Host-specificity and core taxa of seagrass leaf microbiome identified across tissue age and geographical regions | Sanders-Smith, R. & Segovia, B.T.(joint contribution), Forbes, C., Hessing-Lewis, M., Morien, E., Lemay, M.A., O'Connor, M. I., Parfrey, L.W.

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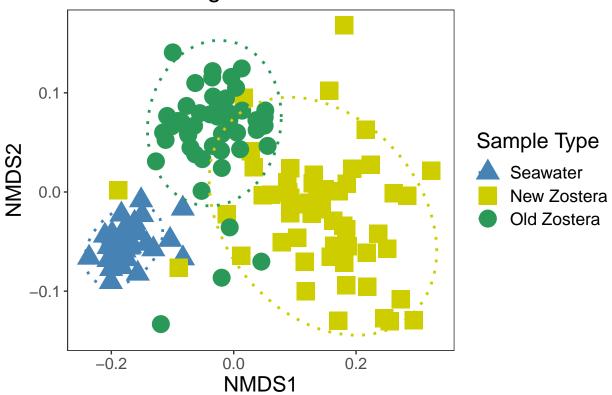
NMDS and PERMANOVAs comparison across samples types

```
#### load packages ####
library(QsRutils)
library(EcolUtils)
library(phyloseq)
library(ggplot2)
library(dplyr)
library(phylosmith)
### importing rarefied phyloseq object ###
phylo_merge_rare <- readRDS("data/phylo_merge_rarefied_16S_paper.rds")</pre>
phylo_merge_rare
## phyloseq-class experiment-level object
## otu_table() OTU Table: [ 1206 taxa and 149 samples ]
## sample_data() Sample Data: [ 149 samples by 4 sample variables ]
## tax_table() Taxonomy Table: [ 1206 taxa by 7 taxonomic ranks ]
                    Phylogenetic Tree: [ 1206 tips and 1204 internal nodes ]
## phy_tree()
### disabling scientific notation ###
options(scipen = 999)
### set.seed for reproducibility ###
set.seed(3245)
### root tree ###
phylo_merge_rare <- root_phyloseq_tree(phylo_merge_rare)</pre>
tree1 = phy_tree(phylo_merge_rare)
```

whole dataset excluding artificial

```
phylo_NO_artificial_1 <- subset_samples(phylo_merge_rare, sample_growth!="artificial")</pre>
### IMPORTANT !!! ###
### Re-order phyloseq object to run PERMANOVA ###
phylo_NO_artificial <- set_sample_order(phylo_NO_artificial_1, c('sample_growth'))</pre>
# check if re-ordering worked, set same order for metadata later
SampleID_reordered <- as.data.frame(sample_data(phylo_NO_artificial)$SampleID)
#View(SampleID reordered) # "seawater", "zostera new", "zostera old"
### ordinate using weighted Unifrac, Bray-Curtis and Jaccard dissimilarity
NMDS_bray_NO_artificial <- ordinate(phylo_NO_artificial, "NMDS", "bray")
NMDS UNI NO artificial <- ordinate(phylo NO artificial, "NMDS", "wunifrac")
NMDS_jacc_NO_artificial <- ordinate(phylo_NO_artificial, "NMDS", "jaccard")</pre>
### NMDS plot graph ###
colours_no_artificial <- c(zostera_new="yellow3",</pre>
                           zostera old="#2a9958",
                           seawater="steelblue")
NMDS_all <- plot_ordination(phylo_NO_artificial, NMDS_UNI_NO_artificial,
                                                            color = "sample_growth",
                                                            shape = "sample growth",
                                                            title = "Weighted Unifrac") +
  geom_point(size=6) +
  scale_colour_manual(name = "Sample Type", values=colours_no_artificial,
                      labels=c("Seawater","New Zostera","Old Zostera")) +
  scale_shape_manual(name = "Sample Type", values=c(17,15,16),
                     labels=c("Seawater","New Zostera","Old Zostera")) +
  stat_ellipse(type = "t", linetype = 3, size = 1) +
  theme_bw() +
  theme (text = element_text(size=16),
         axis.text.x=element_text(size=12),
         axis.text.y = element_text(size=12),
         plot.title = element_text(hjust = 0.5),
         panel.grid.major = element_blank(), #remove major grid
         panel.grid.minor = element_blank())#remove minor grid
NMDS_all
```

Weighted Unifrac



```
##################
### PERMANOVAs ###
##################
#### Importing master table ####
allsamples <- read.csv("data/Bact_3000_metadata_clean.csv", header = T)
### get metadata only ###
metadata <- allsamples[,1:4]</pre>
### Remove artificial seagrass ###
remove artificials <- c("artificial")</pre>
noartificials <- allsamples %>%
  dplyr::filter(!sample_growth %in% remove_artificials)
x <- c("seawater", "zostera_new", "zostera_old")
noartificials <- noartificials %>%
  mutate(sample_growth = factor(sample_growth, levels = x)) %>%
  arrange(sample_growth)
### Get metadata without artificial seagrass ###
metadata_no_artificial <- metadata %>%
  dplyr::filter(!sample_growth %in% remove_artificials)
metadata_no_artificial <- metadata_no_artificial %>%
  mutate(sample_growth = factor(sample_growth, levels = x)) %>%
  arrange(sample_growth)
### Calculate weighted Unifrac, Bray-Curtis and Jaccard dissimilarity ###
phylo_NO_artificial.df <- data.frame(sample_data(phylo_NO_artificial))</pre>
```

```
sampledist_all_bray <- phyloseq::distance(phylo_NO_artificial, method = "bray")</pre>
sampledist_all_UNI <- phyloseq::distance(phylo_NO_artificial, method = "wunifrac")</pre>
sampledist_all_jacc <- phyloseq::distance(phylo_NO_artificial, method = "jaccard")</pre>
### Convert distances from phyloseq objects to matrices ###
bray <- as.matrix(sampledist all bray)</pre>
UNI <- as.matrix(sampledist_all_UNI)</pre>
jacc <- as.matrix(sampledist all jacc)</pre>
dist_bray_all_mtx <- as.dist(as(bray, "matrix"))</pre>
dist_UNI_all_mtx <- as.dist(as(UNI, "matrix"))</pre>
dist_jacc_all_mtx <- as.dist(as(jacc, "matrix"))</pre>
### test for heteroscedasticity in the data ###
homogeneity <-betadisper(dist_bray_all_mtx ,metadata_no_artificial$sample_growth)
permutest(homogeneity, pairwise = TRUE)
## Permutation test for homogeneity of multivariate dispersions
## Permutation: free
## Number of permutations: 999
##
## Response: Distances
              Df Sum Sq Mean Sq
                                     F N.Perm Pr(>F)
               2 0.97793 0.48897 71.88 999 0.001 ***
## Groups
## Residuals 137 0.93194 0.00680
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## Pairwise comparisons:
## (Observed p-value below diagonal, permuted p-value above diagonal)
##
                                  seawater
                                                            zostera_new zostera_old
                                            0.0010000000000000000208167
## seawater
                                                                              0.001
## zostera new 0.0000000000000000000029418
                                                                              0.007
## zostera_old 0.0000000000023608193926018 0.0028683187047296209229441
#boxplot(homogeneity)
homogeneity <-betadisper(dist_UNI_all_mtx ,metadata_no_artificial$sample_growth)
permutest(homogeneity, pairwise = TRUE)
##
## Permutation test for homogeneity of multivariate dispersions
## Permutation: free
## Number of permutations: 999
##
## Response: Distances
             Df Sum Sq Mean Sq
                                     F N.Perm Pr(>F)
              2 0.20542 0.102708 38.308 999 0.001 ***
## Groups
## Residuals 137 0.36731 0.002681
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## Pairwise comparisons:
## (Observed p-value below diagonal, permuted p-value above diagonal)
```

```
zostera_new zostera_old
##
                         seawater
## seawater
                                  0.0010000000000000
                                                           0.001
                                                           0.010
## zostera new 0.000000000038717
## zostera_old 0.0000000002885092 0.0055906415960946
#boxplot(homogeneity)
homogeneity <-betadisper(dist_jacc_all_mtx ,metadata_no_artificial$sample_growth)
permutest(homogeneity, pairwise = TRUE)
## Permutation test for homogeneity of multivariate dispersions
## Permutation: free
## Number of permutations: 999
##
## Response: Distances
              Df Sum Sq Mean Sq
                                     F N.Perm Pr(>F)
## Groups
              2 0.55741 0.278706 81.308
                                           999 0.001 ***
## Residuals 137 0.46961 0.003428
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Pairwise comparisons:
## (Observed p-value below diagonal, permuted p-value above diagonal)
                                  seawater
                                                           zostera_new zostera_old
                                                                             0.001
## seawater
                                           0.0010000000000000000208167
## zostera_new 0.000000000000000000022538
                                                                             0.002
## zostera old 0.0000000000003699179346949 0.0020638498401832337282580
#boxplot(homogeneity)
### Because we have heteroscedasticity, we should have the same n so PERMANOVA is robust ###
# GET LIST OF RANDOMLY SUBSET OF DATA FOR PHYLOSEQ FILTERING / PERMANOVA #
sum(noartificials $sample_growth == "seawater") #35 samples
## [1] 35
sum(noartificials $sample_growth == "zostera_new") #54 samples
## [1] 54
sum(noartificials $sample_growth == "zostera_old") #51 samples
## [1] 51
### Randomly get 35 samples from water and zostera so they have same n: ###
bact_random_noartificials <- noartificials %>%
  group_by(sample_growth) %>%
  sample_n(size = 35)
## See if it worked ###
sum(bact_random_noartificials$sample_growth == "seawater")
## [1] 35
sum(bact_random_noartificials$sample_growth== "zostera_new")
```

[1] 35

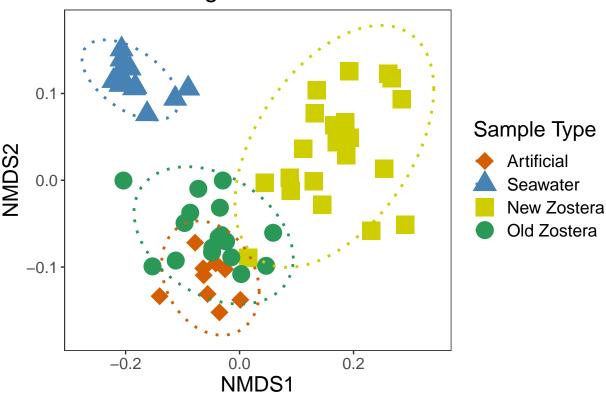
```
sum(bact_random_noartificials$sample_growth == "zostera_old")
## [1] 35
### list of randomly subset samples ###
bact_random_list_noartificials <- bact_random_noartificials$SampleID
length(bact random list noartificials)
## [1] 105
### Now use the list above to subsample data in phyloseq object ###
phylo random noartificials <- subset samples(phylo NO artificial,
                                             sample_names(phylo_NO_artificial) %in%
                                               bact_random_list_noartificials)
### Calculate weighted Unifrac, Bray-Curtis and Jaccard dissimilarity again, but for random samples ###
phylo_random_noartificials.df <- data.frame(sample_data(phylo_random_noartificials))</pre>
sampledist_random_bray <- phyloseq::distance(phylo_random_noartificials,</pre>
                                            method = "bray")
sampledist_random_UNI <- phyloseq::distance(phylo_random_noartificials,</pre>
                                           method = "wunifrac")
sampledist_random_jacc <- phyloseq::distance(phylo_random_noartificials,</pre>
                                           method = "jaccard")
### Convert distances from phyloseq objects to matrices ###
bray_random <- as.matrix(sampledist_random_bray)</pre>
UNI_random <- as.matrix(sampledist_random_UNI)</pre>
jacc random <- as.matrix(sampledist random jacc)</pre>
dist bray random mtx <- as.dist(as(bray random , "matrix"))</pre>
dist_UNI_random_mtx <- as.dist(as(UNI_random, "matrix"))</pre>
dist_jacc_random_mtx <- as.dist(as(jacc_random, "matrix"))</pre>
#### get list of subsamples from metadata ###
noartificials_subsample <- metadata_no_artificial %>%
 dplyr::filter(SampleID %in% bact_random_list_noartificials)
noartificials_subsample$sample_growth <- factor(noartificials_subsample$sample_growth)</pre>
### RANDOM samples (35 samples for each sample type) ###
### with geographical regions as a random effect ###
###################################
#### PERMANOVA Bray Curtis ####
##################################
adonis2(dist_bray_random_mtx ~ noartificials_subsample$sample_growth + region,
       data= noartificials_subsample , permutations=10000, by = "margin")
## Permutation test for adonis under reduced model
## Marginal effects of terms
## Permutation: free
## Number of permutations: 10000
## adonis2(formula = dist_bray_random_mtx ~ noartificials_subsample$sample_growth + region, data = noar
##
                                         Df SumOfSqs
                                                          R2
                                                                  F
                                                                        Pr(>F)
```

```
## noartificials_subsample$sample_growth
                                         2
                                           13.665 0.34418 29.398 0.00009999
## region
                                         3
                                            2.514 0.06331 3.605 0.00009999
## Residual
                                        99
                                            23.009 0.57953
                                       104
## Total
                                            39.704 1.00000
## noartificials subsample$sample growth ***
## region
## Residual
## Total
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
# pairwise comparisons
set.seed(3245)
adonis.pair(dist_bray_random_mtx, noartificials_subsample$sample_growth,
           nper = 10000, corr.method = "BH")
                    combination SumsOfSqs MeanSqs F.Model
                                                                      P. value
       seawater <-> zostera_new 8.527401 8.527401 36.59432 0.3498691 0.00009999
## 1
       seawater <-> zostera_old 8.962933 8.962933 42.11586 0.3824686 0.00009999
## 3 zostera_new <-> zostera_old 3.780699 3.780699 12.40253 0.1542554 0.00009999
    P.value.corrected
## 1
           0.00009999
## 2
           0.00009999
## 3
           0.00009999
#### PERMANOVA Weighted Unifrac ####
adonis2(dist_UNI_random_mtx ~ noartificials_subsample$sample_growth + region,
       data= noartificials_subsample , permutations=10000, by = "margin")
## Permutation test for adonis under reduced model
## Marginal effects of terms
## Permutation: free
## Number of permutations: 10000
##
## adonis2(formula = dist_UNI_random_mtx ~ noartificials_subsample$sample_growth + region, data = noart
                                        Df SumOfSqs
                                                                F
                                                                      Pr(>F)
##
                                                        R2
                                            3.0965 0.55081 70.7544 0.00009999
## noartificials_subsample$sample_growth
## region
                                         3
                                            0.2369 0.04214 3.6091
                                                                      0.0004
## Residual
                                        99
                                            2.1663 0.38535
## Total
                                       104
                                           5.6217 1.00000
## noartificials_subsample$sample_growth ***
## region
## Residual
## Total
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
# pairwise comparisons
set.seed(3245)
adonis.pair(dist UNI random mtx, noartificials subsample$sample growth,
           nper = 10000, corr.method = "BH")
```

```
##
                    combination SumsOfSqs MeanSqs F.Model
## 1
       seawater <-> zostera_new 2.401254 2.401254 104.65861 0.6061592 0.00009999
       seawater <-> zostera old 1.351031 1.351031 79.83888 0.5400398 0.00009999
## 3 zostera_new <-> zostera_old 1.075378 1.075378 34.89440 0.3391283 0.00009999
   P.value.corrected
## 1
           0.00009999
## 2
           0.00009999
## 3
           0.00009999
#############################
#### PERMANOVA Jaccard ####
adonis2(dist_jacc_random_mtx ~ noartificials_subsample$sample_growth + region,
       data= noartificials_subsample , permutations=10000, by = "margin")
## Permutation test for adonis under reduced model
## Marginal effects of terms
## Permutation: free
## Number of permutations: 10000
## adonis2(formula = dist_jacc_random_mtx ~ noartificials_subsample$sample_growth + region, data = noar
                                        Df SumOfSqs
                                                         R2
                                                                        Pr(>F)
                                                                  F
                                             10.223 0.23180 16.2904 0.00009999
## noartificials subsample$sample growth
## region
                                         3
                                              2.462 0.05583 2.6155 0.00009999
## Residual
                                        99
                                            31.064 0.70435
## Total
                                            44.103 1.00000
                                        104
## noartificials_subsample$sample_growth ***
## region
## Residual
## Total
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
# pairwise comparisons
set.seed(3245)
adonis.pair(dist_jacc_random_mtx, noartificials_subsample$sample_growth,
           nper = 10000, corr.method = "BH")
                    combination SumsOfSqs MeanSqs
                                                    F.Model
## 1
       seawater <-> zostera_new 6.356585 6.356585 20.389251 0.23067568
       seawater <-> zostera_old 6.739848 6.739848 22.728630 0.25051222
## 3 zostera_new <-> zostera_old 2.769003 2.769003 7.329975 0.09730489
       P.value P.value.corrected
## 1 0.00009999
                      0.00009999
## 2 0.00009999
                      0.00009999
## 3 0.00009999
                      0.00009999
choked dataset including artificial
```

```
region=="choked")
### IMPORTANT !!! ###
### Re-order phyloseq object to run PERMANOVA ###
phylo_choked <- set_sample_order(phylo_choked, c('sample_growth'))</pre>
# check if re-ordering worked, set same order for metadata later
SampleID_reordered_choked <- as.data.frame(sample_data(phylo_choked)$SampleID)
#View(SampleID_reordered_choked) # "artificial". "seawater", "zostera_new", "zostera_old"
### ordinate using weighted Unifrac, Bray-Curtis and Jaccard dissimilarity
NMDS_bray_CHOKED <- ordinate(phylo_choked, "NMDS", "bray")</pre>
NMDS_UNI_choked <- ordinate(phylo_choked, "NMDS", "wunifrac")</pre>
NMDS jacc choked <- ordinate(phylo choked, "NMDS", "jaccard")
### NMDS plot graph ###
colours_choked <- c(zostera_new="yellow3",</pre>
                           zostera_old="#2a9958",
                           artificial="#D55E00",
                           seawater="steelblue")
NMDS_choked <- plot_ordination(phylo_choked, NMDS_UNI_choked,
                color = "sample_growth",
                shape = "sample_growth",
                title = "Weighted Unifrac") +
  geom\ point(size = 6) +
  scale_colour_manual(name = "Sample Type", values=colours_choked,
                      labels=c("Artificial", "Seawater", "New Zostera", "Old Zostera")) +
  scale_shape_manual(name = "Sample Type", values=c(18,17,15,16),
                     labels=c("Artificial", "Seawater", "New Zostera", "Old Zostera")) +
  stat_ellipse(type = "t", linetype = 3, size = 1) +
  theme bw() +
  theme (text = element_text(size=16),
         axis.text.x=element_text(size=12),
         axis.text.y = element_text(size=12),
         plot.title = element_text(hjust = 0.5),
         panel.grid.major = element_blank(), #remove major grid
         panel.grid.minor = element_blank())
NMDS_choked
```

Weighted Unifrac



```
##################
### PERMANOVAs ###
##################
#### Importing master table ####
allsamples <- read.csv("data/Bact_3000_metadata_clean.csv", header = T)
### get metadata only ###
metadata <- allsamples[,1:4]</pre>
### Remove other regions ###
remove_other_regions <- c("triquet", "mcmullin", "goose")</pre>
metadata_choked <- metadata %>%
  dplyr::filter(!region %in% remove_other_regions)
x <- c("artificial", "seawater", "zostera_new", "zostera_old")
metadata_choked <- metadata_choked %>%
  mutate(sample_growth = factor(sample_growth, levels = x)) %>%
  arrange(sample_growth)
### Calculate weighted Unifrac, Bray-Curtis and Jaccard dissimilarity ###
phylo_choked.df <- data.frame(sample_data(phylo_choked))</pre>
sampledist_choked_bray <- phyloseq::distance(phylo_choked, method = "bray")</pre>
sampledist_choked_UNI <- phyloseq::distance(phylo_choked, method = "wunifrac")</pre>
sampledist_choked_jacc <- phyloseq::distance(phylo_choked, method = "jaccard")</pre>
### Convert distances from phyloseq objects to matrices ###
bray_choked <- as.matrix(sampledist_choked_bray)</pre>
UNI_choked <- as.matrix(sampledist_choked_UNI)</pre>
```

```
jacc_choked <- as.matrix(sampledist_choked_jacc)</pre>
dist_bray_choked_mtx <- as.dist(as(bray_choked, "matrix"))</pre>
dist_UNI_choked_mtx <- as.dist(as(UNI_choked, "matrix"))</pre>
dist_jacc_choked_mtx <- as.dist(as(jacc_choked, "matrix"))</pre>
### test for heteroscedasticity in the data ###
homogeneity <-betadisper(dist_bray_choked_mtx ,metadata_choked$sample_growth)
permutest(homogeneity, pairwise = TRUE)
## Permutation test for homogeneity of multivariate dispersions
## Permutation: free
## Number of permutations: 999
##
## Response: Distances
            Df Sum Sq Mean Sq
                                      F N.Perm Pr(>F)
              3 0.42253 0.140844 11.243
                                           999 0.001 ***
## Groups
## Residuals 55 0.68897 0.012527
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Pairwise comparisons:
## (Observed p-value below diagonal, permuted p-value above diagonal)
                artificial
                              seawater zostera_new zostera_old
                           0.967000000 0.001000000
                                                         0.005
## artificial
## seawater
              0.969629502
                                       0.001000000
                                                         0.004
## zostera new 0.000020651 0.000067519
                                                         0.376
## zostera_old 0.002064833 0.003334703 0.375445337
#boxplot(homogeneity)
homogeneity <-betadisper(dist_UNI_choked_mtx ,metadata_choked$sample_growth)
permutest(homogeneity, pairwise = TRUE)
## Permutation test for homogeneity of multivariate dispersions
## Permutation: free
## Number of permutations: 999
## Response: Distances
                  Sum Sq Mean Sq
                                        F N.Perm Pr(>F)
             3 0.062009 0.0206696 7.6437
                                             999 0.003 **
## Groups
## Residuals 55 0.148727 0.0027041
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## Pairwise comparisons:
## (Observed p-value below diagonal, permuted p-value above diagonal)
              artificial seawater zostera new zostera old
                          0.25900000 0.00600000
## artificial
                                                       0.016
## seawater
              0.27229566
                                      0.00100000
                                                       0.001
                                                       0.675
## zostera_new 0.00812457 0.00031821
## zostera_old 0.02327334 0.00155221 0.68578051
```

```
#boxplot(homogeneity)
homogeneity <-betadisper(dist_jacc_choked_mtx ,metadata_choked$sample_growth)
permutest(homogeneity, pairwise = TRUE)
##
## Permutation test for homogeneity of multivariate dispersions
## Permutation: free
## Number of permutations: 999
## Response: Distances
            Df Sum Sq Mean Sq
                                     F N.Perm Pr(>F)
            3 0.26594 0.088646 12.398
## Groups
                                           999 0.001 ***
## Residuals 55 0.39326 0.007150
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## Pairwise comparisons:
## (Observed p-value below diagonal, permuted p-value above diagonal)
                artificial
                               seawater zostera_new zostera_old
## artificial
                            0.8330000000 0.0010000000
                                                            0.002
## seawater
              0.8445627460
                                         0.0010000000
                                                            0.005
## zostera new 0.0000023338 0.0000309397
                                                            0.302
## zostera_old 0.0016194344 0.0030342657 0.2985231996
#boxplot(homogeneity)
### Because we have heteroscedasticity, we should have the same n so PERMANOVA is robust ###
# GET LIST OF RANDOMLY SUBSET OF DATA FOR PHYLOSEQ FILTERING / PERMANOVA #
sum(metadata_choked$sample_growth == "seawater") #12 samples
## [1] 12
sum(metadata_choked$sample_growth == "zostera_new") #21 samples
sum(metadata_choked$sample_growth == "zostera_old") #17 samples
## [1] 17
sum(metadata_choked$sample_growth == "artificial") # 9 samples
## [1] 9
### Randomly get 9 samples from all sample types so they have same n: ###
bact_random_choked <- metadata_choked %>%
 group_by(sample_growth) %>%
  sample_n(size = 9)
## See if it worked ###
sum(bact_random_choked$sample_growth == "seawater")
sum(bact_random_choked$sample_growth== "zostera_new")
## [1] 9
```

```
sum(bact_random_choked$sample_growth == "zostera_old")
## [1] 9
sum(bact_random_choked$sample_growth == "artificial")
### list of randomly subset samples ###
bact_random_list_choked <- bact_random_choked$SampleID</pre>
length(bact_random_list_choked)
## [1] 36
### Now use the list above to subsample data in phyloseq object ###
phylo_random_choked <- subset_samples(phylo_choked, sample_names(phylo_choked) %in% bact_random_list_c
### Calculate weighted Unifrac, Bray-Curtis dissimilarity ###
phylo_random_choked.df <- data.frame(sample_data(phylo_random_choked))</pre>
sampledist_random_bray <- phyloseq::distance(phylo_random_choked, method = "bray")</pre>
sampledist_random_UNI <- phyloseq::distance(phylo_random_choked, method = "wunifrac")</pre>
sampledist_random_jacc <- phyloseq::distance(phylo_random_choked, method = "jaccard")</pre>
### Convert distances from phyloseq objects to matrices ###
bray_random <- as.matrix(sampledist_random_bray)</pre>
UNI_random <- as.matrix(sampledist_random_UNI)</pre>
jacc_random <- as.matrix(sampledist_random_jacc)</pre>
dist_bray_random_mtx <- as.dist(as(bray_random , "matrix"))</pre>
dist_UNI_random_mtx <- as.dist(as(UNI_random, "matrix"))</pre>
dist_jacc_random_mtx <- as.dist(as(jacc_random, "matrix"))</pre>
#### get list of subsamples from metadata ###
choked subsample <- metadata choked %>%
 filter(SampleID %in% bact_random_list_choked)
choked_subsample$sample_growth <- factor(choked_subsample$sample_growth)</pre>
### RANDOM samples (9 samples for each sample type) ###
####################################
#### PERMANOVA Bray Curtis ####
#################################
adonis2(dist_bray_random_mtx ~ sample_growth,
       data= choked_subsample , permutations=10000, by = "margin")
## Permutation test for adonis under NA model
## Marginal effects of terms
## Permutation: free
## Number of permutations: 10000
##
## adonis2(formula = dist_bray_random_mtx ~ sample_growth, data = choked_subsample, permutations = 1000
                Df SumOfSqs
##
                                 R2
                                               Pr(>F)
## sample_growth 3 6.4184 0.49084 10.283 0.00009999 ***
## Residual
              32 6.6579 0.50916
```

35 13.0763 1.00000

Total

```
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
# pairwise comparisons
set.seed(3245)
adonis.pair(dist_bray_random_mtx, choked_subsample$sample_growth,
           nper = 10000, corr.method = "BH")
##
                    combination SumsOfSqs
                                           MeanSqs
                                                    F.Model
                                                                   R.2
## 1
        artificial <-> seawater 3.2314321 3.2314321 25.927859 0.6183922
## 2 artificial <-> zostera_new 2.2237379 2.2237379 10.559932 0.3975888
## 3 artificial <-> zostera_old 0.8470396 0.8470396 3.960743 0.1984266
## 4
       seawater <-> zostera_new 2.6631364 2.6631364 13.166946 0.4514338
       seawater <-> zostera_old 2.4766169 2.4766169 12.049588 0.4295816
## 6 zostera_new <-> zostera_old 1.3947721 1.3947721 4.785034 0.2302154
       P.value P.value.corrected
## 1 0.00009999
                    0.000149985
## 2 0.00009999
                     0.000149985
## 3 0.00019998
                     0.000239976
## 4 0.00009999
                     0.000149985
## 5 0.00009999
                     0.000149985
## 6 0.00049995
                     0.000499950
#### PERMANOVA Weighted Unifrac ####
adonis2(dist_UNI_random_mtx ~ sample_growth,
       data= choked_subsample , permutations=10000, by = "margin")
## Permutation test for adonis under NA model
## Marginal effects of terms
## Permutation: free
## Number of permutations: 10000
## adonis2(formula = dist_UNI_random_mtx ~ sample_growth, data = choked_subsample, permutations = 10000
                Df SumOfSqs
                               R2
                                        F
## sample_growth 3 1.21427 0.64027 18.985 0.00009999 ***
## Residual
                32 0.68224 0.35973
## Total
                35 1.89651 1.00000
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
# pairwise comparisons
set.seed(3245)
adonis.pair(dist_UNI_random_mtx, choked_subsample$sample_growth,
           nper = 10000, corr.method = "BH")
##
                    combination SumsOfSqs
                                           MeanSqs
                                                   F.Model
## 1
        artificial <-> seawater 0.4789498 0.4789498 44.560071 0.7357995
## 2 artificial <-> zostera_new 0.4854919 0.4854919 22.577663 0.5852522
     artificial <-> zostera_old 0.0686746 0.0686746 3.081452 0.1614894
## 4
       seawater <-> zostera_new 0.6763421 0.6763421 33.229715 0.6749930
       seawater <-> zostera_old 0.3560962 0.3560962 16.847228 0.5128965
## 6 zostera new <-> zostera old 0.3629810 0.3629810 11.381724 0.4156686
       P.value P.value.corrected
## 1 0.00009999
                     0.000149985
```

```
## 2 0.00009999
                     0.000149985
## 3 0.01209879
                     0.012098790
## 4 0.00009999
                     0.000149985
## 5 0.00009999
                     0.000149985
## 6 0.00049995
                     0.000599940
##############################
#### PERMANOVA Jaccard ####
############################
adonis2(dist_jacc_random_mtx ~ sample_growth,
       data= choked_subsample , permutations=10000, by = "margin")
## Permutation test for adonis under NA model
## Marginal effects of terms
## Permutation: free
## Number of permutations: 10000
##
## adonis2(formula = dist_jacc_random_mtx ~ sample_growth, data = choked_subsample, permutations = 1000
##
                Df SumOfSqs
                              R2
                                       F
                                               Pr(>F)
## sample_growth 3
                     5.3671 0.36833 6.2197 0.00009999 ***
                     9.2044 0.63167
## Residual
                32
## Total
                35 14.5714 1.00000
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
# pairwise comparisons
set.seed(3245)
adonis.pair(dist_jacc_random_mtx, choked_subsample$sample_growth,
           nper = 10000, corr.method = "BH")
##
                     combination SumsOfSqs
                                            MeanSqs
                                                      F.Model
                                                                     R2
        artificial <-> seawater 2.6575711 2.6575711 12.486156 0.4383237
## 2 artificial <-> zostera_new 1.8203658 1.8203658 6.185789 0.2788176
## 3 artificial <-> zostera_old 0.8805755 0.8805755 2.976182 0.1568378
## 4
       seawater <-> zostera_new 2.1522277 2.1522277 7.703042 0.3249812
       seawater <-> zostera_old 2.0539797 2.0539797 7.309743 0.3135918
## 6 zostera_new <-> zostera_old 1.1694048 1.1694048 3.226545 0.1678172
       P.value P.value.corrected
## 1 0.00009999
                    0.000149985
## 2 0.00009999
                     0.000149985
## 3 0.00019998
                     0.000239976
## 4 0.00009999
                     0.000149985
## 5 0.00009999
                     0.000149985
## 6 0.00049995
                     0.000499950
new and old growth seagrass leaves
to account for sampling from the same plant
```

```
phylo_NO_artificial

## phyloseq-class experiment-level object
## otu_table() OTU Table: [ 1206 taxa and 140 samples ]

## sample_data() Sample Data: [ 140 samples by 4 sample variables ]
## tax_table() Taxonomy Table: [ 1206 taxa by 7 taxonomic ranks ]
## phy_tree() Phylogenetic Tree: [ 1206 tips and 1205 internal nodes ]
```

```
### Remove seawater samples from phyloseq object ###
phylo_old_new <- subset_samples(phylo_NO_artificial,</pre>
                                 sample growth!="seawater")
### IMPORTANT !!! ###
### Re-order phyloseg object to run PERMANOVA ###
phylo_old_new <- set_sample_order(phylo_old_new, c('sample_growth'))</pre>
# check if re-ordering worked, set same order for metadata later
SampleID_reordered_old_new <- as.data.frame(sample_data(phylo_old_new)$SampleID)
#View(SampleID_reordered_old_new) #"zostera_new", "zostera_old"
### Remove seawater samples from metadata ###
remove_water <- c("seawater")</pre>
metadata_new_old <- metadata_no_artificial %>%
  dplyr::filter(!sample_growth %in% remove_water)
x <- c("zostera_new", "zostera_old")</pre>
metadata_new_old <- metadata_new_old %>%
  mutate(sample_growth = factor(sample_growth, levels = x)) %>%
  arrange(sample_growth)
### Calculate weighted Unifrac, Bray-Curtis dissimilarity ###
phylo new old.df <- data.frame(sample data(phylo old new ))</pre>
sampledist_new_old_bray <- phyloseq::distance(phylo_old_new,</pre>
                                               method = "bray")
sampledist_new_old_UNI <- phyloseq::distance(phylo_old_new,</pre>
                                              method = "wunifrac")
### Convert distances from phyloseq objects to matrices ###
bray_new_old <- as.matrix(sampledist_new_old_bray)</pre>
UNI_new_old <- as.matrix(sampledist_new_old_UNI)</pre>
dist_bray_new_old_mtx <- as.dist(as(bray_new_old , "matrix"))</pre>
dist_UNI_new_old_mtx <- as.dist(as(UNI_new_old, "matrix"))</pre>
### test for heteroscedasticity in the data ###
homogeneity <-betadisper(dist_bray_new_old_mtx ,metadata_new_old$sample_growth)
permutest(homogeneity, pairwise = TRUE)
##
## Permutation test for homogeneity of multivariate dispersions
## Permutation: free
## Number of permutations: 999
## Response: Distances
              Df Sum Sq Mean Sq
                                        F N.Perm Pr(>F)
              1 0.06017 0.060165 9.3292
                                             999 0.003 **
## Groups
## Residuals 103 0.66426 0.006449
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## Pairwise comparisons:
## (Observed p-value below diagonal, permuted p-value above diagonal)
               zostera new zostera old
                                  0.002
## zostera new
## zostera old
                  0.002871
```

```
#boxplot(homogeneity)
homogeneity <-betadisper(dist_UNI_new_old_mtx ,metadata_new_old$sample_growth)
permutest(homogeneity, pairwise = TRUE)
##
## Permutation test for homogeneity of multivariate dispersions
## Permutation: free
## Number of permutations: 999
## Response: Distances
                 Sum Sq
                                       F N.Perm Pr(>F)
                           Mean Sq
             1 0.023806 0.0238060 8.0018
                                              999 0.011 *
## Groups
## Residuals 103 0.306435 0.0029751
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## Pairwise comparisons:
## (Observed p-value below diagonal, permuted p-value above diagonal)
               zostera_new zostera_old
## zostera_new
                                 0.011
               0.0056176
## zostera old
#boxplot(homogeneity)
### Because we have heteroscedasticity, we should have the same n so PERMANOVA is robust ###
# GET LIST OF RANDOMLY SUBSET OF DATA FOR PHYLOSEQ FILTERING / PERMANOVA #
sum(metadata_new_old$sample_growth == "zostera_new") #54 samples
## [1] 54
sum(metadata_new_old$sample_growth == "zostera_old") #51 samples
## [1] 51
###Randomly get 51 samples from new and old zostera so they have same n:
bact_random_new_old <- metadata_new_old %>%
  group_by(sample_growth) %>%
 sample n(size = 51)
## See if it worked ###
sum(bact_random_new_old$sample_growth== "zostera_new")
## [1] 51
sum(bact_random_new_old$sample_growth == "zostera_old")
## [1] 51
### list of randomly subset samples ###
bact_random_new_old_list <- bact_random_new_old$SampleID</pre>
length(bact_random_new_old_list)
## [1] 102
### Now use the list above to subsample data in phyloseq object ###
phylo_random_new_old <- subset_samples(phylo_old_new,</pre>
                                        sample_names(phylo_old_new) %in%
```

```
bact_random_new_old_list)
### Calculate weighted Unifrac, Bray-Curtis dissimilarity ###
phylo_random_new_old.df <- data.frame(sample_data(phylo_random_new_old))</pre>
sampledist_random_new_old_bray <- phyloseq::distance(phylo_random_new_old,
                                                 method = "bray")
sampledist_random_new_old_UNI <- phyloseq::distance(phylo_random_new_old,</pre>
                                                method = "wunifrac")
### Convert distances from phyloseq objects to matrices ###
bray_random_new_old <- as.matrix(sampledist_random_new_old_bray)</pre>
UNI_random_new_old <- as.matrix(sampledist_random_new_old_UNI)</pre>
dist_bray_random_new_old_mtx <- as.dist(as(bray_random_new_old , "matrix"))</pre>
dist_UNI_random_new_old_mtx <- as.dist(as(UNI_random_new_old , "matrix"))</pre>
#### get list of subsamples from metadata ###
new_old_subsample <- metadata_new_old %>%
 filter(SampleID %in% bact_random_new_old_list)
### RANDOM samples (51 samples for each sample type) ###
##################################
#### PERMANOVA Bray Curtis ####
#################################
adonis2(dist_bray_random_new_old_mtx ~ sample_growth + individuals,
       data= new_old_subsample,
       permutations=10000, by = "margin")
## Permutation test for adonis under reduced model
## Marginal effects of terms
## Permutation: free
## Number of permutations: 10000
## adonis2(formula = dist_bray_random_new_old_mtx ~ sample_growth + individuals, data = new_old_subsamp
##
                Df SumOfSqs
                                R2
                                        F
                                              Pr(>F)
                     4.326 0.11857 15.3717 0.00009999 ***
## sample_growth
                1
## individuals
               57 19.049 0.52217 1.1876
                                              0.0032 **
               43 12.100 0.33169
## Residual
## Total
               101 36.481 1.00000
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
#### PERMANOVA Weighted Unifrac ####
adonis2(dist_UNI_random_new_old_mtx ~ sample_growth + individuals,
       data= new_old_subsample,
       permutations=10000, by = "margin")
## Permutation test for adonis under reduced model
## Marginal effects of terms
```

Permutation: free

```
## Number of permutations: 10000

## adonis2(formula = dist_UNI_random_new_old_mtx ~ sample_growth + individuals, data = new_old_subsampl

## sample_growth 1 1.3999 0.29250 49.8261 0.00009999 ***

## individuals 57 1.9229 0.40179 1.2007 0.09199 .

## Residual 43 1.2081 0.25243

## Total 101 4.7858 1.00000

## ---

## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
```