Notes on EGFR and ALK

Bradley Hintze

October 5, 2016

1 Epidermal Growth Factor Receptor (EGFR)

Overview:

Epidermal Growth Factor Receptor (EGFR) is a cell surface receptor that is activated when it binds its ligand causing homodimerization [1]. This dimerization causes activation if the inner-cellular C-terminal kinase activity causing autophosphorylation of C-terminal tyrosines Y992, Y1045, Y1068, Y1148 and Y1173 [2]. This phosphorylation acts as a signal to many pathways involved in cell migration, adhesion, and proliferation [3].

Implications in NSCLC:

Mutations in EGFR have been linked to squamous-cell carcinoma [4]. Some of these mutations have been shown to constitutive activate EGFR [5]. Nearly 90% of lung-cancer—specific EGFR mutations are L858R and deletion 746-750 [6]. Other point mutations have also been observed, such as L861Q [5] and 719 [7]. The drug gefitinib has shown to have a positive clinical response to those with tumors that contain these mutations, but this a small sub-population of those with NSCLC, only 10 to 19 percent of patients with chemotherapy-refractory advanced NSCLC [8, 9]. This suggests that gefitinib is promising as a targeted therapy for those having tumors with EGFR mutations described here.

Structural Observations of EGFR Mutations:

The reason for the L858R mutant causing constitutive activation of EGFR can be explained in structural terms. Leucine 858 is in the activation loop of

the kinase domain. Part of this loop is a helix in the inactive conformation and sits in the active site blocking kinase activity. Leucine 858 is in this inactive helix and makes numerous hydrophobic interactions with the active site thus locking the helix in place and blocking kinase activity. The mutation to arginine completely disrupts this hydrophobic interaction since arginine is a bulky, positively charged amino acid. This disruption makes it impossible for the inactive helix to snugly fit into the active site thus adopting the active conformation causing constitutive activation [10].

The reason the deletion 746-750 causes constitutive activation of EGFR may be due to its interaction with the inactivating helix; the interaction may lock the inactivating helix into place thus helping the inactivity. Obviously, if this is the case, deleting this loop would destabilize the inactivating helix [10].

Facts About Gefitinib

- FDA approved in May 2003
- Gefitinib is an ATP competitor.
- Gefitinib binds 20-fold more tightly to the L858R mutant than to the wild-type enzyme [11].

Facts about EGFR mutations

- The kinase activity of the EGFR L858R mutant is 50-fold more active than the wild-type kinase, and the G719S mutant is approximately 10-fold more active than wild-type [11].
- The T790M mutation increases the affinity for ATP in the L858R context. This causes gefitinib resistance as ATP can actively compete agaist gefitinib [12]. This may explain the genetic dispoint to lung cancer of those with the germ line T790M EGFR mutation [13].

Questions Pending

- Are there published guidelines for using gefitinib or erlotinib?
- What mutations are (and are not) sensitive to gefitinib and erlotinib?

- Paez et al. showed that mutations in EGFR to be more common in Japanese (Asians?) compared to European descendants and women compared to men [14]. Can we show frequency of mutation vs. race/sex?
- Is targeted treatment being misused, i.e. is gefitinib being used on patients lacking the EGFR L858R mutant? If so, are survival rates as expected lower than the group having the L858R mutant?

2 Anaplastic Lymphoma Kinase (ALK)

Bradley needs to do research!

References

- [1] Yosef Yarden and Joseph Schlessinger. Epidermal growth factor induces rapid, reversible aggregation of the purified epidermal growth factor receptor. *Biochemistry*, 26(5):1443–1451, mar 1987.
- [2] J. Downward, P. Parker, and M. D. Waterfield. Autophosphorylation sites on the epidermal growth factor receptor. *Nature*, 311(5985):483– 485, oct 1984.
- [3] Kanae Oda, Yukiko Matsuoka, Akira Funahashi, and Hiroaki Kitano. A comprehensive pathway map of epidermal growth factor receptor signaling. *Mol Syst Biol*, 1(1):E1–E17, may 2005.
- [4] Francine Walker, Laurent Abramowitz, Dalila Benabderrahmane, Xavier Duval, Véronique Descatoire, Dominique Hénin, Thérèse Lehy, and Thomas Aparicio. Growth factor receptor expression in anal squamous lesions: modifications associated with oncogenic human papillomavirus and human immunodeficiency virus. *Human Pathology*, 40(11):1517–1527, nov 2009.
- [5] Thomas J. Lynch, Daphne W. Bell, Raffaella Sordella, Sarada Gurubhagavatula, Ross A. Okimoto, Brian W. Brannigan, Patricia L. Harris, Sara M. Haserlat, Jeffrey G. Supko, Frank G. Haluska, David N. Louis, David C. Christiani, Jeff Settleman, and Daniel A. Haber. Activating mutations in the epidermal growth factor receptor underlying responsiveness of non–small-cell lung cancer to gefitinib. New England Journal of Medicine, 350(21):2129–2139, may 2004.
- [6] Rafael Rosell, Teresa Moran, Cristina Queralt, Rut Porta, Felipe Cardenal, Carlos Camps, Margarita Majem, Guillermo Lopez-Vivanco, Dolores Isla, Mariano Provencio, Amelia Insa, Bartomeu Massuti, Jose Luis Gonzalez-Larriba, Luis Paz-Ares, Isabel Bover, Rosario Garcia-Campelo, Miguel Angel Moreno, Silvia Catot, Christian Rolfo, Noemi Reguart, Ramon Palmero, José Miguel Sánchez, Roman Bastus, Clara Mayo, Jordi Bertran-Alamillo, Miguel Angel Molina, Jose Javier Sanchez, and Miquel Taron. Screening for epidermal growth factor receptor mutations in lung cancer. New England Journal of Medicine, 361(10):958–967, sep 2009.

- [7] T. Kosaka. Mutations of the epidermal growth factor receptor gene in lung cancer: Biological and clinical implications. *Cancer Research*, 64(24):8919–8923, dec 2004.
- [8] Mark G. Kris, Ronald B. Natale, Roy S. Herbst, Jr Thomas J. Lynch, Diane Prager, Chandra P. Belani, Joan H. Schiller, Karen Kelly, Harris Spiridonidis, Alan Sandler, Kathy S. Albain, David Cella, Michael K. Wolf, Steven D. Averbuch, Judith J. Ochs, and Andrea C. Kay. Efficacy of gefitinib, an inhibitor of the epidermal growth factor receptor tyrosine kinase, in symptomatic patients with non-small cell lung cancer. JAMA, 290(16):2149, oct 2003.
- [9] M. Fukuoka. Multi-institutional randomized phase II trial of gefitinib for previously treated patients with advanced non-small-cell lung cancer. *Journal of Clinical Oncology*, 21(12):2237–2246, may 2003.
- [10] Xuewu Zhang, Jodi Gureasko, Kui Shen, Philip A. Cole, and John Kuriyan. An allosteric mechanism for activation of the kinase domain of epidermal growth factor receptor. *Cell*, 125(6):1137–1149, jun 2006.
- [11] Cai-Hong Yun, Titus J. Boggon, Yiqun Li, Michele S. Woo, Heidi Greulich, Matthew Meyerson, and Michael J. Eck. Structures of lung cancer-derived EGFR mutants and inhibitor complexes: Mechanism of activation and insights into differential inhibitor sensitivity. *Cancer Cell*, 11(3):217–227, mar 2007.
- [12] C.-H. Yun, K. E. Mengwasser, A. V. Toms, M. S. Woo, H. Greulich, K.-K. Wong, M. Meyerson, and M. J. Eck. The t790m mutation in EGFR kinase causes drug resistance by increasing the affinity for ATP. Proceedings of the National Academy of Sciences, 105(6):2070–2075, jan 2008.
- [13] Daphne W Bell, Ira Gore, Ross A Okimoto, Nadia Godin-Heymann, Raffaella Sordella, Roseann Mulloy, Sreenath V Sharma, Brian W Brannigan, Gayatry Mohapatra, Jeff Settleman, and Daniel A Haber. Inherited susceptibility to lung cancer may be associated with the t790m drug resistance mutation in EGFR. *Nature Genetics*, 37(12):1315–1316, oct 2005.
- [14] J. G. Paez. EGFR mutations in lung cancer: Correlation with clinical response to gefitinib therapy. *Science*, 304(5676):1497–1500, jun 2004.