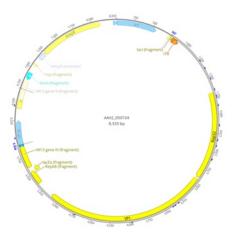
AAV Batch #: AAV2-04172024-scCAG-mgL

Serotype: AAV2

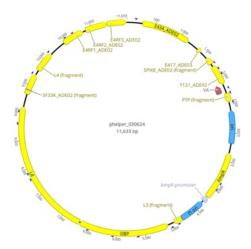
AAV construct: pAAV-scCAG-mGreenLantern

Plasmid maps:

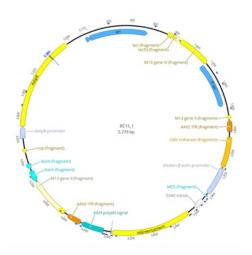
AAV2 (rep/cap):



pHelper:



scCAG-mGreenLantern-BC:

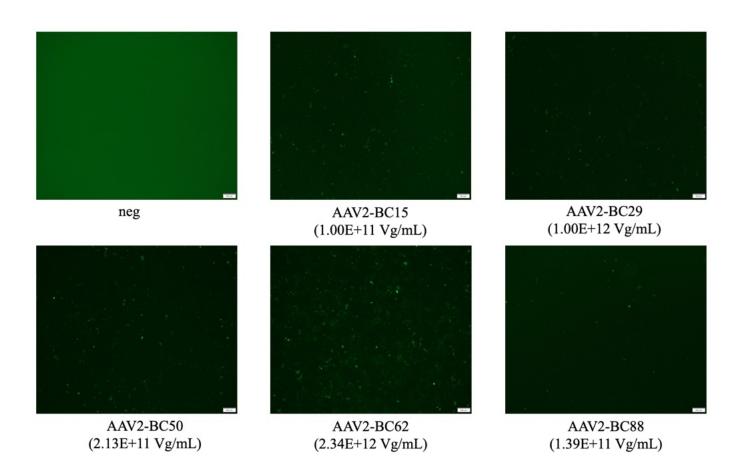


Packaging protocol:

Barcoded AAVs were packaged using a triple transfection method in adherent cells. Seed 0.6E+6 to 0.7E+6 cells/mL into a 125 mL shaker flask containing 30 mL of Expi293 medium (Thermo Fisher – A1435101) the day before transfection. On the following day, prepare the transfection mix of pHelper: AAV2: Barcode (BC)=1.67:1:1. Then, add the FectoVIR reagent to the mix and gently invert the solution (do not vortex) to prevent disintegration of the Fecto/DNA complex. Incubate the mixture for 15 minutes at room temperature. After incubation, add the required volume of the transfection mix to each flask. Finally, incubate the cells at 37°C for 72 hours post-transfection. After three days of culturing, cell lysates were subjected to three freeze-thaw cycles. Cell debris was removed by centrifugation and the supernatants were purified through AAV2-based affinity column. The final viral elute was carried on for titrations.

In vitro infectivity test:

1 uL of packaged virus was used to infect 1 well of a 6-well plate containing HEK293 cells at 70% confluency. The cells were imaged for GFP signal after 48 hrs.



Titering:

The titers were measured using qPCR AAV titer kit (ABM, catalog# G931), following the manufacturer's protocol.

AAV construct	Titer (Vg/mL)
AAV2-CAG-BC15_1 mgL	1.00E+11
AAV2-CAG-BC29_2 mgL	1.00E+12
AAV2-CAG-BC50_1 mgL	2.13E+11
AAV2-CAG-BC62_ mgL	2.34E+12
AAV2-CAG-BC88_1 mgL	1.39E+11

Mass photometry:

Mass photometry was performed on a SamuxMP (Refyn).

	AAV2-BC15	AAV2-BC29	AAV2-BC50	AAV2-BC62 4	AAV2-BC88	
	AAV2 BC15-mGL 13	AAV2 BC29-mGL 80 80 80 80 80 80 80 80 80 8	AAV2 BC50-mGL 10 10 10 10 10 10 10 10 10 1	AAV2 BC62-mGL 15	AAV2 BC88-mGL 24	
% full	5.8	4	6.8	7.8	4.4	
% empty	94.2	96	92.9	92.2	95.1	
% ambiguous	0.0	0.2	0.3	0.1	0.5	

Fungal and bacterial contamination:

 $2~\mu L$ of purified AAV was tested for bacterial and fungal contamination using qPCR-based Femto Bacterial DNA Quantification Kit (catalog # E2006) and Femto Fungal DNA Quantification Kit (catalog # E20067), following manufacturer's instructions. A $C_t > 30$ was considered no contamination.

Test	AAV2 library mix
Fungus (Ct)	39.335
Bacteria (Ct)	34.197

1 μ L of purified AAV was also tested in 100 mL of FTM media, culturing at 37 $^{\circ}$ C for 14 days. Media was clear and free of contamination.



Mycoplasma:

The final library pool was investigated for mycoplasma contamination, using Myco-Sniff-Valid Mycoplasma PCR Detection kit (MP Biomedicals, Catalog# 093050301), following the manufacturer's protocol. No mycoplasma was detected.



Endotoxin:

SAMPLE RESULTS:

The final library pool was tested for endotoxin contamination. The samples were submitted for testing to Charles River Laboratories and analyzed using PTS kinetic LAL analysis.

Serial Number/Bay: MCS 19211061/1 Start/End Temp (°C): 37.0:37.0 Sample Name: AAV2 Library Sample ID: N/A Sample Lot: N/A Dil./Conc.: 1:500 mL/mL Dilution: N/A Sample Comments: N/A Cartridge Lot Number: 4516155 Cartridge Range: 0.5-0.005 EU/mL Calibration Code: 418156542390 Archived Spike Concentration: 0.043 EU/mL Range: 181-965 seconds Y-Intercept: +2.148 Slope: -0.363 Endotoxin Value: 78.3 EU/mL Sample CV Limit: <25% Status: Endotoxin Limit: No Limit Status: N/A Spike CV Limit: <25% Status: VALID Spike Recovery Range: 50-200% Alert Limit: N/A Cartridge Type: LAL Cartridge Performed With: N/A SAMPLE DATA SPIKE DATA Channel CV% Sample Value Channel CV% Time 266 246 0.0490 78.3 EU/mL 2.3% 114%

Dilution curve analysis:

Final titer of the pooled dilution curve was 2.48E+11 vg/mL. Barcodes from the pooled library were amplified by PCR and then subjected to sequencing on an Illumina iSeq100. Quantification of barcodes revealed the following library balance:

X-axis label		dilution (Vg/mL)	counts_1	counts_2	counts_3	Avg counts	Std dev
1	BC62	1.00E+11	576735	437692	479533	497987	71335
2	BC50	1.00E+10	43890	53560	59722	48725	6838
3	BC29	1.00E+09	6218	5153	5316	5562	574
4	BC15	1.00E+08	1798	1437	1489	1575	195
5	BC88	1.00E+07	120	154	157	144	21

