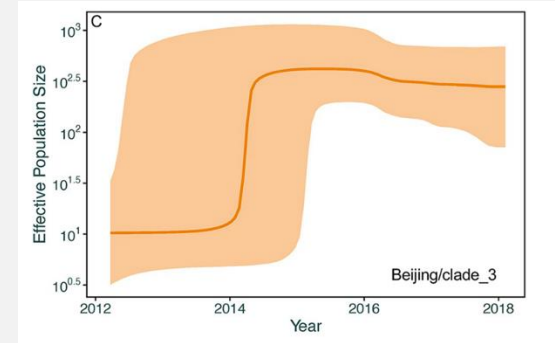
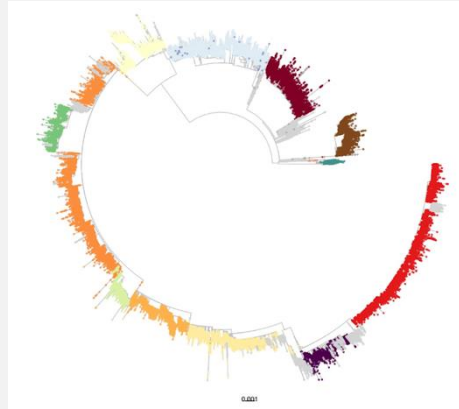


GENOMIC ANALYSIS AND PHYLODYNAMICS

Lecture 2: Variant Calling and Phylogenetic Trees



Instructor: Dr. Ben Sobkowiak

MRC Senior Research Fellow, University College London

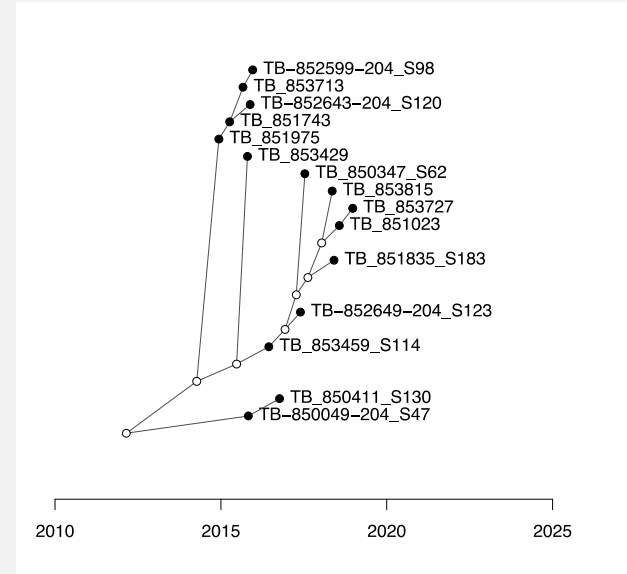
LECTURE 2: VARIANT CALLING AND PHYLOGENETIC TREES

- What can variation within the genome tell us?
- What are genomic variants?
- How do we detect variation in the genome?
- Linking variation to evolutionary relationships using phylogenetic trees

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What can variation in the genome tell us?

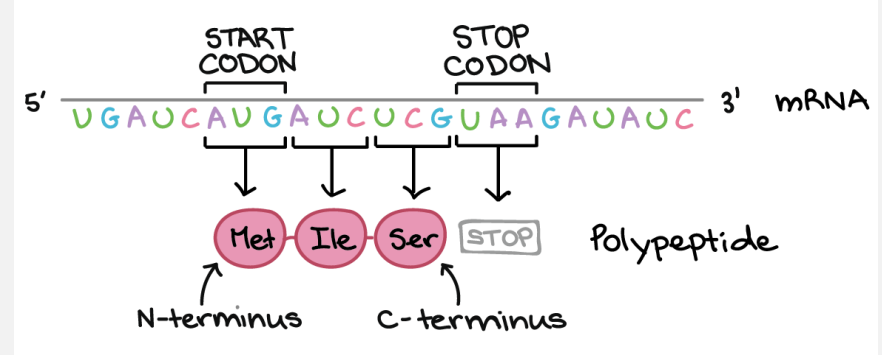
- Insights into the evolution of organisms and selection
 - Variations in the genome contribute to the adaptation of species to their environment
- Specific mutations may alter the protein that is coded for by a gene and change characteristics
- Analysing the amount and patterns of differences can relate to the amount of divergence and common ancestry between individuals
 - Can be linked to evolutionary history and transmission



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How the genome is read

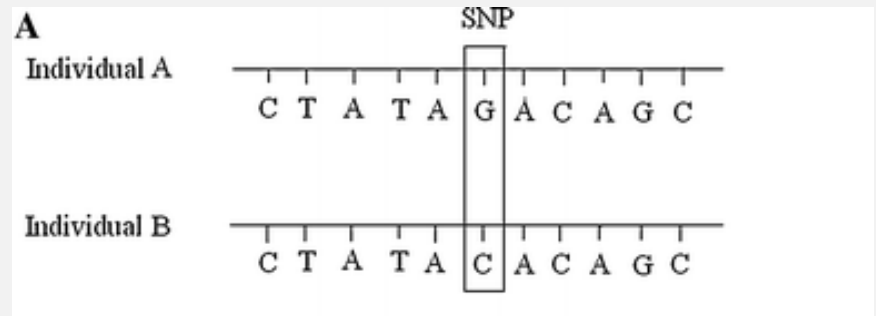
- Process of reading the genome is transcription
- RNA polymerase binds to promoters (upstream of a gene) and then 'reads' the genetic code to make a complimentary RNA strand
- This RNA strand is then translated into proteins by building amino acid blocks coded for by the sequence
- A specific sequence of three nucleotide bases (codon) encodes an amino acid



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Single nucleotide polymorphisms

- Single Nucleotide Polymorphisms, or SNPs, are the most common type of genetic variation
- A point mutation where a single nucleotide base (A, C, G, or T) in the DNA sequence is replaced by one of the other three bases at the same position
- Most common form of genomic variation



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Single nucleotide polymorphisms

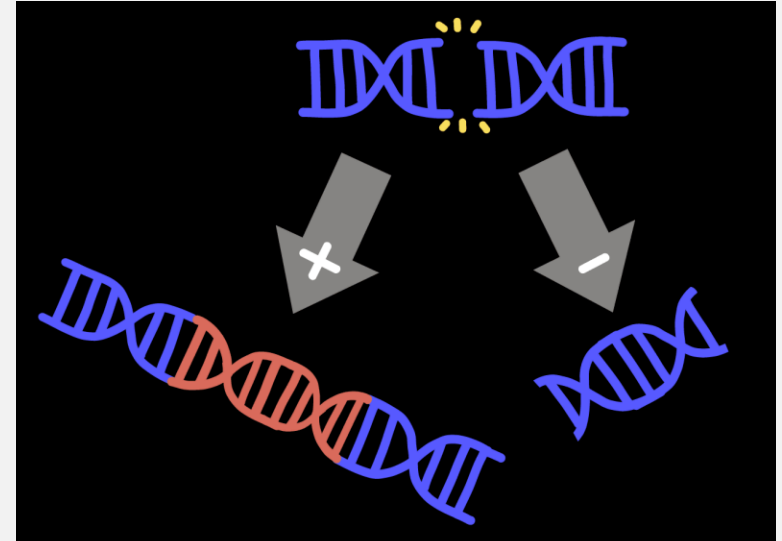
- SNPs can result in a change in the amino acid that is encoded (non-synonymous) or still code for the same amino acid (synonymous)
- There are 64 codons but only 20 amino acids (+ start and stop codons)

		Second Letter									
		U		C		A		G			
1st letter	U	UUU	Phe	UCU	Ser	UAU	Tyr	UGU	Cys	U	3rd letter
	UUC		UCC			UAC		UGC		C	
	UUA	Leu	UCA			UAA	Stop	UGA	Stop	A	
	UUG		UCG			UAG	Stop	UGG	Trp	G	
1st letter	C	CUU	Leu	CCU	Pro	CAU	His	CGU	Arg	U	
	CUC	CCC		CAC		CGC	C				
	CUA	CCA		CAA		Gln	CGA	A			
	CUG	CCG		CAG		CGG	G				
1st letter	A	AUU	Ile	ACU	Thr	AAU	Asn	AGU	Ser	U	
	AUC	ACC		AAC		AGC	C				
	AUA	ACA		AAA		Lys	AGA	A			
	AUG	ACG		AAG		AGG	Arg	G			
1st letter	G	GUU	Val	GCU	Ala	GAU	Asp	GGU	Gly	U	
	GUC	GCC		GAC			GGC			C	
	GUA	GCA		GAA		Glu	GGA			A	
	GUG	GCG		GAG			GGG			G	

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Insertions and deletions (Indels)

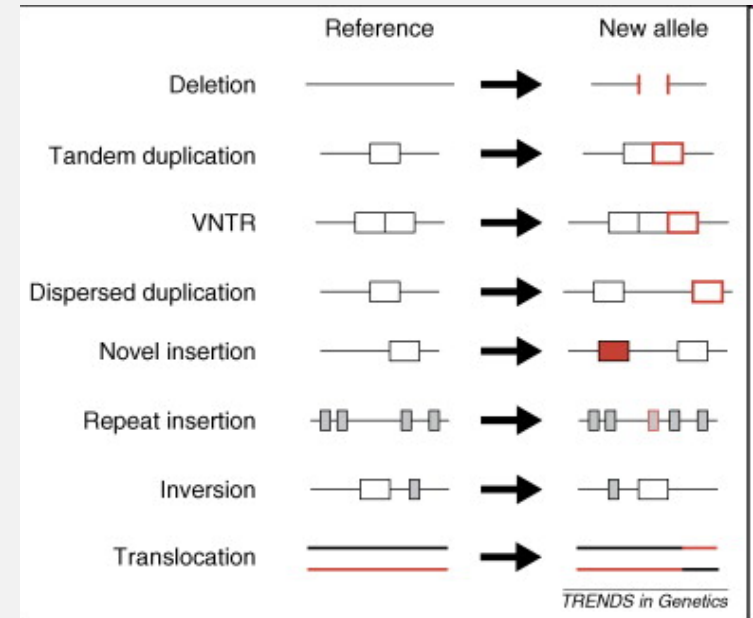
- Often abbreviated as Indels
- The addition or removal of one or more nucleotide bases
- Can have significant effects on the structure and function of genes and can contribute to genetic diversity within populations
- Can have most impact when they are 'frameshift' indels – changes the codon position



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Other genomic variants

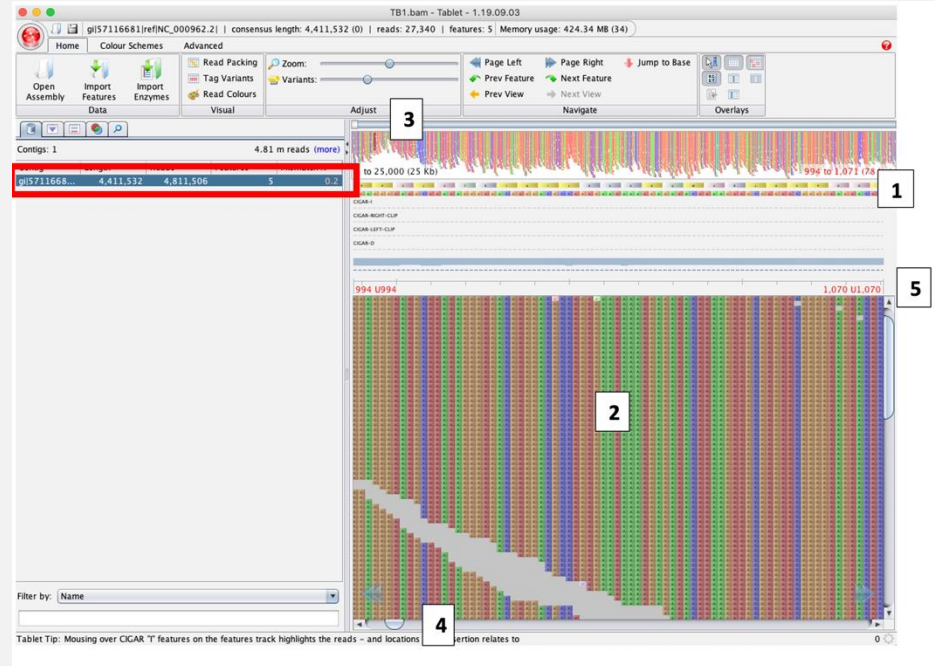
- Gene duplications
- Inversions
- Translocations
- Rarer, and more difficult to detect and analyze



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Assembled/aligned genomes

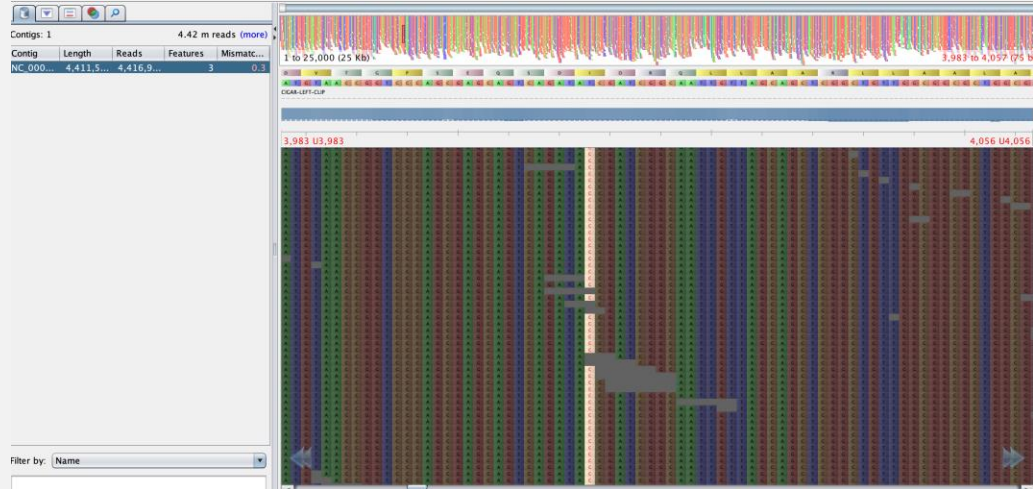
- We now have our assembled or aligned genome
- The file format is called a BAM (or SAM) file
 - A TAB-delimited text format consisting of an optional header and an alignment.
- Minimum format agreed on to report sequencing results, and includes all the data in a *fastq* file



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Assembled/aligned genomes

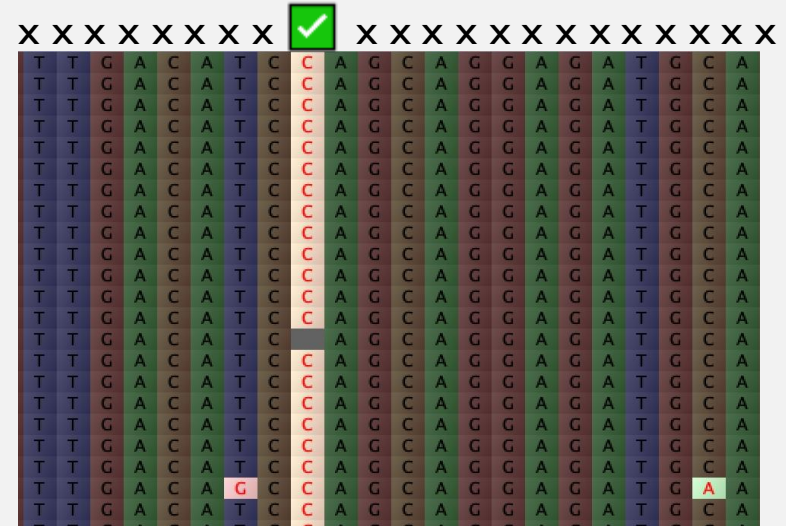
- Could scroll through the alignment file to detect variation
- But, time-consuming and subjective – how do we decide between variation and error?
- We can use tools to read through the BAM file and identify true variation



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Identifying variants

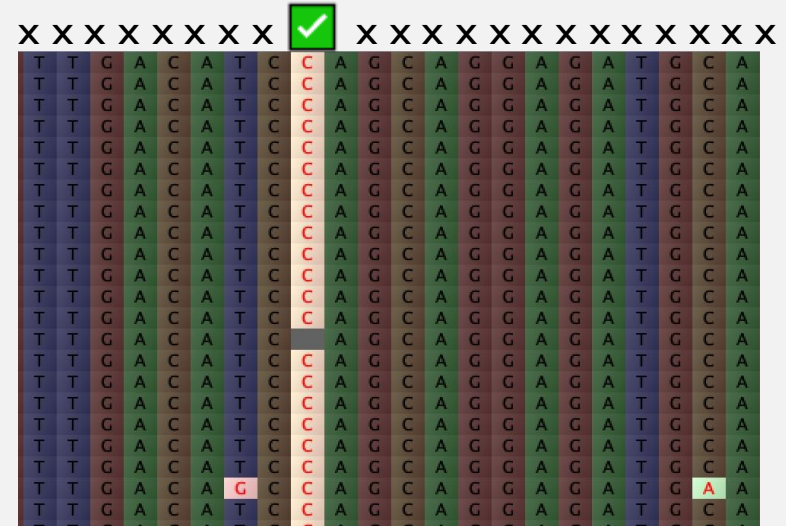
- We use modelling approaches to identify variants between the reference and sequences of samples, and between samples.
- Need to maximize sensitivity/specificity of calls by accounting for sequence error, poor alignment, low quality etc.



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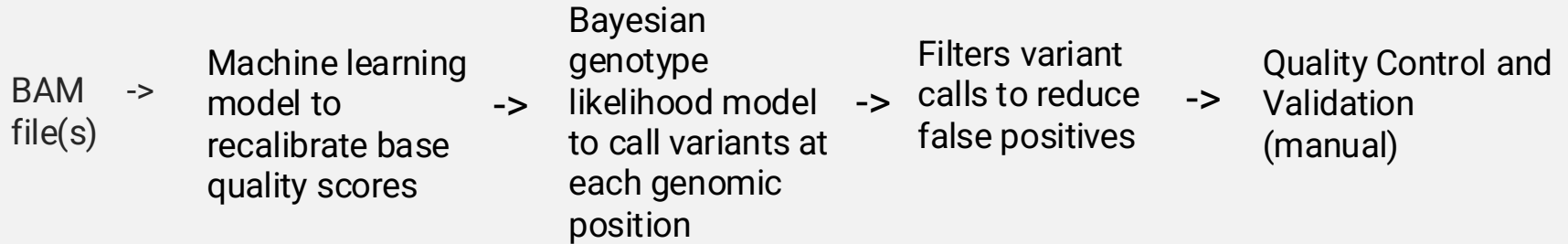
Variant calling software

- Multiple variant calling software (for short-read data) are available including:
 - GATK
 - SAM/BCFtools
 - Delly
 - Pilon
 - FreeBayes
 - DeepVariant
- Have different strengths depending on the type of variants to call



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GATK variant calling



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Variant Call Format (VCF) files

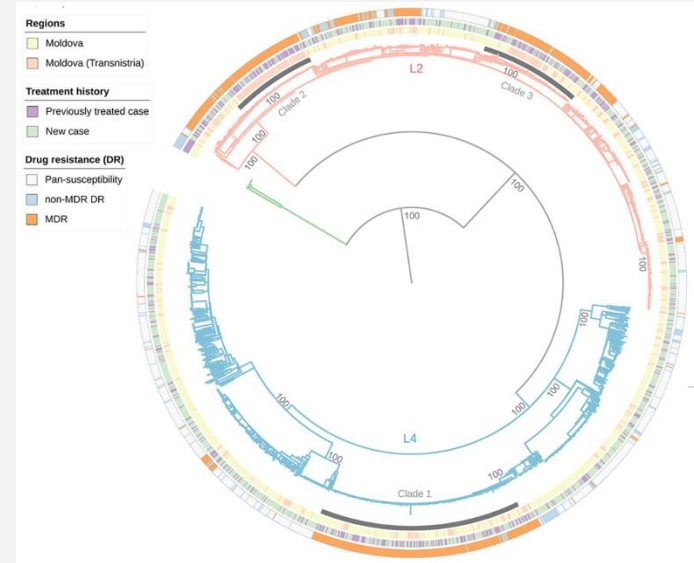
- The Variant Call Format (VCF) is a widely used file format in genomics for storing information about genetic variants, such as single nucleotide polymorphisms (SNPs), insertions, deletions, and other types of genetic variation.
- Important part is the Genotype data - the alleles carried by each individual at the variant site.
 - e.g., "0/0" for homozygous reference, "0/1" for heterozygous, "1/1" for homozygous alternate

```
1 ##fileformat=VCFv4.2
2 ##FILTER=ID=PASS,Description="All filters passed">
3 ##bcftoolsVersion=1.10.2+htslib-1.10.2
4 ##bcftoolsCommand=pileup -b -Q 20 -d 500 -C 50 -Ou -o DP,AD -f Data/H37Rv.fasta Data/TB1.bam Data/TB2.bam
5 ##reference=1/ncgi/57116681/ref/NC_000962.2
6 ##contig=ID=gl|57116681|ref/NC_000962.2,length=4411532>
7 ##ALT=ID=,Description="Represents allele(s) other than observed.">
8 ##INFO=ID=INDEL,Number=0,Type=Flag,Description="Indicates that the variant is an INDEL.">
9 ##INFO=ID=INDV,Number=1,Type=Integer,Description="Maximum number of raw reads supporting an indel">
10 ##INFO=ID=INF,Number=1,Type=Float,Description="Maximum fraction of raw reads supporting an indel">
11 ##INFO=ID=DP,Number=1,Type=Integer,Description="Raw read depth">
12 ##INFO=ID=VD,Number=1,Type=Float,Description="Variant Distance Bias for filtering splice-site artefacts in RNA-seq data (bigger is better)",Version=
13 ##INFO=ID=RPB,Number=1,Type=Float,Description="Mann-Whitney U test of Read Position Bias (bigger is better)">
14 ##INFO=ID=MQB,Number=1,Type=Float,Description="Mann-Whitney U test of Mapping Quality Bias (bigger is better)">
15 ##INFO=ID=QOB,Number=1,Type=Float,Description="Mann-Whitney U test of Base Quality Bias (bigger is better)">
16 ##INFO=ID=MQSB,Number=1,Type=Float,Description="Mann-Whitney U test of Mapping Quality vs Strand Bias (bigger is better)">
17 ##INFO=ID=SQB,Number=1,Type=Float,Description="Segregation based metric">
18 ##INFO=ID=MQF,Number=1,Type=Float,Description="Fraction of MQB reads (smaller is better)">
19 ##FORMAT=ID=PL,Number=6,Type=Integer,Description="List of Phred-scaled genotype likelihoods">
20 ##FORMAT=ID=DP,Number=1,Type=Integer,Description="Number of high-quality bases">
21 ##FORMAT=ID=AD,Number=4,Type=Integer,Description="Allele depths (high-quality bases)">
22 ##FORMAT=ID=GT,Number=1,Type=String,Description="Genotype">
23 ##INFO=ID=ICB,Number=1,Type=Float,Description="Inbreeding Coefficient Binomial test (bigger is better)">
24 ##INFO=ID=HQB,Number=1,Type=Float,Description="Bias in the number of HQBs number (smaller is better)">
25 ##INFO=ID=AC,Number=A,Type=Integer,Description="Allele count in genotypes for each ALT allele, in the same order as listed">
26 ##INFO=ID=AN,Number=1,Type=Integer,Description="Total number of alleles in called genotypes">
27 ##INFO=ID=QA,Number=4,Type=Integer,Description="Number of high-quality ref-forward, ref-reverse, alt-forward and alt-reverse bases">
28 ##INFO=ID=MQ,Number=1,Type=Integer,Description="Average mapping quality">
29 ##bcftools_callVersion=1.10.2+htslib-1.10.2
30 ##bcftools_callCommand=call -mv -Ou Date=Thu Mar 5 15:28:59 2020
31 ##bcftools_viewVersion=1.10.2+htslib-1.10.2
32 ##bcftools_viewCommand=view Data/TB-bcf; Date=Thu Mar 5 15:34:29 2020
33 #CHROM POS ID REF ALT QUAL FILTER INFO FORMAT Data/TB1.bam Data/TB2.bam
34 gl|57116681|ref/NC_000962.2| 1849 . C A 483 . DP=188;VDB=0.620126;SGB=-1.38612;MQSB=0.42763;MQOF=0;AC=4;AN=4;DP4=0,0,104,81;MQ45 G
35 gl|57116681|ref/NC_000962.2| 1977 . A G 483 . DP=252;VDB=0.227948;SGB=-1.38629;MQSB=0.9956;MQOF=0;AC=4;AN=4;DP4=0,0,96,79;MQ45 GT:PL
36 gl|57116681|ref/NC_000962.2| 4013 . T C 483 . DP=380;VDB=0.917995;SGB=-1.38629;MQSB=0.697421;MQOF=0;AC=4;AN=4;DP4=0,0,148,113;MQ45 G
37 gl|57116681|ref/NC_000962.2| 7362 . G C 483 . DP=276;VDB=0.963713;SGB=-1.38629;MQSB=0.98887;MQOF=0;AC=4;AN=4;DP4=0,0,127,125;MQ45 G
38 gl|57116681|ref/NC_000962.2| 7585 . G C 483 . DP=265;VDB=0.893931;SGB=-1.38629;MQSB=0.813234;MQOF=0;AC=4;AN=4;DP4=0,0,109,121;MQ45 G
39 gl|57116681|ref/NC_000962.2| 9304 . G A 483 . DP=271;VDB=0.722358;SGB=-1.38629;RPB=1;MQSB=0.7223;MQOF=0;AC=4;AN=4;DP4=0,0,148,113;MQ45 G
40 gl|57116681|ref/NC_000962.2| 11879 . A G 483 . DP=283;VDB=0.227714;SGB=-1.38629;MQSB=0.88965;MQOF=0;AC=4;AN=4;DP4=0,0,141,106;MQ41 G
41 gl|57116681|ref/NC_000962.2| 14785 . T C 483 . DP=382;VDB=0.582188;SGB=-1.38629;MQSB=0.329038;MQOF=0;AC=4;AN=4;DP4=0,0,157,116;MQ42 G
42 gl|57116681|ref/NC_000962.2| 14861 . G T 483 . DP=319;VDB=0.277102;SGB=-1.38629;MQSB=0.880851;MQOF=0;AC=4;AN=4;DP4=0,0,167,122;MQ42 G
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LECTURE 2: VARIANT CALLING AND PHYLOGENETIC TREES

Phylogenetic trees

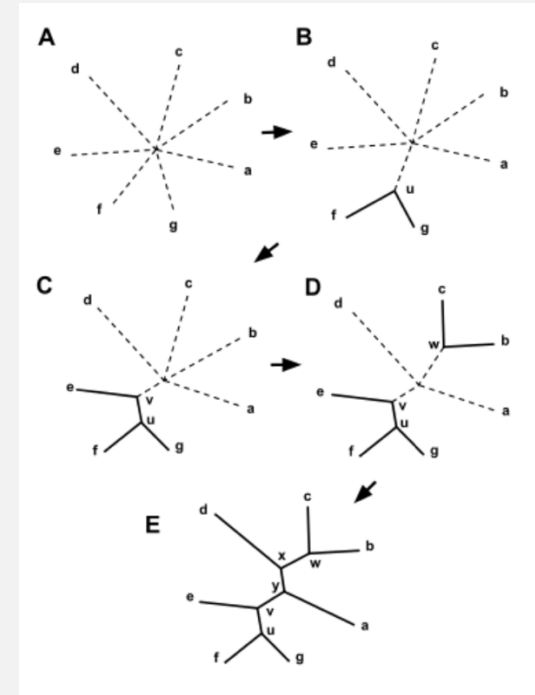
- A phylogenetic tree is a diagram that represents evolutionary relationships among organisms
- They illustrate the ancestral lineage and divergence of species, genes, or other taxonomic units.
- These trees help in understanding evolutionary history, inferring patterns of descent, and clarifying the timing of evolutionary events.



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Simple methods (Neighbour-Joining)

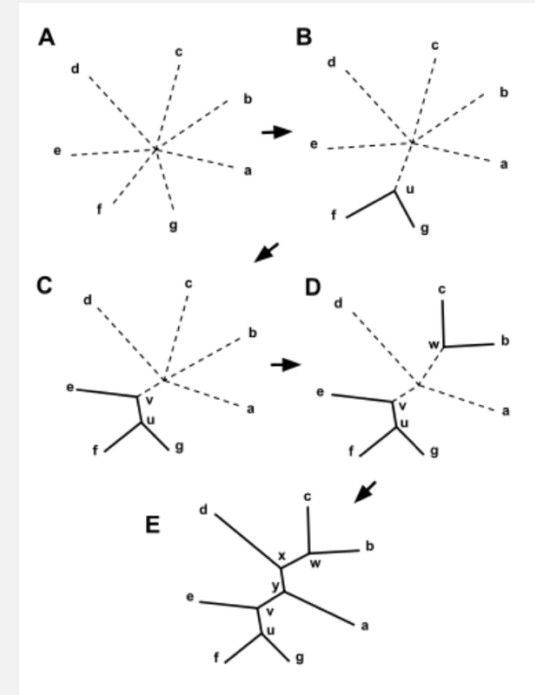
- Neighbor-Joining (NJ) is a distance-based method used to construct phylogenetic trees
- Starts with a star-like tree, where all entities are connected to a central node (A)
- The Neighbor-Joining algorithm iteratively joins entities (nodes) in the tree while minimizing the total branch length



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Simple methods (Neighbour-Joining)

- Neighbor-Joining is a relatively efficient and versatile method for constructing phylogenetic trees
- However, it does not explicitly model the underlying evolutionary processes, such as substitutions, insertions, deletions, or other events
- This limitation makes it less suitable for analyzing complex evolutionary scenarios



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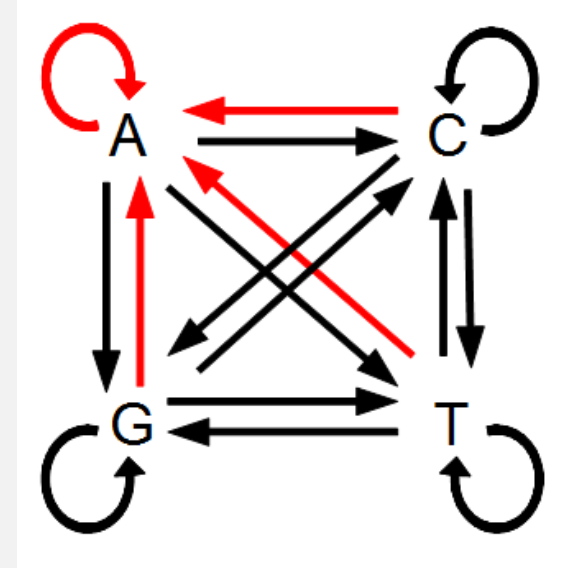
Evolutionary models

- More sophisticated tree building methods can include models:
 - Nucleotide substitution models
 - Molecular clock models
 - Population models
 - Coalescent models

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Nucleotide substitution models

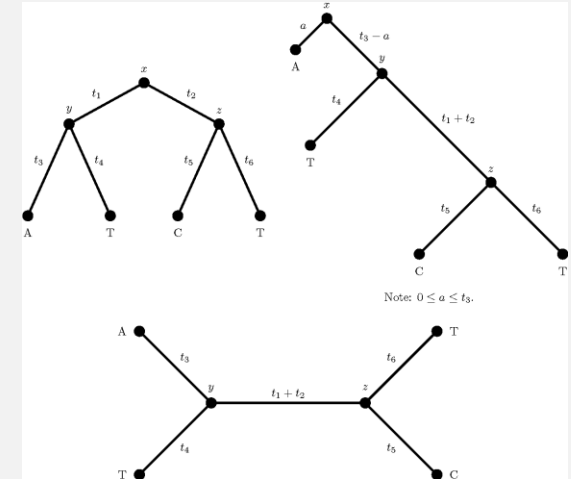
- Describe the rates at which nucleotides in DNA sequences change over time
- Provide a framework for estimating the likelihood of observed DNA sequence data on a phylogenetic tree
- Common models include:
 - Jukes-Cantor (JC) Model
 - Hasegawa-Kishino-Yano (HKY) Model
 - General Time Reversible (GTR) Model



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More complex methods (Maximum Likelihood)

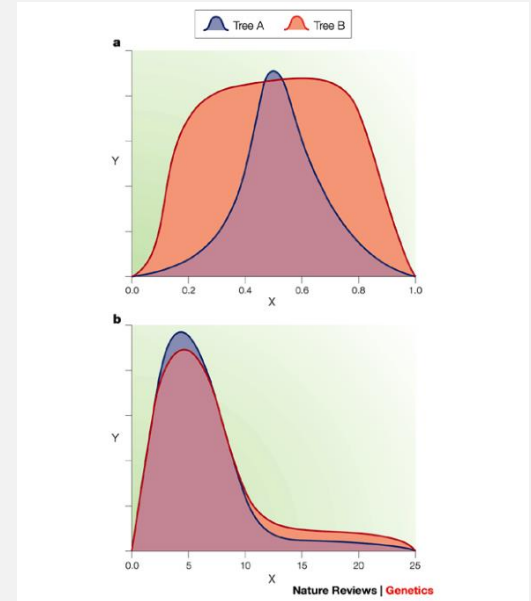
- ML tree construction aims to find the tree topology and branch lengths that maximize the likelihood of the observed sequence data under a specified evolutionary model
- ML is powerful for its statistical rigor and ability to handle complex models of evolution
- Generally considered less flexible in handling complex models compared to Bayesian methods



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Bayesian phylogenies

- Uses Bayesian statistics to estimate the posterior probabilities of different phylogenetic trees, incorporating the observed data and prior knowledge
- Calculates the probabilities of different trees by combining the likelihood of the observed data with the prior probability distributions over tree space, based on certain models of evolution
- Require careful selection of priors and are computationally demanding.

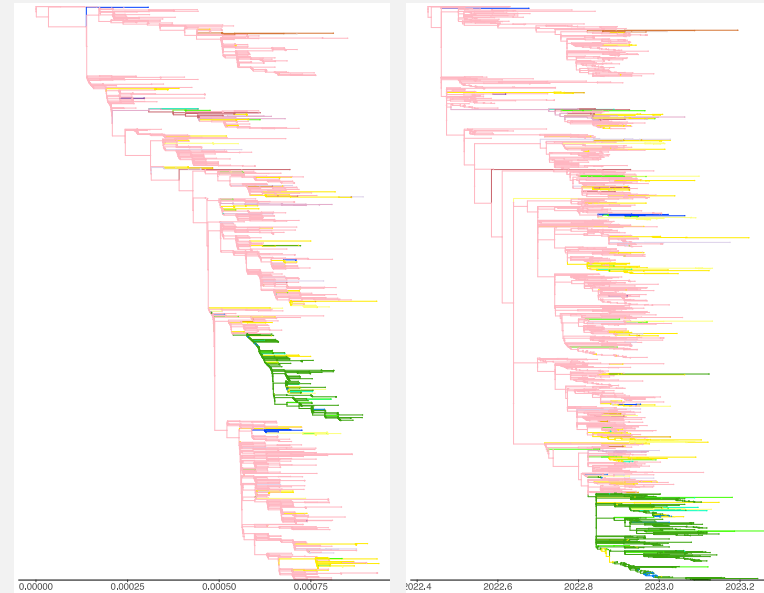


From Holder & Lewis, 2003

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Timed vs untimed trees

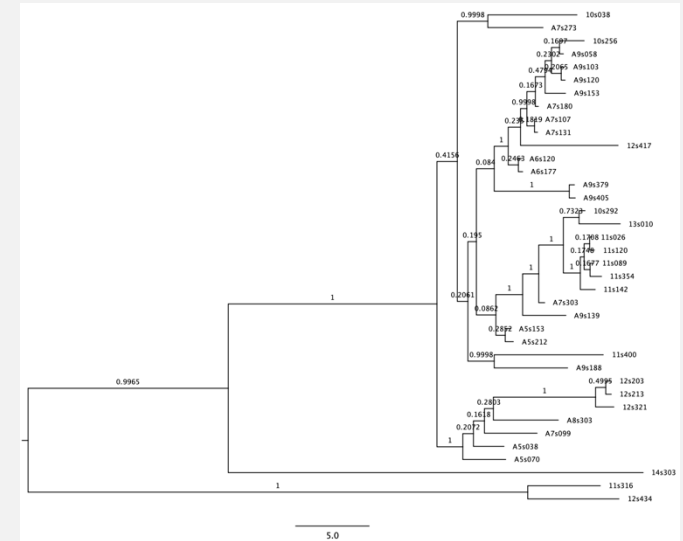
- In untimed trees, branch lengths typically represent genetic or sequence divergence
- Help to understand the evolutionary ancestry and genetic relatedness among taxa
- Timed trees incorporate the estimated timing of evolutionary events, branches are scaled to unit time
- Useful for estimating divergence times, studying temporal changes in evolutionary processes, and reconstructing the evolutionary history of lineages



LECTURE 2: VARIANT CALLING AND PHYLOGENETIC TREES

Assessing phylogenetic trees

- Bootstrapping is a resampling technique used in phylogenetics to assess the robustness of the inferred phylogenetic tree topology
- It can estimate the reliability of the branching patterns in a phylogenetic tree
- Resamples data and builds multiple trees to assign support values to branches and nodes
- High bootstrap support values indicate the inferred relationships are likely to be accurate



PRACTICAL 2: VARIANT CALLING AND MAXIMUM LIKELIHOOD TREES

1. Variant calling and VCFs
2. Building consensus sequences
3. Aligning consensus sequences
4. Maximum Likelihood trees