

# class09\_structural

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## The PDB database

It's easier to import through **readr** package using the **read\_csv()** function.

```
library(readr)
```

```
data <- read_csv("Data_Export_Summary.csv")
```

```
Rows: 6 Columns: 9
-- Column specification -----
Delimiter: ","
chr (1): Molecular Type
dbl (4): Integrative, Multiple methods, Neutron, Other
num (4): X-ray, EM, NMR, Total

i Use `spec()` to retrieve the full column specification for this data.
i Specify the column types or set `show_col_types = FALSE` to quiet this message.
```

```
data
```

```
# A tibble: 6 x 9
`Molecular Type` `X-ray`    EM    NMR Integrative `Multiple methods` Neutron
<chr>           <dbl>    <dbl> <dbl>      <dbl>           <dbl>    <dbl>
1 Protein (only) 176378  20438 12709      342        221     83
2 Protein/Oligosacch~ 10284   3396   34        8        11      1
3 Protein/NA       9007    5931   287       24        7      0
4 Nucleic acid (only) 3077    200   1554       2        15      3
5 Other            174     13    33        3        0      0
6 Oligosaccharide (o~ 11      0     6        0        1      0
# i 2 more variables: Other <dbl>, Total <dbl>
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

The percentage of X-ray is 81.43% and 12.27% for Electron Microscopy.

Q2: What proportion of structures in the PDB are protein?

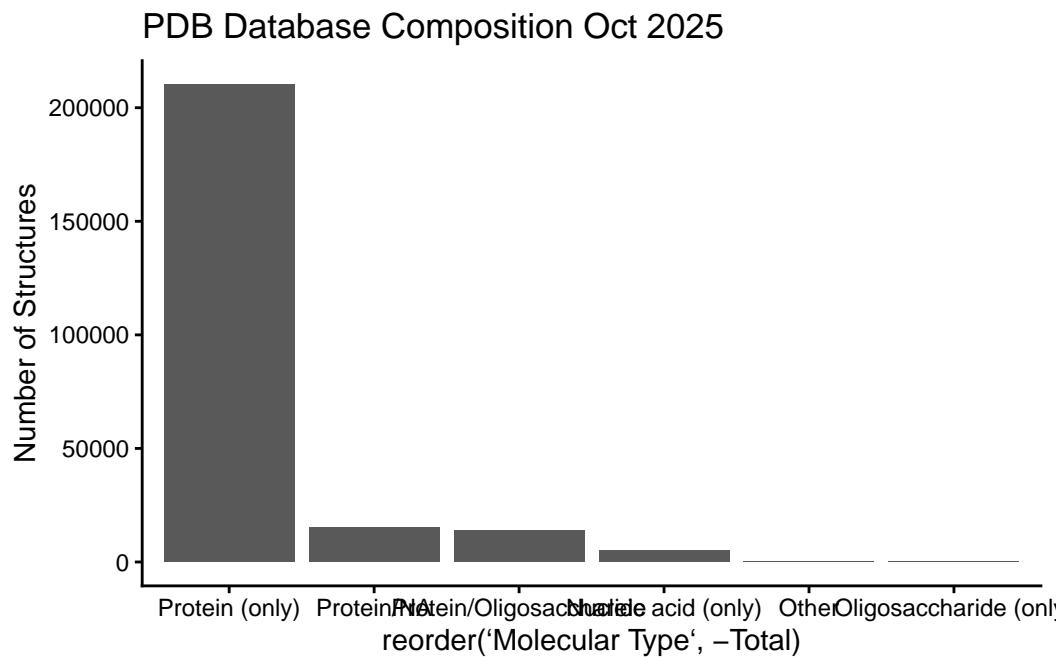
0.86

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

868

```
library(ggplot2)
library(tidyr)

ggplot(data) +
  aes(reorder(`Molecular Type`, -Total), Total) +
  geom_col() +
  labs(title = "PDB Database Composition Oct 2025", y = "Number of Structures") +
  theme_classic()
```



## Visualizing structure data

Using Mol\*

I can insert any figure or image file using markdown format: ! [Description] (name\_of\_image)



Figure 1: This is the HIV-Pr dimer with bound inhibitor

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

To reduce the complexity of the model.

Q5: There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have

yes, it is water 308.

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend “Ball & Stick” for these side-chains). Add this figure to your Quarto document.

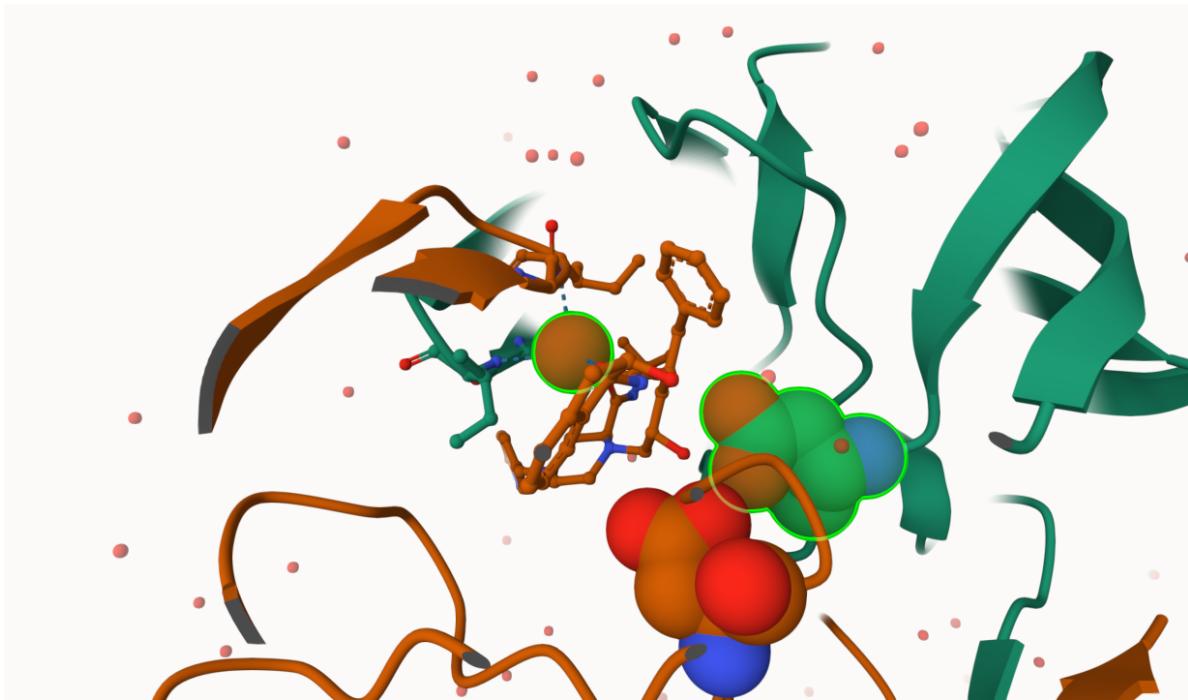


Figure 2: The catalytic ASP25 and

### Bio3D package for structural bioinformatics

We can use the bio3D package to read and analyze biomolecular data in R:

```
library(bio3d)  
  
hiv <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
hiv
```

```

Call: read.pdb(file = "1hsg")

Total Models#: 1
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]

Protein sequence:
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMIGGIGGFVKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWPKMIGGIGGFVKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF

+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call

```

Q7: How many amino acid residues are there in this pdb object?

198

Q8: Name one of the two non-protein residues?

MK1

Q9: How many protein chains are in this structure?

2

```
head(hiv$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N <NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10	
2	ATOM	2	CA <NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62	
3	ATOM	3	C <NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64	
4	ATOM	4	O <NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40	
5	ATOM	5	CB <NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87	
6	ATOM	6	CG <NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40	
	segid	elesy	charge										
1	<NA>	N	<NA>										
2	<NA>	C	<NA>										

```

3 <NA>      C  <NA>
4 <NA>      O  <NA>
5 <NA>      C  <NA>
6 <NA>      C  <NA>

```

Let's get the sequence

```
pdbseq(hiv)
```

```

 1   2   3   4   5   6   7   8   9   10  11  12  13  14  15  16  17  18  19  20
"P" "Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G" "G" "Q" "L" "K"
21  22  23  24  25  26  27  28  29  30  31  32  33  34  35  36  37  38  39  40
"E" "A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M" "S" "L" "P" "G"
41  42  43  44  45  46  47  48  49  50  51  52  53  54  55  56  57  58  59  60
"R" "W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V" "R" "Q" "Y" "D"
61  62  63  64  65  66  67  68  69  70  71  72  73  74  75  76  77  78  79  80
"Q" "I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L" "V" "G" "P" "T"
81  82  83  84  85  86  87  88  89  90  91  92  93  94  95  96  97  98  99  1
"P" "V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T" "L" "N" "F" "P"
 2   3   4   5   6   7   8   9   10  11  12  13  14  15  16  17  18  19  20  21
"Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G" "G" "Q" "L" "K" "E"
22  23  24  25  26  27  28  29  30  31  32  33  34  35  36  37  38  39  40  41
"A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M" "S" "L" "P" "G" "R"
42  43  44  45  46  47  48  49  50  51  52  53  54  55  56  57  58  59  60  61
"W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V" "R" "Q" "Y" "D" "Q"
62  63  64  65  66  67  68  69  70  71  72  73  74  75  76  77  78  79  80  81
"I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L" "V" "G" "P" "T" "P"
82  83  84  85  86  87  88  89  90  91  92  93  94  95  96  97  98  99
"V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T" "L" "N" "F"

```

Let's trim to chain A and get just it's sequence

```
chainA <- trim.pdb(hiv, chain="A")
chainA.seq <- pdbseq(chainA)
```

Let's blast

# SO ANYWAY I STARTED



# BLASTING

```
blast <- blast.pdb(chainA.seq)
```

Searching ... please wait (updates every 5 seconds) RID = GGU5HCFV014

..

Reporting 249 hits

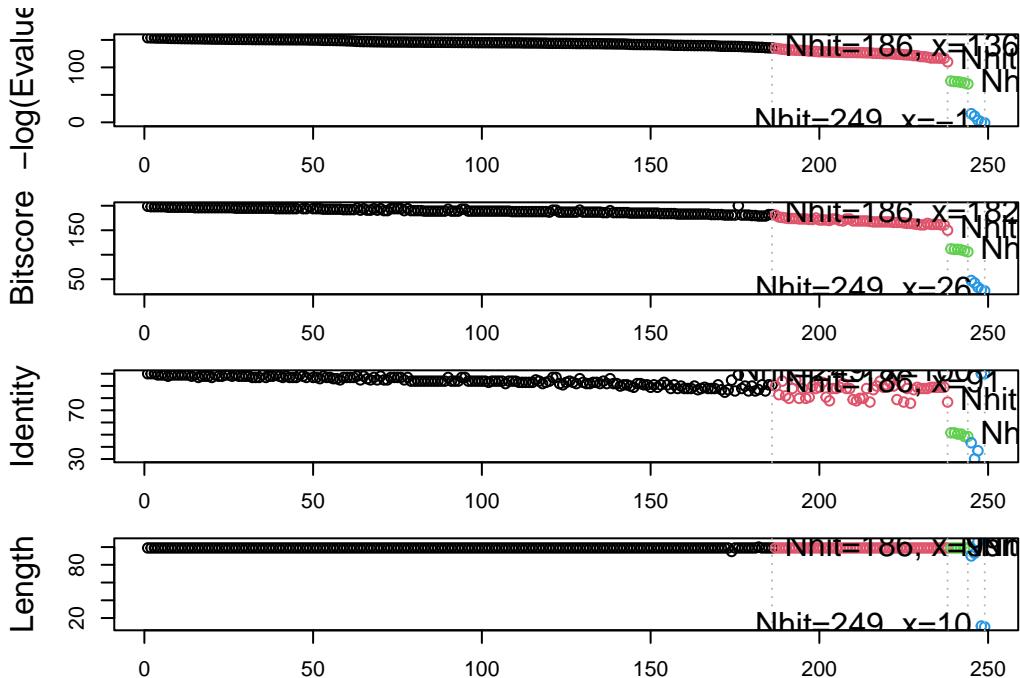
```
head(blast$hit.tbl)
```

	queryid	subjectids	identity	alignmentlength	mismatches	gapopens	q.start	q.end	s.start	s.end	evalue	bitscore	positives	mlog.evalue	pdb.id	acc
1	Query_2110573	1W5V_A	100.00		99	0	0		12	110	1.38e-67	199	100	153.9511	1W5V_A	1W5V_A
2	Query_2110573	2FDE_A	100.00		99	0	0		2	100	1.70e-67	198	100	153.7426	2FDE_A	2FDE_A
3	Query_2110573	1AJV_A	100.00		99	0	0		1	99	1.99e-67	198	100	153.5851	1AJV_A	1AJV_A
4	Query_2110573	2R38_A	98.99		99	1	0		1	99	2.50e-67	198	100	153.3569	2R38_A	2R38_A
5	Query_2110573	2R3T_A	98.99		99	1	0		1	99	2.50e-67	198	100	153.3569	2R3T_A	2R3T_A
6	Query_2110573	1HXB_A	98.99		99	1	0		1	99	2.50e-67	198	100	153.3569	1HXB_A	1HXB_A

```
hits <- plot(blast)
```

```
* Possible cutoff values: 135 110 69 -2
Yielding Nhits: 186 238 244 249

* Chosen cutoff value of: 69
Yielding Nhits: 244
```



```
hits$pdb.id
```

```
[1] "1W5V_A" "2FDE_A" "1AJV_A" "2R38_A" "2R3T_A" "1HXB_A" "1BV9_A" "1AAQ_A"
[9] "1AXA_A" "1HVS_A" "1ZP8_A" "2QHC_A" "1A8G_A" "204L_A" "5COK_A" "1TCX_A"
[17] "2Z54_A" "1D4S_A" "1BV7_A" "1BWA_A" "1A9M_A" "2FLE_A" "1ODY_A" "1GNN_A"
[25] "1GNM_A" "5YRS_B" "1HEF_E" "10DX_A" "4QGI_A" "1BVE_A" "2AZ8_A" "1A30_A"
[33] "6DH6_A" "6DHO_A" "2I4D_A" "600S_A" "1RL8_A" "5YRS_A" "1ZSF_A" "2Q64_A"
[41] "6DH3_A" "2NPH_A" "2Q63_A" "1LZQ_A" "1FB7_A" "1G6L_A" "1HIV_A" "600U_A"
[49] "1HVC_A" "2I4V_A" "2AZ9_A" "600T_A" "2P3B_B" "5KAO_A" "2WLO_A" "6OPT_A"
[57] "1IZI_A" "1MRX_A" "2PYM_A" "2PYN_A" "1DMP_A" "4K4P_A" "1LV1_A" "1AID_A"
[65] "1LV1_A" "1ZBG_A" "3TKG_A" "1HVC_A" "5YOK_A" "1G6L_A" "1FGC_C" "3K4V_A"
[73] "3KT5_A" "3KT5_A" "4QLH_A" "4QLH_A" "2F3K_A" "4Q5M_A" "2AOC_A" "3B80_A"
[81] "3VF5_A" "2AVQ_A" "1DW6_C" "1KZK_A" "2HS1_A" "1K6C_A" "1MTB_A" "4Q1X_A"
[89] "4Q1W_A" "4Q5M_A" "3D1X_A" "2AVM_A" "3PWM_A" "3KT2_A" "3KT2_A" "1SDV_A"
[97] "3JWV_A" "3OY4_A" "1A94_A" "2HS2_A" "4EJ8_A" "2FGU_A" "2AVV_A" "3JW2_A"
[105] "3BVA_A" "1FFF_C" "3S43_B" "2NXD_A" "1FG6_C" "1EBK_C" "4Q1Y_A" "3EL4_A"
[113] "1F7A_A" "1K2B_A" "2FGV_A" "1Z8C_A" "2G69_A" "3EL9_A" "30XV_A" "1BDR_A"
[121] "3N3I_A" "3N3I_A" "30XW_A" "3S43_A" "3EM3_A" "3CYW_A" "5KQX_A" "2B60_A"
[129] "7DOZ_A" "1K2C_A" "1MT7_A" "3EM4_A" "4QJ9_A" "1BDL_A" "3LZS_A" "5T84_A"
[137] "4DQB_A" "7DOZ_A" "4QJ2_A" "3LZV_A" "1SGU_A" "2FXE_A" "1BDQ_A" "3U71_A"
[145] "2R5P_A" "4OBD_A" "7MAS_A" "3IXO_A" "3D3T_A" "5YOJ_A" "3LZU_A" "4NJS_A"
[153] "3EKP_A" "1B6J_A" "3EKQ_A" "2RKF_A" "1C6X_A" "7MAR_A" "4DQF_A" "1RPI_A"
```

```
[161] "3OU1_B" "3PJ6_A" "2P3A_A" "60GQ_A" "30Q7_A" "5KR1_A" "30QD_A" "4RVI_A"
[169] "30QA_A" "1B6K_A" "30UD_B" "6MK9_A" "3S09_A" "1Q9P_A" "6I45_A" "7SEP_A"
[177] "4NJT_A" "3BXR_A" "4YOA_A" "4DQC_A" "2FDD_A" "2RKG_A" "4DQH_A" "2P3C_A"
[185] "4EP2_A" "4EP2_A" "4EQO_A" "4NPT_A" "6OPU_A" "4NPU_A" "3U7S_A" "3HAW_A"
[193] "2AZB_A" "3TTP_A" "3HBO_A" "3GGU_A" "7N6T_A" "60PV_A" "4EQO_A" "60PX_A"
[201] "204N_A" "5T2E_A" "3UCB_A" "3KA2_A" "3FSM_A" "60PW_A" "2AZC_A" "3FSM_A"
[209] "3HLO_A" "2P3D_A" "3T3C_A" "7MYP_A" "6054_X" "60PY_A" "4Z4X_A" "60PZ_A"
[217] "2JE4_A" "1DAZ_C" "7MAP_A" "7MAQ_A" "1K1U_A" "2B7Z_A" "3MWS_A" "1K1T_A"
[225] "8DCH_A" "3I2L_A" "6P9A_A" "2FXD_A" "2J9J_A" "3DCK_A" "2J9J_B" "3NXE_A"
[233] "2040_A" "2040_A" "3NXE_A" "3KA2_A" "3HLO_A" "5B18_A" "1SIP_A" "2SAM_A"
[241] "1AZ5_A" "1SIV_A" "1HII_A" "1IVP_A"
```

## Prediction of functional motions

We can run a Normal Analysis (NMA) to predict large scale motions/flexibility/dynamics of any biomolecules that we can read into R.

Let's look at ADK and chain A only.

```
adk <- read.pdb("1ake")
```

```
Note: Accessing on-line PDB file
PDB has ALT records, taking A only, rm.alt=TRUE
```

```
adk_A <- trim.pdb(adk, chain="A")
adk_A
```

```
Call: trim.pdb(pdb = adk, chain = "A")

Total Models#: 1
Total Atoms#: 1954, XYZs#: 5862 Chains#: 1 (values: A)

Protein Atoms#: 1656 (residues/Calpha atoms#: 214)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 298 (residues: 242)
Non-protein/nucleic resid values: [ AP5 (1), HOH (241) ]

Protein sequence:
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLVT
```

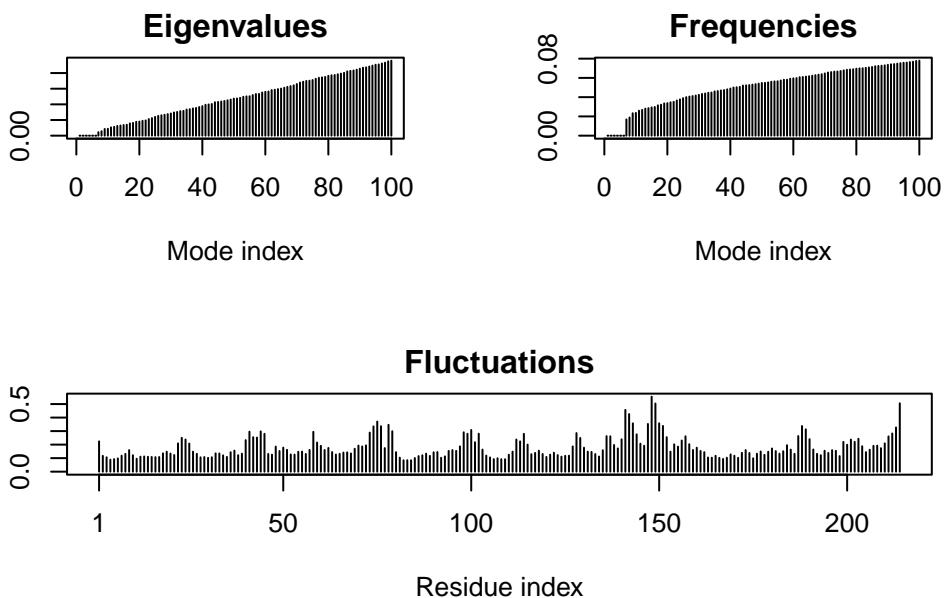
```
DELVIALVKERIAQEDCRNGFLLDGFPRTRIPQADAMKEAGINVVDYVLEFDVPDELIVDRI  
VGRRVHAPSGRKYHVKFNPPKVEGKDDVTGEELTRKDDQEETVRKRLVEYHQMTAPLIG  
YYSKAEAGNTKYAKVDGTPVAEVRADLEKILG
```

```
+ attr: atom, helix, sheet, seqres, xyz,  
       calpha, call
```

```
m <- nma(adk_A)
```

```
Building Hessian...      Done in 0.009 seconds.  
Diagonalizing Hessian... Done in 0.216 seconds.
```

```
plot(m)
```



Let's write out a “trajectory” of predicted motion

```
mktrj(m, file="adk_nma.pdb")
```

## Play with 3D viewing in R

We can use the new **bio3dviewer** package, which is not yet on CRAN, to render interactive 3d views in R and HTML quarto output reports.

To install from GitHub, we can use the **pak** package.

```
library(bio3dview)  
  
#view.pdb(adk)
```

Q10. Which of the packages above is found only on BioConductor and not CRAN?

msa

Q11. Which of the above packages is not found on BioConductor or CRAN?:

bio3d-view

Q12. True or False? Functions from the devtools package can be used to install packages from GitHub and BitBucket?

TRUE

Q13. How many amino acids are in this sequence, i.e. how long is this sequence?

214