Help:

RNAlien is a tool for automatic construction of RNA family models from a single sequence. The sections of this help document cover the use of the webservice and the commandline tool, as well as details of the pipeline used in the backend.

If you look for source code or a installation guide for the tool please refer to the Tool subpage.

Please note that constructions can take up to 24h, if you want to construct multiple families please use the tool instead of the webservice.

This manual is included with the Tool as manual.pdf. (https://github.com/eggzilla/RNAlien/raw/master/manual.pdf)

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Webservice:

Input:

RNAlien Webservice accepts a fasta file with a single sequence as obligatory input. Example GlmS fasta file content:

>AARQ02000011.1/391-585

AAUUGAAUAGAAGCGCCAGAACUGAUUGGGACGAAAAUGCUUGAAGGUGAAAUCCCUGAA AAGUAUCGAUCAGUUGACGAGGAGGAGAUUAAUCGAAGUUUCGGCGGGAGUCUCCCGGCU GUGCAUGCAGUCGUUAAGUCUUACUUACAAAUCAUUUGGGUGACCAAGUGGACAGAGUAG UAAUGAAACAUGCUU The organism where this sequence originates from can either be supplied via the autocomplete function, as name or via the NCBI Taxononomy id. In out current example that would be the organism (Listeria monocytogenes FSL N3-165) or Taxononomy id(393124). The id can be retrieved as described here (Retrieve Taxonomy id).

Output:

RNAlien collects potential family members in multiple iterations. While the construction process continues, the progress and intermediate results are summarized in a table. Each iteration goes through multiple states (loading, sequence search, sequence retrieval, candidate alignment, candidate filtering, query selection, model calibration, done).

Progress:

Iteration	Upper taxonomic treshold id	Aligned Sequences	Alignment Link	Covariance Model Link	Status
0	205922	0	loading	loading	done
1	205922	0	stockholm-format	covariance-model	done
2	294	3	stockholm-format	covariance-model	done
3	136843	8	stockholm-format	covariance-model	model calibration

Once the construction is finished all result files are available in the result table. The organisms of found sequences are visualized in the taxonomic overview.

Results:

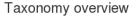
Loa	Facta	Stockholm	Alignment	Covariance	Model	Dnaz	Output	ometat	Output	Zin	Archivo	
Log	rasta	Stockholli	Alignment	Covariance	woder	milaz	Output	Cilistat	Output	Zip	Archive	

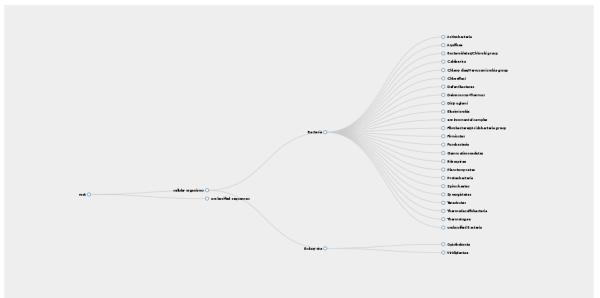
The evaluation results for the constructed covariance model and the stockholm alignment used to build it are summarized in the evaluation table.

Evaluation Results

CMstat statistics for result.	cm	RNAz statistics for resul	t alignment:
Sequence Number	1604	Mean pairwise identity	70.09
Effective Sequences	15.75	Shannon entropy	0.52075
Consensus length	336	GC content	0.5253
Expected maximum hit-length	742	Mean single sequence minimum free energy	-121.74
Basepairs	67	Consensus minimum free energy	-79.69
Bifurcations	5	Energy contribution	-73.3
Modeltype	"cm"	Covariance contribution	-6.39
Relative Entropy CM	0.59	Combinations pair	1.62
Relative Entropy HMM	0.46	Mean z-score	-0.4
		Structure conservation index	0.65
		Background model	dinucleotide
		Decision model	structural RNA alignment quality
		SVM decision value	-0.64
		SVM class propability	0.149028
		Prediction	OTHER

The taxonomnic tree of the organsims the included sequences originated from can be found in the zoom and collapsable taxonomy tree





Finally all included sequences are listed in the included sequences table. Included Sequences

Taxonomy Id	Included in Iteration	Entry Header
136843	1	gb CP008896.1 :c4840158-4839764 Pseudomonas fluorescens strain UK4, complete genome
86264	1	gi 559137252:1-396 TPA: Pseudomonas brassicacearum subsp. brassicacearum NFM421 tmRNA gene Psmon_brass_NFM421
294	1	gi 559137242:1-394 TPA: Pseudomonas fluorescens NZI7 tmRNA gene Psmon_prote_Pf5
380021	2	gi 610523943:c953216-952821 Pseudomonas protegens Cab57 DNA, complete genomecmS_3_396_'+'
200451	2	gb CP004045.1 :3880333-3880731 Pseudomonas poae RE*1-1-14, complete genomecmS_3_399_'+'
136843	2	gb CP011507.1 :c4107953-4107555 Pseudomonas trivialis strain IHBB745, complete genomecmS_3_399_'+'
29442	2	gi 559137212:1-236 TPA: Pseudomonas tolaasii NCPPB 2192 tmRNA gene Psmon_fluor_SBW25cmS_3_236_'+'
294	2	gi 559137242:1-394 TPA: Pseudomonas fluorescens NZI7 tmRNA gene Psmon_prote_Pf5cmS_1_394_'+'
1232139	3	gi 652789639:3292883-3293275 Pseudomonas pseudoalcaligenes genome assembly Ppseudo_Pac, chromosome : cmS_2_393_'+'
587851	3	gb AY911521.1 :1-348 Pseudomonas chlororaphis tmRNA (ssrA) gene, partial sequencecmS_1_346_'+'
587753	3	gb CP009290.1 :5735684-5736076 Pseudomonas chlororaphis subsp. aurantiaca strain JD37, complete genomecmS_2_393_'+
578833	3	gb CP007441.1 :c964480-964104 Pseudomonas stutzeri strain 28a24, complete genomecmS_2_377_'+'
312306	3	gi 559136889:1-392 TPA: Pseudomonas entomophila L48 tmRNA gene Psmon_entom_L48cmS_1_392_'+'
251722	3	gi 559137038:1-387 TPA: Pseudomonas syringae pv. aesculi str. 0893_23 tmRNA gene Psmon_syrin_1448AcmS_1_387_'+'
208964	3	gb CP008749.1 :c901873-901516 Pseudomonas aeruginosa PAO1H2O genomecmS_2_354_'+'
136845	3	gb AY911522.1 :1-349 Pseudomonas putida tmRNA (ssrA) gene, partial sequencecmS_1_347_'+'
136842	3	gb CP011110.1 :5753906-5754298 Pseudomonas chlororaphis strain PCL1606, complete genomecmS_2_393_'+'
136841	3	gb CP012679.1 :4779561-4779918 Pseudomonas aeruginosa strain PA1RG, complete genomecmS_2_354_'+'
103796	3	gb CP011972.1 :5087900-5088288 Pseudomonas syringae pv. actinidiae ICMP 18884, complete genomecmS_3_389_'+'
86264	3	gb CP002585.1 :c970721-970325 Pseudomonas brassicacearum subsp. brassicacearum NFM421, complete genomecmS_3_397_'+'

Tool:

Input:

The RNAlien command line utility accepts several command line parameters(see table). Please note that RNAlien requires a active and uninterrupted internet connection, which it also verifies before starting.

Parameter	Switch	Required	Description	Default value
inputFastaFilePath	-i	yes	Path to input fasta file	
outputPath	-0	yes	Path to output directory	
inputTaxId	-t	no	NCBI taxonomy ID number of input RNA organism	
			Only candidate sequences with a nSCI higher than	

inputnSCICutoff	-Z	no	this value are accepted.	1.0
inputEvalueCutoff	-е	no	Evalue cutoff for cmsearch filtering.	0.001
inputBlastDatabase	-b	no	Specify name of blast database to use.	nt
coverageFilter	-a	no	Filter blast hits by coverage of at least 80%.	True
singleHitperTax	-S	no	Only the best blast hit per taxonomic entry is considered.	False
blastsoftmasking	-f	no	Toggles blast softmasking, meaning exclusion of low complexity (repetative) regions in lookup table.	True
inputqueryselectionmethod	-m	no	Method for selection of queries (filtering,clustering).	filtering
threads	-c	no	Number of available cpu slots/cores.	1
taxonomyRestriction	-r	no	Restrict search space to taxonomic kingdom (bacteria,archea,eukaryia).	
sessionIdentificator	-d	no	Optional session id that is used instead of automatically generated one.	
Help	-?	no	Print this help	

Example call for RNAlien:

RNAlien -i /home/user/newrna.fa -c 5 -t 562 -o /home/user/temp/ -d construction1

This command starts RNAlien with the input fasta file /home/user/newrna.fa, 5 cores, the taxonomy id set to E. coli. RNAlien will create a directory called construction1 in /home/user/temp/. On how to obtain the taxonomy id for the organism the sequence originates from see Retrieve Taxonomy id.

Output:

RNAlien creates a output folder which contains all files used in the construction and log files. The most relevant output files and the logfile are directly deposited in the main folder. Each step of model expansion has an own subfolder. Following is a description of the output folder contents, of the different iteration directories that are created during model construction (empty,initial model construction, model expansion), the blast log directory that is contained in each iteration directory and the Log file content.

+RNAlien output folder

|--Log file: Summary for construction process, tool versions, iteration info, evaluation results (see Log file)

|--result.cm: Result covariance model

|--result.stockholm: Result stockholm alignment

|--result.fa: Result fasta

|--result.csv

|--evaluation: contains RNAz and cmstat output for result-files

|--log: contains non-iteration specific log files

|--1 iteration directories (see description below)

|--2 ..

|--3 ..

+iterationdirectory: initial model construction

|--model.cm: Result covariance model

|--model.stockholm: Result stockholm alignment

|--model.fa: Result fasta

|--log: Raw and processed blast hits and accepted /rejected candidates (see iteration log directory)

|--input.fa: The input fasta sequence |--input.fold: RNAfold output of the input fasta sequence |--1.fa: fasta

file for first candidate |--1.alifold: RNAalifold file for the input sequence and the first candidate

|--1.fold: RNAfold output for the first candidate

|--2.fa: Fasta file for the second candidate

|-..

+iterationdirectory: modelexpansion

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|--model.cm: Result covariance model

|--model.stockholm: Result stockholm alignment

|--model.fa: Result fasta

|--log: Directory that contains raw and processed blast hits (see blastdirectory)

|--1.fa: fasta file for first candidate

|--1.cmsearch

|--2.fa

|--2.cmsearch

|--..

+iteration log directory

1

- |--1_1blastOutput: Raw blast output for first query, indicated by leading 1_
- |--1 2blastHits: Parsed blast output
- |--1_3blastHitsFilteredByLength: Blasthits filtered by exceeding 3* query length
- |--1_3ablastHitsFilteredByLength: Blasthits filtered by having >80% coverage
- |--1_4blastHitsFilteredByParentTaxId: Only one blasthit per parent taxid
- |--1_5filteredBlastResult: Only one blasthit per taxid
- |--1_6requestedSequenceElements: Blasthit derived sequences requested from Entrez
- |--1_10afullSequencesWithSimilars: Sequences retrieved from Entrez
- |--1_10fullSequences: fullSequencesWithSimilars filtered for only containing unique sequences
- |--2_1blastOutput: Raw blast output for second query, indicated by leading 2_

|--..

- |--11candidates: All query specific sequences merged
- |--12candidatesFilteredByCollected: Filter for sequences not identical with collected
- |--13selectedCandidates: Sequences selected either by nSCI or cmsearch for inclusion in model by set evalue cutoff
- |--14rejectedCandidates: Sequences that were rejected
- |--15potentialCandidates: Sequences that are within a 10^3 interval of the set evalue cutoff,

will be reevaluated at end of modelconstruction

Log file

Benchmark

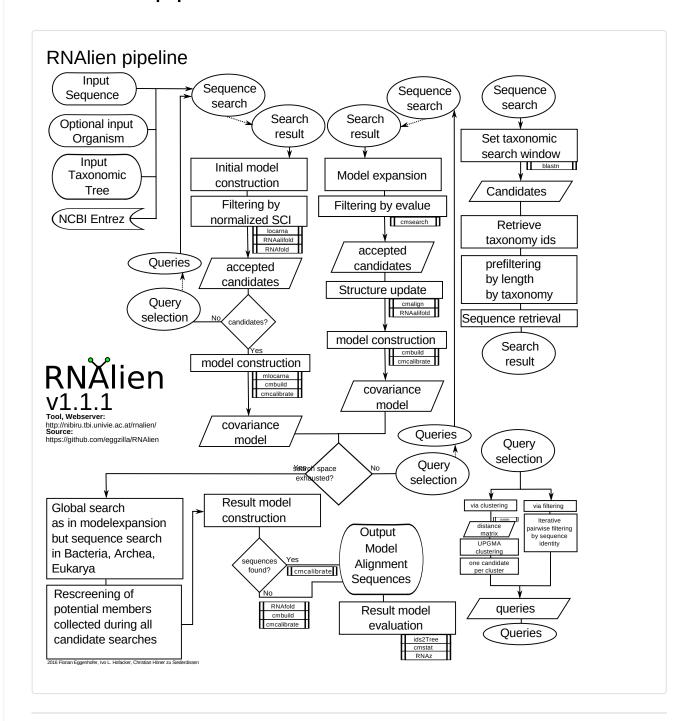
RNAfamilies constructed for benchmark are available as archive:

Rfam id Family name	Archive link (.tar.gz)
RF000015S rRNA	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/1.tar.gz)
RF000025_8S rRNA	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/2.tar.gz)
RF00003U1	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/3.tar.gz)
RF00004U2	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/4.tar.gz)

RF00005tRNA	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/5.tar.gz)
RF00008 Hammerhead_3	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/6.tar.gz)
RF00010 RNaseP_bact_a	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/7.tar.gz)
RF00011 RNaseP_bact_b	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/8.tar.gz)
RF00017 Metazoa_SRP	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/9.tar.gz)
RF00023tmRNA	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/10.tar.gz)
RF00026U6	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/11.tar.gz)
RF00028 Intron_gpl	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/12.tar.gz)
RF00029 Intron_gpII	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/13.tar.gz)
RF00032 Histone3	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/14.tar.gz)
RF00037 IRE_I	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/15.tar.gz)
RF00044 Phage_pRNA	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/16.tar.gz)
RF00050 FMN	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/17.tar.gz)
RF00059TPP	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/18.tar.gz)
RF00114S15	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/19.tar.gz)
RF00162SAM	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/20.tar.gz)
RF00164s2m	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/21.tar.gz)
RF00167 Purine	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/22.tar.gz)
RF00168 Lysine	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/23.tar.gz)
RF00169 Bacteria_small_SRF	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/24.tar.gz)
RF00174 Cobalamin	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/25.tar.gz)
RF00175 HIV-1_DIS	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/26.tar.gz)
RF00177 SSU_rRNA_bacteria	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/27.tar.gz)
RF00207K10_TLS	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/28.tar.gz)
RF00209 IRES_Pesti	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/29.tar.gz)
RF00234glmS	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/30.tar.gz)
RF00374 Gammaretro_CES	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/31.tar.gz)
RF00380ykoK	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/32.tar.gz)
RF00458 IRES_Cripavirus	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/33.tar.gz)
RF00480 HIV_FE	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/34.tar.gz)
RF00500 TCV_H5	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/35.tar.gz)
RF00504 Glycine	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/36.tar.gz)
DE00942 mir 220	
RF00843 mir-228	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/37.tar.gz)
RF00843 mir-228	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/37.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/38.tar.gz)
RF00871 mir-689	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/38.tar.gz)
RF00871 mir-689 RF01051 c-di-GMP-I	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/38.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/39.tar.gz)
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RF00871 mir-689 RF01051 c-di-GMP-I RF01054 preQ1-II RF01073 GP_knot1	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/38.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/39.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/40.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/41.tar.gz)
RF00871 mir-689 RF01051 c-di-GMP-I RF01054 preQ1-II RF01073 GP_knot1 RF01118 PK-G12rRNA	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/38.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/39.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/40.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/41.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/42.tar.gz)
RF00871 mir-689 RF01051 c-di-GMP-I RF01054 preQ1-II RF01073 GP_knot1 RF01118 PK-G12rRNA RF01380 HIV-1_SD	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/38.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/39.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/40.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/41.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/42.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/44.tar.gz)
RF00871 mir-689 RF01051 c-di-GMP-I RF01054 preQ1-II RF01073 GP_knot1 RF01118 PK-G12rRNA RF01380 HIV-1_SD RF01510 MFR	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/38.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/39.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/40.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/41.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/42.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/44.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/45.tar.gz)
RF00871 mir-689 RF01051 c-di-GMP-I RF01054 preQ1-II RF01073 GP_knot1 RF01118 PK-G12rRNA RF01380 HIV-1_SD RF01510 MFR RF01689 AdoCbl-variant	archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/38.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/39.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/40.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/41.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/42.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/44.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/45.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/46.tar.gz) archive (http://nibiru.tbi.univie.ac.at/rnalien_tmp/benchmark/46.tar.gz)
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Values for the specificity and recall plots were computed with the RNAlienStatistics executable and the rnalienstatistics.pl (https://github.com/eggzilla/RNAlien/blob/master/scripts/alienresultstatistics.pl) script included in RNAlien.

RNAalien pipeline



Retrieve Taxnonomy id

To obtain the Taxonomy id go to NCBI Taxonomy (http://www.ncbi.nlm.nih.gov/taxonomy). Enter the organisms name in the Taxonomy field (e.g Escherichia coli) and click search. You are then

redirected to a Summary page (http://www.ncbi.nlm.nih.gov/taxonomy/?term=Escherichia+coli). Click the organism name. Depending on how specific your search was you can either select from a list of organisms (http://www.ncbi.nlm.nih.gov/Taxonomy/Browser/wwwtax.cgi?id=562) or you are directed to a organism page Escherichia coli str. K-12 substr. MG1655star (http://www.ncbi.nlm.nih.gov/Taxonomy/Browser/wwwtax.cgi? mode=Info&id=879462&Ivl=3&Iin=f&keep=1&srchmode=1&unlock). You can find the taxonomy id right below the organisms name (Escherichia coli str. K-12 substr. MG1655star Taxonomy ID: 879462).

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