

Class 10: Structural Bioinformatics Pt 1

Bernice Lozada (A16297973)

What's in the PDB Database?

The main repository of biomolecular structure info is the PDB <www.rcsb.org>.

This database contains:

```
pdb_csv <- read.csv("Class10.csv", row.names = 1)
pdb_csv
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	161,663	12,592	12,337	200	74	32
Protein/Oligosaccharide	9,348	2,167	34	8	2	0
Protein/NA	8,404	3,924	286	7	0	0
Nucleic acid (only)	2,758	125	1,477	14	3	1
Other	164	9	33	0	0	0
Oligosaccharide (only)	11	0	6	1	0	4
Total						
Protein (only)	186,898					
Protein/Oligosaccharide	11,559					
Protein/NA	12,621					
Nucleic acid (only)	4,378					
Other	206					
Oligosaccharide (only)	22					

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

First need to get rid of the commas - can you find a function to get rid of the commas?

```
x <- pdb_csv$X.ray
as.numeric(sub(",", "", x))
```

```
[1] 161663    9348    8404    2758    164     11
```

Let's write a function for this and then use `apply()` to work on the entire table of data.

```
sumcomma <- function(x) {  
  sum(as.numeric(sub(",", "", x)))  
}  
apply(pdb_csv, 2, sumcomma)
```

X.ray	EM	NMR	Multiple.methods
182348	18817	14173	230
Neutron	Other	Total	
79	37	215684	

```
(apply(pdb_csv, 2, sumcomma))/sumcomma(pdb_csv$Total) *100
```

X.ray	EM	NMR	Multiple.methods
84.54405519	8.72433746	6.57118748	0.10663749
Neutron	Other	Total	
0.03662766	0.01715473	100.00000000	

84.5% of structures are solved by X-ray and 8.7% are solved by electron microscopy.

Q2: What proportion of structures in the PDB are protein?

```
pdb_total <- sumcomma(pdb_csv$Total)  
(as.numeric(sub(",", "", pdb_csv[1, "Total"]))) / pdb_total
```

```
[1] 0.8665362
```

0.86 of the structures in the PDB are protein.

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

There are about 0.0075118 structures in the PDB.

Visualizing the HIV-1 protease structure



I want to insert my image from Mol* here.

Here, I insert my image with the Asp in both Chain A and B as spacefill into the program.



Working with the bio3d package

```
library(bio3d)  
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
pdb
```

Call: `read.pdb(file = "1hsg")`

```

Total Models#: 1
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]

```

```

Protein sequence:
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF

```

```

+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call

```

```

pdbseq(pdb)[25]

```

```

25
"D"

```

Predicting functional motions of a single structure

We can do a bioinformatics prediction of functional motions (i.e. flexibility/dynamics):

```

pdb_1 <- read.pdb("6s36")

```

```

Note: Accessing on-line PDB file
PDB has ALT records, taking A only, rm.alt=TRUE

```

```

# Normal mode analysis = NMA
m <- nma(pdb_1)

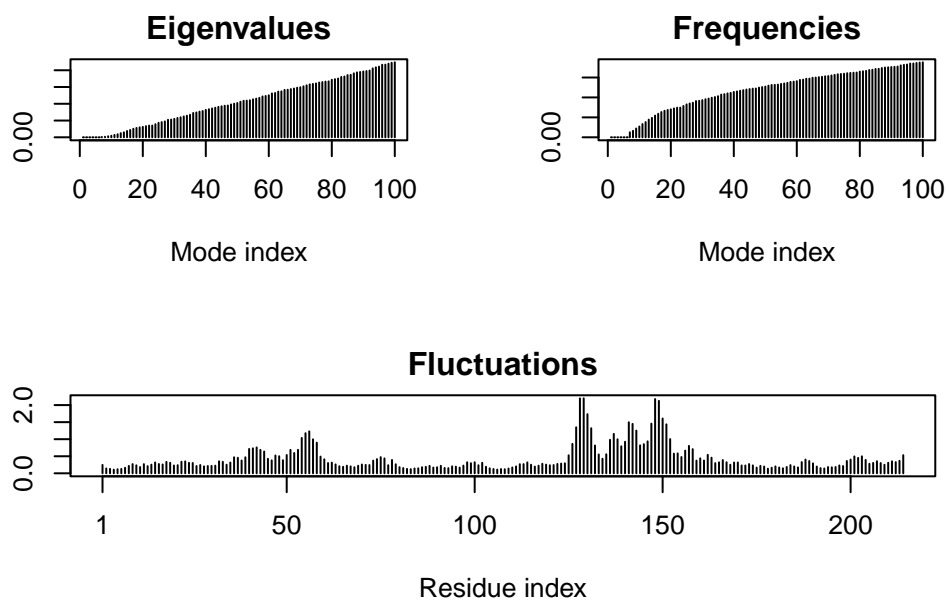
```

```

Building Hessian...      Done in 0.063 seconds.
Diagonalizing Hessian... Done in 0.581 seconds.

```

```
plot(m)
```



```
# Writes a pdb file to working directory  
mktrj(m, file="adk_m7.pdb")
```