# Project2

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2023-01-13

## Separating noisy and clean ecgs

### Loading the data

Loading the ECG data from the two noisy folders and the training 2017 as clean data. Iterating through every file, calculating the spectral power density with pwelch and the autocorrelation using acf. Since noise in ECG data is mostly low frequency baseline wandering, power line artifacts and white noise, we particularly selected the amplitudes from 0-5 Hz, 50 Hz, and an average across all frequencies. For the correlation we limited it to a maximum lag of 30 to limit the dimensions and data. Finally we compiled a single data frame, labeling as "noisy" and "clean". To avoid having to repeat the data processing every time, we saved that dataframe to a csy.

```
# load noisy data from two folders
f_noisy1 <- list.files("./EcgNOISY1")</pre>
f_noisy2 <- list.files("./ECGNOISY2")</pre>
f_noisy <- append(f_noisy1, f_noisy2)</pre>
# load clean data
f_clean <- list.files("./training2017", pattern="*.mat")</pre>
# iterate through data, making pwelch 0-5 and 50 and acf lag.max=30
noisy_data_p <- data.frame()</pre>
noisy_data_a <- data.frame()</pre>
for (f in f_noisy1){
  path <- paste("./EcgNOISY1/", f, sep = "")</pre>
  ecg <- readMat(path)$newval[1,]</pre>
  p \leftarrow pwelch(ecg, fs = 300)
  a <- acf(ecg, lag.max = 30, plot = FALSE)
  spec <- append(p$spec[1:6], p$spec[51])</pre>
  spec <- append(spec, mean(p$spec))</pre>
  noisy data p <- rbind(noisy data p, spec)</pre>
  noisy_data_a <- rbind(noisy_data_a, a$acf[2:31])</pre>
}
for (f in f_noisy2){
  path <- paste("./ECGNOISY2/", f, sep = "")</pre>
  ecg <- readMat(path)$newval[1,]</pre>
  p \leftarrow pwelch(ecg, fs = 300)
  a <- acf(ecg, lag.max = 30, plot = FALSE)
  spec <- append(p$spec[1:6], p$spec[51])</pre>
  spec <- append(spec, mean(p$spec))</pre>
  noisy_data_p <- rbind(noisy_data_p, spec)</pre>
  noisy_data_a <- rbind(noisy_data_a, a$acf[2:31])</pre>
}
```

```
noisy_data_p <- na.omit(noisy_data_p)</pre>
colnames(noisy_data_p) <- c("Hz0", "Hz1", "Hz2", "Hz3", "Hz4", "Hz5", "Hz50", "WhiteNoise")
colnames(noisy_data_a) <- c(1:30)</pre>
noisy data p$class <- "noisy"
noisy_data_a$class <- "noisy"</pre>
# same for clean data
clean data p <- data.frame()</pre>
clean data a <- data.frame()</pre>
for (f in f_clean){
  path <- paste("./training2017/", f, sep = "")</pre>
  ecg <- readMat(path)$val[1,]</pre>
  p \leftarrow pwelch(ecg, fs = 300)
  a <- acf(ecg, lag.max = 30, plot = FALSE)
  spec <- append(p$spec[1:6], p$spec[51])</pre>
  spec <- append(spec, mean(p$spec))</pre>
  clean_data_p <- rbind(clean_data_p, spec)</pre>
  clean_data_a <- rbind(clean_data_a, a$acf[2:31])</pre>
}
colnames(clean_data_p) <- c("Hz0", "Hz1", "Hz2", "Hz3", "Hz4", "Hz5", "Hz50", "WhiteNoise")
colnames(clean data a) <- c(1:30)
clean_data_p$class <- "clean"</pre>
clean_data_a$class <- "clean"</pre>
# concat data
p_data <- rbind(na.omit(clean_data_p), na.omit(noisy_data_p))</pre>
p_data$class <- as.factor(p_data$class)</pre>
a_data <- rbind(na.omit(clean_data_a), na.omit(noisy_data_a))</pre>
a_data$class <- as.factor(a_data$class)</pre>
write.csv(p_data, "./p_data.csv", row.names = FALSE)
write.csv(a_data, "./a_data.csv", row.names = FALSE)
```

#### Training classifier

To train the classifier we first split the data randomly into a 70/30 split. We train two distinct linear sym, one on the spectral density data, one on the autocorrelation data. After training the performance is evaluated on the test data and a confusion matrix is calculated.

```
a_training <- a_data[split, ]</pre>
a_test <- a_data[!split, ]</pre>
# train predictor
trctrl <- trainControl(method = "repeatedcv", number = 10, repeats = 3)</pre>
p_knn <- train(class ~ .,</pre>
             data = p_training,
             method = "svmLinear",
             trControl = trctrl,
             tuneLength = 10)
a_knn <- train(class ~ .,</pre>
               data = a_training,
               method = "svmLinear",
               trControl = trctrl,
               tuneLength = 10)
# test
p_knn_test <- predict(p_knn, p_test)</pre>
confusionMatrix(table(p_knn_test, p_test$class))
## Confusion Matrix and Statistics
##
##
## p_knn_test clean noisy
        clean 2403
##
                       719
##
        noisy
                 20 1659
##
##
                   Accuracy : 0.8461
                     95% CI : (0.8356, 0.8562)
##
       No Information Rate: 0.5047
##
##
       P-Value [Acc > NIR] : < 2.2e-16
##
##
                      Kappa : 0.6913
##
    Mcnemar's Test P-Value : < 2.2e-16
##
##
               Sensitivity: 0.9917
               Specificity: 0.6976
##
##
            Pos Pred Value: 0.7697
##
            Neg Pred Value: 0.9881
##
                Prevalence: 0.5047
            Detection Rate: 0.5005
##
##
      Detection Prevalence: 0.6503
##
         Balanced Accuracy: 0.8447
##
##
          'Positive' Class : clean
a_knn_test <- predict(a_knn, a_test)</pre>
confusionMatrix(table(a_knn_test, a_test$class))
## Confusion Matrix and Statistics
##
##
```

```
## a_knn_test clean noisy
        clean 2570
##
                      280
##
        noisy
                 94
                     2222
##
##
                  Accuracy: 0.9276
                    95% CI: (0.9202, 0.9345)
##
       No Information Rate: 0.5157
##
       P-Value [Acc > NIR] : < 2.2e-16
##
##
##
                     Kappa: 0.8547
##
    Mcnemar's Test P-Value : < 2.2e-16
##
##
##
               Sensitivity: 0.9647
##
               Specificity: 0.8881
##
            Pos Pred Value: 0.9018
            Neg Pred Value: 0.9594
##
##
                Prevalence: 0.5157
##
            Detection Rate: 0.4975
##
      Detection Prevalence: 0.5517
##
         Balanced Accuracy: 0.9264
##
##
          'Positive' Class : clean
##
```

# Classifying ECG data based on heart rythm

### Preprocessing

Firstly we define functions of the preprocessing the ECG data. To filter the data and be able to extract features of the QRS complex, we apply a simplified model of the pan-tompkins algorithm. This consists of first applying a passband filter, trying to remove noise. As the R peak is one of the clearest mark of the heartbeat in ECG data, we focus on exaggerating those marks and extract them. This is done by applying a derivative filter, squaring the values to achieve only positive values and the integrating using a moving window average to restore the dynamics of the original signal. Once we have identified the R-peaks, we extract the sequence and calculate the features from it.

```
pan_tompkins <- function(t_series, sampling_freq = 300){
    # passband filter
    fil_N <- 8
    fs <- 200 #sampling rate
    nf <- fs/2
    fc <- c(5, 32) # remove frequencies from 5Hz to 32Hz
    fc_norm <- fc/nf

re_series <- resample(t_series, fs, sampling_freq)

fir_filter <- fir1(fil_N, fc_norm, type="pass")
    f_x <- filtfilt(fir_filter, re_series)

# derivative filter
    d_x <- c()
    for (i in 3:(length(f_x)-2)){
        d_x[i] <- (1/(8/nf)) * (-f_x[i-2] - 2*f_x[i-1] + 2*f_x[i+1] + f_x[i+1])
}</pre>
```

```
# integration filter by moving window average
  m \times \leftarrow c()
  for (i in 1:length(d_x)){
    m \times [i] \leftarrow mean(d \times [i:i+8])
  # determine peaks
  peaks <- findpeaks(m x, minpeakheight = mean(m x, na.rm = TRUE),</pre>
                       minpeakdistance = 50)
  peaks_d <- data.frame(peaks)</pre>
  colnames(peaks_d) <- c("height", "loc", "start", "end")</pre>
  no_peaks <- m_x
  for (i in 1:length(peaks_d$loc)){
    p_start <- peaks_d$start[i]</pre>
    p_end <- peaks_d$end[i]</pre>
    no_peaks[p_start: p_end] <- c(NA * (p_end - p_start))</pre>
  npk <- sd(no_peaks, na.rm = TRUE)</pre>
  spk <- mean(peaks_d$height)</pre>
  threshold1 \leftarrow npk + 0.25 * (spk - npk)
  threshold2 <- 0.5 * threshold1
  peaks_d <- peaks_d[peaks_d$height > threshold1,]
  peaks d$loc <- peaks d$loc
  return(sort(peaks_d$loc) * 5)
nn_features <- function(peak_seq){</pre>
  nn <- peak_seq[2:length(peak_seq)] - peak_seq[1:length(peak_seq) - 1]</pre>
  succ_dif <- nn[2:length(nn)] - nn[1:length(nn)-1]</pre>
  sdnn <- sd(nn)
  sdsd <- sd(succ_dif)</pre>
  pnn50 = length(succ_dif[abs(succ_dif) > 50])/length(succ_dif)
  pnn20 = length(succ_dif[abs(succ_dif) > 20])/length(succ_dif)
  features <- data.frame(sdnn, sdsd, pnn50, pnn20)</pre>
  return(features)
}
```

#### Loading the data

We load the data from the folder and prepare a data frame with the features. Again we save the data frame to a csv to be able to avoid this time intensive step when not necessary.

```
help("list.files")
dir <- "./training2017/"
files <- list.files(dir, pattern = "*.mat")
labels <- read.csv("./REFERENCE.csv", header = FALSE)
colnames(labels) <- c("file", "class")
ecg_set <- data.frame()
columns <- c("file", "class", "sdnn", "sdsd", "pnn50", "pnn20")
ecg_dataset = data.frame(matrix(nrow = 0, ncol = length(columns)))
for (i in 1:length(files)){
    # get file name
    recording <- labels$file[i]
    path <- paste(dir, recording, ".mat", sep = "")</pre>
```

```
series <- readMat(path)$val[1,]
r_peaks <- pan_tompkins(series)
features <- nn_features(r_peaks)
labels$sdnn[i] <- features$sdnn
labels$sdsd[i] <- features$sdsd
labels$pnn50[i] <- features$pnn50
labels$pnn20[i] <- features$pnn20
}
write.csv(labels, "./feature_data.csv")</pre>
```

### Training and testing

After again splitting into training/test at 70/30, we train a k-nearest neighbor algorithm on the data and evaluate on the test set using a confusion matrix. Since in this case we have lower dimensionality than in the autocorrelation case before, we decided on a knn approach rather than a linear-sym, since lower dimensional data is often harder to separate linearly, where in turn higher dimensional data is often sparse.

```
## Confusion Matrix and Statistics
##
##
## test_pred
                      Α
                           N
                                 0
                                 0
##
                 2
                      0
                           Λ
           Α
                 6
                     38
                           7
                                53
##
           N
               42
                     77 1353
                              419
##
           0
               24
                              281
##
                   102
                         156
##
## Overall Statistics
##
                   Accuracy : 0.6539
##
                     95% CI : (0.6351, 0.6723)
##
##
       No Information Rate: 0.5922
##
       P-Value [Acc > NIR] : 8.176e-11
##
##
                      Kappa : 0.3
##
##
    Mcnemar's Test P-Value : < 2.2e-16
##
## Statistics by Class:
##
```

```
##
                     Class: ~ Class: A Class: N Class: O
## Sensitivity
                     0.0270270 0.17512 0.8925 0.3732
## Specificity
                     1.0000000 0.97183 0.4847
                                                0.8439
## Pos Pred Value
                     1.0000000 0.36538 0.7155 0.4991
## Neg Pred Value
                     0.9718530 0.92712 0.7564 0.7636
## Prevalence
                     0.0289063 0.08477 0.5922 0.2941
## Detection Rate
                     0.0007813 0.01484 0.5285 0.1098
## Detection Prevalence 0.0007813 0.04063 0.7387
                                                0.2199
                     0.5135135  0.57347  0.6886  0.6086
## Balanced Accuracy
```