# **Technical Report**

This is a technical report about the analyses performed as part of the African genomic diversity based of the workplan in Fig 1.

#### Initial workplan prep Generate a text file with individual and population ID mappings. Remove variants with < 6X coverage.</li> - Annotate ancestral allele for each variant and remove variants with unknown ancestral allele. Data - Annotate VCF with GERP scores and SNPEff. - Calculate derived allele frequencies per population - Construct SFS (site frequency spectrum) per population. - Extract number of singletons per population. Extract number of novel For each pair-wise population comparison, calculate HDVs. variants per population. Proportion. of functional SNPs Extract singletons and in singletons vs. doubletons per pop. in other variants Calculate mean freq. of novel If possible, perform variants per pop and Validate with sampling Determine sharing GO analysis. compare across populations. with replacement. across pops

Fig 1. Initial workplan proposed by Laura.

Rare variants analysis

# Data preparation

Data to be used by our stream consist of Baylor data only. This dataset was recalled together with sanger and trypanogen, and the phased using Eagle.

Novel variants analysis

Shared variants analysis

The files shared with the genomic diversity analysis group were per chromosome (1 to 22) in VCF format as Eagle.baylor.\${CHRM}.vcf.annoted.vcf.hg19\_multianno.vcf.gz located in /popdata/gapw/GAPW\_DATA/baylor/PHASED/

## Generate a text file with individual and population IDs

- Input: /popdata/gapw/GAPW\_DATA/baylor/UNPHASED/BAYLOR\_UNPHASED.sample
- Output: /spaces/gapw/diversity/mamana/baylor\_sample\_info\_ind\_pop\_id\_only.tsv
- · Command:

```
> awk '{print $1"\t"$2}' ${input} > ${ouput}
```

# Remove variants with < 6X coverage

- Input: /spaces/gapw/diversity/mamana/VCF/Eagle.baylor.\$ {CHRM}.vcf.annoted.vcf.hg19\_multianno.vcf.gz
- Output: /spaces/gapw/diversity/mamana/VCF\_FILTERED/baylor\_phased\_chr\${CHRM}
   \_dp6.vcf.gz
- Command:

```
> bcftools view -i 'DP>6' ${input} | bgzip -c > ${output}
```

# Annotate ancestral allele for each variant and remove variants with unknown ancestral allele

- Input: /spaces/gapw/diversity/mamana/VCF\_FILTERED/baylor\_phased\_chr\${CHRM}
   \_dp6.vcf.gz
- Output: /spaces/gapw/diversity/mamana/VCF\_FILTERED/baylor\_phased\_chr\${CHRM}
   \_dp6\_anc\_f.vcf.gz
- Command:
  - Annotate the AA using house-made python script from Laura:

```
> ../scripts/add-ANC-to-vcf_new.py -g --in {input} --out
{output=output1} --genomedata genomedata_path
```

Filter for sites with AA only:

```
> bcftools view -i 'AA!="." & AA!="-" & AA!
="N"' {input=output1} | bgzip -c > {output}
```

# Annotate VCF with GREP scores and snpEff

GREP scores where not annotate for anymore as there were already annotated for by Emile. We only annotated using snpEff and using snpSift for dbSNP IDs.

- Input: /spaces/gapw/diversity/mamana/VCF\_FILTERED/baylor\_phased\_chr\${CHRM}
   \_dp6\_anc\_f.vcf.gz
- Output: /spaces/gapw/diversity/mamana/VCF\_ANN/baylor\_phased\_chr\${CHRM}
   \_dp6\_anc\_f\_dbsnp.vcf.gz and /spaces/gapw/diversity/mamana/VCF\_ANN/baylor\_phased\_chr\$
   {CHRM}\_dp6\_anc\_f\_dbsnp\_snpeff.vcf.gz
- Command:
  - Annotate for dbSNP IDs using snpSift:

```
> snpSift annotate {dbsnp db} {input} > {output=output1} -v
```

Annotate using snfEff:

```
> snpEff -v {human_db} -stats {output}.html -csvStats
{output}.csv -dataDir {snpEff_database} {input=output1} >
{output=output2}
```

- dbSNP database (dbsnp\_db): dbSNP\_human\_9606\_b150\_GRCh37p13.vcf
- human db: hq19
- snpEff\_database: location of your snpEff database if not in default snpEff path

### Calculate derived allele frequencies per population

1. Split sample file in population

- Input: /spaces/gapw/diversity/mamana/baylor sample info ind pop id only.tsv
- Output (pop\_file): ../samples/{POP}.sample, for POP in Benini, Botswana, Burkina, Cameroon, Ghana, Mali, Nigeria, Zambia
- Command:

```
def split sample list per pop(POP, input file, output file):
     Read baylor sample info ind pop id only.tsv and
     split it by population
     i = 1 #line no
     data = []
     message("Extracting "+POP+' to '+output file)
     for line in open (input file):
           if i > 1:
                line = line.strip().split()
                POP = line[1].strip()
                if POP == POP:
                     data.append('\t'.join(line)+'\n')
           i += 1
     if len(data) > 0:
           out = open(output_file, "w")
           for elt in data:
                out.writelines(elt)
           out.close()
```

#### 2. Split into population

- Input: /spaces/gapw/diversity/mamana/VCF\_ANN/baylor\_phased\_chr\${CHRM}
   \_dp6\_anc\_f\_dbsnp\_snpeff.vcf.gz
- Output: /spaces/gapw/diversity/mamana/VCF\_POP/{POP}\_phased\_chr\${CHRM}
   \_dp6\_anc\_f\_dbsnp\_snpeff.vcf.gz
- Command:

```
> vcftools \
    --gzvcf {input} \
    --keep {pop_file} \
    --recode --recode-INFO-all --mac 0 -c | \
    bgzip -c > {output}
```

## 3. Calculate derived allele frequencies

- Input: /spaces/gapw/diversity/mamana/VCF\_POP/{POP}\_phased\_chr\${CHRM}
   \_dp6\_anc\_f\_dbsnp\_snpeff.vcf.gz
- Output: /spaces/gapw/diversity/mamana/VCF\_POP/{POP}\_phased\_chr\${CHRM}
   \_dp6\_anc\_f\_dbsnp\_snpeff.daf.frq
- Command:

```
> vcftools \
    --gzvcf {input} \
    --freq --derived \
    --out {output}
```