with special reference to some vectors of malaria”. In: *Monograph* *series World Health Organisation* (1962).
8. B. L. Dickens and H. L. Brant. “Eﬀects of marking methods and fluorescent dusts on *Aedes aegypti* survival”. In: *Parasites & Vectors* 7.1 (2014), p. 1.
9. A. Fox and R. Brust. “How do dilatations form in mosquito ovarioles?” *Parasitology Today* 10.1 (1994), pp. 19–23.
10. B Gates. *The Deadliest Animal in the World*. https://www.gatesnotes.com/Health/Most-Lethal-Animal-Mosquito-Week. 2014.
11. A. Gelman and D. B. Rubin. “Inference from iterative simulation using multiple sequences”. In: *Statistical Science* (1992), pp. 457–472.
12. M. Gillies and T. Wilkes. “A study of the age-composition of populations of *Anopheles gambiae* Giles and A. funestus Giles in North-Eastern Tanzania”. *Bulletin of Entomological Research* 56.02 (1965), pp. 237–262.
13. C. A. Guerra et al. “A global assembly of adult female mosquito mark-release-recapture data to inform the control of mosquito-borne pathogens”. *Parasite & Vectors* 7.1 (2014), p. 276.
14. L. C. Harrington et al. “Age-dependent survival of the dengue vector *Aedes aegypti* (Diptera: Culicidae) demonstrated by simultaneous release–recapture of diﬀerent age cohorts”. *Journal of Medical Entomology* 45.2 (2008), pp. 307–313.
15. T. Hoc and T. Wilkes. “The ovariole structure of *Anopheles gambiae* (Diptera: Culicidae) and its use in determining physiological age”. *Bulletin of Entomological Research* 85.01 (1995), pp. 59–69.
16. L. E. Hugo et al. “Adult survivorship of the dengue mosquito *Aedes aegypti* varies seasonally in central Vietnam”. *PLoS Neglected Tropical Diseases* 8.2 (2014), e2669.
17. L. E. Hugo et al. “Evaluations of mosquito age grading techniques based on morphological changes”. *Journal of Medical Entomology* 45.3 (2008), 353–369.
18. B. Kay. “Age structure of populations of *Culex annulirostris* (Diptera: Culicidae) at Kowanyama and Charleville, Queensland”. *Journal of* *Medical Entomology* 16.4 (1979), pp. 309–316.
19. B. Lambert. *A Student’s Guide to Bayesian Statistics*. Sage, 2018.
20. B. Lambert et al. “Monitoring the Age of Mosquito Populations Using Near-Infrared Spectroscopy”. *Scientific reports* 8.1 (2018), p. 5274.
21. A. Lange and T. Hoc. *Abortive oogenesis and physiological age in blood-sucking mosquitoes (Diptera: Culicidae). Meditsinskaya Parazitologiya i Parasitarnye Bolezni 50, 48–56*. 1981.
22. G. Macdonald et al. *The epidemiology and control of malaria.* London, Oxford University Press, 1957.
23. V. S. Mayagaya et al. “Non-destructive determination of age and species of *Anopheles gambiae* sl using near-infrared spectroscopy”. *The American Journal of Tropical Medicine and Hygiene* 81.4 (2009), pp. 622–630.
24. G. Müller, A Junnila, and Y Schlein. “Eﬀective control of adult *Culex* *pipiens* by spraying an attractive toxic sugar bait solution in the vegetationnear larval habitats”. *Journal of Medical Entomology* 47.1 (2010), 63–66.
25. G. C. Müller, V. D. Kravchenko, and Y. Schlein. “Decline of Anopheles sergentii and Aedes caspius populations following presentation of attractive toxic (spinosad) sugar bait stations in an oasis”. *Journal of the American* *Mosquito Control Association* 24.1 (2008), pp. 147–149.
26. G. C. Müller et al. “Field experiments of Anopheles gambiae attraction to local fruits/seedpods and flowering plants in Mali to optimize strategies for malaria vector control in Africa using attractive toxic sugar bait methods”. *Malaria journal* 9.1 (2010), p. 262.
27. G. C. Müller et al. “Successful field trial of attractive toxic sugar bait (ATSB) plant-spraying methods against malaria vectors in the *Anopheles* *gambiae* complex in Mali, West Africa”. In: *Malaria Journal* 9.1 (2010), 210.
28. C. Murdock et al. “Complex eﬀects of temperature on mosquito immune function”. *Proc. R. Soc. B* (2012), rspb20120638.
29. J. Nedelman. “A negative binomial model for sampling mosquitoes in a malaria survey”. *Biometrics* (1983), pp. 1009–1020.
30. V. Polovodova. “The determination of the physiological age of female *Anopheles* by the number of gonotrophic cycles completed”. *Meditsin-skaia Parazitologiia Parazitar Bolezni* 18 (1949), pp. 352–355.
31. R. C. Russell. “Population age composition and female longevity of the arbovirus vector *Culex annulirostris skuse* near Echua, Victoria, in the Murray Valley of southeastern Austria 1979-1985”. *Australian Journal* *of Experimental Biology & Medical Science* 64.6 (1986).
32. M. Service. “Studies on sampling larval populations of the *Anopheles* *gambiae* complex”. In: *Bulletin of the World Health Organisation* 45.2(1971), p. 169.
33. M. Sikulu et al. “Evaluating RNAlater® as a preservative for using near-infrared spectroscopy to predict *Anopheles gambiae* age and species”. *Malaria Journal* 10.1 (2011), p. 186.
34. J. B. Silver. *Mosquito ecology: field sampling methods*. Springer Science & Business Media, 2007.
35. M. E. Sinka et al. “The dominant *Anopheles* vectors of human malaria in Africa, Europe and the Middle East: occurrence data, distribution maps and bionomic précis”. *Parasites & Vectors* 3.1 (2010), p. 117.
36. L. M. Styer et al. “Mosquitoes do senesce: departure from the paradigm of constant mortality”.*The American Journal of Tropical Medicine and* *Hygiene* 76.1 (2007), pp. 111–117.
37. N. O. Verhulst, J. A. Loonen, and W. Takken. “Advances in methods for colour marking of mosquitoes”. *Parasites & Vectors* 6.1 (2013), p. 1.
38. World Health Organisation et al. “WHO statement on the first meeting of the International Health Regulations (2005) - Emergency Committee on Zika virus and observed increase in neurological disorders and neonatal malformations”. In: 37.3 (2016), pp. 332–333.
39. World Health Organization et al. “Global report on insecticide resistance in malaria vectors: 2010–2016”. WHO (2018).
40. H. Yang et al. “Assessing the eﬀects of temperature on the population of *Aedes aegypti*, the vector of dengue”. *Epidemiology and Infection* 137.08 (2009), pp. 1188–1202.