CS481/CS583: Bioinformatics Algorithms

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KMER DATA STRUCTURES

Baseline problem

In-memory representation of a large set of short k-mers:

e.g.

ACTGAT

GTATGC

ATTAAA

GAATTG

. . .

Applications

- Genome assembly
- Error-correction of DNA sequencing data
- Detection of similarity between sequences
- Detection of distances between datasets
- Alignment
- Pseudoalignment / quasi-mapping
- Detection of taxonomy
- Indexing large collections of sequencing datasets
- Quality control
- Detection of events (e.g., SNPs, indels, CNVs, alt. transcription)
- ...

k-mers

Sequences of k consecutive letters, e.g. ACAG or TAGG for k=4

N.G. de Bruijn (1946), de Bruijn sequences ¹



C. Shannon (1948), information theory ²



Framing the problem

Large set of k-mers: 10⁶ - 10¹¹ elements

k in [11; 10³]

Operations to support

- Construction (from a disk or stream)
- Membership ("is X in the set?")
- **Iteration** (enumerate all elements in the set)

- ...

Extensions:

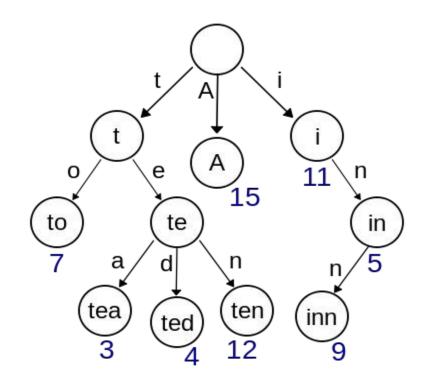
- Associate value(s) to k-mers (e.g. abundance)
- Navigate the de Bruijn graph

Membership test: Tries

Worst case

- O(k) insertion
- O(k) deletion
- O(k) search

Also supports indexing

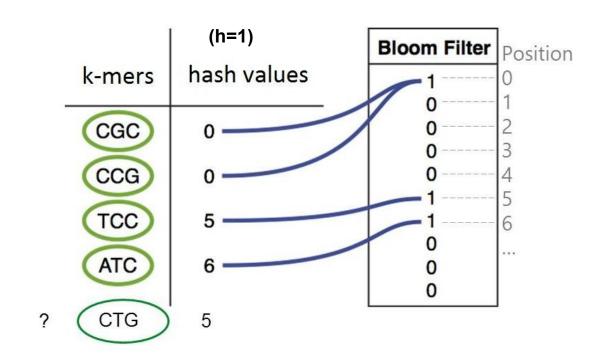


Membership test: Bloom filters

Init a bit array
Take h hash
functions
Insertion: put 1's at
positions given by
hash functions

Query: are there 1's at all positions given by hash functions?

- O(hk) insertion
- (O(hk) deletion)
- O(hk) query

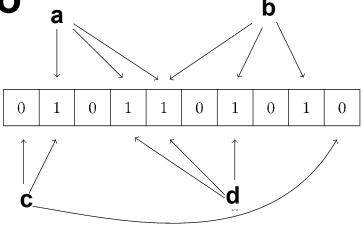


Bloom filter

- Bloom filter encodes a set of k-mers
- Uses a bit array B of length m and d hash functions
 - \Box to insert x, we set B[h_i(x)] = 1, for i=1,...,d
 - to query y, we check if B[h_i(y)] all equal 1, for i=1,...,d
- Need an estimate for n, the number of k-mers to insert

Bloom filter example

- a and b are inserted in to a Bloom filter with m = 10, n=2, d=3
- c is not in the set, since
 some bits are 0



- d has not been inserted, but is still reported in the set, a false positive
- Bloom filters have no false negatives

Bloom filter

Storing n k-mers in m bit array with d hash functions has a false positive rate of

$$\approx (1-e^{-d n/m})^d$$

- Given n and m, the optimal d is ≈m/n ln(2)
- Example m = 8n, d=5 gives 2.16% fpr
 m = 6n, d=4 gives 5.6% fpr
 m = 4n, d=3 gives 14.6% fpr
- m=8n, corresponds to storing 1 byte per k-mer

Space: $m = 1.44n \log 2(\epsilon)$ where ϵ is the false positive rate

Counting k-mers

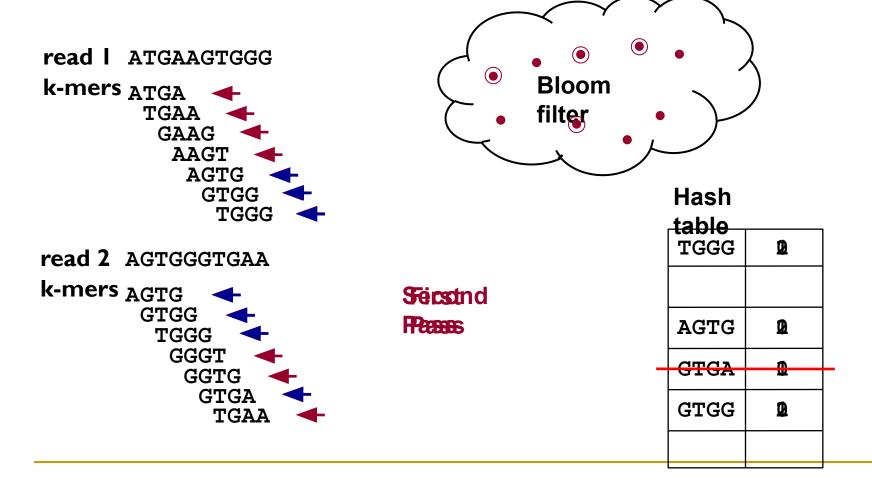
Simple method

Store each k-mer in a hash table with a counter

- Memory needed
 - store canonical k-mers
 - 2 bits for each of A,C,G,T
 - k/4 bytes per k-mer (k=31, 8 bytes)
 - 1-2 bytes per counter
 - +10% hash table overhead
- For a genome of size G, expect to see up to G distinct k-mers (2.5-3 billion for Human)
- ~ 36 Gb of memory

Memory-efficient k-mer counting

Use a Bloom filter and a hash table



Algorithm

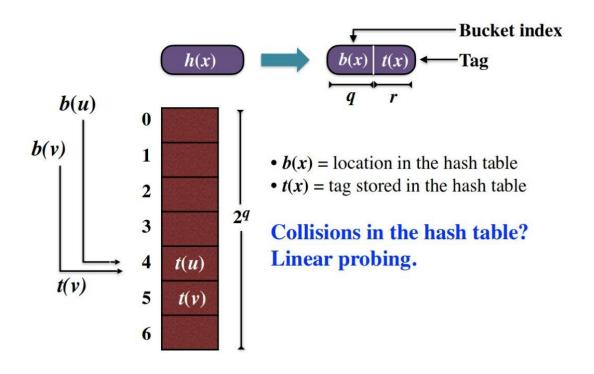
- This scheme guarantees
 - k-mers seen twice will be in the hash table
 - some unique k-mers will slip through
 - second pass gives accurate counts and allows to discard false positives
- Memory usage
 - full for k-mers in hash table (~ 9 bytes)
 - minimal for k-mers in bloom filter (~ .5-1 bytes)

Results whole genome

- 25-mers in 36 bp reads
- 2.37 billion distinct 25-mers in hg18
- 12.18 billion 25-mers in the sequencing data
 - 9.35 billion unique
 - 2.83 billion with coverage 2 or greater

Program	Time (hrs)	Memory (G)
BFCounter	23.82	42
Naïve	> 26.83	>128

Approximate membership test: Counting Quotient Filter



Hybrid between a compact hash table and a Bloom Filter.

Approximate membership

- O(k) insertion
- O(k) deletion
- O(k) query

Fig: P. Pandley, SIGMOD 2017 https://pdfs.semanticscholar.org/6bde/f4a86108309086de4071c9d28d97565a84a 4.pdf

Pandley, Bender, Johnson, Patro, *SIGMOD* 2017

Multi-set Membership Test: Sequence Bloom Trees

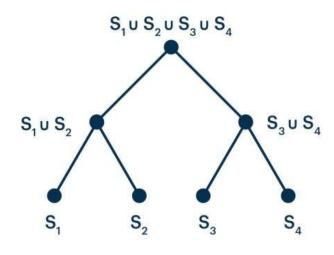


Fig: https://www.sevenbridges.com/sequence-bloom-trees-principles/

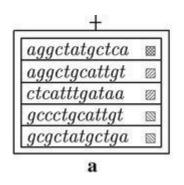
Application: fast sequence search in 1000's of RNA-seq experiments

Leaf: Bloom Filter of a sequencing dataset Internal nodes: Bitwise union of children BF's

- Representation of sets of k-mer sets
- Approximate membership across all datasets in O(hits) instead of O(datasets)
- No k-mer iteration
- Insertion/deletion of complete datasets
- Whole structure resides on disk Solomon, Kingsford, *Nat Biotech 2017* Sun, Harris, Chikhi, Medvedev, *RECOMB 2017* Solomon, Kingsford, *RECOMB 2017*

Multi-set Membership Test: Bloom Filter Tries





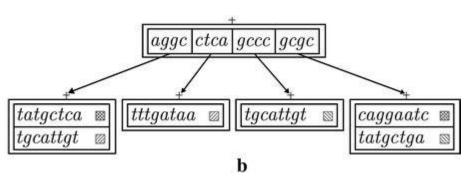


Fig:BFT article

Principle: cut k-mers into chunks, insert in a burst trie, Bloom Filters added for speed

- Representation of sets of k-mer sets
- Tailored to pan-genomes: a single k-mer belongs to many sets
- Explicit dBG operations support

Application: indexing and compression of pan-genomes

Holley, Wittler, Stoye, WABI 2016

Alternative: colored de Bruijn graphs

Slide by Rayan Chikhi

De Bruijn Graphs

- n-dimensional directed graph of m symbols
 - mⁿ vertices: all possible length-n sequences of m symbols
 - Edges between vertices v and w if sequence(w) can be generated by shifting sequence(v) by one character and add one new character
 - $S = \{s_1, s_2, ..., s_m\}$
 - $V = S^n = \{(s_1, ..., s_1, s_1), (s_1, ..., s_1, s_2), ..., (s_m, ..., s_m, s_m)\}$
 - $= \{((v_1, v_2, ..., v_n), (w_1, w_2, ..., w_n)): v_2 = w_1, v_3 = w_2, ..., v_n = w_{n-1}\}$

De Bruijn Graph for DNA Assembly

- m = 4 (A, C, G, T)
- n = k (k-mer size)
- 4^k potential vertices
 - In reality if k is sufficiently large, upper bound is genome size
 - Twin vertices: vertices with sequences that are reverse-complement of each other
 - AAAA twin of TTTT

De Bruijn Assemblers

- Currently the most common for short read seq: Euler, ALLPATHS-LG, Velvet, ABySS, SOAPdenovo
- Divide reads into k-mers
 - Build graph from k-mers
 - Put an edge if there is k-1 bp prefix-suffix match
 - Error correction
 - Eulerian path
- The first parts (graph construction & correction) is essentially common to all these assemblers, with a few implementation differences (e.g., parallelization/distributed computing in ABySS)

TAGTCGAGGCTTTAGATCCGATGAGGCTTTAGAGACAG

AGTCGAG CTTTAGA CGATGAG CTTTAGA GTCGAGG TTAGATC ATGAGGC GAGACAG GAGGCTC ATCCGAT AGGCTTT GAGACAG AGTCGAG TAGATCC ATGAGGC TAGAGAA TAGTCGA CTTTAGA CCGATGA TTAGAGA CGAGGCT AGATCCG TGAGGCT AGAGACA TAGTCGA GCTTTAG TCCGATG GCTCTAG TCGACGC GATCCGA GAGGCTT AGAGACA TAGTCGA TTAGATC GATGAGG TTTAGAG GTCGAGG TCTAGAT ATGAGGC TAGAGAC AGGCTTT ATCCGAT AGGCTTT GAGACAG AGTCGAG TTAGATT ATGAGGC AGAGACA GGCTTTA TCCGATG TTTAGAG CGAGGCT TAGATCC TGAGGCT GAGACAG AGTCGAG TTTAGATC ATGAGGC TTAGAGA GAGGCTT GATCCGA GAGGCTT GAGACAG

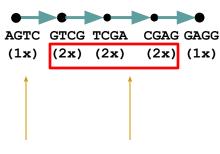
AGTCGAG CTTTAGA CGATGAG CTTTAGA GTCGAGG TTAGATC ATGAGGC GAGACAG GAGGCTC ATCCGAT AGGCTTT GAGACAG AGTCGAG TAGATCC ATGAGGC TAGAGAA TAGTCGA CTTTAGA CCGATGA TTAGAGA CGAGGCT AGATCCG TGAGGCT AGAGACA TAGTCGA GCTTTAG TCCGATG GCTCTAG TCGACGC GATCCGA GAGGCTT AGAGACA TAGTCGA TTAGATC GATGAGG TTTAGAG GTCGAGG TCTAGAT ATGAGGC TAGAGAC AGGCTTT ATCCGAT AGGCTTT GAGACAG AGTCGAG TTAGATT ATGAGGC AGAGACA GGCTTTA TCCGATG TTTAGAG CGAGGCT TAGATCC TGAGGCT GAGACAG AGTCGAG TTTAGATC ATGAGGC TTAGAGA GAGGCTT GATCCGA GAGGCTT GAGACAG

First read: GTCGAGG



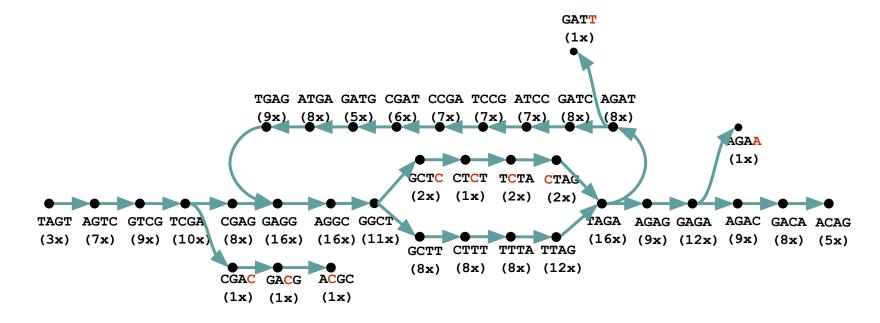
First read: GTCGAGG

Second read: AGTCGAG

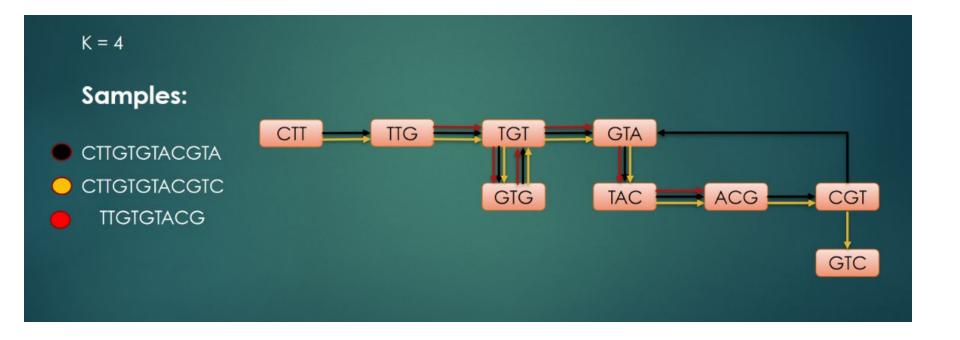


insert increment counter

All the others...



Colored de Bruijn Graphs



For more

- "Data structures based on k-mers for querying large collections of sequencing data sets", Camille Marchet, Christina Boucher, Simon J. Puglisi, Paul Medvedev, Mikaël Salson and Rayan Chikhi
 - Genome Res, 2020: https://genome.cshlp.org/content/early/2020/12/16/gr.260604.119.abstract
 - bioRxiv:
 - https://www.biorxiv.org/content/10.1101/866756v3